Note

Synthesis and characterisation of methyl 2-O-(β -D-glucopy-ranosyl)-6-O-(a-L-rhamnopyranosyl)-a-D-glucopyranoside^{*}

Chris F. Hansmann

Fruit and Fruit Technology Research Institute, Private Bag X5013, Stellenbosch 7600 (Republic of South Africa)

The branched trisaccharide, 2- $O(\beta$ -D-glucopyranosyl)-6-O(a-L-rhamnopyranosyl)-D-glucopyranose (2), is regarded as the sugar moiety of an anthocyanin pigment isolated from the fruits and flowers of certain *Begonia*, *Clivia*, *Rubus*, *Prunus*, and *Ribis* species^{1,2}. The cyanidin-3-glycoside of 1 has been identified as one of the major pigments of Montmorency cherries (*Prunus cerasus*)³. It has also been isolated as quercetin and kaempherol glycosides from the flowers of potato (*Solanum tuberosum*)⁴. Synthesis of the methyl glycoside of 1, methyl 2- $O(\beta$ -D-glucopyranosyl)-6-O(a-L-rhamnopyranosyl)-a-D-glucopyranoside (2) is reported.

RESULTS AND DISCUSSION

The target methyl glycoside (2) was prepared in an overall yield of 21.8% starting from methyl 4,6-O-benzylidene-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (3, ref. 5).

Methyl 2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)- α -D-glucopyranoside (4) was obtained from 3 (89%) utilising 90% aqueous trifluoroacetic acid⁶ for debenzylidenation. Methyl 3,4-di-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-6-O-trityl- α -D-glucopyranoside (5) was obtained by treating 4 with chlorotriphenylmethane in pyridine, followed by acetylation.

Detritylation of 5 gave 6 (70% yield) which may be a generally useful intermediate in the synthesis of methyl 2-O-(β -D-glucopyranosyl)-6-O-(glycosyl)-a-D-glucopyranosides. Compound 8 was synthesised (63% yield) by condensation of 6 with 2,3,4-tri-Oacetyl-a-L-rhamnopyranosyl bromide (7, ref. 7) under Helferich–Zirner conditions. Deacetylation was effected by transesterification with sodium methoxide⁸ to yield the methyl glycopyranoside 2 in 76% yield.

0008-6215/90/\$ 03.50 © 1990 Elsevier Science Publishers B.V.

^{*} Part of an M.Sc. Food Science thesis submitted to the University of Stellenbosch. The research was done under the guidance of the late Prof. B. H. Koeppen.



The structure of 2 was verified as follows: The methyl glucopyranoside (2) was permethylated⁹, the product hydrolysed¹⁰ with 2M HCl, and the hydrolysate analysed by g.l.c. after trimethylsilylation, according to the Kovats' retention-index system¹¹ (Table I). Samples of per-O-Me₃Si derivatives of 2,3,4-tri-O-methyl-L-rhamnopyranose (9), 2,3,4,6-tetra-O-methyl-D-glucopyranose (10), and 3,4-di-O-methyl-D-glucopyranose (11) were used as reference compounds. The close agreement observed between the retention indexes of derivatives obtained from 2 and the reference compounds supports the expected structure of 2.

¹³C-N.m.r. data for 2, methyl 2-O- β -D-glucopyranosyl-a-D-glucopyranoside (12), and 6-O-a-L-rhamnopyransyl-D-glucopyranose (13) are summarised in Table II. Resonances in the spectrum of 2 were assigned by comparison with signals in the spectra of 12 and 13. Resonances in the spectra of 12 and 13 were assigned according to the multiplicity (in the case of methoxyl and 6-glycosyl carbons) observed in the singlefrequency proton off-beat decoupled spectra and by comparison with the spectra of suitable reference compounds in the literature¹³⁻¹⁶. The spectrum of 13, complicated by the occurrence of both the *a*- and β -anomeric forms, is in close agreement with that¹³ measured in D₂O.

TABLE I

G.l.c. retention indexe	s for the trimethylsily	l derivatives obtained	from target and	reference compounds
-------------------------	-------------------------	------------------------	-----------------	---------------------

Compound	Chromatogra	phic conditions	Retention indexes		
	Temperature (°)	Column			
	110	1% OV1	1394.2		
2	110	1% OV1	1535.5; 1552.4		
	140	2.5% OV1	1719.8; 1781.1		
2,3,4-Tri-O-methyl-L-rhamnopyranose (10)	110	1% OV1	1389.7		
2.3.4.6-Tetra-O-methyl-a-D-glucopyranoside (9)	110	1% OV1	1537.6; 1550.3		
3,4-Di-O-methyl-D-glucopyranose (11)	140	2.5% OV1	1719.6; 1781.6		

TABLE II

¹³C-N.m.r. data^{*a*} for the methyl 2-O- β -D-glucopyranosyl-*a*-D-glucopyranoside (12), 6-O-*a*-L-rhamnopyranosyl-D-glucopyranose (13) and 2

Compound	Anomeric configuration ^b	Chemical shift						
		C-1	C-2	C-3	C-4	C-5	C-6	OCH ₃
12	a	98.78	81.44	72.24ª	70.07	71.81ª	60.73	54.25
	β	104.85	73.58	76.73	69.72	76.28	61.13	
13	a	92.06	72.09	72.92	70.25ª	70.52ª	66.89	
	в	96.67	74.61	76.46	70.25ª	75.03	66.89	
	a"	100.43	70.25 [*]	70.52 ^b	71.91	68.14	17.75	
		100.29						
2	a	98.76	81.27	71.84ª	70.09	70.99	66.37	54.27
	ß	104.84	73.59	76.30	69.80	76.30	61.14	
	a″	100.50	70.31	70.65*	71.98"	68.22	17.81	_

^a Chemical shifts in p.p.m. downfield from tetramethylsilane. Assignments marked with identical alphabetical characters may be reversed. ^b The unprimed, single-primed and double-primed anomeric prefixes refer to the *a*-D-glucopyranosidic (reducing D-glucopyranose in the case of 13), β -D-glucopyranosyl and *a*-L-rhamnopyranosyl units in compounds 2, 12, and 13 respectively.

No conclusions could be made by ¹³C-n.m.r. spectroscopy regarding the anomeric configuration of the rhamnopyranosyl linkage, as the anomeric carbon atoms in the spectra of α - and β -L-rhamnopyranosides exhibit practically identical chemical shifts^{13,15}. This is attributed to the close proximity of three dipoles in the β -L-rhamnopyranosides and -rhamnopyranoses¹⁶.

Anomeric configuration of glycosidic bonds may be assigned¹² on the basis of molecular rotation $[M_D]$. Calculated $[M]_D$ values of $+118.4^\circ$ and $+453.6^\circ$ were obtained for **8** and for the isomeric methyl 2-O-(β -D-glucopyranosyl)-6-O-(β -L-rhamnopy-

ranosyl)-a-D-glucopyranoside nona-acetate, respectively. The experimental $[M]_D$ value found for 8 (+96.9°) supports the a-L configuration for the rhamnosidic linkage.

EXPERIMENTAL

General methods. — Melting points were determined on a Kofler block. Optical rotations were determined with a Perkin–Elmer 241 polarimeter. ¹H-N.m.r. spectra were recorded at 60 MHz for solutions in CDCl₃. Chemical shifts are reported in p.p.m. downfield from Me₄Si. Gas–liquid chromatography was performed on a Hewlett–Packard 5831A gas chromatograph equipped with dual flame-ionisation detectors, operated in the single detector mode. Silanised glass columns (3 m × 2 mm i.d.) packed with OV-1 (Table II) on acid-washed, dimethylsilane-treated Chromosorb W (60–80 mesh) were utilised. Nitrogen was used as carrier gas at a flow rate of 15 mL/min. Injection and detector blocks were maintained at 270°. Hydrogen and air flow-rates to the detector were 42.5 and 255 mL/min, respectively. Retention indexes were determined relative to suitable *n*-alkanes. ¹³C-N.m.r. spectra for solutions in Me₂SO-d₆ were recorded at 20 MHz using 5-mm tubes under the following conditions: acquisition time: 0.819 sec.; pulse delay: 0.000 sec.; pulse width: $6-9 \times 10^{-6}$ sec.; number of data points: 8192–9000 depending on the sample; pulse angle: 45–67° depending on the sample; resolution: 1.221 Hz per data point.

Methylation of the target compound 2. — Compound 2 (\sim 50 mg) was methylated as described by Brimacombe⁹.

Hydrolysis of the methylated target compound. — The methylation product of **2** was hydrolysed with 2M HCl¹⁰, transferred to CHl₃ and dried *in vacuo*.

2,3,4-Tri-O-methyl-L-rhamnopyranose (9) and 2,3,4,6-tetra-O-methyl-D-glucopyranose (10). — Compounds 9 and 10 were obtained by methylation followed by hydrolysis¹⁷.

Formation of Me_3Si derivatives. — The residue obtained after hydrolysis of the permethylated target compound, and also 3,4-di-O-methyl-D-glucopyranose (11), 9, and 10 were silylated by using 5:1:1 pyridine–HMDS–TMCS¹⁸. The silylation products were transferred to *n*-hexane¹⁹. In the case of 11, the available sample was extremely small and the silylation products were injected directly. Because of the severe tailing of pyridine observed on the 1% OV-1 column, a 2.5% OV-1 column was used.

6-O-a-L-Rhamnopyranosyl-D-glucopyranose (13). — Compound 13 was available from a previous synthesis; $[a]_D^{20} - 6.5^\circ$ (c 1.5, MeOH)²⁰.

Methyl 2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyanosyl)-a-D-glucopyranoside (4). — Compound 3 (ref. 5, 1.94 g, 3.2 mmol) was dissolved in 90% (v/v) aq. CF₃CO₂H (5 mL)⁶. After 20 min, diethyl ether (15 mL) was slowly added with stirring. The crystals

^{*} The unprimed, single-primed and double-primed numbers refer to atoms in the *a*-D-glucopyranosidic, β -D-glucopyranosyl, and *a*-L-rhamnopyranosyl units, respectively.

were filtered off, dried in a vacuum desiccator over paraffin wax, and recrystallised from EtOAc (1.49 g, 89%), m.p. 162–163°, $[a]_D^{17}$ + 55.50° (c 1.9, CHCl₃).

Anal. Calc. for C₂₁H₃₂O₁₅: C, 48.09; H, 6.15. Found: C, 48.2; H, 6.05.

Methyl 3,4-di-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-6-Otrityl-a-D-glucopyranoside (5). — A solution of 4 (2.6 g, 5 mmol) and chlorotriphenylmethane (1.3 g, 3.6 mmol) in anhydrous pyridine (13 mL) were stirred for 12 h. Additional chlorotriphenylmethane (0.5 g, 1.8 mmol) was added, followed by Ac₂O (15 mL) after another 2 h. The mixture was heated to boiling point and allowed to cool. After 30 min, it was processed conventionally, and crystallised from MeOH to give 5 (2.9 g, 68%), m.p. 140–141°, $[a]_{D}^{17}$ + 71.10° (c 3, CHCl₃); ¹H-n.m.r.: δ 7.1–7.6 (m, 15 H, aromatic), 4.5–5.6 (m, 7 H), 4.1–4.4 (m, 2 H, Ha-6^{*}, Hb-6'), 3.6–4.1 (m, 3 H), 3.43 (s, 3 H, methoxyl), 2.8–3.25 (m, 2 H, Ha-6, Hb-6), 1.9–2.15 (m, 15 H, acetoxyl), 1.69 (s, 3 H, acetoxyl).

Anal. Calc. for C44H50O17: C, 62.11; H, 5.92. Found: C, 62.2; H, 5.78.

Methyl 3,4-di-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-a-D-glucopyranoside (6). — Compound 5 (1.0 g, 1.2 mmol) was dissolved in AcOH (18 mL) and cooled to ~10°. Hydrogen bromide [1.6 mL, 40% (m/v) in AcOH, ~8 mmol] was added and the mixture shaken vigorously for 45 sec. The mixture was processed conventionally and the product crystallised from EtOH to give 6 (0.5 g, 70%), m.p. 160–162°, $[a]_D^{21} + 49.7^\circ$ (c 1.2, CHCl₃); ¹H-n.m.r.: δ 4.5–5.7 (m, 7 H), 4.1–4.6 (m, 2 H, Ha-6', Hb-6'), 3.5–4.0 (m, 5 H), 3.42 (s, 3 H, methoxyl), 2.47 (s, 1 H, 6-OH), 1.98–2.12 (m, 18 H, acetoxyl).

Anal. Calc. for C₂₅H₃₆O₁₇: C, 49.34; H, 5.96. Found: C, 49.2; H, 6.12.

Methyl 3,4-di-O-acetyl-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-6-O-(2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl)- α -D-glucopyranoside (8). — A mixture of 6 (0.5 g, 0.8 mmol), Hg(CN)₂ (0.32 g, 1.2 mmol) and HgBr₂ (0.45 g, 1.2 mmol) in MeCN (10 mL) was stirred while tri-O-acetyl-L-rhamnopyranosyl bromide (8, 0.46 g, 1.2 mmol) was added. After 45 min, the mixture was processed conventionally, and 8 crystallised from EtOH (0.46 g, 63.6%), m.p. 167–169°, $[a]_D^{21} + 11.0°$ (c 1.1, CHCl₃); ¹H-n.m.r.: δ 3.5–5.6 (m, 17 H), 3.43 (s, 3 H, methoxyl), 1.98–2.13 (m, 27 H, acetoxyl), 1.20 (d, 3 H, Ha-6", Hb-6", Hc-6").

Anal. Calc. for C₃₇H₅₁O₂₄: C, 50.51: H, 5.84. Found: C, 50.6; H, 5.71.

Methyl 2-O-(β -D-glucopyranosyl)-6-O-(a-L-rhamnopyranosyl)-a-D-glucopyranoside (2). — Compound 2 was prepared by deacetylation of 8 (64 mg)⁸ and was obtained as a pure (t.l.c., silica gel, 3:17 acetone-benzene) amorphous powder, which failed to crystallise from various solvents. (28 mg, 76%), $[a]_{21}^{21} + 13.1^{\circ}$ (c 3, MeOH).

Methyl 2-O- β -D-glucopyranosyl-a-D-glucopyranoside (12). — Compound 12 was prepared by deacetylation of 4 (140 mg)⁸. The residue (81 mg, 85%) was crystallised from EtOH, m.p. 254–256°.

The author is indebted to Mr. H. C. Spies, Department of Chemistry, University of Stellenbosch, for recording the n.m.r. spectra. A donation of 3,4-di-O-methyl-D-glucopyranose by Dr. E. Merrifield, Department of Chemistry, University of Cape Town, is appreciated.

REFERENCES

- 1 J. B. Harborne and E. Hall, Biochem. J., 88 (1963) 41p-42p.
- 2 J. B. Harborne and E. Hall, Phytochemistry, 3 (1964), 453-463.
- 3 R. R. Fisher and J. H. von Elbe, J. Milk Food Technol., 33 (1970), 481-483.
- 4 J. B. Harborne, Comparative Biochemistry of the Flavanoids. Academic Press, New York, 1967, 64.
- 5 B. Coxon and H. G. Fletcher, Jr., J. Org. Chem., 26 (1961) 2892-2894.
- 6 J. E. Christensen and L. Goodman, Carbohydr. Res., 7 (1968) 510-512.
- 7 E. Fischer, M. Bergmann, and A. Rabe, Ber., 53 (1920), 2362-2388.
- 8 A. Thompson and M. L. Wolfrom, Methods Carbohydr. Chem., 2 (1963), 215-220.
- 9 J. S. Brimacombe, Methods Carbohydr. Chem., 6 (1972), 376-378.
- 10 E. B. Rathbone, A. M. Stephen, and K. G. R. Pachler, Carbohydr. Res., 20 (1971), 141-150.
- 11 L. S. Ettre, Chromatographia, 6 (1973), 489-495.
- 12 W. Klyne, Biochem. J., 47 (1950), xli-xlii.
- 13 C. Lafitte, A. N. Phuoc Du, F. Winternitz, R. Wylde, and F. Pratviel-Sosa, Carbohydr. Res., 67 (1978), 105-115.
- 14 T. Usui, N. Yamaoka, K. Matsuda, and K. Tuzimura, J. Chem. Soc., Perkin Trans. 1, (1973) 2425-2432.
- 15 R, Kasai, M. Okihara, J. Asakawa, K. Mizutani, and O. Tanaka, Tetrahedron, 35 (1979), 1427-1432.
- 16 E. Breitmaier and W. Voelter, ¹³C NMR spectroscopy. Methods and Applications in Organic Chemistry., Verlag Chemie, Weinheim, 1978, 247–263.
- 17 E. S. West, and R. F. Holden, Org. Synth. 3 (1955), 800-803.
- 18 A. E. Pierce, Silylation of Organic Compounds. Pierce Chemical Company, Rockford, Illinois, 1978, 20.
- 19 R. D. Wood, P. K. Raju, and R. Reiser, J. Am. Oil Chemists' Soc., 42 (1965), 161.
- 20 M. J. Theron, M. Sc. Thesis, University of Stellenbosch, (1974).