

FIVE DITERPENES AND OTHER CONSTITUENTS FROM
NINE *BACCHARIS* SPECIES*FERDINAND BOHLMANN,† CHRISTA ZDERO,† MICHAEL GRENZ,† AUTAR K. DHAR,† HAROLD ROBINSON‡ and
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Key Word Index—*Baccharis* spp.; Compositae; diterpenes; new clerodanes; new labdanes; new flavanones; new aromatics.

Abstract—The investigation of nine *Baccharis* species afforded, in addition to known compounds, three new clerodane derivatives related to bacchotricuneatin B, two *ent*-labdane derivatives, two 3-acetoxyflavanones, a benzofuran derivative and a chromene. The overall picture of the chemistry of this large genus is still not very clear.

INTRODUCTION

The results on the chemistry of the large genus *Baccharis* (tribe Astereae, Compositae) so far available do not show a clear picture. In addition to simple acetylenes, typical for large parts of the tribe [1], clerodanes [2–7] and the triterpene baccharis oxide [6, 8, 9, 16] are widespread. Also the presence of flavones and flavanones have been reported several times [3, 5, 6, 10–14], while from one species unusual macrolide sesquiterpenes [15] and from a few, in addition to widespread ones, some unusual *p*-hydroxyacetophenone derivatives were isolated [6, 16]. We now have investigated nine further Brazilian species. Again a very complex picture was obtained.

RESULTS AND DISCUSSION

The aerial parts of *Baccharis cassinaefolia* DC. afforded germacrene D, bicyclgermacrene, squalene, lupenone, euphol acetate, spathulenol [17], lachnophyllum ester (1) [1], the chromene 2 [18] and the aldehyde 5 [16]. Furthermore, baccharisoxide (25) [8], as well as a mixture of diterpenes were present. Even by HPLC this mixture could be separated only in part. The ¹H NMR data indicated that the compounds were diesters differing in the ester residues only, which were angelate, senecioate and 2-methylbutyrate. The MS, however, showed no molecular ions; only the M – RCO₂H peaks were detected. Partial saponification led to the formation of two monoesters, both with an additional methoxy group, obviously introduced by the solvent. Careful ¹H NMR studies led to the structures 14 and 15 (Table 1). In connection with the ¹H NMR data of the diesters the formation of 14 and 15 could be explained only if an elimination–addition reaction were assumed. Treatment of the diesters with potassium hydroxide first led to the elimination products 13a and b, which by addition of

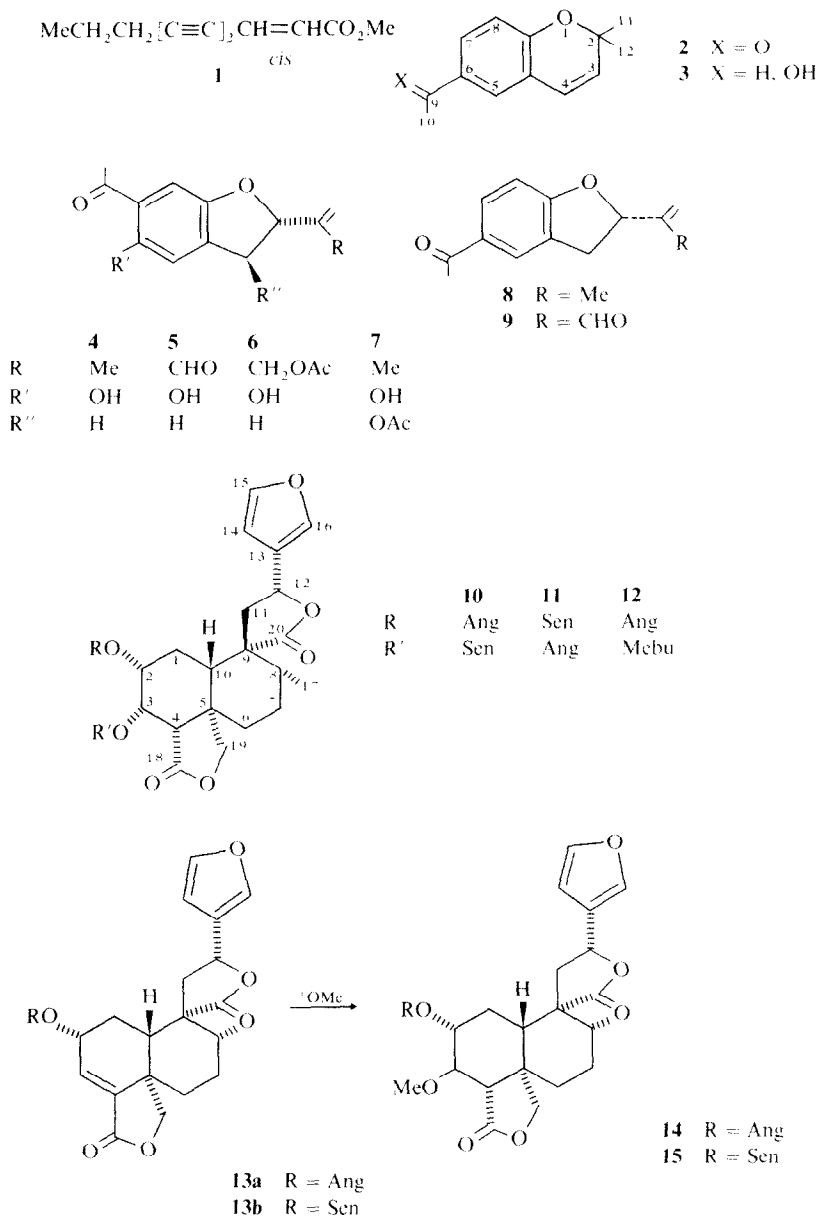
methanol were transformed to the methoxy compounds 14 and 15. As could be visualized from the 3-H couplings, the stereochemistry at C-3 had changed from an axial to an equatorial oxygen function. The ¹H NMR data of one of the diesters separated by HPLC showed the presence of an angelate and a 2-methylbutyrate residue. As the latter is missing in the methoxy compounds, obtained from the whole mixture, the 2-methylbutyrate must be placed at C-3; consequently the structure of this compound is 12 and those of the two other diesters 10 and 11, as both methoxy compounds were obtained from the mixture of these two diester, which could not be separated. Compounds 10–12 are closely related to bacchotricuneatin B, where the stereochemistry was established by X-ray [5]. The ¹H NMR data are very similar to those of 10–12 and consequently the stereochemistry most probably is the same at C-5, C-8 to C-10 and C-12. We have therefore named the new compounds as diacyloxy-3,4βH-bacchotricuneatin B.

The roots again contain germacrene D, squalene, 1, 2, 5 and 25 as well as 8 [19], 9 [20] and the carbinol 3, its structure following from the spectral data which were identical with those of the reduction product of 2 (see Experimental).

The aerial parts of *B. intermixta* Gardn. afforded germacrene D, β-farnesene, *ent*-kaurenic acid (18), grandifloric acid (20) [21], sakuranetin (27) and two diterpenes, the diacetate 16a and the triacetate 17a, which could be purified only after saponification to 16b and 17b. 17b already has been isolated previously from *Achyrocline alata* and was transformed to the triacetate 17a [22]. The structures clearly followed from the ¹H NMR data of the alcohols (Table 2). The roots afforded germacrene D, 5 and 6 [16] and 17a.

The aerial parts of *B. calvenscens* DC. afforded germacrene D, bicyclgermacrene, caryophyllene, squalene, friedelin, oleanolic acid, lupeol, spathulenol [17], 2, 5, the angelate 24 [23] and the clerodane derivative 23 [7], while the roots contain squalene, lachnophyllum ester (1), 2, 4, 5, 8, 25 and the acetate 7, which has not been

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reported previously. The structures clearly followed from the ^1H NMR spectrum (see Experimental).

The aerial parts of *B. varians* Gardn. afforded benzylbenzoate, naringenin (**26**), eriodictyol (**28**) and two further flavanones, the acetates **29** and **30**. **29** was transformed to the acetate **31**. The structures clearly followed from the ^1H NMR data (Table 3), which also indicated the *trans* stereochemistry at C-2 and C-3. The roots only gave baccharis oxide (**25**).

The aerial parts of *B. serraluta* (Lam.) Pers. afforded germacrene D, bicylogermacrene, squalene, caryophyllene epoxide and sakuranetin (**27**), while the roots gave baccharis oxide (**25**).

The aerial parts of *B. reticularia* DC. afforded germacrene D, bicylogermacrene, squalene, lupenone, lupeol, baccharis oxide (**25**) and eriodictyol (**28**) [24], while the roots gave **5** and **25**.

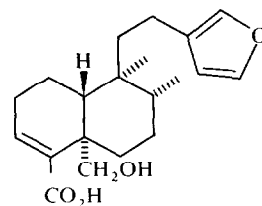
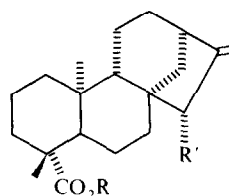
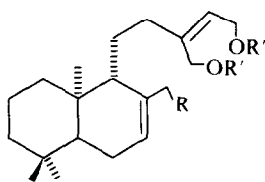
The aerial parts of *B. polyphylla* Gardn. afforded germacrene D, γ -humulene, bicylogermacrene, *ent*-kaurenic acid (**18**), grandifloric acid (**20**), large quantities of its methyl ester (**21**) and the angelate **22a** [16] as well as the cinnamic acid derivatives **32** [25] and **33** [25]. The roots gave bicylogermacrene and **25** only.

The aerial parts of *B. ramosissima* Gardn. afforded germacrene D, bicylogermacrene, lupeol, lupenone, **2**, **5**, **25** and a triterpene alcohol, its structure could not be established. All data agree with an isomer of euphol with an α -hydroxy group and a 9,11-double bond. The stereochemistry, however, could not be determined.

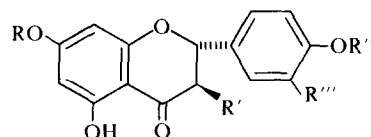
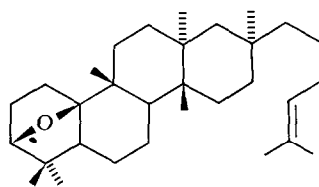
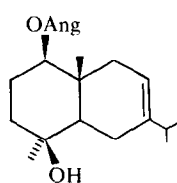
The aerial parts of *B. salzmännii* DC. afforded germacrene D, bicylogermacrene, caryophyllene epoxide, spathulenol, *ent*-kaurenic acid (**18**), the tiglate **22b** [26], naringenin (**26**) and three triterpenes, a ketone, an acetate and an alcohol, closely related to that from *B.*

Table 1. ^1H NMR spectral data of compounds **10–12**, **14** and **15** (270 MHz, CDCl_3 , TMS as int. standard)

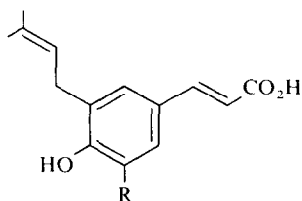
	10	11	12	14†	15‡
1-H		2.21 <i>ddd</i>		2.20 <i>m</i>	2.33 <i>ddd</i>
1'-H					1.42 <i>m</i>
2-H		5.42 <i>m</i>		5.40 <i>m</i>	4.94 <i>ddd</i>
3-H	6.52 <i>dd</i>	6.49 <i>dd</i>	6.45 <i>dd</i>		3.57 <i>dd</i>
4-H		1.95 <i>br. s*</i>			2.29 <i>d</i>
11-H		2.60 <i>dd</i>	2.61 <i>dd</i>	2.48 <i>d</i>	
11'-H		2.52 <i>dd</i>	2.53 <i>dd</i>		2.47 <i>d</i>
12-H		5.48 <i>br. t</i>	5.45 <i>br. t</i>		5.42 <i>dd</i>
14-H		6.38 <i>dd</i>	6.36 <i>dd</i>		6.39 <i>dd</i>
15-H		7.43 <i>dd</i>			7.42 <i>dd</i>
16-H		7.47 <i>br. s</i>	7.43 <i>br. s</i>		7.46 <i>dd</i>
17-H		1.15 <i>d</i>	1.14 <i>d</i>		1.13 <i>d</i>
19-H	4.68 <i>d</i>	4.67 <i>d</i>	4.68 <i>d</i>	4.53 <i>d</i>	4.50 <i>d</i>
19'-H	4.06 <i>dd</i>	4.05 <i>dd</i>	4.06 <i>dd</i>	4.38 <i>dd</i>	4.37 <i>dd</i>

* In C_6D_6 , † OMe 3.50 *s*, ‡ OMe 3.48 *s*.OAng: 6.13 *qq*, 1.96 *dq*, 1.87 *dq*, [J (Hz): 3',4' = 7; 3',5' = 4',5' = 1.3]; OSen: 5.65 *qq*, 2.15 *d*, 1.93 *d* [J (Hz): 3',4' = 3',5' = 1.3]; OMebu: 2.36 *iq*, 1.60 *ddq*, 1.42 *ddq*, 0.86 *t*, 1.13 *d* [J (Hz): 2',3' = 2',5' = 3',4' = 7; 3',3'_2 = 14]. J (Hz): 1,1' = 13; 1,2 = 7.5; 1',2 = 7; 1,10 = 3.5; 2,3 = 3,4 ~ 1.5 (**14/15**: 2,3 = 7; 3,4 = 6.5); 6,19' = 1.5; 8,17 = 7; 11,12 = 8; 12,12' = 15; 14,15 = 15,16 ~ 1.5; 14,16 = 1; 19,19' = 10.

16a	16b	17a	17b	18	19	20	21	22a	22b	23
R	H	H	OH	R	H	H	H	H	H	
R'	Ac	H	H	R'	H	OH	OH	OAng	OTigl	



	26	27	28	29	30	31
R	H	Me	H	H	H	Ac
R'	H	H	H	OAc	OAc	OAc
R''	H	H	H	H	H	Ac
R'''	H	H	OH	OH	H	OAc



32 R = H
33 R = $\text{CH}_2\text{CH}=\text{CMe}_2$

Table 2. ^1H NMR spectral data of compounds **16a** and **17a**

	16a	17a
7-H	5.40 <i>br. s</i>	5.75 <i>br. d</i>
12-H	2.38 <i>br. ddd</i>	2.49 <i>br. ddd</i>
12'-H	2.07 <i>m</i>	2.08 <i>m</i>
14-H	5.65 <i>br. t</i>	5.62 <i>br. t</i>
15-H	4.24 <i>br. d</i>	4.18 <i>br. d</i>
16-H	4.24 <i>d</i>	4.18 <i>d</i>
16'-H	4.18 <i>d</i>	4.00 <i>d</i>
17-H	1.70 <i>br. s</i>	4.18 <i>br. s</i>
18-H	0.87 <i>s</i>	0.89 <i>s</i>
19-H	0.86 <i>s</i>	0.87 <i>s</i>
20-H	0.77 <i>s</i>	0.77 <i>s</i>

J (Hz): 6,7 = 5; 11,12 = 11; 11',12 = 5; 12,12' = 13; 14,15 = 7; 16,16' = 12.

ramoissima. The structures could not be estimated. The roots yielded bicyclogermacrene, γ -humulene, squalene, **5**, **18** and baccharis oxide (**25**).

Comparing the chemistry of the *Baccharis* species investigated now, it is obvious that some species can be characterized by the presence of baccharis oxide, while others have typical clerodane derivatives. These are sometimes replaced by *ent*-kaurenes, while so far from one species only labdanes were isolated. Another group can be characterized by flavanones, which often co-occur with baccharis oxide. Probably the benzofurans **4**–**7** are also of chemotaxonomic importance as the unusual position of the acetyl group seems to be very rare. Surely further investigations are necessary to get a clearer picture.

EXPERIMENTAL

^1H NMR: 270 MHz, TMS as int. stand; MS: 70 eV, direct inlet. The air-dried plant material, collected in north-eastern

Brazil, was extracted with Et_2O -petrol, 1:2 and the extracts obtained were separated first by column chromatography (SiO_2 , act. grade II) and further by repeated TLC (SiO_2 , GF 254). The mixture of **10**–**12** could be separated by HPLC (reversed phase, MeOH– H_2O , 7:3) in part. Known compounds were identified by comparing the IR and ^1H NMR data with those of authentic material.

Baccharis cassinaefolia (voucher RMK 8003). The roots (900 g) afforded 5 mg germacrene D, 80 mg squalene, 80 mg **1**, 20 mg **2**, 8 mg **3** (Et_2O -petrol, 1:3), 50 mg **5**, 8 mg **8**, 8 mg **9** and 100 mg **25**, while the aerial parts (800 g) yielded 10 mg germacrene D, 30 mg squalene, 2 mg bicyclogermacrene, 5 mg lupenone, 15 mg euphol acetate, 2 mg spathulenol, 2 mg **1**, 4 mg **2**, 3 mg **5**, 20 mg **10** and **11** (ca 1:1, Et_2O -petrol, 3:1), 7 mg **12** (Et_2O -petrol, 3:1) and 10 mg **25**.

Baccharis intermixta (voucher RMK 8139). The roots (400 g) afforded 20 mg germacrene D, 4 mg **5**, 4 mg **6** and 65 mg **17a** (Et_2O -petrol, 1:1), while the aerial parts (800 g) yielded 260 mg germacrene D, 130 mg β -farnesene, 0.2 g **16a** (Et_2O -petrol, 1:1), 2 g **17a**, 500 mg **18**, 220 mg **20** and 300 mg **27**.

Baccharis calvescens (voucher RMK 7989). The roots (150 g) afforded 10 mg squalene, 2 mg **1**, 1 mg **2**, 10 mg **4**, 10 mg **5**, 1 mg **7** (Et_2O -petrol, 1:1), 2 mg **8** and 20 mg **25**, while the aerial parts (1 kg) yielded 50 mg squalene, 20 mg germacrene D, 5 mg bicyclogermacrene, 20 mg caryophyllene, 30 mg friedelin, 100 mg lupeol, 50 mg oleanolic acid, 3 mg spathulenol, 1 mg **2**, 10 mg **5**, 20 mg **23** and 3 mg **24**.

Baccharis varians (voucher RMK 8089). The roots (25 g) afforded 25 mg **25**, while the aerial parts (100 g) yielded 60 mg benzyl benzoate, 4 mg **26**, 16 mg **28**, 20 mg **29** (Et_2O) and 6 mg **30** (Et_2O).

Baccharis serrulata (voucher RMK 8058). The roots (40 g) afforded 18 mg **25** and the aerial parts (200 g) 18 mg germacrene D, 2 mg bicyclogermacrene, 40 mg squalene, 2 mg caryophyllene epoxide and 105 mg **27**.

Baccharis reticularia (voucher RMK 8106). The roots (100 g) afforded 3 mg **5**, 80 mg **25** and the aerial parts (1 kg) 50 mg germacrene D, 5 mg bicyclogermacrene, 30 mg squalene, 30 mg lupenone, 80 mg lupeol, 50 mg **25** and 30 mg **28**.

Baccharis polyphylla (voucher RMK 8108). The roots (100 g) afforded 10 mg bicyclogermacrene and 50 mg **25**, while the aerial

Table 3. ^1H NMR spectral data of compounds **29**–**31**

	29 ($(\text{D}_3\text{C})_2\text{CO}$)	30 ($(\text{D}_3\text{C})_2\text{CO}$)	31 (CDCl_3)
2-H	5.36 <i>d</i>	5.43 <i>d</i>	5.42 <i>d</i>
3-H	5.84 <i>d</i>	5.88 <i>d</i>	5.75 <i>d</i>
6-H	6.00 <i>d</i>	6.00 <i>d</i>	6.79 <i>d</i>
8-H	6.03 <i>d</i>	6.04 <i>d</i>	6.61 <i>d</i>
2'-H	7.08 <i>br. s</i>	7.42 <i>d</i>	7.31 <i>d</i>
3'-H	6.90 <i>m</i> }	6.91 <i>d</i>	7.28 <i>d</i>
5'-H			
6'-H			
OH	1.98 <i>s</i>	1.98 <i>s</i>	2.08 <i>s</i> 2.30 <i>s</i> 2.32 <i>s</i> 2.32 <i>s</i>
OH	11.58 <i>s</i>	11.59 <i>s</i>	11.28 <i>s</i>

J (Hz): 2,3 = 12; 6,8 = 1.7; 2',3' = 8; 2',6' = 1.5.

parts (350 g) gave 150 mg germacrene D, 600 mg γ -humulene, 50 mg bicyclogermacrene, 400 mg **18**, 100 mg **20**, 1.5 g **21**, 50 mg **22a**, 40 mg **32** and 50 mg **33**.

Baccharis ramoississima (voucher RMK 8176). The aerial parts (300 g) afforded 20 mg germacrene D, 10 mg bicyclogermacrene, 30 mg lupeol, 20 mg lupenone, 2 mg **2**, 3 mg **5**, 10 mg **25** and 30 mg of a triterpene alcohol, whose structure could not be fully established.

Baccharis salzmännii (voucher RMK 8056). The roots (360 g) afforded 5 mg γ -humulene, 5 mg bicyclogermacrene, 40 mg squalene, 5 mg **5**, 300 mg **18** and 520 mg **25**, while the aerial parts (820 g) yielded 100 mg germacrene D, 3 mg bicyclogermacrene, 20 mg caryophyllene epoxide, 3 mg spathulenol, 180 mg **18**, 5 mg **22b**, 500 mg **26**, 10 mg of a triterpene ketone, 10 mg of a triterpene acetate and 10 mg of a triterpene alcohol, their structures not being established.

[1-*Hydroxyethyl*]-2,2-dimethylchromene (**3**). Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3620 (OH), 1495, 1270, 1160, 965; MS m/e (rel. int.): 204.115 (M^+ , 11) ($C_{13}H_{16}O_2$), 189 ($M - \text{Me}$, 100), 171 (189 – H_2O , 12), 161 (189 – CO, 7); 1H NMR ($CDCl_3$): 5.62 (*d*, 3-H), 6.31 (*d*, 4-H), 6.73 (*d*, 8-H), 7.08 (*dd*, 7-H), 6.99 (*d*, 5-H), 4.78 (*q*, 9-H), 1.45 (*d*, 10-H), 1.42 (*s*, 11,12-H) ($J = (\text{Hz}): 3.4 = 9.5; 5.7 = 2; 7.8 = 8; 9.10 = 7$). $LiAlH_4$ reduction of **2** afforded a compound, its spectral data being identical with those of **3**.

3β -Acetoxy-6-acetyl-2 α -isopropenyl-5-hydroxy-2,3H-benzofuran (**7**). Colourless oil, 3500–2700, 1660 (chelated hydroxy ketone), 1755 (OAc); MS m/e (rel. int.): 276.100 (M^+ , 80) ($C_{15}H_{16}O_5$), 216 ($M - \text{HOAc}$, 100), 201 (216 – Me , 95); 1H NMR ($CDCl_3$): 4.98 *d*, 2-H), 6.12 (*d*, 3-H), 7.06 (*s*, 4-H), 7.23 (*s*, 7-H), 2.63 (*s*, 9-H), 5.09 and 4.98 (*br. s*, 11-H), 1.76 (*br. s*, 12-H).

2 α -Angeloyloxy-3 α -seneciolyoxy and 2 α -seneciolyoxy-3 α -angeloyloxy-3,4 β H-bacchotricuneatin (**10** and **11**). Colourless, crystalline mixture, which could not be separated, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1775, 1765 (lactone), 1725, 1710 (CO_2R), 1595, 880 (furan); MS m/e (rel. int.): 440.184 ($M - C_4H_7CO_2H$, 2) ($C_{25}H_{38}O_7$), 422 (440 – H_2O , 2), 382 (440 – CH_2CO_2 , 1), 340 (440 – $C_4H_7CO_2H$, 3), 310 (340 – CH_2O , 3), 83 ($C_4H_7CO^+$, 100), 55 (83 – CO, 39).

$$[\alpha]_{24}^D = \frac{589}{-42.6} \quad \frac{578}{-44.7} \quad \frac{546}{-51.6} \quad \frac{436 \text{ nm}}{-97.9}$$

($c = 0.95$, $CHCl_3$).

2 α -Angeloyloxy-3 α -[2-methylbutyryloxy]-3,4 β H-bacchotricuneatin (**12**). Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1775, 1765 (lactone), 1730, 1710 (CO_2R), 880 (furan); MS m/e (rel. int.): 442 ($M - C_4H_7CO_2H$, 3), 440 ($M - C_4H_9CO_2H$, 3), 340 (440 – $C_4H_7CO_2H$, 6), 310 (340 – CH_2O , 9), 85 ($C_4H_9CO^+$, 24), 83 ($C_4H_7CO^+$, 100), 57 (85 – CO, 48).

Partial saponification of **10–12**. To 15 mg **10–12** in 2 ml MeOH 200 mg KOH in 0.5 ml H_2O were added. After 1 min dil. H_2SO_4 was added. TLC (Et_2O) afforded 6 mg **14**, colourless crystals (Et_2O –petrol), mp 195, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1785, 1768 (lactone), 1720 ($C = CCO_2R$), 880 (furan); MS m/e (rel. int.): 472.210 (M^+ , 48) ($C_{26}H_{32}O_8$), 442 ($M - CH_2O$, 5), 440 ($M - \text{MeOH}$, 2), 372 ($M - C_4H_7CO_2H$, 22), 343 (372 – CHO, 8), 312 (343 – OMe, 45), 83 ($C_4H_7CO^+$, 90), 55 (83 – CO, 100). The second fraction still was a mixture of **14** and **15** as shown by the 1H NMR spectrum (Table 1).

15,16-Diacetoxy-ent-labda-6,14-diene (**16a**). Colourless gum, purified as its diol **16b** ($LiAlH_4$ in Et_2O , room temp.), colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3620 (OH), 860 ($CH=C$); MS m/e (rel. int.): 306.256 (M^+ , 4) ($C_{20}H_{34}O_2$), 288 ($M - H_2O$, 5), 273

(288 – Me , 5), 204 ($M - C_5H_{10}O_2$, 100).

$$[\alpha]_{24}^D = \frac{589}{+1.8} \quad \frac{578}{+1.9} \quad \frac{546}{+2.4} \quad \frac{436 \text{ nm}}{+2.9}$$

($c = 0.34$, $CHCl_3$).

15,16,17-Triacetoxy-ent-labda-6,14-diene (**17a**). IR and 1H NMR data identical with those of the triacetate prepared previously [16]. $LiAlH_4$ reduction afforded the triol **17b**, colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3620 (OH), 860 ($CH=C$); MS m/e (rel. int.): 304 ($M - H_2O$, 0.3), 286.230 (304 – H_2O , 15) ($C_{20}H_{36}O$), 273 (288 – Me , 5), 220 ($C_{15}H_{24}O$, 25), 109 ($C_8H_{13}^+$, 100).

3β -Acetoxyeriodictyol (**29**). Colourless gum, MS m/e (rel. int.): 346 (M^+ , 1), 286 ($M - \text{HOAc}$, 5), 65 (100). 10 mg **29** in 0.2 ml Ac_2O were heated for 2 hr at 70. Evaporation and TLC (Et_2O –petrol, 3:1) afforded 10 mg **31**, Colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 1790 (PhOAc), 1665, 1635, 1580; MS m/e (rel. int.): 472.101 (M^+ , 16) ($C_{23}H_{20}O_{11}$), 430 ($M - \text{ketene}$, 22), 412 ($M - \text{HOAc}$, 15), 370 (430 – HOAc , 24), 57 (100).

3β -Acetoxyneringenin (**30**). Colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3600 (OH), 1715 ($C\equiv$); MS m/e (rel. int.): 330.074 (M^+ , 20) ($C_{17}H_{14}O_7$), 270 ($M - \text{HOAc}$, 55), 153 (100), 136 (72).

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