640 Communications Synthesis

oxidoreductions^{11,12} and the diamond lattice section of the active site of HLADH has been successfuly applied^{13,14,15} in these reactions.

We describe here the reduction of methyl (\pm) -5-chloro-2-oxobicyclo[2.2.1]heptane-7-anti-carboxylic $[(\pm)$ -1; readily available from norbornadiene⁴] with baker's yeast (Method A) and by an optimized procedure using Candida utilis (Method B).

The reduction of racemic 1 with baker's yeast under usual laboratory conditions gave a mixture of the isomeric hydroxyesters 2a and 2b. These hydroxy compounds were converted into a mixture of the diastereoisomeric benzoates 3a and 3b which could be separated by flash chromatography. Cleavage of the isolated benzoates 3a and 3b was achieved by treatment with methanolic potassium benzoate at ambient temperature and the resultant pure hydroxy compounds 2a and 2b were oxidized with chromium(VI) oxide/sulfuric acid in acetone (Jones oxidation) to give the pure compounds (+)-1 and (-)-1, respectively (Method A).

The Enantioselective Microbial Reduction of Methyl (\pm)-5-Chloro-2-oxobicyclo[2.2.1.]heptane-7-anti-carboxylate

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Methyl(+)- and (-)-5-chloro-2-oxobicyclo[2.2.1]heptane-7-anti-carboxylates, useful intermediates in prostaglandin synthesis, are prepared on a preparative scale from the racemic compound by two routes using enant-ioselective microbial reduction as a key-step.

Substituted derivatives of bicyclo[2.2.1]heptane-2-one are useful intermediates in various syntheses of prostaglandins¹⁻⁴, the availability of optically active compounds of this type being of major importance. Several methods for the microbial synthesis of chiral compounds have been developed⁵⁻⁸. The known reduction of bicyclic ketones using baker's yeast has also been applied in the field of prostaglandins^{9,10}. The bicyclo[2.2.1]heptane-2-one system has been studied with respect to stereoselective HLAD-catalyzed

August 1986 Communications 641

In order to improve the somewhat laborious procedure of Method A, we have optimized the reduction using fermentation procedures under carefuly controlled conditions and various microorganisms**. The best results were obtained using a strain of Candida utilis (CCY 29-38-18) and stopping the reaction at 52-53% conversion of racemic 1. Separation of the mixture of 2 and (-)-1 thus obtained is easy. The isolated crude compound 2 is oxidized with Jones' reagent to give the pure compound (+)-1 in 23% overall yield (Method B). The "unnatural" compound (-)-1 is isolated in 33% yield. Comparable results are obtained when the reaction is performed on a 501 fermentation scale.

The structures of compounds **2a**, **2b**, **3a**, and **3b** were elucidated by N.M.R.-spectral data (Tables 1 and 2). A discussion of the ¹H- and ¹³C-N.M.R. spectra will be published separately.

Table 1. ¹H-N.M.R. (CDCl₃/TMS_{int}) Chemical Shifts of Compounds 2a, 2b, 3a, and 3b

Proton	δ [ppm]					
	2a	2b	3a	3b		
1-H	2.83 br. d	2.78 m	2.94 br. d	3.08 tt		
2-H	3.83 ddt	4.19 m	4.80 ddd	5.13 dddd		
3-endo-H	1.87 ddd	2.21 ddd	1.94 ddd	2.44 ddd		
3-exo-H	2.29 dddd	0.94 dd	1.85 dddd	1.21 dd		
4-H	2.57 dp	2.71 tt	2.91 dp	2.91 br. d		
5-H	3.77 br. dt	4.03 ddd	3.95 dd	4.09 ddd		
6-endo-H	1.70 br. dd	2.77 ddd	2.08 ddd	2.75 ddd		
6-exo-H	1.56 dddd	2.20 ddt	2.43 dddd	2.36 ddt		
7-H	3.05 p	2.54 p	3.04 p	2.68 p		
OCℍ ₃	3.69 s	3.68 s	3.71 s	3.72 s		
ОН	1.70 br. s			1944		
$O-CO-C_6H_5$			7.40-7.48 m	7.42~7.50 m		
0-5				(2H)		
			7.53-7.61 m	7.54~7.62 m		
				(1 H)		
			7.97-8.02 m	7.98-8.04 m		
				(2H)		

Method A:

Mixture of Methyl (2S,5R,7S)- and (2S,5S,7R)-5-Chloro-2-hydroxybicyclo[2.2.1]heptane-7-carboxylates (2a + 2b): Commercial baker's yeast (20 g) is suspended in water (200 m).

Commercial baker's yeast (20 g) is suspended in water (200 ml), sacharose (20 g) is added, and mixture is stirred at 30–32 °C for 45

Table 2. Coupling Constants $J_{H,H}$ of Compounds **2a**, **2b**, **3a**, and **3b**

H _i , H _j	J [Hz]			
	2a	2b	3a	3b
1,2	1,0	3.6	0.9	4.3
1,4	1.2	1.5	1.2	1.4
1,6 _{endo}	1.0	0.	1.6	0.
$1,6_{exy}$	4.8	4.8	5.0	4.3
1,7	1.4	1.3	1.4	1.4
2.3_{endo}	7.8	10.0	6.6	10.1
2.3_{exo}	3.2	3.6	2.9	3.4
2,7	1.4	1.3	0.	1.6
$3_{endo}, 3_{exo}$	14.7	13.8	14.6	14.3
Sendo, 4	0.	5.03	0.	5.1
enda, 7	1.4	0.	1.4	0.
e_{xo} , 4	5.0	0.	4.6	0.
exo, 5	0.	0.	1.2	0.
,5	1.5	1.1	1.0	1.2
$,6_{exo}$	1.2	1.0	1.1	1.3
l,7	1.4	1.3	1.4	1.4
,6 _{enda}	6.9	8.0	7.8	7.8
$6,6_{exo}$	2.5	3.4	3.2	3.6
ondo, 6exo	14.0	14.5	14.9	14.5
endo: 7	0.	1.6	0.	1.7
o_{exo} , 7	0.	0.	1.4	0.

min. Racemic methyl 5-chloro-2-oxobicyclo[2.2.1]heptane-7-anticarboxylate (1; 2.0 g, 0.01 mol) is added in one portion and the mixture is stirred at 28°C for 2-3 days. The progress of reaction is monitored by T.L.C. (silicagel G; benzene/ethyl acetate/dioxan 90/3/7). The cells are separated by centrifugation or filtration through Celite¹⁰⁰, and are washed with water (100 ml). The oil left after evaporation in vacuo is taken up in 1,2-dichloroethane (90 ml) and this solution is dried with sodium sulfate, filtered, and evaporated. The crude product (2.4 g) is distilled in vacuo to give a mixture of 2a and 2b; yield: 1.8 g, 90%); b. p. 105-7°C/31 Pa.

C₉H₁₃ClO₃ calc. C 52.82 H 6.40 (204.6) found 52.76 6.31

Methyl (2R,5R,7S)-2-Benzoyloxy-5-chlorobicyclo[2.2.1]heptane-7-carboxylate (3a) and Methyl (2S,5S,7R)-2-Benzoyloxy-5-chlorobicyclo[2.2.1]heptane-7-carboxylate (3b):

The isomer mixture 2a + 2b (3.458 g, 16.9 mmol) is dissolved in pyridine (30 ml) and benzoyl chloride (6.0 g, 43 mmol) is added with stirring. The reaction is monitored by T.L.C. (silica gel; benzene/acetone 95/5). The mixture is left at ambient temperature overnight, then diluted with ether (250 ml). This solution is washed succesively with water (50 ml), hydrochloric acid (1/1; 2×50 ml), and water (2×50 ml), and is dried with sodium sulfate. The solvent is evaporated and the remaining crude oil is flash chromatographed (silica gel Merck H; benzene/acetone 99/1) to give the individual isomers 3a and 3b.

Diastereoisomer 3a; yield: 1.88 g (36%); m.p. 76.5–77.5°C; $[\alpha]_D^{25}$: $\pm 13.4^{\circ}$ (c = 0.7, methanol).

Diastereoisomer 3b; yield: 2.30 g (44%); m.p. 107–108°C; $[\alpha]_D^{25}$: + 25.3° (c = 0.4, methanol).

Methyl (2S,5R,7S)-5-Chloro-2-hydroxybicyclo[2.2.1]heptane-7-carboxylate (2a) and Methyl (2S,5S,7R)-5-Chloro-2-hydroxybicyclo[2.2.1]heptane-7-carboxylate (2b):

The benzoate 3a or 3b (5.907 g, 18.5 mmol) is dissolved in methanol (150 ml), dry potassium carbonate (0.1 g) is added, and the mixture is stirred for 12 h. DOWEX $50W^{**}$ (0.5 g) is then added with stirring. The mixture is filtered, and the ion-exchange resin thoroughly washed with methanol (2 × 50 ml). The combined organic phases are evaporated, the crude residue is triturated with heptane (100 ml),

642 Communications SYNTHESIS

and the product then crystallized from heptane (250 ml) to pure 2a (3.43 g) or 2b. An additional amount of 2a (0.417 g) or 2b is obtained by flash chromatography of the mother liquor.

C₉H₁₃ClO₃ calc. C 52.82 H 6.40 (204.6) found for **2a** 52.71 6.31 found for **2b** 52.77 6.34

Isomer 2a: yield: 3.85 g (98%); m.p. 89~91°C; $[\alpha]_D^{25}$: +6.9° (c = 0.4, methanol).

Isomer **2b**; yield: 3.84 g (98%); m.p. 89-91 °C; $[\alpha]_D^{25}$: -20.9° (c = 0.4, methanol).

Methyl (+)- and (-)-5-Chloro-2-oxobicyclo[2.2.1]heptane-7-anticarboxylate [(+)-1 or (-)-1, respectively]:

A 2.6 molar solution of chromium(VI) oxide in 8 molar sulfuric acid (4.5 ml) is added dropwise to a stirred solution of compound 2a or 2b (2.04 g, 10 mmol) in acetone (100 ml) and stirring is continued for 15 min. Excess oxidizing agent is then destroyed by the addition of 2-propanol (0.5 ml). The green precipitate is filtered off and washed with acetone (50 ml). The combined acetone solutions are evaporated in vacuo to give the pure product (+)-1 or (-)-1, respectively.

Isomer (+)-1; yield: 1.91 g (95%); m.p. 105–106°C (from water); $[\alpha]_D^{20}$: + 31.9° (c = 0.5, methanol).

Isomer (-)-1; yield: 1.89 g (95%); m.p. 105–106°C (from water); $[\alpha]_D^{20}$: -34° (c = 0.4, methanol).

Method B:

Methyl (+)- and (-)-5-Chloro-2-oxobicyclo[2.2.1]heptane-7-anti-carboxylate [(+)- 1 and (-)-1] by Partial Microbial Reduction:

Microorganism paste of Candida utilis (CCY 29-38-18; 100 g), water (2300 ml), and standard nutrient broth (60 ml) are placed in a 5000 ml fermentor. The pH value is adjusted to 4 with aqueous ammonia. During the fermentation, the ethanol content is held at 0.1 % by the gradual addition of 96 % ethanol. The suspension is aerated in such a manner that an oxygen saturation of 1.1 % at 30 °C is maintained for 1 h. The racemic ester (\pm) -1 (15 g, 74 mmol) is then added in one portion and the progress of the reaction is monitored by G.L.C. (SE-30). After $\sim 3-4$ h (consumption of 52-53% of 1), the reaction is quenched by the addition of acetone (2000 ml). The microorganisms are and washed with acetone (100 ml) and the combined filtrate is evaporated in vacuo (50°C). The remaining crude oil (22 g) is diluted with water (10 ml), and extracted with ethyl acetate (3×50 ml). The combined organic phases are evaporated in vacuo. The residual crude oily product [containing 50.1% (-)-1 and 40.8 % 2a] is flash-chromatographed (silica gel; chloroform and chloroform/methanol 95/5) to give the crude product (-)-1 [yield: 9 g; m. p. 75–80 °C; $[\alpha]_D^{20}$: -23.1° (c = 0.82, methanol)] and the crude product **2a** [yield: 7.6 g; $[\alpha]_D^{20}$: +4.24° (c = 0.82, methanol)]. The crude compound 2a (7.6 g) is dissolved in acetone (80 ml) and a 2.6 molar solution of chromium(VI) oxide in 8 molar sulfuric acid (2 ml) is added dropwise, with stirring. Excess oxidizing agent is then destroyed by the addition of 2-propanol (3 ml). The green precipitate is filtered off and washed with acetone (80 ml). The combined acetone solutions are evaporated in vacuo to give the pure product (+)-1 which is further purified by recrystallization from water (110 ml); yield: 3.50 g [23 % based on (\pm)-1]; m.p. 105 °C; [α]_D²⁰: $+ 33.0^{\circ}$ (c = 0.8, methanol).

The crude product (-)-1 is purified by three recrystallizations from water; yield: 5 g [33%, based on (\pm)-1]; m.p. 105-106°C; [α]_D²⁰: -33.5° (c = 0.8, methanol).

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