

Direct photolysis and singlet photosensitization of **1** and **2** occur with absorption of $\sim 82\text{--}90$ kcal/mol of energy. Electronically excited singlets **1-2*** and/or possibly excited **15*** are thus highly energetic and their conversions into **4*** and **5*** are spin allowed. Rearrangement to **5*** is now extensive and the product does not undergo alteration as occurs when derived from vibrationally excited intermediates in the gas phase.^{7c} Finally, triplet photosensitization of diazo compounds and diazirines as in the present systems may give advantage over thermolysis and direct photolysis for specific synthesis.

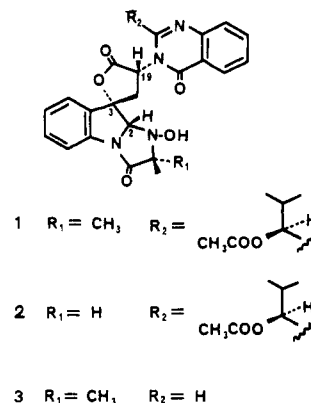
References and Notes

- (12) In diglyme containing lithium *tert*-butoxide, **2** photolyzes at 20 °C to **4** (51.6%), **5** (46.6%), and **7** (1.8%).
- (13) Extension of above general theory of this manuscript raises the question that such gas-phase decompositions involve electronically excited intermediates.
- (14) 10-Thioxanthone effects efficient photosensitization of larger ring azo analogues of **1** and **3**: P. S. Engel, *J. Am. Chem. Soc.*, **91**, 6903 (1969).
- (15) (a) Hanovia 679A lamp, Corning 7380 filter; 10-thioxanthone, ϵ 2800 (349 nm). (b) Reaction mixtures were samples at <20% conversion. (c) Hanovia 679A lamp, potassium chromate filter.
- (16) Neopentane (~1%) formed upon photosensitizations in cumene possibly arises by hydrogen abstraction by **12** and then the neopentyl radical.
- (17) Isomerizations of **12** with spin preservation to **13** and **14** apparently are higher energy demanding processes.^{2d}
- (18) (a) P. F. Zittel, G. B. Ellison, S. V. O'Neil, E. Herbert, W. C. Lineberger, and W. P. Reinhardt, *J. Am. Chem. Soc.*, **98**, 3731 (1976).^{18b} (b) Prior experimental values for the triplet-singlet energy gap for methylene range from 6 to 10 kcal/mol.^{18c} (c) H. M. Frey, *J. Chem. Soc., Chem. Commun.*, 1024 (1972); W. L. Hase, R. J. Phillips and J. W. Simons, *Chem. Phys. Lett.*, **12**, 161 (1971); F. S. Rowland, C. McKnight, and E. K. C. Lee, *Ber. Bunsenges.*, **72**, 236 (1968).
- (19) (a) R. C. Friedmann of this laboratory has observed that photosensitization of 4-methyl-1-pyrazoline with benzophenone and with 10-thioxanthone in tetrahydrofuran at -78 to 25 °C yields **7** (97.0–98.0%) and **8** (1.5–2.8%). Direct photolysis of 4-methyl-1-pyrazoline in tetrahydrofuran at -78 to 25 °C results in **7** (84.4–88.0%), **8** (10.0–15.4%), and 1-butene (0.3–1.0%). The behavior of 4-methyl-1-pyrazoline and its presumed photolytic intermediates,^{19b} triplet and singlet 2-methyl-1,3-propane diradicals, is quite different from that for photosensitization, thermolysis, and photolysis of **3** and argues strongly (1) against triplet and singlet β -C–H abstraction reactions for 2-methyl-1-propylidenes from **3** as a major source of **8** (and probably **7**) and (2) for spin inversion of triplet to singlet 2-methyl-1-propylidene and then to **8** (and probably **7**). Further, triplet photosensitization of 4,4-dimethyl-1-pyrazoline to give **4** (94–98%) and 2-methyl-1-butene (trace) as the only intramolecular products supports the conversion of **12** from **1** into **11** and then **5** (and probably **4**). (b) For summary of the experimental results and the theory of decomposition of various 1-pyrazolines, see G. Koga, N. Koga, and J.-P. Anselme and R. J. Dreier in "The Chemistry of the Hydrazo, Azo, and Azoxy Groups, Part 2", S. Patai, Ed., Wiley, New York, 1978, Chapters 19 and 20.

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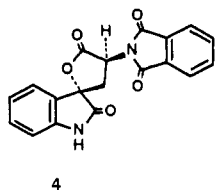
Total Synthesis of Tryptoquivaline G

A strain of the fungus *Aspergillus clavatus* collected from mold damaged rice produced a group of toxic, tremor inducing metabolites with novel structures. Tryptoquivaline (**1**) was found to be the major metabolite, and a transformation product containing a δ -lactone ring was used to determine its structure and relative configuration by X-ray crystallography.¹ Comparison of circular dichroism and ¹H NMR spectra with those of nortryptoquivaline² suggested structure **2** for this companion metabolite. Tryptoquivaline G (**3**) is a representative of a more recently discovered group of mycotoxins produced by *Aspergillus fumigatus*.³⁻⁵ It, as well as tryptoquivaline L (**17**), an artefact, lacks the isobutyl side chain. The total synthesis of tryptoquivaline G (**3**) outlined here confirms the proposed

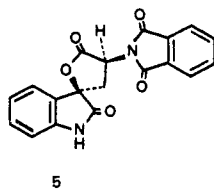


structure and establishes both relative and absolute configuration.

The synthesis relied heavily on a new method for the conversion of *N*-acyltryptophans to spirolactones and the steric course of the reaction was explored with a model compound.⁶ Oxidation of *N*-phthalimido-L-tryptophan with 2 equiv of trichloromethanesulfonyl chloride–dimethyl sulfoxide⁷ (CH₂Cl₂, –20 °C) gave a 65% yield of two diastereomeric lactones (**4** and **5**) in a ratio of 7:3. The major isomer **4**, mp



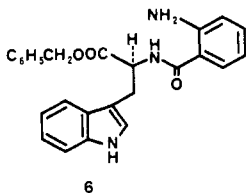
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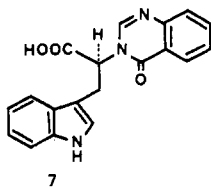
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275–277 °C, [α]_D²⁵ –133° (*c* 1.7, acetone), was recovered unchanged after treatment with sodium hydride or imidazole in dimethylformamide. The minor isomer **5**, mp 279–281 °C, [α]_D²⁴ –206° (*c* 1.4, acetone), under the same conditions was converted into the *enantiomer* of **4**, [α]_D²⁵ +132° (*c* 1.4, acetone), demonstrating different configurations at the spiro atom in the two diastereomers. Since the epimerization of tryptoquivaline G (**3**) to tryptoquivaline L (**17**) is accompanied by a large negative shift in optical rotation, the absolute configurations at C₃ and C₁₉ should be opposite those of the model compound **5**. The major, and thermodynamically more stable, epimer formed in the oxidative lactonization thus has the correct stereochemistry at the spiro center.

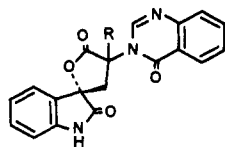
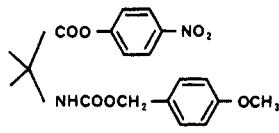
Intermediate **6** was prepared by condensation of L-tryptophan with *o*-nitrobenzoyl chloride to give the amide, mp



6



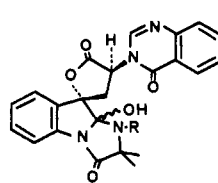
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8 R = α –H9 R = β –H

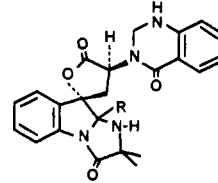
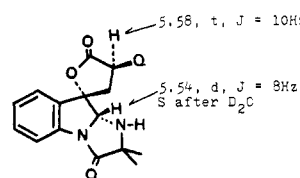
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212–214 °C, esterification with phenyldiazomethane to the benzyl ester, mp 148–150 °C, and reduction with iron (HCl, ethanol, reflux). The formamide, mp 140–142 °C (HCOOH, benzene, reflux), on dehydration (TsOH, xylene, reflux) gave a quinazolinone which was hydrogenolyzed to the acid **7**, mp 238–240 °C, over palladium on carbon in ethanol (overall yield from L-tryptophan was 50%). Oxidation of **7** with 2 equiv of methanesulfonyl anhydride–dimethyl sulfoxide (CH₂Cl₂, –20 °C, 5 h) gave 56–66% spirolactone **8**, mp 321–322 °C dec, [α]_D²⁵ –377° (*c* 0.07, CH₃CN), in addition to <10% epimer differing in configuration at the spiro center. To confirm the stereochemistry, **8** was exposed to potassium hydride in THF–DMF (20 °C, 3 min), and the resulting enolate was protonated with 1% HCl in THF (–70 °C). Product **9** (40%), mp 315–318 °C dec, [α]_D²⁵ +320° (*c* 0.02, CH₃CN), was the *enantiomer* of the minor product formed in the oxidative lactonization. Owing to the exceptional lability of **9** to base, the

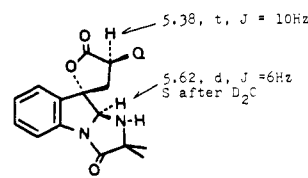
synthesis was continued with **8**. Derivative **10**, mp 82–83 °C, was prepared from α -methylalanine and *p*-methoxybenzyl-8-quinolyl carbonate⁸ with the aid of triethylamine, followed by esterification with *p*-nitrophenol and dicyclohexylcarbodiimide. Lactone **8** was then silylated with bis(trimethylsilyl)acetamide⁹ and the crude product condensed with **10** in DMF containing tetramethylammonium chloride. Treatment of the crude imide with triethylamine gave the highly insoluble cyclol **11**, mp 243–245 °C, in 81% overall yield. Transforma-

11 R = COOCH₂–C₆H₄–OCH₃

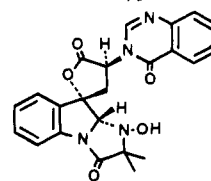
12 R = H

13 R = β –H14 R = α –H

15 Q = quinazolinyl



16



17

tion to the amins **15** and **16** could be accomplished by consecutive treatment of **11** with sodium cyanoborohydride, DDQ, and trifluoroacetic acid, but the two epimers were obtained in a ratio of 1:1. In a more stereoselective sequence, the *p*-methoxybenzyloxycarbonyl group was first removed by treatment of a suspension of the cyclol **11** in ethyl acetate with trifluoroacetic acid in the presence of anisole (0 °C, 30 min).¹⁰ The deprotected cyclol **12**, mp 268–270 °C (76%), was reduced with sodium cyanoborohydride (THF, H₂O, HCl, pH ~3, 20 °C, 1 h) to give a 4:1 mixture of dihydroquinazolinones **13**, mp 248–250 °C, and **14**, mp 243–245 °C. Reoxidation to the corresponding quinazolinones **15**, mp 250–251 °C (acetone), [α]_D²⁵ –260° (*c* 0.01, acetone), and **16**, mp 172–174 °C (AcOEt–hexane), [α]_D²⁵ –67° (*c* 0.12, CHCl₃), was accomplished in 89% and 80% yield, respectively, with DDQ in chloroform. ¹H NMR spectra of the two isomers were used to establish the configurations at C₂, and the major isomer on oxidation with *m*-chloroperbenzoic acid² gave tryptoquivaline L (**17**, 85%), mp 275–277 °C dec, [α]_D²⁵ –230° (*c* 0.02, acetone), with spectral properties in accord with those published.^{4,5} Contrathermodynamic epimerization, as in the conversion of **8** into **9**, gave tryptoquivaline G identical (melting point, [α]_D, IR, ¹H NMR and chromatographic behavior) with material isolated from *A. fumigatus*.¹¹ The tryptoquivalines are thus derived from D-tryptophan.

The configuration of nortryptoquivaline (**2**) was established by X-ray analysis¹² and reduction of this metabolite with zinc followed by hydrolysis which yielded L-alanine.^{3,5} The absolute configuration of the metabolite (**2**) thus agrees with that of tryptoquivaline G (**3**) determined by synthesis.

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General Acid Catalysis in the Hydrolysis of Benzo[a]pyrene 7,8-Diol 9,10-Epoxides

Sir:

The most studied of the carcinogenic hydrocarbons is the ubiquitous environmental contaminant benzo[a]pyrene (BP). Studies from several laboratories¹ have recently allowed the identification of the metabolite (+)-7 β ,8 α -dihydroxy-9 α ,10 α -epoxy-7,8,9,10-tetrahydro BP ((+)-**2**, Figure 1) as an ultimate carcinogen of BP in newborn mice.² Two diastereomeric 7,8-diol 9,10-epoxides of BP are metabolically possible, each capable of existing in enantiomeric forms. Although (\pm)-**1** is >30-fold more hydrolytically reactive in the physiological pH range³ and more susceptible to attack by nucleophiles in nonaqueous solution relative to (\pm)-**2**,⁶ only (+)-**2** is tumorigenic in newborn mice. Both **1** and **2** alkylate the phosphate backbone of nucleic acid and effect the strand scission of DNA.⁷ Because alkylation at phosphate may play an important role in the mutagenic and carcinogenic activities of **1** and **2**, the mechanisms of reaction of **1** and **2** with hydrogen phosphate species (and other general acids) in aqueous dioxane solutions were determined and are reported in this study.⁸

The rates of reaction of **1** and **2** in 10% dioxane-water solutions at a given pH exhibit a marked first-order dependence on concentration of phosphate buffer (Figure 2).^{9,10} The rate data for series of serially diluted buffer solutions of constant pH were fit to

$$k_{\text{obsd}} = k_{\text{HA}}[\text{HA}] + k_{\text{H}+\text{aH}+} + k_0 \quad (1)$$

where $k_{\text{H}+\text{aH}+}$ and k_0 represent contributions by the acid catalyzed and spontaneous hydrolysis mechanisms, respectively,⁹ and HA refers to the dihydrogen phosphate ion (H_2PO_4^-). Values of $k_{\text{H}_2\text{PO}_4^-}$ for hydrolysis of **1** and **2**, obtained from plots of k_{obsd} vs. $[\text{H}_2\text{PO}_4^-]$ for solutions at a given pH, were found to be constant within experimental error over

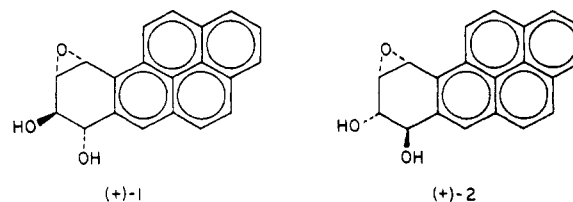


Figure 1. Diastereomeric BP 7,8-diol 9,10-epoxides.³ The enantiomers of each diastereomer shown are those found bound to nucleic acid when BP is applied to mouse skin.⁴

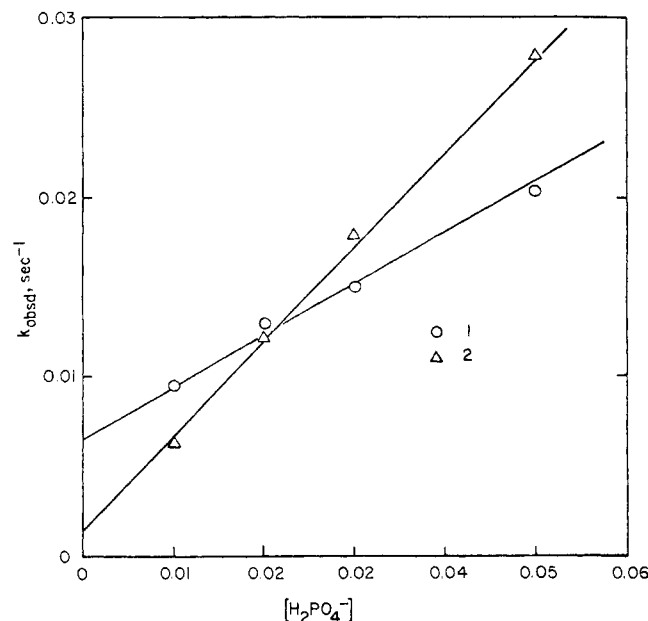


Figure 2. Plots of k_{obsd} vs. $[\text{H}_2\text{PO}_4^-]$ for the hydrolysis of **1** and **2** in 10% dioxane-water solutions at 25 °C, ionic strength 0.2 (NaClO_4), pH 7.02.

Table I. Values of k_{HA} for the General Acid Catalyzed Hydrolysis of **1** and **2** in 10% Dioxane-Water^a at 25 °C^b

HA	pH	$k_{\text{HA}}(\text{1}),$ $\text{M}^{-1} \text{s}^{-1}$	$k_{\text{HA}}(\text{2}),$ $\text{M}^{-1} \text{s}^{-1}$
HOAc	4.81	0.72 ± 0.06	1.32 ± 0.02
	5.11	0.71 ± 0.04	1.33 ± 0.03
	6.34	0.31 ± 0.01	0.48 ± 0.03
	6.70	0.28 ± 0.03	0.49 ± 0.01
	7.02	0.27 ± 0.01	0.48 ± 0.02
H_2PO_4^-	7.10	0.28 ± 0.01	0.54 ± 0.02
	7.60	0.25 ± 0.01	0.49 ± 0.02
	7.94	0.055 ± 0.006	0.026 ± 0.001
	8.54	0.056 ± 0.005	0.020 ± 0.002
	8.68	0.048 ± 0.004	0.0043 ± 0.0003
Tris-H^+	8.94	0.048 ± 0.002	0.0044 ± 0.0004
	9.22	0.19 ± 0.02	0.046 ± 0.003
phenol	9.58	0.17 ± 0.01	0.046 ± 0.002

^a Volume/volume; ionic strength, 0.2 (NaClO_4). ^b Rates were monitored by observing the absorbance change of the reaction solutions at 348 nm in the thermostated cell compartment (25.0 ± 0.1 °C) of a Gilford 2400 spectrophotometer. Rate constants were determined from least-squares plots of k_{obsd} vs. $[\text{HA}]$ for solutions of constant pH but varied buffer concentrations.

the pH range studied (6.3-7.6, Table I). These data indicate that H_2PO_4^- is the only catalytic or reactive phosphate species in the hydrolysis of **1** and **2** under these conditions.

Several kinetically indistinguishable mechanisms might account for the H_2PO_4^- term in eq 1. One possibility is the specific acid-general base mechanism outlined in Scheme I (arbitrarily shown for the reaction of **1**). This mechanism involves attack of hydrogen phosphate ion (HPO_4^{2-}) on the protonated epoxide **3** to yield phosphate ester **4**.