

Synthesis and Antitumor Activity of 3- and 5-Hydroxy-4-methylpyridine-2-carboxaldehyde Thiosemicarbazones

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Received April 21, 1992

To develop an α -(N)-heterocyclic carboxaldehyde thiosemicarbazone with clinical utility as an anticancer agent, two analogues, 3-hydroxy-4-methylpyridine-2-carboxaldehyde thiosemicarbazone (3-HMP) and 5-hydroxy-4-methylpyridine-2-carboxaldehyde thiosemicarbazone (5-HMP), of 5-hydroxypyridine-2-carboxaldehyde thiosemicarbazone (5-HP) have been designed and synthesized by two different methods. 3-HMP and 5-HMP both showed better antitumor activity than their respective parent compounds, 3-hydroxypyridine-2-carboxaldehyde thiosemicarbazone and 5-HP, in mice bearing the L1210 leukemia.

The α -(N)-heterocyclic carboxaldehyde thiosemicarbazones (HCTs) constitute, as a class, the most potent known inhibitors of ribonucleoside diphosphate reductase. The reductive conversion of ribonucleotides to their deoxyribonucleotide counterparts is a particularly critical step in the synthesis of DNA, since deoxyribonucleotides are present in extremely low levels in mammalian cells, and Corey and Chiba¹ have presented arguments that an inhibitor of ribonucleotide reductase could be more effective than an inhibitor of DNA polymerase in blocking DNA synthesis. Thus, it seems reasonable that a strong inhibitor of ribonucleotide reductase would be a useful weapon in the therapeutic armamentarium against cancer. 5-Hydroxypyridine-2-carboxaldehyde thiosemicarbazone (5-HP) is the only member of the HCT series that has been administered to man as part of a phase 1 study. The selection of 5-HP for clinical trial was due to (a) its activity against a spectrum of transplanted tumors and spontaneous dog lymphomas and (b) its ease of parenteral administration as the sodium salt. The results of two independent phase 1 studies^{2,3} showed that transient decreases in blast counts occurred in 6 of 25 patients with leukemia, while no antitumor effects were observed in 18 patients with solid tumors. Administration of relatively large doses of drug was limited primarily by gastrointestinal toxicity. In addition, the most aggressive drug regimens also produced myelosuppression, hemolysis, anemia, hypertension, and hypotension. The exceedingly weak antileukemic activity of 5-HP that was observed in the phase 1 trial was attributed to the relatively short biological half-life of 5-HP in humans, which was due to the rapid formation and elimination of the *O*-glucuronide conjugate. Thus, the $t_{1/2}$ of 5-HP in the blood of mice was 15 min, while the drug had a $t_{1/2}$ in humans of 2.5 to 10.5 min, depending upon the patient. Twenty percent of a therapeutic dose of 5-HP was excreted in the urine of the mouse within 24 h; whereas, a therapeutic dose of 5-HP was

excreted 2- to 3.5-times faster in man. Approximately 75% of the material found in the urine of patients was in the form of an *O*-glucuronide,² which had no activity against ribonucleotide reductase. In an attempt to circumvent this problem, our laboratory designed and synthesized 5-amino-4-methylisoquinoline-1-carboxaldehyde thiosemicarbazone, an isoquinoline derivative containing a 5-amino function to permit formulation as an acid salt and a 4-methyl group, which we have shown provides steric protection of the 5-NH₂ substituent from enzymatic acetylation, a reaction that eliminates anticancer activity.^{4,5} Unfortunately, this promising agent was judged to not be sufficiently water soluble to permit adequate formulation for use in man. For this reason, we have synthesized hydroxy-substituted pyridine thiosemicarbazones, which are significantly more soluble as sodium salts.

Chemistry

3-Hydroxy-4-methylpyridine-2-carboxaldehyde thiosemicarbazone (14, 3-HMP) and 5-hydroxy-4-methylpyridine-2-carboxaldehyde thiosemicarbazone (17, 5-HMP) were synthesized by well-documented methodology⁶⁻⁸ as shown in Schemes I and II, respectively. Compounds 2-7 were synthesized by minor modifications to the procedures described by Furukawa.⁹ 2,4-Lutidine (1) was nitrated to give the two isomers, 2,4-dimethyl-3- and 5-nitropyridine (2 and 3, respectively), in approximately equal amounts. Catalytic hydrogenation of compounds 2 and 3 over 5% Pd/C in absolute ethanol gave the corresponding amino derivatives 4 and 5. Diazotization of compounds 4 and 5

(1) Cory, J. G.; Chiba, P. Combination Chemotherapy Directed at the Components of Nucleoside Diphosphate Reductase. In *Inhibitors of Ribonucleoside Diphosphate Reductase Activity*; Cory, J. G., Cory, A. M., Eds.; Pergamon Press: Oxford, 1989; pp 245-264.

(2) DeConti, R. C.; Toftness, B. R.; Agrawal, K. C.; Tomchick, R.; Mead, J. A. R.; Bertino, J. R.; Sartorelli, A. C.; Creasey, W. A. Clinical and Pharmacological Studies with 5-Hydroxy-2-formylpyridine Thiosemicarbazone. *Cancer Res.* 1972, 32, 1455-1462.

(3) Krakoff, I. H.; Etcubanas, E.; Tan, C.; Mayer, K.; Bethune, V.; Burchenal, J. H. Clinical Trial of 5-Hydroxypicolinaldehyde Thiosemicarbazone (5-HP; NSC-107392) With Special Reference to its Iron-Chelating Properties. *Cancer Chemother. Rep.* 1974, 58, 207-212.

(4) Agrawal, K. C.; Schenkman, J. B.; Denk, H.; Mooney, P. D.; Moore, E. C.; Wodinsky, I.; Sartorelli, A. C. 4-Methyl-5-amino-1-formylisoquinoline Thiosemicarbazone, a Second-Generation Antineoplastic Agent of the α -(N)-Heterocyclic Carboxaldehyde Thiosemicarbazone Series. *Cancer Res.* 1977, 37, 1692-1696.

(5) Agrawal, K. C.; Mooney, P. D.; Sartorelli, A. C. Potential Antitumor Agents. 13. 4-Methyl-5-amino-1-formylisoquinoline Thiosemicarbazone. *J. Med. Chem.* 1976, 19, 970-972.

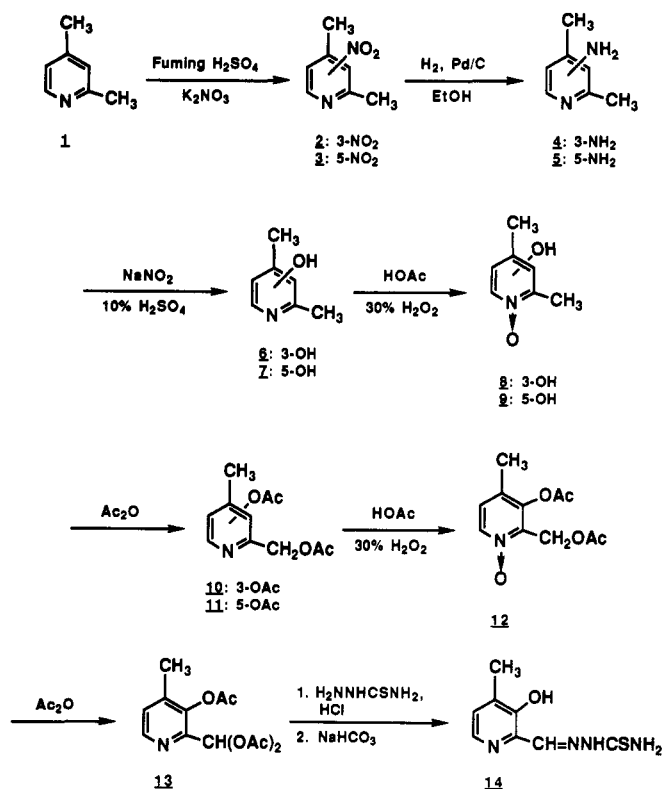
(6) Blanz, E. J., Jr.; French, F. A. The Carcinostatic Activity of 5-Hydroxy-2-formylpyridine Thiosemicarbazone. *Cancer Res.* 1968, 28, 2419-2422.

(7) Agrawal, K. C.; Lin, A. J.; Booth, B. A.; Wheaton, J. R.; Sartorelli, A. C. Potential Antitumor Agents. 9. 2-Formyl(*m*-amino)phenylpyridine Thiosemicarbazones. *J. Med. Chem.* 1974, 17, 631-635.

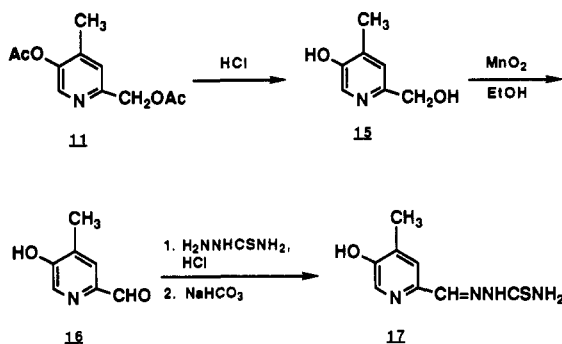
(8) Agrawal, K. C.; Booth, B. A.; DeNuzzo, S. M.; Sartorelli, A. C. Potential Antitumor Agents. 14. 4-Substituted 2-Formylpyridine Thiosemicarbazones. *J. Med. Chem.* 1976, 19, 1209-1214.

(9) Furukawa, S. Synthesis of Pyridine Derivatives. VII. Nitration of 2,4-Lutidine. *J. Pharm. Soc. Jpn.* 1956, 76, 900-902.

Scheme I

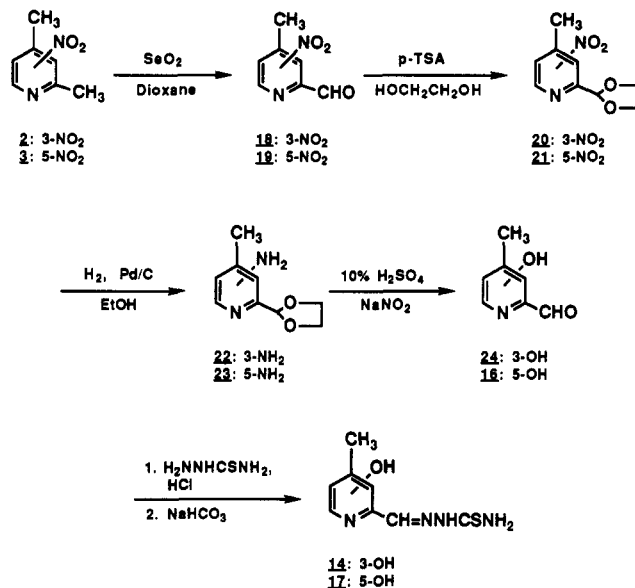


Scheme II



with sodium nitrite in 10% sulfuric acid, followed by hydrolysis of the resulting products, gave the respective hydroxy compounds 6 and 7. Treatment of 6 and 7 with 30% hydrogen peroxide in glacial acetic acid produced the *N*-oxides 8 and 9, which were then refluxed with acetic anhydride to give the acetates 10 and 11. A repeat of the *N*-oxidation procedure with compound 10, followed by rearrangement of the resulting *N*-oxide 12 in refluxing acetic anhydride, yielded the corresponding 2-pyridinealdehyde diacetate derivative 13. Treatment of 13 with thiosemicarbazide in the presence of hydrochloric acid⁶ produced 3-hydroxy-4-methylpyridine-2-carboxaldehyde thiosemicarbazone (14, Scheme I). Hydrolysis of the acetate 11 with hydrochloric acid gave 5-hydroxy-2-(hydroxymethyl)-4-methylpyridine (15). Oxidation of 15 with manganese oxide in ethanol yielded the corresponding aldehyde 16, which was then condensed with thiosemicarbazide to afford the desired compound 17^{7,8} (Scheme II). Conversion of 3-hydroxy-2-(hydroxymethyl)-4-methylpyridine, the isomer of 5-hydroxy-2-(hydroxymethyl)-4-methylpyridine (15), to the corresponding aldehyde by oxidation with manganese oxide, however, has not been successful, probably because 3-hydroxy-4-methylpyridine-

Scheme III



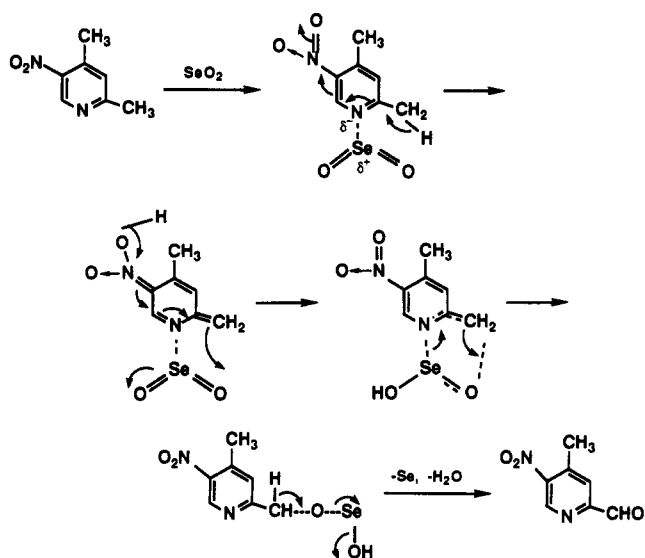
2-carboxaldehyde is not stable under these oxidation conditions. The overall yields of the syntheses of 3-HMP (14) and 5-HMP (17) were 0.42% and 8.3%, respectively, based upon the corresponding 3- and 5-nitrolutidines. Because these low overall yields were unsatisfactory, especially for the synthesis of 3-HMP, another more efficient synthetic route has been devised (Scheme III). Selective oxidation¹⁰ of 2,4-dimethyl-3- and 5-nitropyridines (2 and 3, respectively) with selenium dioxide in dioxane gave the corresponding aldehydes, 18 and 19, which were then refluxed in toluene with ethylene glycol and *p*-toluenesulfonic acid to yield the corresponding 1,3-dioxolanes (20 and 21).⁸ These protected intermediates were reduced by catalytic hydrogenation in the presence of 10% Pd/C to produce the respective amino derivatives, 22 and 23, which were then converted to the 3- and 5-hydroxy-4-methylpyridine-2-carboxaldehydes, 24 and 16, by treatment with sodium nitrite in 10% sulfuric acid. Condensation of 24 and 16 with thiosemicarbazide afforded the desired compounds 14 and 17 in overall yields of 4.8% and 21%, respectively. French and Blanz¹¹ also reported the synthesis of 3-HMP by a different methodology; however, no synthetic procedure nor any spectroscopic data to confirm the structure of this compound were given. Furthermore, in contrast to our test results, which showed that 3-HMP had antitumor activity against the L1210 leukemia, they reported that this agent was inactive against this tumor cell line.¹¹

It is interesting that the 2-methyl groups in compounds 2 and 3 were considerably more sensitive to selenium dioxide oxidation than their 4-methyl counterparts. 4-Methyl-3-nitropyridine-2-carboxaldehyde (18) and 4-methyl-5-nitropyridine-2-carboxaldehyde (19) were isolated in 20% and 55% yields, respectively, by silica gel column chromatography after the oxidation. In addition to unreacted starting material, an amount of 4-methyl-3- and 5-nitro-2-pyridinecarboxylic acid was also isolated.

(10) Dodd, R. H.; Ouannes, C.; Robert-Gero, M.; Potier, P. Hybrid Molecules: Growth Inhibition of *Leishmania donovani* Promastigotes by Thiosemicarbazones of 3-Carboxy- β -carbonylones. *J. Med. Chem.* 1989, 32, 1272-1276.

(11) French, F. A.; Blanz, E. J., Jr. α -(*N*-Formylheteroaromatic Thiosemicarbazones. Inhibition of Tumor-Derived Ribonucleoside Diphosphate Reductase and Correlation with *in Vitro* Antitumor Activity. *J. Med. Chem.* 1974, 17, 172-181.

Scheme IV



When the reaction time was prolonged, the amount of the acid byproducts was increased; however, no detectable amounts of 2-methyl-3- and 5-nitropyridine-4-carboxaldehydes were found. A cyclic mechanism is proposed for the oxidation reaction of 5-nitro-2,4-lutidine (Scheme IV), which is analogous to the mechanism proposed by Corey and Schaefer¹² for the oxidation of 7-methylquinoline, except that a cyclic transition state is suggested. Such an intermediate may account for the selective oxidation of the 2-methyl group. A similar, but more hindered, cyclic transition state may be formed for the oxidation of 3-nitro-2,4-lutidine, which might explain why the 2-methyl group in the 3-nitro derivative is more difficult to oxidize than its 5-nitro counterpart.

Biological Results and Discussion

The tumor-inhibitory properties of 3-HMP (14) and 5-HMP (17) were compared with those of 3-hydroxypyridine-2-carboxaldehyde thiosemicarbazone (3-HP) and 5-HP by measuring their effects on the survival time of CD_2F_1 female mice bearing the L1210 leukemia. Compounds were administered at daily dosage levels of from 10 to 60 mg/kg by intraperitoneal (ip) injection to groups of 5 tumor-bearing mice once a day for 6 consecutive days by methodology described previously.¹³ The prolongation of life span produced by the maximum effective daily dose of each compound is shown in Table I. The 4-methyl-substituted derivatives, 3- and 5-HMP, were both equivalent to or more effective than their corresponding parent compounds, 3-HP and 5-HP, when administered following solubilization in DMSO or in suspension, respectively. The greater antitumor activity of the agents when administered in suspension presumably derives from their slow solubilization in the peritoneal cavity which provides a long-lasting effect. The greater activity of 3-HMP and 5-HMP than their non-methylated counterparts is consistent with the previous finding that the addition of methyl or other hydrophobic groups onto the 3, 4, or 5 carbon atoms of the pyridine ring increased activity as inhibitors of ribonu-

Table I. Comparative Effects of 3-HP, 5-HP, 3-HMP, and 5-HMP on Mice Bearing the L1210 Leukemia

compd	injection form	optimum daily dosage ^a (mg/kg)	Av Δ wt ^b (%)	T/C ^c (%)
3-HP	DMSO solution	40	+1.5	114
5-HP	DMSO solution	40	+1.8	132
3-HMP (14)	DMSO solution	40	+0.5	135
5-HMP (17)	DMSO solution	40	-7.4	138
5-HP	suspension	60	+4.6	146
3-HMP (14)	suspension	50	+0.9	168
5-HMP (17)	suspension	40	-3.4	186

^a Administered once daily for six consecutive days, beginning 24 h after tumor implantation. ^b Average weight change of mice from onset to termination of drug treatment. ^c % T/C represents the ratio of the survival time of treated to control mice $\times 100$.

cleotide reductase, probably due to a hydrophobic binding region in the target enzyme molecule.¹⁴

Experimental Section

Melting points were determined with a Thomas-Hoover Unimelt apparatus and are uncorrected. ¹H NMR spectra were recorded at 90 MHz on a Varian EM-390 or at 500 MHz on a Bruker WM-500 spectrometer with Me_4Si as the internal reference. High-resolution mass spectra (HRMS) were recorded on a VG ZAB-SE mass spectrometer equipped with a VG 11-250 data system. The fast atom bombardment (FAB) spectrum was produced using the standard VG IONTECH LTD field gun with xenon gas at 8 kV anode potential. Accurate masses were calculated interactively with the data system using the peaks from poly(ethylene glycol) as reference masses. TLC was performed on EM precoated silica gel sheets containing a fluorescent indicator. Elemental analyses were carried out by the Baron Consulting Co., Orange, CT. Where analyses are indicated only by symbols of the elements, the analytical results for those elements were within $\pm 0.4\%$ of the theoretical value.

2,4-Dimethyl-3- and 5-nitropyridine (2 and 3).⁹ Fuming sulfuric acid (1500 g, 15.3 mol) was added slowly to 2,4-lutidine (165 mL, 1.43 mol) and cooled in an ice bath with stirring. Potassium nitrate (262.5 g, 2.60 mol) was then added slowly. The reaction mixture was gradually heated to 100 °C and maintained at this temperature for 8 h. The reaction mixture was then heated at 120 °C for an additional 8 h. After cooling to room temperature, the reaction mixture was poured onto ice (2.5 kg). The solution was neutralized to pH 7 using potassium carbonate and extracted with chloroform (3 \times 4 L). The organic layer was dried over anhydrous Na_2SO_4 , and the solvent was evaporated; the remaining solution was distilled under reduced pressure. 2,4-Dimethyl-3-nitropyridine [(2; 41.7 g, 0.27 mol, 19%, 37 °C (0.24 mm Hg)], 2,4-dimethyl-5-nitropyridine [(3; 38.2 g, 0.25 mol, 18%, 44 °C (0.17 mm Hg)], and a mixture of 2,4-dimethyl-3- and 5-nitropyridine (13.74 g, 0.09 mol) were obtained. Compound 2: ¹H NMR (90 MHz, CDCl_3) δ 2.33 (s, 3 H, 4- CH_3), 2.53 (s, 3 H, 2- CH_3), 7.02 (d, 1 H, 5-H, $J_{5,6} = 4.5$ Hz), 8.35 (d, 1 H, 6-H, $J_{5,6} = 4.5$ Hz). Compound 3: ¹H NMR (90 MHz, CDCl_3) δ 2.70 (s, 6 H, 2- and 4- CH_3), 7.17 (s, 1 H, 3-H), 9.10 (s, 1 H, 6-H).

3-Amino-2,4-dimethylpyridine (4). To a solution of 2,4-dimethyl-3-nitropyridine (2; 31.4 g, 0.21 mol) in 200 mL of absolute ethanol was added 5% Pd/C (2 g). The mixture was hydrogenated under 60 psi of pressure for 2 h. The solution was filtered, and the solvent was evaporated in vacuo to give a solid (24.0 g, 98%): mp 48–50 °C (lit.⁹ mp 51–53 °C). The product appeared homogeneous on TLC and by NMR analysis and was used without further purification: ¹H NMR (90 MHz, CDCl_3) δ 2.17 (s, 3 H, 4- CH_3), 2.33 (s, 3 H, 2- CH_3), 3.60 (s, 2 H, 3- NH_2 , D_2O exchangeable), 6.85 (d, 1 H, 5-H, $J_{5,6} = 4.5$ Hz), 7.85 (d, 1 H, 6-H, $J_{5,6} = 4.5$ Hz).

5-Amino-2,4-dimethylpyridine (5). This compound was synthesized by methodology used for 4 except the starting

(12) Corey, E. J.; Schaefer, J. P. Studies on the Mechanism of Oxidation of Ketones by Selenium Dioxide (Part I). *J. Am. Chem. Soc.* 1960, 82, 918–929.

(13) Lin, T. S.; Xu, S. P.; Zhu, L. Y.; Cosby, L. A.; Sartorelli, A. C. Synthesis of 2,3-Diaziridinyl-1,4-naphthoquinone Sulfonate Derivatives as Potential Antineoplastic Agents. *J. Med. Chem.* 1989, 32, 1467–1471.

(14) Sartorelli, A. C.; Agrawal, K. C.; Moore, E. C. Mechanism of Inhibition of Ribonucleoside Diphosphate Reductase by α -(*N*)-Heterocyclic Aldehyde Thiosemicarbazones. *Biochem. Pharmacol.* 1971, 20, 3119–3123.

material was 3: yield 24.1 g (98%); mp 62–64 °C (lit.⁹ mp 66–68 °C); ¹H NMR (90 MHz, CDCl₃) δ 2.10 (s, 3 H, 4-CH₃), 2.37 (s, 3 H, 2-CH₃), 3.33 (s, 2 H, 3-NH₂, D₂O exchangeable), 6.70 (s, 1 H, 3-H), 7.79 (s, 1 H, 6-H).

2,4-Dimethyl-3-hydroxypyridine (6). To a solution of 3-amino-2,4-dimethylpyridine (4; 25.0 g, 0.21 mol) in 10% sulfuric acid (400 mL) cooled to 0 °C by dry ice in acetone with stirring was added a solution of sodium nitrite (16.2 g, 0.23 mol) in 160 mL of water dropwise at 0–5 °C over a period of 7 min. The solution was maintained at 0 °C for an additional 15 min and then heated in a steam bath for 15 min. After cooling to room temperature, the solution was neutralized with K₂CO₃ to pH 7. The product was then extracted with chloroform (3 × 500 mL). The organic layer was dried over anhydrous Na₂SO₄ and the solvent was removed in vacuo. The product was recrystallized from acetone and the mother liquid was purified by silica gel column chromatography (EtOAc) to afford an additional amount of the pure product. The total yield was 12.7 g (51%) as a colorless solid: mp 105–106 °C (lit.⁹ mp 99–101 °C); ¹H NMR (90 MHz, CDCl₃) δ 2.25 (s, 3 H, 4-CH₃), 2.50 (s, 3 H, 2-CH₃), 6.97 (d, 1 H, 5-H, *J*_{5,6} = 4.5 Hz), 7.95 (d, 1 H, 6-H, *J*_{5,6} = 4.5 Hz), 11.20 (s, 1 H, 3-OH, D₂O exchangeable).

2,4-Dimethyl-5-hydroxypyridine (7). This compound was synthesized by methodology used for 6 except the starting material was 5: yield 12.6 g (51%) as a colorless solid; mp 146–148 °C (lit.⁹ mp 144–146 °C); ¹H NMR (90 MHz, CDCl₃) δ 2.20 (s, 3 H, 4-CH₃), 2.47 (s, 3 H, 2-CH₃), 6.87 (s, 1 H, 3-H), 7.97 (s, 1 H, 6-H), 11.43 (s, 1 H, 5-OH, D₂O exchangeable).

2,4-Dimethyl-3-hydroxypyridine *N*-Oxide (8). To a stirred solution of 2,4-dimethyl-3-hydroxypyridine (6; 23.7 g, 0.19 mol) in 130 mL of glacial acetic acid was added dropwise 36 mL of 30% hydrogen peroxide. The reaction mixture was heated to 80 °C and two additional portions of 30% hydrogen peroxide (36 mL) were added at 3-h intervals. The solution was maintained at 80 °C for a total of 9 h and the solvent was removed under reduced pressure. The residue was purified by silica gel column chromatography (EtOAc/MeOH, 7:3, v/v) to give 10.3 g (38%) of product: mp 134–136 °C; ¹H NMR (90 MHz, Me₂SO-*d*₆) δ 2.17 (s, 3 H, 4-CH₃), 2.32 (s, 3 H, 2-CH₃), 6.94 (d, 1 H, 5-H, *J*_{5,6} = 6 Hz), 7.72 (s, 1 H, 6-H, *J*_{5,6} = 6 Hz); HRMS (FAB) *m/z* calcd for C₇H₉NO₂, 140.0711; found, 140.0707. Anal. (C₇H₉NO₂) C, H, N.

2,4-Dimethyl-5-hydroxypyridine *N*-Oxide (9). This compound was synthesized by methodology used for 8 except the starting material was 7: yield 10.0 g (37%); mp 229 °C dec; ¹H NMR (90 MHz, Me₂SO-*d*₆) δ 2.10 (s, 3 H, 4-CH₃), 2.22 (s, 3 H, 2-CH₃), 7.07 (s, 1 H, 3-H), 7.70 (s, 1 H, 6-H); HRMS (FAB) *m/z* calcd for C₇H₉NO₂, 140.0711; found, 140.0722.

3-Acetoxy-2-(acetoxymethyl)-4-methylpyridine (10). A mixture of 2,4-dimethyl-3-hydroxypyridine *N*-oxide (8; 11.3 g, 81 mmol) and acetic anhydride (200 mL) was heated at 110 °C with stirring for 2.5 h. After cooling, the solvent was evaporated under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc/hexane, 1:1, v/v) to yield 13.5 g (74%) of product as a slightly yellow oil: ¹H NMR (90 MHz, CDCl₃) δ 2.20 (s, 3 H, 4-CH₃), 2.37 (s, 6 H, 2 OCOCH₃), 5.17 (s, 2 H, 2-CH₂), 7.15 (d, 1 H, 5-H, *J*_{5,6} = 4.5 Hz), 8.35 (d, 1 H, 6-H, *J*_{5,6} = 4.5 Hz); HRMS (FAB) *m/z* calcd for C₁₁H₁₃NO₄, 224.0923; found, 224.0935. Anal. (C₁₁H₁₃NO₄) C, H, N.

5-Acetoxy-2-(acetoxymethyl)-4-methylpyridine (11). This compound was synthesized by methodology used for 10 except the starting material was 9: yield 9.85 g (54%) as a yellow oil; ¹H NMR (90 MHz, CDCl₃) δ 2.15 and 2.25 (two s, 6 H, two OCOCH₃), 2.35 (s, 3 H, 4-CH₃), 5.13 (s, 2 H, 2-CH₂), 7.23 (s, 1 H, 3-H), 8.23 (s, 1 H, 6-H); HRMS (FAB) *m/z* calcd for C₁₁H₁₃NO₄, 224.0923; found, 224.0943. Anal. (C₁₁H₁₃NO₄) C, H, N.

3-Acetoxy-2-(acetoxymethyl)-4-methylpyridine *N*-Oxide (12). To a solution of 3-acetoxy-2-(acetoxymethyl)-4-methylpyridine (10; 13.5 g, 60 mmol) in 74 mL of glacial acetic acid was added dropwise with stirring 21 mL of 30% hydrogen peroxide. The mixture was heated to 80 °C and two additional portions of 30% hydrogen peroxide (21 mL) were added at 3-h intervals. The solution was maintained at 80 °C for a total of 9 h. The solvent was evaporated in vacuo, and the residue was purified by silica gel column chromatography (EtOAc/MeOH, 7:3, v/v) to give 2.62 g (18%) of product: mp >360 °C. The product was used immediately for the next step.

3-Acetoxy-2-(diacetoxymethyl)-4-methylpyridine (13). A mixture of 3-acetoxy-2-(acetoxymethyl)-4-methylpyridine *N*-oxide (12; 2.77 g, 11.6 mmol) and 54 mL of acetic anhydride was heated with stirring at 110 °C for 2.5 h. After cooling, the solvent was evaporated under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc/hexane, 1:1, v/v) to yield 1.54 g (47%) of product as a yellow oil: ¹H NMR (90 MHz, CDCl₃) δ 2.10–2.40 (m, 12 H, 4-CH₃, three OCOCH₃), 5.17 (s, 2 H, 2-CH₂), 7.20–7.38 (m, 1 H, 5-H), 8.37–8.52 (m, 1 H, 6-H); HRMS (FAB) *m/z* calcd for C₁₃H₁₅NO₆, 282.0978; found, 282.0990. Anal. (C₁₃H₁₅NO₆) H, N; C: calcd, 55.51; found, 56.01.

3-Hydroxy-4-methylpyridine-2-carboxaldehyde Thiosemicarbazone (14). **Method A.** To a slurry of thiosemicarbazide (0.26 g, 2.9 mmol) in 5 mL of concentrated HCl and 15 mL of ethanol was added a solution of 13 (0.8 g, 2.9 mmol) in 10 mL of ethanol. The reaction mixture was stirred at 50 °C for 2 h and the precipitate was filtered after cooling. The yellow solid was recrystallized from aqueous ethanol solution (1:1, v/v) containing 5% concentrated HCl to afford 0.25 g (35%) of product as the hydrochloride salt: mp 243 °C dec; ¹H NMR (500 MHz, Me₂SO-*d*₆) δ 2.52 (s, 3 H, 4-CH₃), 3.80 (br s, 1 H, 3-OH, D₂O exchangeable), 7.73 (d, 1 H, 5-H, *J*_{5,6} = 4.5 Hz), 8.27 (d, 1 H, 6-H, *J*_{5,6} = 4.5 Hz), 8.35 (s, 1 H, 2-CH), 8.66 and 8.88 (two s, 2 H, NH₂, D₂O exchangeable), 12.07 (s, 1 H, NH, D₂O exchangeable); HRMS (FAB) *m/z* calcd for C₈H₁₀N₄OS, 211.0654; found, 211.0651. Anal. (C₈H₁₀N₄OS·HCl·H₂O) C, H, N.

The hydrochloride was stirred in 10% sodium bicarbonate to yield the free base: mp 227–228 °C dec (lit.¹¹ mp 223–224 °C); ¹H NMR (500 MHz, Me₂SO-*d*₆) δ 2.23 (s, 3 H, 4-CH₃), 4.80 (br s, 1 H, 3-OH, D₂O exchangeable), 7.26 (d, 1 H, 5-H, *J*_{5,6} = 5 Hz), 8.05 (d, 1 H, 6-H, *J*_{5,6} = 5 Hz), 8.20 (s, 2 H, NH₂, D₂O exchangeable), 8.35 (s, 1 H, 2-CH), 11.80 (s, 1 H, NH, D₂O exchangeable).

Method B. To a solution of 3-amino-2-(1,3-dioxolan-2-yl)-4-methylpyridine (22; 0.6 g, 3.3 mmol) in 15 mL of 10% H₂SO₄ at 0 °C (ice bath) with stirring was added dropwise a solution of NaNO₂ (0.38 g, 5.5 mmol) in 3 mL of water. The mixture was stirred at 0 °C for 15 min and then heated in a steam bath for 30 min. The resulting solution was evaporated at room temperature under reduced pressure to yield 3-hydroxy-4-methylpyridine-2-carboxaldehyde (24) as a syrup, which was dissolved in 15 mL of water, decolorized with charcoal, and filtered. To the filtrate was added a solution of thiosemicarbazide (0.31 g, 3.3 mmol) in 5 mL of 5% concentrated HCl. The mixture was refluxed for 30 min and then cooled, and the yellow precipitate was filtered, washed with water, and recrystallized from aqueous ethanol solution (1:1, v/v) containing 5% concentrated HCl to afford 0.21 g (30%) of product: the melting point and all spectroscopic data were identical with those obtained by Method A.

5-Hydroxy-2-(hydroxymethyl)-4-methylpyridine (15). A mixture of 5-acetoxy-2-(acetoxymethyl)-4-methylpyridine (11; 6.2 g, 4.5 mmol) and 200 mL of concentrated HCl was refluxed for 1 h. After cooling, the reaction mixture was evaporated to dryness under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc/MeOH, 7:3, v/v) to give 3.8 g (97%) of product: mp 161–162 °C; ¹H NMR (90 MHz, Me₂SO-*d*₆) δ 2.33 (s, 3 H, 4-CH₃), 4.70 (s, 2 H, 2-CH₂), 7.67 (s, 1 H, 3-H), 8.22 (s, 1 H, 6-H); HRMS (FAB) *m/z* calcd for C₇H₉NO₂, 140.0711; found, 140.0736.

5-Hydroxy-4-methylpyridine-2-carboxaldehyde Thiosemicarbazone (17). **Method A.** To a solution of 15 (3.9 g, 28 mmol) in 100 mL of ethanol was added MnO₂ (10.0 g, 0.12 mol) and the reaction mixture was heated to reflux for 2 h with stirring. The mixture was filtered and the filtrate was concentrated under reduced pressure to 80 mL. Because the aldehyde (16) is unstable, concentrated HCl (8 mL) was added immediately. Thiosemicarbazide (1.5 g, 17 mmol) was added to the aldehyde solution with stirring, and the reaction mixture was heated to reflux for 30 min. The precipitate was filtered upon cooling and recrystallized in aqueous ethanol solution (1:1, v/v) containing 5% concentrated HCl to afford 3.3 g (81%) of product: mp 229 °C; ¹H NMR (500 MHz, Me₂SO-*d*₆) δ 2.33 (s, 3 H, 4-CH₃), 4.01 (br s, 1 H, 5-OH, D₂O exchangeable), 8.02 (s, 1 H, 3-H), 8.20 (s, 1 H, 6-H), 8.22 (s, 1 H, 2-CH), 8.58 (s, 2 H, NH₂, D₂O exchangeable), 12.0 (s, 1 H, NH, D₂O exchangeable); HRMS (FAB) *m/z* calcd

for $C_8H_{10}N_4OS$, 211.0654; found, 211.0671. Anal. ($C_8H_{10}N_4OS \cdot HCl \cdot H_2O$) C, H, N.

The hydrochloride was stirred in 10% sodium bicarbonate to yield the free base: mp 220–222 °C dec; 1H NMR (500 MHz, Me_2SO-d_6) δ 2.15 (s, 3 H, 4- CH_3), 7.95 (s, 1 H, 3-H), 7.97 (s, 1 H, 6-H), 8.01 (s, 1 H, 2-CH), 8.04 and 8.18 (two s, 2 H, NH_2 , D_2O exchangeable), 10.1 (s, 1 H, 5-OH, D_2O exchangeable), 12.0 (s, 1 H, NH, D_2O exchangeable).

Method B. This compound was also prepared from the corresponding 5-amino derivative 5-amino-2-(1,3-dioxolan-2-yl)-4-methylpyridine (23) via the aldehyde 16 by the same procedure described for the synthesis of compound 14 (method B): yield 0.32 g (46%); the melting point and all spectroscopic data were identical with those obtained by method A.

4-Methyl-3-nitropyridine-2-carboxaldehyde (18). A mixture of 2,4-dimethyl-3-nitropyridine (2; 5.0 g, 33 mmol) and selenium dioxide (4.5 g, 42 mmol) in anhydrous 1,4-dioxane (100 mL) was refluxed under an atmosphere of nitrogen for 35 h. The reaction mixture was cooled and filtered to remove the precipitated black selenium. The filtrate was evaporated in vacuo to dryness, and the residue was chromatographed on a silica gel (120 g) column ($CH_2Cl_2/EtOAc$, 10:1, v/v; R_f 0.65) to afford 1.1 g (20%) of white crystals: mp 101–102 °C; 1H NMR (90 MHz, $CDCl_3$) δ 2.35 (s, 3 H, 4- CH_3), 7.47 (d, 1 H, 5-H, $J_{5,6}$ = 4.5 Hz), 8.72 (d, 1 H, 6-H, $J_{5,6}$ = 4.5 Hz), 9.95 (s, 1 H, 2-CHO). Anal. ($C_7H_6N_2O_3$) C, H, N.

4-Methyl-5-nitropyridine-2-carboxaldehyde (19). This compound was prepared from the nitro derivative 3 by the same procedure described for the synthesis of compound 18, except the reaction time was 4 h: yield 6.0 g (55%); mp 82–83 °C (lit.⁶ mp 81–82 °C); TLC, R_f 0.86 ($CH_2Cl_2/EtOAc$, 3:2, v/v); 1H NMR (90 MHz, $CDCl_3$) δ 2.70 (s, 3 H, 4- CH_3), 7.90 (s, 1 H, 3-H), 9.20 (s, 1 H, 6-H), 10.10 (s, 1 H, 2-CHO).

2-(1,3-Dioxolan-2-yl)-4-methyl-3-nitropyridine (20). To 0.75 g (14 mmol) of compound 18 in 100 mL of toluene was added 40 mg of *p*-toluenesulfonic acid monohydrate and 2 mL of ethylene glycol. The reaction mixture was refluxed with stirring, and a Dean-Stark trap was used to remove the water formed during condensation until complete disappearance of the starting material was observed. The mixture was cooled and then washed with 25 mL of 10% $NaHCO_3$ solution, followed by 25 mL of

water. The toluene layer was dried over anhydrous $MgSO_4$, and the solvent was removed under reduced pressure. The residue was chromatographed on a silica gel (120 g) column ($CH_2Cl_2/EtOAc$, 10:1, v/v; R_f 0.42) to afford 1.1 g (85%) of white crystals: mp 46–48 °C; 1H NMR (90 MHz, $CDCl_3$) δ 2.40 (s, 3 H, 4- CH_3), 4.07 (s, 4 H, CH_2CH_2), 6.05 (s, 1 H, 2-CH), 7.30 (d, 1 H, 5-H, $J_{5,6}$ = 4.5 Hz), 8.60 (d, 1 H, 6-H, $J_{5,6}$ = 4.5 Hz). Anal. ($C_9H_{10}N_2O_4$) C, H, N.

2-(1,3-Dioxolan-2-yl)-4-methyl-5-nitropyridine (21). This compound was synthesized by methodology used for 20 except the starting material was 19: yield 2.3 g (91%); mp 77–79 °C; (lit.¹⁰ mp 77 °C) TLC, R_f 0.74 ($CH_2Cl_2/EtOAc$, 3:2, v/v); 1H NMR (90 MHz, $CDCl_3$) δ 2.65 (s, 3 H, 4- CH_3), 4.10 (s, 4 H, CH_2CH_2), 5.85 (s, 1 H, 2-CH), 7.50 (s, 1 H, 3-H), 9.12 (s, 1 H, 6-H).

3-Amino-2-(1,3-dioxolan-2-yl)-4-methylpyridine (22). The nitro derivative 20 (1.1 g, 5.2 mmol) was dissolved in 200 mL of ethanol and hydrogenated in a Parr apparatus under 50 psi of pressure in the presence of 10% Pd/C (200 mg) for 20 h. After filtration, the filtrate was evaporated under reduced pressure to give the product (0.9 g, 94%) as a syrup: ninhydrin positive; 1H NMR (90 MHz, $CDCl_3$) δ 2.12 (s, 3 H, 4- CH_3), 4.05 (m, 4 H, CH_2CH_2), 4.10 (br s, 2 H, 3- NH_2 , D_2O exchangeable), 5.76 (s, 1 H, 2-CH), 6.92 (d, 1 H, 5-H, $J_{5,6}$ = 4.5 Hz), 7.86 (d, 1 H, 6-H, $J_{5,6}$ = 4.5 Hz). Anal. ($C_9H_{12}N_2O_2$) C, H, N.

5-Amino-2-(1,3-dioxolan-2-yl)-4-methylpyridine (23). This compound was synthesized by methodology used for 22 except the starting material was 21: yield 1.2 g (92%); mp 79–80 °C; 1H NMR (90 MHz, $CDCl_3$) δ 2.15 (s, 3 H, 4- CH_3), 3.70 (br s, 2 H, 5- NH_2 , D_2O exchangeable), 4.10 (m, 4 H, CH_2CH_2), 5.70 (s, 1 H, 2-CH), 7.15 (s, 1 H, 3-H), 8.00 (s, 1 H, 6-H). Anal. ($C_9H_{12}N_2O_2$) C, H, N.

Acknowledgment. This investigation was supported by U.S. Public Health Service Grant CA-53340. We wish to thank Ms. Regina Loomis for her excellent technical assistance. We also acknowledge the support of the Northeast NMR Facility at Yale University for the high-resolution NMR spectra, made possible by a grant from the Chemical Division of the National Science Foundation (Grant No. CHE-7916210).