Synthesis and antibronchospastic activity of theophylline thioacetal derivatives

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theophylline derivatives / antibronchospastic mucolytic

Introduction

Theophylline and its derivatives [1, 2, 3] are well-known for their bronchodilator activity and consequent efficacy in the treatment of bronchial asthma.

Related N₇-substituted theophylline derivatives such as proxyphylline [4], bamiphylline [5], diphylline [6], and doxophylline [7, 8] are also in therapeutic use. Among these, doxophylline (Ansimar[®]), was shown to be effective in reducing bronchial muscle spasm, its activity being greater than that of theophylline, showing very poor side effects on cardiovascular and central nervous systems [9] due to a lack of affinity between this drug and A1 and A2 adenosine receptors [10–12].

As a part of our research on antibronchospastic compounds, we prepared some doxophylline thioanalogues with the aim of synthesizing a compound showing 2 complementary pharmacodynamic activities, *i.e.* mucolytic and antibronchospastic. Indeed the dithiocycloalkyl side chain could bestow the mucolytic activity on the theophylline molecule, as has previously been demonstrated for other sulfur containing compounds [13-15].

Chemistry

The general synthetic methods employed to prepare compounds II-IV involved cyclothioacetalyzation (Scheme 1) of 7-theophyllineacetaldehyde (I).

While dithioacetals **II**, **III** were obtained under BF₃etherate catalysis, compound **IV** was prepared in the presence of ZnCl₂. The 7-theophylline-1,3-dithiolane-1oxide diastereomers (**V**, **VI**) were synthesized in good yields form **II** by H_2O_2 oxidation in acetic acid and the separation of the 2 diastereomers was achieved by using flash-chromatography. Other cyclic sulfoxide diastereomers were separated in this way [17].



Scheme 1.

The stereochemical assignment was obtained according to the following evidence: i) the *trans* isomer exhibited the signals of methylene group ($>N-CH_2-$) at 4.65 and 4.45 δ upfield with respect of signals of *cis* isomer. This is in good agreement with the observations of Carey [18]. ii) the *cis* isomer, upon heating in aprotic solvent (dimethylsulfoxide), was smoothly transformed into the corresponding dihydro-1,4-dithiin derivative whereas the *trans* isomer was not. This is in accord with Chen's results [19].

The structures of all mentioned new products were confirmed by analytical IR, UV, NMR, and MS spectral data.

Results and Discussion

The data obtained from the pharmacological studies indicate compound \mathbf{II} as the most interesting of the tested compounds.

In fact it showed a antihistaminic action equivalent to that of aminophylline in the guinea-pig trachea test (ED₅₀ 164 μ g/ml, Table I) and greater antibronchospastic

New products

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activity (ED₅₀ 5.1 mg/kg, Table II) than compounds III, IV, V, VI and reference compounds.

Furthermore, compound **II** significantly increased the production of tracheobronchial mucus in the rabbit after oral treatment, its potency being markedly higher than that of other known mucokinetic drugs, used as reference (bromhexyne, N-acetylcysteine, Table III). On the contrary doxophylline, the 1,3 dioxolane theophylline derivative, was completely devoid of this effect.

Table I. In vitro antibronchospastic activity of compounds II-VI and reference compounds on guinea-pig isolated trachea.

Chemicals Compound Confidential IC_{50} $(\mu \tilde{g} / ml)$ limits 92-294 II 164.7 m 496.4 350 - 502The reported melting points are uncorrected. 300.2 250 - 380IV Spectral studies 352.6 246 - 504280 - 484VI 384.2 Doxophylline MS-80 (Electron impact). 215.4 105 - 312Aminophylline 206.1 117-363 Syntheses

Due to the lack of free thiol group in the molecule, this compound probably behaved as an indirectly acting (blocked thiol group) mucolytic, its target being the syaliltransferases enzymes [24].

All the tested compounds did not present significant oral toxicity with LD >900 mg / kg as shown in Table IV.

Compound II was therefore selected for further pharmacological and clinical studies.

Experimental protocols

Chemistry

7-Theophyllineacetaldehyde was prepared from 7-(2,3-dihydroxy-propyl)-theophylline according to Toffoli *et al.* [16]. 1,2-ethandithiol, 1,3-Propandithiol, 1,2-mercaptoethanol, p-toluensulfonic acid, boron trifluoride etherate, 35% hydrogen peroxide and dioxane were from E. Merck (Darmstadt, FRG). Analytical grade chemicals and solvents were used as received without purification.

¹H NMR spectra were recorded in CHCl₃ on a FT JEOL GX270 spectrometer using tetramethylsilane as internal standard.

IR and UV spectra were recorded on 781 Perkin-Elmer and U-3200 double-beam Hitachi, respectively. Mass spectra were from a Kratos

7-(1,3-Dithiolan-2-vlmethyl)theophylline **II.** 1,2-Ethandithiol 5 ml

| Table | II. Inh | ibitory | effect | of theo | phylline | thioacetal | derivatives | on acet | vlcholine | -induced | bronchos | pasm in | anaesthetized | guinea- | pigs |
|-------|---------|---------|--------|---------|----------|------------|-------------|---------|-----------|----------|----------|---------|---------------|---------|--------|
| | | | | | | | | | | | | | | 0 | c . o. |

| Compound | Dose (mg/kgi.v.) | % Inhibition (mean ± SD) | ED ₅₀ (mg/kgi.v.) |
|---------------|----------------------------|---|---------------------------------|
| Π | 2.5 5.0 10.0 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | 5.35 |
| m | 2.5 5.0 10.0 15.0 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | >15.00 |
| IV (* 1977) | 5.0 10.0 15.0 | 14.5 ± 3.2 34.7 ± 7.7 47.3 ± 7.1 | 15.20 |
| V | 2.5 5.0 10.0 15.0 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | >15.00 |
| VI | 2.5 5.0 10.0 15.0 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | >15.00 |
| Doxophylline | 2.5 5.0 10.0 | $\begin{array}{rrrr} 6.5 \ \pm \ 3.5 \\ 26.7 \ \pm \ 5.7 \\ 47.4 \ \pm \ 6.4 \end{array}$ | 10.85 |
| Aminophylline | 2.5 5.0 10.0 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | 10.90 |

(5.6 g, 0.059 mol) and 10 ml boron trifluoride etherate were added to a warm solution of 7-theophyllineacetaldehyde (10 g, 0.045 mol) in 350 ml dry dioxane. After 5 h of heating at 60°C under stirring, the dioxane was evaporated under reduced pressure and 400 ml CHCl₃ were added. The solution was thoroughly washed with 5% aqueous NaOH (2×200 ml) and shaken with saturated brine (2×200 ml).

The organic phase was then dried over anhydrous sodium sulphate and the solvent evaporated. The crude product was purified by flash-chromatography on silica gel eluting with CH_2Cl_2 and then with CH_2Cl_2 /EtOH (99/1) to give 6.6 g II (50% yield). mp: 125–127°C from methanol (white needles).

¹H NMR (270 MHz) (CDCl₃, δ): 3.25 (s, 4H, CH₂CH₂), 3.40 (s, 3H, N₃-CH₃), 3.59 (s, 3H, N₁-CH₃), 4.20 (d, 2H, CH₂CH, *J* = 7 Hz), 4.93 (t, 1H, CH₂CH, *J* = 7 Hz), 7.66 (s, 1H, N=CH). IR (CHCl₃): 1705, 1660, 1550 cm⁻¹. UV (CH₃OH): $\lambda_{max} = 275$, 204 nm. MS (EI): *m/z*: 298 (M⁺, 12.4), 180 (35.5), 119 (56.1), 118 (69.5), 105 (100).

7-(1,3-Dithian-2-ylmethyl)theophylline III. The synthesis was performed as described for II. mp: 154-156°C from methanol (white needles).

¹H NMR (270 MHz) (CDCl₃, δ): 2.00 (m, 2H, CH₂CH₂CH₂), 2.74–2.92 (m, 4H, 2CH₂–S), 3.40 (s, 3H, N₃–CH₃), 3.58 (s, 3H, N₁–CH₃), 4.44 (t, 1H, CH₂CH, J = 7.4 Hz), 4.65 (t, 2H, CH₂CH, J = 7.4 Hz), 7.66 (s, 1H, N=CH). IR (CHCl₃): 1705, 1660, 1605, 1550 cm⁻¹. UV (CH₃OH): $\lambda_{max} = 275$, 205 nm. MS (EI): m/z: 312 (M⁺, 20.6), 180 (23.2), 134 (15.2), 133 (32.5), 132 (100), 119 (78.9).

7-(1,3-Oxathiolan-2-ylmethyl)theophylline IV. The synthesis was performed as described for compound II using $ZnCl_2$ instead of boron

Table IV. Acute toxicity in mice following oral administration of theophylline derivatives.

| Compound | LD ₅₀ (mg⁄kg) | |
|----------|-----------------------------|--|
| п | 1200 | |
| ш | 900 | |
| IV | 980 | |
| v | 900 | |
| VI | 900 | |
| | | |

¹H NMR (270 MHz) (CDCl₃, δ): 2.86–3.04 (m, 2H, SCH₂), 3.38 (s, 3H, N₃–CH₃), 3.57 (s, 3H, N₁–CH₃), 3.80–3.89 (m, 1H, OCH), 4.24–4.31 (m, 1H, OCH), 4.52 (dd, 1H, CH_a–CH_c, JH_aH_b = 14 Hz, JH_aH_c = 6.6 Hz), 4.66 (dd, 1H, CH_b–CH_c, JH_bH_c = 3.3 Hz, JH_aH_b = 14 Hz), 5.43 (dd, 1H, CH₂–CH_c, JH_aH_c = 3.3 Hz, JH_bH_c = 6.6 Hz), 7.64 (s, 1H, N=CH). IR (CHCl₃): 1705, 1660, 1605, 1550 cm⁻¹. UV (CH₃OH): $\lambda_{max} = 275$, 205 nm. MS (EI): m/z:282 (M⁺, 13.2), 194 (33.1), 193 (6.5), 180 (31.7), 103 (83.6), 89 (100).

7-(1,3-Dithiolan-2-ylmethyl)theophylline-1S-oxide V, VI. 2 ml 35% H₂O₂ were added to a solution of 7-(1,3-dithiolan-2-ylmethyl)theophylline **II** (7 g, 0.031 mol) in 75 ml acetic acid.

The mixture was stirred for 5 h at room temperature then 500 ml diethyl ether was added to induce precipitation. After overnight storage at -30° C, the precipitate was filtered and the

After overnight storage at -30° C, the precipitate was filtered and the 2 diastereomers were separated by flash-chromatography on a silica-gel column: elution with CH₂Cl₂/CH₃OH 97.5/2.5 gave the *cis* isomer, followed by the *trans* isomer on eluting with CH₂Cl₂/CH₃OH (96.5/3.5) mixture (64% yield, 55:45 *trans:cis*).

The 2 diastereomers melted with decomposition at 204-206 °C (CH₃OH) for the *trans* isomer and at 200-202 °C (CH₃OH) for the *cis* isomer.

trans-isomer: ¹H NMR (270 MHz) (CDCl₃, δ): 2.98 (m, 1H, 1,3 dithiolane ring), 3.41 (s, 3H, N₃-CH₃), 3.56 (m, 2H, 1,3 dithiolane ring), 3.61 (s, 3H, N₁-CH₃), 3.80 (m, 1H, 1,3 dithiolane ring), 4.45 (dd, 1H, CH_a-CH_c), 4.65 (m, 2H, CH_b-CH_c), 7.73 (s, 1H, CH=N). IR (CHCl₃): 1700, 1655, 1550 cm⁻¹. UV (CH₃OH): $\lambda_{max} = 275, 207$ nm. MS (EI): m/z: 314 (M⁺, 76.9), 298 (12.2), 297 (79.1), 286 (56.6), 238 (36.1), 237 (11.9), 206 (26.8), 205 (100), 193 (14.3), 181 (17.7), 180 (81.0).

cts-isomer: ¹H NMR (270 MHz) (CDCl₃, δ): 2.84 (m, 1H, 1,3-dithiolane ring), 3.41 (s, 3H, N₃-CH₃), 3.45 (m, 1H, overlapped, 1,3-dithiolane ring), 3.61 (s, 3H, N₁-CH₃), 3.83 (m, 1H, 1,3-dithiolane ring), 4.64 (dd, 1H, CH_a-CH_c, JH_aH_b = 14 Hz, JH_aH_c = 8.6 Hz), 4.82 (dd, 1H, CH_b-CH_c, JH_aH_b = 14 Hz, JH_bH_c = 6 Hz), 5.0 (dd, 1H, CH_a+_b-CH_c, JH_aH_c = 8.6 Hz), 7.75 (s, 1H, CH=N). IR (CHCl₃): 1700, 1655, 1550 cm⁻¹. UV (CH₃OH) λ_{max} = 275, 207 nm. MS (EI): *m* / *z*: 314 (M⁺, 87.4), 298 (13.2), 297 (85.3), 286 (59.4), 238 (37.8), 237 (12.4), 206 (26.5), 205 (100), 181 (17.3), 180 (82.3)

Thermal rearrangement of V0.5 g of V were dissolved in 20 ml of anhydrous dimethylsulfoxide and heated at 100°C for 64 h. The reaction was then diluted with 100 ml H₂O and extracted with CHCl₃ (3 × 60 ml). The residue was purified by preparative TLC with CHCl₃ as eluant.

¹H NMR (270 MHz) ($CDCl_3$, δ): 3.33 (2H, m, $-CH_2-S-$), 3.42 (3H, s, N₃- CH_3), 3.46 (2H, m, $-CH_2-S-$), 3.60 (3H, s, N₁- CH_3), 6.56 (1H, s, -CH=C), 7.66 (1H, s, -CH=N-). MS (EI): m/z: 296 (M⁺, 100), 263 (38.2), 181 (71.9), 180 (68.1), 116 (56.3).

| Table III. | . Mucoprod | uction ir | ı rabbits | before and | after oral | treatment | with II , | doxophylline, | bromhexyne and | N-acetylcysteine. |
|------------|------------|-----------|-----------|------------|------------|-----------|------------------|---------------|----------------|-------------------|
|------------|------------|-----------|-----------|------------|------------|-----------|------------------|---------------|----------------|-------------------|

| Substance | n | Dose (mg/kg) | Mucoproduction (mg/h (mean ± SD) |) | % Variation with respect to its own | Statistical significance with respect to | |
|------------------|----------------|-----------------|--|--|---|--|--|
| | | | -4-0 h (before treatment) | 0-4 h (after treatment) | base | its own base | |
| Control | 10 | _ | 29.50 ± 8.32 | 31.25 ± 5.93 | + 5.93 | N.S. | |
| П | 10 10 10 | 10 30 100 | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | $\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$ | +13:95 +26.95 +57.27 | N.S. <i>P</i> <0.05 <i>P</i> <0.01 | |
| Doxophylline | 10 | 100 | 29.80 ± 5.80 | 32.10 ± 8.26 | + 7.71 | N.S. | |
| Bromhexyne | 10 | 400 | 28.90 ± 7.82 | 39.01 ± 4.34 | +34.98 | <i>P</i> <0.05 | |
| N-acetylcysteine | 10 | 400 | 27.70 ± 5.50 | 32.19 ± 3.89 | +16.21 | <i>P</i> <0.05 | |

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Pharmacology

In vitro activity on guinea-pig trachea smooth muscle

Male guinea-pig trachea was allowed to function in vitro in 50 ml Krebs' solution, at 37°C under suitable oxygenation.

The lower part of the trachea was closed, while the upper part was connected to a small polyethylene tube leading to a transducer to measure volume variations. Both the trachea and the polyethylene tube were filled with Krebs' solution and the responses to histamine (5 μ g/ml) were determined before and after the addition of theophylline derivatives at increasing concentrations.

In vivo studies on smooth muscles

The method of Konzett and Rossler [20] was utilized with minor modifications [21]. Male guinea-pigs weighing 300-450 g, under urethane anaesthesia (1.0 g/kg, i.p.), were used

The jugular vein of each animal was cannulated for intravenous administration of drugs, while the trachea was connected to pump for artificial respiration (Palmer pump for small animals: stroke frequency 70 insufflations per minute).

The pneumogram was recorded on polygraph (Ugo Basile, Milano, Italy)

When the animals were stabilized, a bronchospasm was stimulated with acetylcholine (0.2 mg/kg i.v.).

After 2 similar responses to spasm inducing injections, theophylline derivatives were administered (0.02-10 mg/kg, i.v.) followed, 2 to 5 min later, by the bronchoconstrictor drug.

The effects of the tested compounds were evaluated with reference to the percentage reduction of the induced bronchoconstriction.

Acute toxicity

Male Swiss mice weighing 18-22 g were used. Test compounds suspended in 0.5% carboxymethylcellulose, were administered orally.

The LD₅₀ values were calculated according Litchfield and Wilcoxon [22], 10 days after test compounds were administered.

The maximal toxicity was observed after 24 h, when the animals showed decreased muscle tone and laboured respiration signs.

Mucolytic activity

For this test a modified method described by Perry and Boyd [23], was used. Groups of 10 New Zealand male, while rabbits, weighing 2.8-3.5 kg, were used.

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