

STRUCTURES OF MOMORDICOSIDES F<sub>1</sub>, F<sub>2</sub>, G, I, K AND L,  
 NOVEL CUCURBITACINS IN THE FRUITS OF MOMORDICA CHARANTIA L.

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**Abstract:** Two bitter cucurbitacins, momordicosides K and L, and four non-bitter cucurbitacins, momordicosides F<sub>1</sub>, F<sub>2</sub>, G and I, were isolated from the immature fruits of Momordica charantia L. (Cucurbitaceae) and their structures were elucidated.

Cucurbitacins comprise a group of triterpenes, most of which are bitter principles commonly distributed in the cucurbitaceous plants. The fruits of Momordica charantia L. (Cucurbitaceae) taste bitter, and had been expected to contain cucurbitacins.

The triterpenoid constituents in this plant have been extensively investigated in this laboratory, and we reported the isolation and structures of five non-bitter cucurbitacins designated as momordicosides A~E isolated from the seeds.<sup>1)</sup> They are the second example of the rare cucurbitacins which have no oxygen function at C<sub>11</sub>.<sup>2)</sup>

In this paper, we wish to report the structures of two bitter glycosides, momordicosides K and L, and four non-bitter glycosides, momordicosides F<sub>1</sub>, F<sub>2</sub>, G and I, isolated from the immature fruits, which are the novel cucurbitacins differ from ones hitherto reported.

The MeOH extractive of the fresh fruits was washed with water, and from the water-insoluble material, momordicosides were isolated in crystalline form by column chromatography.

|  |             |                          |                                   |              |
|--|-------------|--------------------------|-----------------------------------|--------------|
| F <sub>1</sub> : C <sub>37</sub> H <sub>60</sub> O <sub>8</sub> •2.5H <sub>2</sub> O | mp 198-203° | [α] <sub>D</sub> -111.0° | [θ] <sub>200</sub> -82800° (MeOH) | yield 0.003% |
| F <sub>2</sub> : C <sub>36</sub> H <sub>58</sub> O <sub>8</sub> •H <sub>2</sub> O    | 155-158°    | [α] <sub>D</sub> -96.5°  |                                   | 0.0003       |
| G: C <sub>37</sub> H <sub>60</sub> O <sub>8</sub> •2H <sub>2</sub> O                 | 183-187°    | [α] <sub>D</sub> -107.3° |                                   | 0.0009       |
| I: C <sub>36</sub> H <sub>58</sub> O <sub>8</sub> •H <sub>2</sub> O                  | 210-216°    | [α] <sub>D</sub> -110.2° |                                   | 0.0006       |
| K: C <sub>37</sub> H <sub>60</sub> O <sub>9</sub> •1/2H <sub>2</sub> O               | 236-237°    | [α] <sub>D</sub> +63.3°  | [θ] <sub>208</sub> +58500° (MeOH) | 0.0018       |
| L: C <sub>36</sub> H <sub>58</sub> O <sub>9</sub> •2H <sub>2</sub> O                 | 227-232°    | [α] <sub>D</sub> +57.3°  |                                   | 0.0002       |

Enzymatic hydrolysis of F<sub>1</sub> using the crude hesperidinase provided F<sub>1</sub>-aglycone (C<sub>31</sub>H<sub>50</sub>O<sub>3</sub>, mp 139-140°, [α]<sub>D</sub> -90.6°, [θ]<sub>199</sub> -43500° (MeOH)) and D-glucose. <sup>1</sup>H NMR data<sup>3)</sup> [δ 1.31 (6H, s), 3.22 (3H, s, OCH<sub>3</sub>), 3.60 (H, m), 3.55, 3.67 (1H each, d, J=9 Hz), 5.60 (2H, br. s), 5.61 (H, dd, J=10, 4 Hz), 6.14 (H, dd, J=10, 2 Hz)] and <sup>13</sup>C NMR data<sup>3)</sup> (Table) of F<sub>1</sub>-aglycone and <sup>13</sup>C NMR data of F<sub>1</sub> suggested that F<sub>1</sub> is a mono-β-D-glucopyranoside of a tetracarboxylic five-ring triterpene having partial structures presented below.

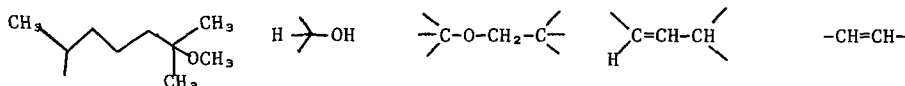
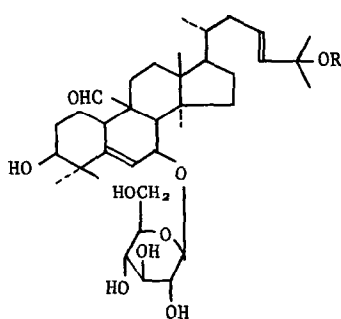
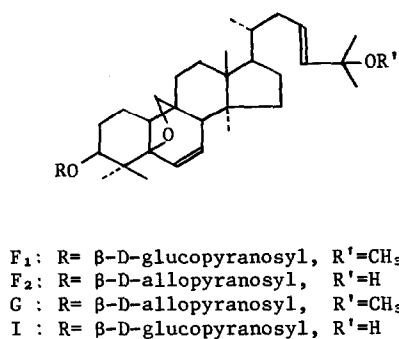


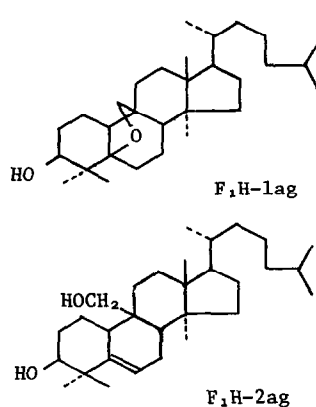
Table  $^{13}\text{C}$  NMR Chemical Shifts of Momordicosides and Their Aglycones

[ (a): aglycone moiety, (s): sugar moiety ]

| momordicosides           | quat. C                  | oxygen-bearing C  | olefinic C                       |
|--------------------------|--------------------------|---|----------------------------------|
| F <sub>1</sub>           | 39.0, 45.2<br>45.4, 48.8 | (a) 52.2q, 74.8s, 80.0t, 85.4d, 85.8s<br>(s) 63.0t, 71.7d, 75.7d, 78.3d(x2)<br>106.8d     | 128.3d, 129.9d<br>134.1d, 137.6d |
| G                        | 38.9, 45.1<br>45.3, 48.7 | (a) 52.1q, 74.7s, 79.9t, 84.9d, 85.7s<br>(s) 63.1t, 69.0d, 72.3d, 72.8d, 75.8d<br>103.6d  | 128.0d, 129.6d<br>133.8d, 137.3d |
| F <sub>1</sub> -aglycone | 37.5, 45.4<br>45.6, 48.7 | 52.1q, 74.7s, 76.2d, 79.8t, 87.5s   | 128.3d, 131.2d<br>132.4d, 137.6d |
| F <sub>2</sub>           | 39.0, 45.2<br>45.4, 48.8 | (a) 69.7s, 80.1t, 85.1d, 85.9s<br>(s) 63.3t, 69.2d, 72.4d, 73.0d, 76.1d<br>103.8d         | 124.1d, 129.9d<br>134.1d, 141.7d |
| I                        | 39.0, 45.2<br>45.3, 48.8 | (a) 69.7s, 80.0t, 85.4d, 85.8s<br>(s) 63.0t, 71.8d, 75.7d, 78.3d(x2)<br>106.8d            | 124.1d, 129.9d<br>134.1d, 141.6d |
| I-aglycone               | 37.6, 45.4<br>45.6, 48.8 | 69.7s, 76.2d, 79.8t, 87.6s  | 124.1d, 131.3d<br>132.4d, 141.7d |
| K*                       | 41.8<br>45.6, 47.9       | (a) 50.0q, 71.5d, 74.7s, 74.7d, 207.0d<br>(s) 62.7t, 71.7d, 75.4d, 78.3d, 78.5d<br>101.5d | 121.9d, 128.0d<br>137.3d, 147.1s |
| L*                       | 41.8<br>45.5, 47.9       | (a) 69.6s, 71.5d, 74.7d, 206.9d<br>(s) 62.8t, 71.5d, 75.4d, 78.5d(x2)<br>101.4d           | 122.0d, 123.8d<br>141.3d, 147.2s |
| F <sub>1</sub> H-lag     | 38.6, 46.1<br>48.5, 49.9 | 76.8d, 78.5t, 89.9s   |                                  |
| F <sub>1</sub> H-2ag     | 39.6, 41.7<br>46.4, 49.4 | 65.5t, 76.0d  | 120.4d, 142.9s                   |

\* The 4th quaternary carbon signal may overlap probably on the signal at  $\delta$ 41.8.

L: R = H



When F<sub>1</sub>-acetate was hydrogenated over PdO in EtOH, two products (F<sub>1</sub>H-1 and F<sub>1</sub>H-2) having no methoxyl group were obtained.<sup>4)</sup> They were separately saponified and methanolized to give respective aglycones, F<sub>1</sub>H-lag (C<sub>30</sub>H<sub>52</sub>O<sub>2</sub>, mp 133-134°, [ $\alpha$ ]<sub>D</sub> +31.4°) and F<sub>1</sub>H-2ag (C<sub>30</sub>H<sub>52</sub>O<sub>2</sub>•H<sub>2</sub>O, mp 169-170°, [ $\alpha$ ]<sub>D</sub> +40.5°). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra indicated that F<sub>1</sub>H-lag is a saturated alcohol in which the ether linkage is retained. F<sub>1</sub>H-2ag is a diol which has a primary hydroxyl group and a trisubstituted double bond which were derived by reductive cleavage of the ether ring and migration of the double bond. The chemical shift of the olefinic proton ( $\delta$ 5.76, br.d, J=6 Hz) and its signal pattern are similar to those ( $\delta$ 5.61, br.d, J=5.5 Hz) of cucurbit-5-ene-3 $\beta$ ,22(S),23(R),24(R),25-pentaol (A-aglycone)<sup>1)</sup> and the <sup>13</sup>C NMR chemical shift of olefinic and quaternary carbons are almost the same with those of A-aglycone<sup>1)</sup> (olefinic C: 119.1d and 143.0s. quaternary C: 34.8, 41.6, 46.8 and 49.2) except for the chemical shift of the quaternary carbon to which the hydroxymethyl group is linked.

The keto-aldehyde (C<sub>30</sub>H<sub>48</sub>O<sub>2</sub>, EI-MS:m/z 440 (M<sup>+</sup>), mp 123-125°, IR(KBr): 1718, 1725(sh.) cm<sup>-1</sup>) obtained by oxidation of F<sub>1</sub>H-2ag with pyridinium chlorochromate in CH<sub>2</sub>Cl<sub>2</sub> showed a negative CD spectrum ([ $\theta$ ]<sub>280</sub> -3400°(dioxane))<sup>5)</sup>. These data indicate that F<sub>1</sub>H-2ag is cucurbit-5-ene-3 $\beta$ ,19-diol, and hence, F<sub>1</sub> is a 3-O- $\beta$ -D-glucopyranoside of 5,19-epoxy-25-methoxy-5 $\beta$ -cucurbita-6,23-dien-3 $\beta$ -ol, and this formulation is well supported by CD curves of F<sub>1</sub> and F<sub>1</sub>-aglycone which showed [ $\theta$ ]<sub>280</sub>-82800° and [ $\theta$ ]<sub>199</sub>-43500°, respectively.<sup>6)</sup>

G exhibited almost the same <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra with those of F<sub>1</sub> except for signals of the sugar moiety. When G-permethyrate was hydrogenated and methanolized, F<sub>1</sub>H-lag and a methylated sugar were provided. The latter was identified as methyl 2,3,4,6-tetra-O-methyl  $\beta$ -D-allopyranoside by its specific rotation (-14.7°)<sup>7)</sup> and by examination of the <sup>1</sup>H NMR spectrum [ $\delta$  4.60(H<sub>1</sub>, d, J=8 Hz), 2.99(H<sub>2</sub>, dd, J=8, 3 Hz), 4.00 (H<sub>3</sub>, t, J=3, 2 Hz), 3.22 (H<sub>4</sub>, dd, J=2, 12 Hz), 3.80 (H<sub>5</sub>, m)]. The J-value (8 Hz) of the anomeric proton of the sugar moiety of G-permethyrate indicates that D-allopyranose is in  $\beta$ -configuration.

Thus, G is the 3-O- $\beta$ -D-allopyranoside of F<sub>1</sub>-aglycone.

F<sub>2</sub> and I are considered to be the C<sub>25</sub>-OH derivatives corresponding to G and F<sub>1</sub>, respectively, because they showed similar <sup>1</sup>H NMR spectra to those of G and F<sub>1</sub>, except that they exhibited a deshielded signal ( $\delta$ 1.53) due to two methyl groups on C<sub>25</sub> and no methoxyl signal, and also because the <sup>13</sup>C NMR signal of C<sub>25</sub> was shifted up-field ca 5 ppm from those of G and F<sub>1</sub>.

Their relationship to G and F<sub>1</sub> were established by converting them into permethylates of G and F<sub>1</sub>, respectively. Therefore, F<sub>2</sub> is the 3-O- $\beta$ -D-allopyranoside of 5,19-epoxy-5 $\beta$ -cucurbita-6,23-diene-3 $\beta$ ,25-diol, and I is its 3-O- $\beta$ -D-glucopyranoside.

K is suggested by <sup>1</sup>H NMR and <sup>13</sup>C NMR data to be a mono- $\beta$ -D-glucopyranoside of a tetracyclic triterpene having a tertiary formyl group ( $\delta$ 10.43, s), a disubstituted and a trisubstituted double bonds, two secondary hydroxyl groups, and a tertiary methoxyl group at the terminal of the side chain, and L to be the corresponding C<sub>25</sub>-OH derivative. The same permethylate was obtained when K and L were separately methylated.

The aglycone (Lag)(C<sub>30</sub>H<sub>48</sub>O<sub>4</sub>, mp 190-193°) of L showed <sup>1</sup>H NMR signals of one carbinyl proton (H,  $\delta$  4.36, br.d, J=5 Hz) coupled with an olefinic proton (H,  $\delta$  6.25, br.d, J=5 Hz), one carbinyl proton (H,  $\delta$  3.81, br.s) and two olefinic protons (2H,  $\delta$  5.91, br.s). The <sup>1</sup>H NMR spectrum of L also depicted the signal of a carbinyl proton at  $\delta$  3.81 and this signal was

shifted down-field ( ca  $\delta$  5.0 ) by acetylation. This fact indicates that the glucose is linked not to the isolated hydroxyl group, but to the one allylic to the trisubstituted double bond.

$\text{NaBH}_4$  reduction of K provided a primary alcohol Kred ( mp 195-197°,  $[\alpha]_D +60.1^\circ$ ,  $[\theta]_{20.5} +87900^\circ$  (MeOH)), which gave  $F_1$ -aglycone on treatment with 50% AcOH in MeOH at 38°.

When Kred was hydrogenated over PdO, the glucopyranosyl group was eliminated and two compounds identical with  $F_1$ H-lag and  $F_1$ H-2ag were yielded.  $F_1$ -aglycone is not the genuine aglycone of Kred, and it is considered to be derived from the glycoside by bond fission between the aglycone-C and the glucose-O giving an intermediate  $C_7$  carbonium cation, subsequent migration of the double bond and then formation of the linkage between  $C_5$  carbonium cation and the oxygen of the primary hydroxyl group. Formation of  $F_1$ H-lag by hydrogenation is also supposed to be initiated by cleavage of  $C_7$ -O bond, followed by migration of the double bond and formation of the ether ring, reductive opening of which leads to  $F_1$ H-2ag.

The orientation of  $C_7$ -hydroxyl group was determined as  $\beta$  from the positive CD curve of Kred<sup>6)</sup> and from the J-values (5 Hz) of  $C_6$ -H and  $C_7$ -H of Lag.<sup>8)</sup>

These data and consideration have led us to the conclusion that Kred is a 7-O- $\beta$ -D-glucopyranoside of 25-methoxy-cucurbita-5,23-diene-3 $\beta$ ,7 $\beta$ ,19-triol, and K and L are 7-O- $\beta$ -D-glucopyranosides of 19-oxo-25-methoxy-cucurbita-5,23-diene-3 $\beta$ ,7 $\beta$ -diol and its  $C_{25}$ -OH derivative, respectively.

Momordicosides  $F_1$ ,  $F_2$ , G and I are the first cucurbitacins having 5 $\beta$ -cucurbitane skeleton occurred in nature, and K and L are noted as the new cucurbitacins having  $C_9$ -formyl and 7-O- $\beta$ -D-glucopyranosyl groups. Among momordicosides, G and  $F_2$  are the first members of cucurbitacins having D-allose as a component sugar.

#### References and Notes

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- 5) R. Tschesche, G. Biernoth and G. Snatzke, Ann., **674**, 196 (1964)
- 6) A. I. Scott and A. D. Wrixon, Tetrahedron, **27**, 4787 (1971)
- 7)  $\beta$ -anomer:  $-19.0^\circ$ ,  $\alpha$ -anomer:  $+148.5^\circ$  ( unpublished data reported by K. Mihashi et al. of this university at the 101st Annual Meeting of The Pharmaceutical Society of Japan, Kumamoto, April, 1981)
- 8) The signal of  $C_6$ -H of 3 $\beta$ ,7 $\alpha$ ,11 $\beta$ -triacetoxy-cucurbit-5-ene is reported as a broad singlet (  $W_{1/2}$  4 Hz ) and that of  $C_7$ -H as a broad doublet (  $J=7$  Hz ).

(Received in Japan 4 September 1981)