

NATURAL (-)-FISSETINIDOL-(4,8)-(-)-EPICATECHIN PROFISSETINIDINS

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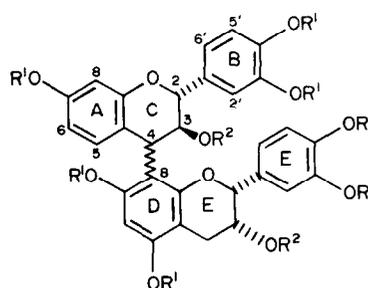
Key Word Index—*Guibourtia coleosperma*; *Colophospermum mopane*, *Baikiaea plurijuga*, Leguminosae; heartwood, profisetinidins, (-)-fisetinidol-(4 α ,8) and (4 β ,8)-(-)-epicatechins.

Abstract—The natural occurrence of the first profisetinidins associated with (-)-epicatechin is demonstrated in the heartwood of three legumes. ¹H NMR spectroscopy at 300 MHz permits assessment of the relative abundance of the predominant rotamers in each of the heptamethyl ether diacetates of the (-)-fisetinidol-(4 α ,8) and (4 β ,8)-(-)-epicatechins at ambient temperatures.

(-)-Epicatechin [(2*R*,3*R*)-2,3-*cis*-flavan-3,3',4',5,7-pentaol] figures prominently both as 'upper' and 'terminal' units in procyanidin- [1-3] and as 'terminal' units in proguibourtinidin- [4-6] condensed tannins. In contrast to the ubiquitous (-)-fisetinidol-(+)-catechin† profisetinidins- [7, 8], natural counterparts of the latter class of condensed tannins associated with (-)-epicatechin are hitherto unknown. Our search for tannin prototypes in the flora of southern Africa has now revealed the presence of (-)-fisetinidol-(4 α ,8) and (4 β ,8)-(-)-epicatechin **1** and **3** in the heartwoods of three species of the Caesalpinioideae, *Guibourtia coleosperma* (Rhodesian copalwood) [9], *Colophospermum mopane* (mopane) [9] and *Baikiaea plurijuga* (Rhodesian teak) [9].

The ethyl acetate extract of the heartwood of *G. coleosperma* afforded both the (4 α ,8)- and (4 β ,8)-(-)-fisetinidol-(-)-epicatechins **1** and **3**. Only the (4 α ,8)-isomer **1** was found in the heartwood extracts (methanol) of *C. mopane* and *B. plurijuga*. Owing to the complexity of the phenolic mixtures of these sources, the novel metabolites **1** and **3** were identified as heptamethyl ether diacetates **2** and **4**. Their ¹H NMR spectra (300 MHz) (Table 1) at ambient temperatures exhibit duplicated signals characteristic of the effects of dynamic rotational isomerism [10].

At these temperatures the heterocyclic regions each display an AMX-($J_{2,3} = J_{3,4} = 10.0$ Hz for **2**; $J_{2,3} = 7.0$, $J_{3,4} = 5.5$ Hz for **4**) and ABMX-system ($J_{2,3} = ca 1.0$ Hz for both **2** and **4**) indicative of 2,3-*trans*-3,4-*trans*: 2,3-*cis*- and 2,3-*trans*-3,4-*cis*: 2,3-*cis* relative configuration for **2** and **4** respectively. Spin decoupling experiments using the 2- (for ABX-systems of rings B and E) and 4-H (for ABX-system of ring A) resonances as reference signals, facilitated definition of the constitution of the 'upper' (-)-fisetinidol and 'lower' (-)-epicatechin moieties. In addition to the aforementioned ABX-systems, the aromatic region of each spectrum exhibited a one-proton singlet



- 1** $\text{---} \text{---} \text{---} \text{---} \text{---} \text{---}$, $R^1 = R^2 = H$
2 $\text{---} \text{---} \text{---} \text{---} \text{---} \text{---}$, $R^1 = Me, R^2 = Ac$
3 $\text{---} \text{---} \text{---} \text{---} \text{---} \text{---}$, $R^1 = R^2 = H$
4 $\text{---} \text{---} \text{---} \text{---} \text{---} \text{---}$, $R^1 = Me, R^2 = Ac$

(δ 6.18, 6.15 for **2** and **4** respectively). The chemical shifts of these were reminiscent of the proton of a C-8 substituted (-)-epicatechin unit [7, 11]. This was unequivocally confirmed by the strong NOE association of the 'residual' proton with both 5-(10.7, 10.8% for **2** and **4** respectively) and 7-OMe (10.1, 12.7% for **2** and **4** respectively) of the (-)-epicatechin 'lower' units. Confirmation for the structures of **2** and **4** and thus also the 2*R*,3*S*,4*S* (C-ring); 2*R*,3*R* (F-ring) and 2*R*,3*S*,4*R* 2*R*,3*R* absolute configuration of **1** and **3** respectively, was obtained by comparison of the ¹H NMR and CD data of **2** and **4** with those of the corresponding derivatives of their synthetic counterparts [7]. As ¹H NMR spectroscopy at 80 MHz only permitted partial assignment of signals for synthetic **2** and **4**, detailed analysis is included in this communication (Table 1).

At room temperature the ¹H NMR spectra of **2** and **4** indicated two predominant rotameric forms, i.e. 41:9 and 39:11 for **2** and **4** respectively. The magnitude of the rotational barrier is such that in DMSO-*d*₆ at 453 K, the upper limit of the Bruker AM-300 system, the spectra of both **2** and **4** still displayed severe line-broadening in the aromatic and heterocyclic regions. This contrasts with the isomeric (-)-fisetinidol-(+)-catechin pair [7] where

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† (-)-Fisetinidol is (2*R*,3*S*)-2,3-*trans*-flavan-3,3',4',7-tetraol and (+)-catechin its 5-*oxy* analogue

Table 1 ^1H NMR (300 MHz) peaks (ppm) of (–)-fisetimidol-(4 α ,8) and (4 β ,8)-(–)-epicatechin heptamethyl ether diacetates **2** and **4** in CDCl_3 at 298 K

Ring	H	2*	4*
A	5	6.74 (<i>d</i> , 8.5)	6.60 (<i>d</i> , 8.5)
	6	6.43 (<i>dd</i> , 2.5, 8.5)	6.11 (<i>dd</i> , 2.5, 8.5)
	8	6.47 (<i>d</i> , 2.5)	5.90 (<i>d</i> , 2.5)
B	2	6.57 (<i>d</i> , 2.0)	6.92 (<i>d</i> , 2.0)
	5	6.68 (<i>d</i> , 8.0)	6.78 (<i>d</i> , 8.0)
	6	6.75 (<i>dd</i> , 2.0, 8.0)	6.93 (<i>dd</i> , 2.0, 8.0)
C	2	4.90 (<i>d</i> , 10.0)	5.47 (<i>d</i> , 7.0)
	3	6.17 (<i>t</i> , 10.0)	5.62 (<i>dd</i> , 5.5, 7.0)
	4	4.88 (<i>d</i> , 10.0)	4.80 (<i>d</i> , 5.5)
D	6	6.18 (<i>s</i>)	6.15 (<i>s</i>)
E	2	6.54 (<i>d</i> , 2.0)	6.77 (<i>d</i> , 2.0)
	5	6.66 (<i>d</i> , 8.0)	6.68 (<i>d</i> , 8.0)
	6	6.42 (<i>dd</i> , 2.0, 8.0)	6.50 (<i>dd</i> , 2.0, 8.0)
F	2	4.98 (<i>br s</i> , <i>ca</i> 1.0)	4.32 (<i>br s</i> , <i>ca</i> 1.0)
	3	5.25 (<i>m</i>)	5.29 (<i>m</i>)
	4 _{ax}	2.81 (<i>dd</i> , 1.5, 18.5)	2.84 (<i>dd</i> , 1.5, 18.5)
	4 _{eq}	2.94 (<i>dd</i> , 4.5, 18.5)	2.95 (<i>dd</i> , 5.5, 18.5)
	OMe	3.41, 3.70, 3.74, 3.79, 3.82, 3.83 (5-D), 3.86 (7-D) (each <i>s</i>)	3.39, 3.74, 3.77 (7-D), 3.82, 3.83, 3.84 (5-D), 3.85 (each <i>s</i>)
	OAc	1.54, 1.72 (each <i>s</i>)	1.84, 1.87 (each <i>s</i>)

*Peaks of the predominant rotamer only,
Splitting patterns and *J*-values (Hz) are given in parentheses

only the (4 β ,8)-derivative exhibited comparable restriction to rotation. Such differences in activation energy to induce 'free rotation' about the interflavanyl bond in the (–)-fisetimidol-(–)-epicatechin- and (+)-catechin analogues are presumably attributable to the effect of the axial 3-OAc(F) in the former group. The considerable higher energy requirements of the 3,4-*cis* analogues, e.g. **4**, may originate from significant contributions of A-conformers of the C-ring [12], these being evident from small $J_{2,3(C)}$ values [7.0 and 6.8 Hz for the (–)-epicatechin and (+)-catechin homologues respectively]. ^1H NMR experiments aimed at defining the preferred conformation about the interflavanyl bond [13] invariably failed as a result of the 'slow exchange' of rotamers at ambient temperatures. Such a phenomenon was evident from the prominent NOE association of 7-OMe(D) with H-4(C) in both **2** (1.1%) and **4** (1.6%).

Although efforts to identify the (4,6)-regio-isomers of **1** and **3** in the above sources have hitherto failed, we have indirect evidence of their natural occurrence from related tetrahydropyrano[2,3-*f* and *g*]chromenes [14]. The extremely low proportions in which the (4,6)-analogues were formed *in vitro* [7] may reflect similar minor concentrations in Nature hence complicating their isolation.

EXPERIMENTAL

^1H NMR spectra were recorded at 300 MHz in CDCl_3 (298 K) or $\text{DMSO}-d_6$ (453 K) with TMS as reference. CD spectra were determined in MeOH. Media used for the separation of components were Sephadex LH-20/ethanol or Fractogel TSK HW-40(S)/EtOH for CC, DC-plastikfolin 60 F₂₅₄ for TLC and Kieselgel PF₂₅₄ (1 mm \times 20 \times 20 cm) for prep. TLC. TLC bands were located under UV and/or with H_2SO_4 -HCHO (40/1) spray reagent and compounds recovered from the absorbent with

Me_2CO . Methylations were performed with excess CH_2N_2 over 48 hr at -15° and acetylations in Ac_2O -pyridine.

(–)-Fisetimidol-(4 α ,8) and (4 β ,8)-(–)-epicatechins **1** and **3** from *G. coleosperma* Heartwood drillings (6 kg) were extracted (EtOAc) and fractionated (Craig counter-current assembly followed by CC on Sephadex LH-20 EtOH) according to the procedures previously described [14]. Methylation of a portion (4 g) of fraction 2F (10.89 g) followed by prep. TLC (C_6H_6 - Me_2CO , 4/1, \times 2) afforded six bands, 2F₁ (R_f 0.62, 186 mg), 2E₂ (R_f 0.58, 253 mg), 2E₃ (R_f 0.54, 222 mg), 2E₄ (R_f 0.48, 268 mg), 2E₅ (R_f 0.45, 251 mg) and 2E₆ (R_f 0.36, 678 mg). Fraction 2E₆ was acetylated and resolved by prep. TLC (C_6H_6 - Me_2CO , 9/1) to give two bands at R_f 0.62 (120 mg) and 0.50 (90 mg). The former band afforded 3-*O*-acetyl-tri-*O*-methyl-(–)-fisetimidol-(4 β ,8)-3-*O*-acetyl-tetra-*O*-methyl-(–)-epicatechin **4** as a white solid [7]. ^1H NMR data (Table 1), MS and CD data [7]. The R_f 0.50 band gave the (4 α ,8)-isomer **2** as an amorphous white solid [7]. ^1H NMR data (Table 1), MS and CD data [7].

(–)-Fisetimidol-(4 α ,8)-(–)-epicatechin **1** from *B. plurijuga* Heartwood drillings (4.3 kg) were extracted (MeOH), enriched (Craig procedure) and fractionated (CC on Sephadex LH-20/EtOH) according to the procedures described earlier [14]. A portion (1.5 g) of fraction 3H (2.64 g) was subjected to CC (Fractogel TSK HW-40(S) EtOH) to give three fractions 3H₁ (172 mg), 3H₂ (342 mg) and 3H₃ (638 mg) in order of increasing R_f . Methylation of fraction 3H₃ and prep. TLC (hexane- Me_2CO -EtOAc, 11/6/3, \times 2) afforded a main band at R_f 0.46 (262 mg). This was acetylated and subsequently resolved by prep. TLC (CHCl_3 -EtOAc, 9/1) to two bands at R_f 0.51 (119 mg) and 0.33 (68 mg). The R_f 0.51 fraction consisted of the known [7] (–)-fisetimidol-(4 α ,8)-(+)-catechin heptamethyl ether diacetate and the R_f 0.33 band of the (–)-fisetimidol-(4 α ,8)-(–)-epicatechin heptamethyl ether diacetate **2**, with physical data identical to those of the compound from *G. coleosperma*.

(–)-Fisetimidol-(4 α ,8)-(–)-epicatechin **1** from *C. mopane* The

fractionation procedure for the methanol extract of the heartwood leading to fraction 43 has previously been described [15, 16]. This fraction (417 mg) was methylated and resolved by prep TLC (C_6H_6 - Me_2CO - $MeOH$, 90:9:1, $\times 3$) to give two bands at R_F 0.17 (23.8 mg) and 0.09 (145.8 mg). The latter band was acetylated and the mixture resolved by prep TLC (hexane- Me_2CO - $EtOAc$, 7:2:1, $\times 9$) to four fractions at R_F 0.33 (8.5 mg), 0.29 (51.1 mg), 0.25 (20.5 mg) and 0.09 (16.7 mg). The R_F 0.25 fraction afforded the (-)-fisetinidol-(4 α ,8)-(-)-epicatechin heptamethyl ether diacetate **2** with spectroscopic data identical to those from *G. coleosperma* and *B. pluriyuga*.

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REFERENCES

- Weinges, K., Kaltenhauser, W., Marx, H-D., Nader, E., Nader, F., Perner, J. and Seiler, D. (1968) *Justus Liebig's Ann Chem.* **711**, 184.
- Haslam, E. (1977) *Phytochemistry* **16**, 1625.
- Foo, L. Y. and Porter, L. J. (1978) *J. Chem. Soc., Perkin Trans I* 1186.
- Pelter, A., Amenechi, P. I., Warren, R. and Harper, S. H. (1969) *J. Chem. Soc. C* 2572.
- du Preez, I. C., Rowan, A. C. and Roux, D. G. (1970) *J. Chem. Soc., Chem. Commun.* 492.
- Ferreira, D., du Preez, I. C., Wijnmaalen, J. C. and Roux, D. G. (1985) *Phytochemistry* **24**, 2415.
- Botha, J. J., Ferreira, D. and Roux, D. G. (1981) *J. Chem. Soc., Perkin Trans I* 1235.
- Tindale, M. D. and Roux, D. G. (1974) *Phytochemistry* **13**, 829.
- Palgrave, K. C. (1983) *Trees of Southern Africa* (Moll, E. J., ed.) pp. 267, 268, 278. C. Struik, Cape Town.
- du Preez, I. C., Rowan, A. C. and Roux, D. G. (1971) *Chem. Commun.* 315.
- Hundt, H. K. L. and Roux, D. G. (1981) *J. Chem. Soc., Perkin Trans I* 1227.
- Porter, L. J., Wong, R. Y., Benson, M. and Chan, B. G. (1986) *J. Chem. Res. M* **830**, S86.
- Brandt, E. V., Young, D. A., Ferreira, D. and Roux, D. G. (1987) *J. Chem. Soc., Perkin Trans I* 2353.
- Steynberg, J. P., Burger, I. F. W., Young, D. A., Brandt, E. V., Steenkamp, J. A. and Ferreira, D. (1988) *J. Chem. Soc., Perkin Trans I* 3323.
- Malan, J. C. S., Steenkamp, J. A., Steynberg, J. P., Young, D. A., Brandt, E. V. and Ferreira, D. (1989) *J. Chem. Soc., Perkin Trans I* (in press).
- Malan, J. C. S., Young, D. A., Steynberg, J. P. and Ferreira, D. (1989) *J. Chem. Soc., Perkin Trans I* (in press).