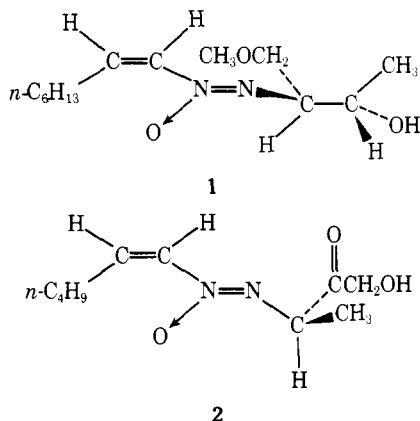


Communications to the Editor

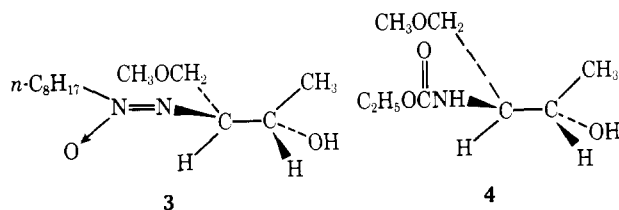
The Synthesis of Elaiomycin, a Naturally Occurring Azoxyalkene

Sir:

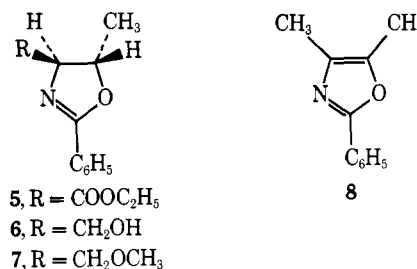
Two of the five naturally occurring azoxy compounds, macrozamin¹ and cycasin,² are glycosides of "methylazoxy-methanol", which has been prepared (as the acetate) from azoxymethane.³ However, syntheses of the mutually related, more complicated, proximal⁴ α,β -(*cis*)-unsaturated azoxyalkenes, elaiomycin (**1**)⁵ and LL-BH872 α (**2**),⁶ require a general synthesis of azoxyalkanes and specific methods for the configurationally controlled introduction of unsaturation.⁷ Despite a recent quickening of interest in azoxyalkanes,^{8,9} routes to **1** or **2** have remained obscure, although the biological significance of **1**¹⁰ (an antibiotic and a carcinogen) and **2**^{6a} (an antifungal agent) makes syntheses highly desirable.¹¹



We are therefore pleased to report the total synthesis of **1** from D-threonine by a three-phase synthetic approach: (A) construction of the distal moiety of **1**, including the azoxy function; (B) elaboration of a proximal *trans*-octenyl group; (C) isomerization to the *cis*-octenyl group. This approach was based on key synthetic methods discovered in our laboratory.¹²⁻¹⁶ A detailed description follows.



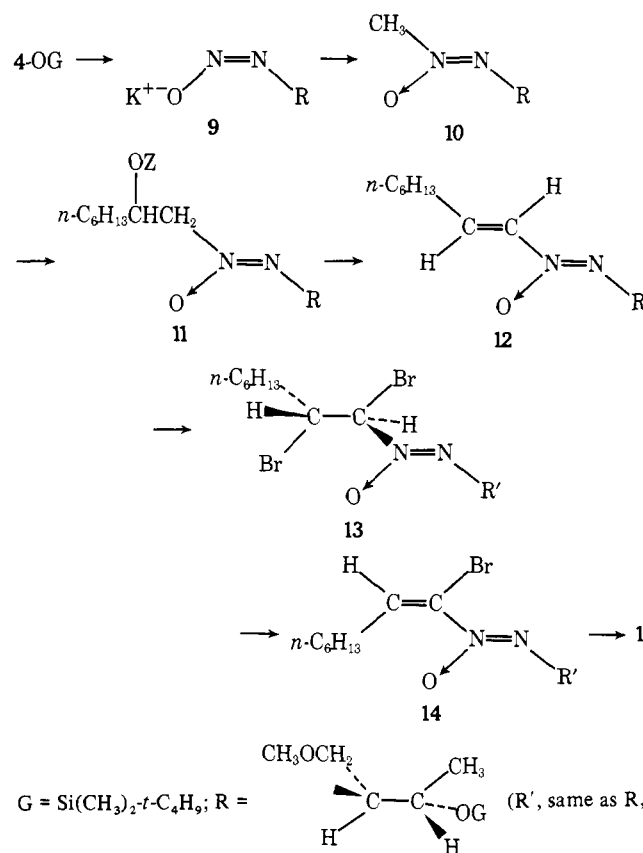
(A) The distal group.¹²⁻¹⁴ As in the synthesis of dihydroelaiomycin, **3**,¹⁴ D-threonine was converted to pivotal urethane **4** by extension of Stevens' method for synthesis of the corresponding amine. The hydrochloride salt of D-threonine ethyl ester was reacted with ethyl benzimidate to yield oxazoline **5** (52%). Reduction of **5** (LiAlH₄, 94%) gave **6**,¹⁷ which was converted (92%) to methyl ether **7** using NaH/CH₃I in THF.¹⁸ Ether **7** was identical with a sample prepared from **6**-OTs and NaOCH₃/CH₃OH,^{5d,14} but the yield was higher in the NaH procedure, and oxazole **8** (a by-product of the methoxide procedure) was not formed. Hydrolysis of **7** (refluxing 6 N HCl, 5 h, then 25 °C, 12 h) gave benzoic acid (96%) and the aqueous amine-hydrochloride, which was neutralized (Na₂CO₃) and converted in situ (ClCOOC₂H₅,



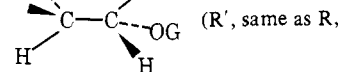
93%) to **4**, identical with a previously prepared sample.^{14,19} Treatment of **4** with *t*-C₄H₉(CH₃)₂SiCl (imidazole, DMF, 25 °C, 17 h)²⁰ quantitatively afforded protected urethane **4**-OG (see Chart 1). Its NMR spectrum resembled that of **4**,¹⁴ but showed δ 0.87 (s, 9 H, *t*-C₄H₉) and 0.03 (s, 6 H, Si(CH₃)₂). Quantitative conversion of **4**-OG to the *N*-nitrosourethane ($\Delta\delta^{\text{CCl}_4}$ OCH₂CH₃ = 0.42)²¹ with ethereal N₂O₄ was followed by cleavage to diazotate **9** using KO_t-C₄H₉ in ether.^{14,21,22} Treatment of an HMPA solution of **9** with excess CH₃I (25 °C, 12 h, 29% based on nitrosourethane) afforded azoxyalkane **10**, which was purified by repetitive TLC²³ (3:1 hexane/ether): NMR δ 4.03 (s, 3 H, CH₃N(O)=N);^{24,25} IR (neat) 1500 cm⁻¹ (azoxy);¹² exact mass ($M^+ - 15$), calcd 261.1633, found 261.1648.

(B) Elaboration of the *trans*-octenyl moiety.¹⁵ Azoxyalkane **10** was converted to its proximal α -carbanion ((*i*-Pr)₂NLi, THF, 0–5 °C, 30 min),¹⁵ which was quenched with excess

Chart 1



G = Si(CH₃)₂-*t*-C₄H₉; R =



but G = H). For conditions and reagents, see text.

n-heptaldehyde (0–5 °C, 1 h) quantitatively affording crude azoxy alcohol **11** (*Z* = H): NMR, δ 4.06 (m, 3 H, $\text{CHN}=\text{N}(\text{O})\text{CH}_2$);^{12,24} IR (neat), 3450 (OH), 1490 (azoxy) cm^{-1} . Without purification, this was converted ($\text{CH}_3\text{SO}_2\text{Cl}$, pyridine, 25 °C, 23 h, 94%) to mesylate **11** (*Z* = SO_2CH_3); NMR δ 2.86 (s, 3 H, CH_3SO_3).²⁴ The crude mesylate, under reflux in toluene containing excess triethylamine (20 h), gave protected *trans*-elaiomycin, **12**, which was purified by repetitive TLC (9:1 hexane/ether). The yield of **12** was 13% from **11** (*Z* = H): NMR δ 6.90 (m, 2 H, vinyl), 4.23 (m, 2 H, distal α -H + SiOCH), 3.58 (m, 2 H, CH_2OCH_3), 3.28 (s, 3 H, OCH_3), 2.25 (m, 2 H, allyl), 1.40 (m, C_5H_{11}) + 1.11 (d, J = 6 Hz, CHCH_3) + 0.91 (s, t - C_4H_9), total \sim 23 H, 0.08 (s, 6 H, $\text{Si}(\text{CH}_3)_2$); IR (neat) 1640 ($\text{C}=\text{C}$), 1460 (azoxy), 950 (*trans*-disubstituted $\text{C}=\text{C}$) cm^{-1} . The spectral properties of **12** coincide with corresponding data for **3**,¹⁴ *trans*- $\text{CH}_3\text{CH}=\text{CHN}(\text{O})=\text{N}-2-\text{C}_8\text{H}_{17}$,¹⁵ and *trans*- $n-\text{C}_6\text{H}_{13}-\text{CH}=\text{CHN}(\text{O})=\text{N}-2-\text{C}_4\text{H}_9$.^{26,27}

(C) Isomerization.¹⁶ Bromine (CCl_4 , 25 °C, 30 min, 100%) added to **12** yielding the corresponding *erythro*-dibromide, whence deprotection²⁰ ($\text{CH}_3\text{COOH}:\text{H}_2\text{O}:\text{THF}$, 3:1:1, 25 °C, 18 h, 95%) gave *erythro*-dibromoelaiomycin, **13**, which was purified by repetitive TLC (3:1 hexane/ether): NMR, δ 5.96 (d, J = 11 Hz, 1 H, proximal α -H), 4.65 (m, 1 H, proximal β -H), 4.11 (m, 2 H, distal α -H + HOCH), 3.58 (m, 2 H, CH_2OCH_3), 3.28 (s, 3 H, OCH_3), 2.21 (br s, 1 H, OH), 1.71–0.65 (m, residual alkyl); IR (neat), 3450 (OH), 1495 (azoxy) cm^{-1} .²⁷ For comparison, the α - and β -proximal protons of *erythro*- $\text{CH}_3\text{CHBrCHBrN}(\text{O})=\text{N}-2-\text{C}_8\text{H}_{17}$ appear at δ 5.85 (d, J = 11 Hz) and 4.73 (m); its distal α -H appears at δ 4.00 (m).¹⁶

Anti elimination of HBr from **13** (DBU, 25 °C, 30 min, 75%)²⁸ gave crude α -bromoelaiomycin, **14**: NMR (CCl_4 , Me_4Si), δ 5.92 (t, J = 8 Hz, 1 H, vinyl);²⁴ IR (neat), 3400 (OH), 1620 ($\text{C}=\text{C}$), 1460 (azoxy) cm^{-1} . For comparison, the vinyl proton of *E*- $\text{CH}_3\text{CH}=\text{CBrN}(\text{O})=\text{N}-2-\text{C}_8\text{H}_{17}$ appears at δ 5.96 (q, J = 7.5 Hz).¹⁶ Crude **14** was debrominated with powdered zinc (Mallinkrodt AR grade ether,²⁹ containing 4 vol % of 30 wt % aqueous CH_3COOH , 25 °C, 24 h, 52%); repetitive TLC (3:1 hexane/ether) afforded elaiomycin, **1**, as well as unreacted **14**.³⁰

Synthetic **1** contained a trace of carbonyl impurity (1740 cm^{-1}), but its IR spectrum was otherwise identical with the published spectrum^{5a} of natural **1**, including bands at 3450 (OH), 1650 ($\text{C}=\text{C}$), 1455 (azoxy), and 785 (*cis* disubstituted $\text{C}=\text{C}$?) cm^{-1} . The UV spectrum gave $\lambda_{\text{max}}^{\text{CH}_3\text{OH}}$ 235, ϵ 1.0 $\times 10^4$ (lit.^{5a,b} 237.5, 1.1×10^4). The NMR spectrum (CCl_4 , Me_4Si) was persuasive: δ 6.83 ("d", J \sim 9 Hz, 1 H, proximal α -H),³¹ 5.70 (q, J \sim 9 Hz, 1 H, proximal β -H), 4.17 (m, 2 H, distal α -H + CHOH), 3.58 (m, 2 H, CH_2OCH_3), 3.33 (s, 3 H, OCH_3), 2.70 (m, 2 H, allyl), 2.13 (m, 1 H, OH), 1.78–0.60 (m, residual alkyl). Both natural **1** and **2** exhibit vinyl doublets, J = 9 Hz, at δ 6.83,^{6a} and **2** exhibits a quartet at δ 5.83, J = 9 Hz.^{6a} In *cis*- $\text{CH}_3\text{CH}=\text{CHN}(\text{O})=\text{N}-2-\text{C}_8\text{H}_{17}$, the corresponding vinyl signals appear at δ 6.70 ("d", J = 9 Hz) and 5.73 (quintet, J = 8 Hz).¹⁶ Other NMR signals of synthetic **1** are in accord with structural expectation.^{6a,14,32}

Reduction of synthetic **1** (5% Rh/ Al_2O_3 , 1 atm of H_2 , CH_3OH , 1 h) gave dihydroelaiomycin, identical in NMR spectrum¹⁴ and TLC behavior with an authentic sample produced via alkylation of **9** (*G* = tetrahydropyranyl) with *n*- $\text{C}_8\text{H}_{17}\text{I}$.¹⁴

Synthetic **1** had $[\alpha]_D^{24} +24.0^\circ$ (*c* 2.8, ethanol), 62.5% of the rotation of natural **1**.^{5a} It is possible that the apparent loss of optical activity is due to the presence of a trace of highly levorotatory impurity in the synthetic **1**.³¹ Alternatively, a dextrorotatory impurity may have been present in natural **1**.^{5a,33}

The overall yield for the 18-step conversion of D-threonine

to **1** was only 0.55%, but we have not optimized the key low-yield steps **9** \rightarrow **10** and **11**-OMs \rightarrow **12**, so that an enhanced yield should be attainable. This initial synthesis of elaiomycin employs strategies which are applicable to **2** and synthetic analogues. Moreover, the crucial sequences substantially broaden the scope of azoxyalkane chemistry.³⁴

Acknowledgments. We thank the Public Health Service (Grant CA-14912 from the National Cancer Institute) and the National Science Foundation for financial support. Helpful discussions with Professors P. F. Hudrlik and R. R. Ruden were much appreciated.

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- (23) All preparative TLC used EM Laboratories silica gel 60 F-254, 2.0-mm plates.
- (24) Characteristic bands are cited for most spectra; other spectral features were in accord with structure. Citations of NMR multiplet positions refer to the multiplet's center or most prominent feature. The NMR solvent was CCl_4 , CHCl_3 , unless otherwise noted.
- (25) The corresponding resonance of $2-\text{C}_8\text{H}_{17}\text{N}=\text{N}(\text{O})\text{CH}_3$ appears at δ 3.94.¹⁵ Despite TLC homogeneity, **10** was not pure; singlets appeared at δ 3.32 and 3.00, perhaps due to the isomeric *N*-methyl-*N*-nitrosoamine.
- (26) R. A. Moss and R. C. Nahas, unpublished work.
- (27) Despite TLC homogeneity, two separately prepared and purified samples gave C, H microanalyses which were \sim 1% high in C; residual traces of hexane acquired during extensive TLC may have been responsible. All intermediates after **9** were oils, and difficult to purify, but their spectra leave little structural uncertainty.
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- (29) The ether must contain a radical inhibitor.¹⁶
- (30) The reaction was stopped prior to completion to avoid overreduction of **1**.
- (31) A trace of *trans*-**1** (<5%) may be present as evidenced by a minor absorption at δ 6.90; cf. 12.

- (32) The NMR spectrum of synthetic **1** was identical with the spectrum of an authentic (natural) sample. We thank Dr. W. J. McGahren for the comparison spectrum.
- (33) More involved explanations are possible. For example, epimerization might have occurred at the distal α -carbon during the conversion of **10** to its proximal α -carbanion with $(i\text{-Pr})_2\text{NLi}$. However, we have shown that similar reactions with optically active 2-octyl-*NNO*-azoxymethane *do not* result in significant racemization.¹⁵ Moreover, as pointed out by a referee, epimerization at the distal α -carbon (epimerization at the hydroxyl-bearing, distal β -carbon is unlikely) would afford a mixture of diastereomers. If such epimerization occurred at the most sensitive step (**10** \rightarrow **11** requires the most strongly basic conditions, see above), then a mixture of (*S,S*)-**11** and (*2S,3R*)-**11** would have been generated. We feel that it is unlikely that the (*2S,3R*) diastereomer would have survived the repetitive TLC purifications applied to **12**, **13**, and synthetic **1**.
- (34) This report is Alkane Diazotates, **24**; for part **23**, see ref 16.
- (35) Fellow of the A. P. Sloan Foundation.
- (36) Postdoctoral Fellow on leave from Sumitomo Chemical Co.

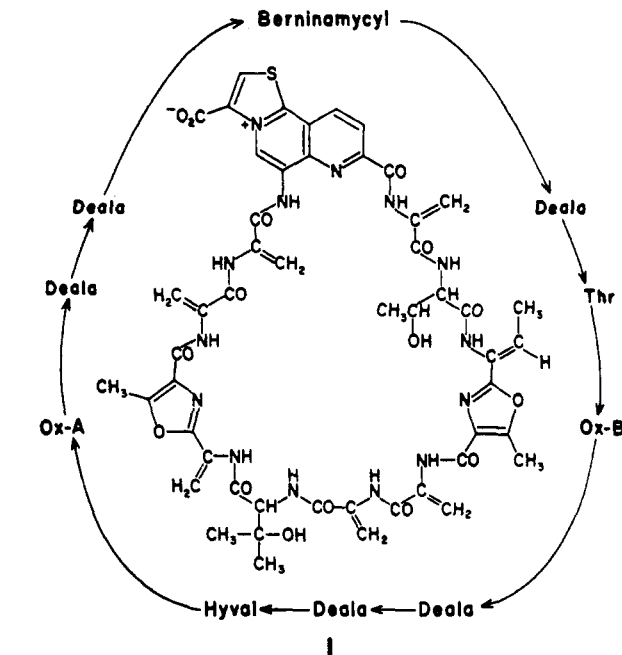
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 Received August 24, 1976

Berninamycin. 3. Total Structure of Berninamycin A^{1,2}

Sir:

In earlier reports¹⁻³ from this laboratory we have described the results of initial structural studies on the novel, sulfur-containing antibiotic berninamycin A, which is a potent inhibitor of bacterial protein synthesis. Degradation products obtained from acidic hydrolysis, methanolysis, and acetolysis of berninamycin A allowed the assignment of the structural subunits shown in the top row of Figure 1,² which account for the total composition of the antibiotic. In the present communication, we assign the total structure of berninamycin A as **1**, based upon new compounds obtained by trifluoroacetylation of the intact antibiotic and its sodium borohydride-reduced and catalytically hydrogenated derivatives.

Treatment of berninamycin A with trifluoroacetic acid at room temperature for 18 h afforded three major compounds (Figure 2). The least polar compound was identified as the previously reported **2**.² A second compound (mp 109–110 °C; C₁₅H₂₀N₄O₆)^{4a} was assigned structure **3**. As previously discussed,² the residues (Deala, Thr, Hyval, Ox-A, Ox-B, Berninamycyl) which comprise berninamycin A have unique ¹H NMR resonances which allow their identification in degra-



dation products formed from the intact antibiotic. The ¹H NMR spectrum of **3** contains the resonances assignable² to the Hyval (1.40 ppm, s, 3 H; 1.50, s, 3 H; 5.49, d, 7 Hz, 1 H) and Ox-A (2.63, s, 3 H; 2.04, s, 3 H) residues and to a pyruvyl unit (2.42 ppm, s, 3 H).

The pyruvyl residue (which results from cleavage of a Deala residue)² can only occupy the N-terminal position, and a structure including the sequence Ox-A \rightarrow Hyval is eliminated by subunit a of Figure 1. Thus, the expected structure for the second trifluoroacetolysis product would be pyruvyl \rightarrow Hyval \rightarrow Ox-A \rightarrow NH₂ (**4**), a structural isomer of **3**. The 1,3-tetrahydrooxazine ring of **3** results from intramolecular addition of the hydroxyl group of Hyval to the enamine of Ox-A in **4** during trifluoroacetolysis. Combination of the sequence of **4** with subunit a allows the assignment of c (Figure 1) as a sequence in the intact antibiotic.

The most polar compound from trifluoroacetolysis of **1** is assigned structure **5** (mp 153 °C dec; C₂₇H₂₆N₈O₈S).^{4a} The ¹H NMR spectrum of **5** has resonances assignable² to Thr, Ox-B, Deala, and Berninamycyl (Figure 1). These residues,

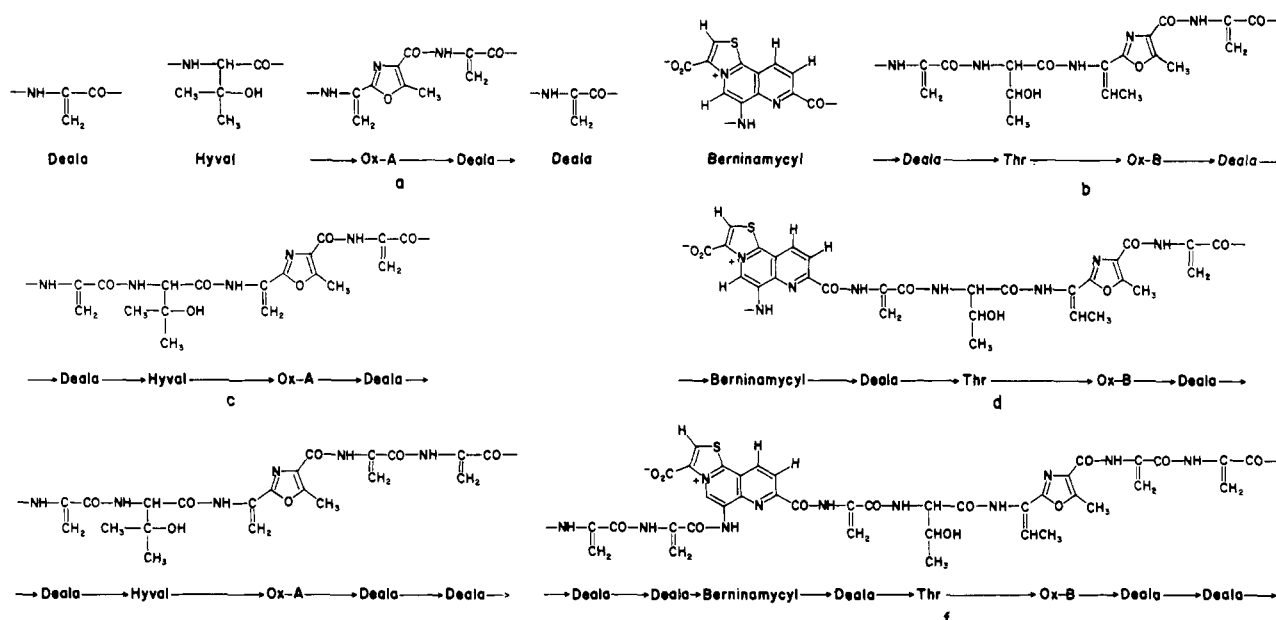


Figure 1. Subunit sequences found in berninamycin A. Subunits shown in the top line were established earlier.²