

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 14 (2004) 5937-5941

1,2,3,4-Tetrahydroquinoline-containing $\alpha_V \beta_3$ integrin antagonists with enhanced oral bioavailability

Shyamali Ghosh,^a Rosemary J. Santulli,^a William A. Kinney,^{a,*} Bart L. DeCorte,^a Li Liu,^a Joan M. Lewis,^a Jef C. Proost,^b Gregory C. Leo,^a John Masucci,^a William E. Hageman,^a Andrew S. Thompson,^c Ian Chen,^c Reiko Kawahama,^c Robert W. Tuman,^a Robert A. Galemmo, Jr.,^a Dana L. Johnson,^a Bruce P. Damiano^a and Bruce E. Maryanoff^a

^aDrug Discovery, Johnson & Johnson Pharmaceutical Research & Development, L.L.C., Welsh & McKean Roads, Spring House, PA 19477-0776, USA

^bDrug Discovery, Johnson & Johnson Pharmaceutical Research & Development, L.L.C., 2340 Beerse, Belgium ^cJ-Star Research, Inc., 3001 Hadley Road, Suite 3, South Plainfield, NJ 07080, USA

> Received 27 May 2004: accepted 31 August 2004 Available online 7 October 2004

Abstract—Reduction of the quinoline ring in an $\alpha_{v}\beta_{3}$ antagonist yielded a 1,2,3,4-tetrahydro derivative as two diastereomers, the four isomers of which were separated by sequential chiral HPLC. Two isomers had significant $\alpha_V \beta_3$ antagonist activity with improved oral bioavailability, relative to the corresponding quinoline derivative. © 2004 Elsevier Ltd. All rights reserved.

Interactions between the $\alpha_V \beta_3$ integrin and its extracellular matrix ligands are a basis of cell adhesion events, leading to the migration, invasion, proliferation, and survival of cells. The $\alpha_V \beta_3$ integrin is expressed in several cell types, such as osteoclasts, endothelial cells, vascular smooth muscle cells, and some tumor cells. Although the normal level of expression is typically low, expression is greatly enhanced in pathological conditions involving tissue remodeling and growth. Hence, this integrin has attracted attention as a therapeutic target for the treatment of cancer, osteoporosis, rheumatoid arthritis, and diabetic retinopathy.¹ Monoclonal antibodies to $\alpha_V \beta_3$ (Vitaxin, CNTO 95) and peptide antagonists based on the Arg-Gly-Asp (RGD) motif have been the most studied to date and have shown good efficacy in various animal models.^{1,2} Interesting efficacy for orally bioavailable, small-molecule $\alpha_V \beta_3$ integrin antagonists3 in models of osteoporosis4 and arthritis5 has been reported.

Starting from the nipecotamide-based $\alpha_{IIb}\beta_3$ antagonist, elarofiban (1, Fig. 1),⁶ we identified the isonipecotamide structure 2 ($\alpha_V \beta_3$ IC₅₀ = 3.6 nM, $\alpha_{IIb} \beta_3$ IC₅₀ = 170 nM)



Figure 1. Evolution of series from nipecotamide (1) to isonipecotamides (2 and 3) to carba-analog (4).

with reversed selectivity for $\alpha_V \beta_3$ relative to $\alpha_{IIb} \beta_3$.^{7,8} This compound represented a good starting point, but it was deficient in showing only moderate selectivity and low oral bioavailability (F = 4%, Table 1). In this case, antagonism of the integrin $\alpha_{IIb}\beta_3$ would be an undesirable activity as it causes blockade of platelet aggregation. Recently, we reported⁸ the introduction of the tetrahydronaphthyridine group⁹ in 3 and the

^{*}Corresponding author. Tel.: +1 215 628 5908; fax: +1 215 628 4985; e-mail: wkinney1@prdus.jnj.com

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter © 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2004.08.067

	Table 1.	Binding	studies at	: α _V β ₃ ,	$\alpha_V \beta_5$	and	$\alpha_{\text{IIb}}\beta_3$	integrins	by	quinoline	based	derivatives	(IC_{50})	1
--	----------	---------	------------	-----------------------------------	--------------------	-----	------------------------------	-----------	----	-----------	-------	-------------	-------------	---

N (CH ₂)m		H ₂) _n OH Ring O
H	ö	

No.	Isomer designation $([\alpha]_D^{23})$	Structural description			$\alpha_V\beta_3 \; (nM)^b$	$\alpha_V\beta_5 \; (nM)^b$	$\alpha_{IIb}\beta_3 \ (nM)^c$	F ^d (%)	
			п	Ring					
2	S			See figure	3.6 ± 0.9	470 ± 110	170 ± 30	4	
3	S			See figure	2.6 ± 0.6	270 ± 50	2900		
27	<i>R</i> , <i>S</i>	1	1	3-Quinoline	0.8 ± 0.2	2.2 ± 0.7	1100	4	
28	<i>R</i> , <i>S</i>	1	1	3-(5,6,7,8-THQ) ^e	1.6 ± 0.7	11 ± 4	3000	0	
29	f	1	1	3-(1,2,3,4-THQ)	2.5 ± 0.4	17 ± 2	14,000		
29a	a (+16°)	1	1	3-(1,2,3,4-THQ)	26 ± 8	1000	>50,000		
29d	d (-15°)	1	1	3-(1,2,3,4-THQ)	2.7 ± 0.7	8.7 ± 2.1	4700	22	
29b	b (+24°)	1	1	3-(1,2,3,4-THQ)	1.2 ± 0.1	7.8 ± 1.4	13,000	8	
29c	c (-25°)	1	1	3-(1,2,3,4-THQ)	52 ± 14	2200	>50,000		
33	f	1	1	3-(1-Me-1,2,3,4-THQ)	49 ± 2	180 ± 20	>50,000		
34	<i>R</i> , <i>S</i>	1	1	3-(6-MeOPy) ^g	0.7 ± 0.2	2.7 ± 0.8	6100	8	
34a	$a (-22^{\circ})^{h}$	1	1	3-(6-MeOPy)	0.4 ± 0.1	3.1 ± 1.5	1800	2	
34b	b (+22°) ^h	1	1	3-(6-MeOPy)	16 ± 6	98 ± 39	>50,000		
30	<i>R</i> , <i>S</i>	2	0	3-Quinoline	0.5 ± 0.1	36 ± 9	870	1	
31	f	2	0	3-(1,2,3,4-THQ)	3.7 ± 1.5	260 ± 60	15,000	100	
31a	a (+20°) ⁱ	2	0	3-(1,2,3,4-THQ)	1.6 ± 0.5	44 ± 12	13,000	46	
31d	d (-20°) ^j	2	0	3-(1,2,3,4-THQ)	5.4 ± 0.2	160 ± 60	17,000	52	
31b	b (+17°) ⁱ	2	0	3-(1,2,3,4-THQ)	70 ± 7	2500	>50,000		
31c	c (-26°) ^j	2	0	3-(1,2,3,4-THQ)	100 ± 40	1500	>50,000		
SB 265123 ^k					4.0 ± 1.3	2.9 ± 0.5	1200	100 ^k	

^a 29a and 29d, 29b and 29c, 34a and 34b, 31a and 31d, 31b and 31c are enantiomeric pairs.

^b Inhibition of human vitronectin binding to immobilized $\alpha_V \beta_3$ and $\alpha_V \beta_5$ ($N \ge 3$).¹

^c Inhibition of biotinylated fibrinogen binding to immobilized $\alpha_{IIb}\beta_3$ ($N \ge 2$).¹⁶

^d See Ref. 18.

^e THQ = tetrahydroquinoline.

^f Mixture of two diastereomers (four isomers).

^g MeOPy = methoxypyridine.

^h Separated by chiral HPLC on Chiralcel[®] OJ (96/4hexane/EtOH elution) at same Boc-protected intermediate stage as 15.

ⁱ Prepared from isomers **18a** ($[\alpha]_D^{23}$ +43) and **18b** ($[\alpha]_D^{23}$ +53), which were separated from **18c/18d** on a Chiralpak[®] OD column (MeOH elution), and from each other on Chiralpak[®] AD column (EtOH elution) as described above.¹³ ^j Prepared from isomers **18c** ($[\alpha]_D^{23}$ -62) and **18d** ($[\alpha]_D^{23}$ -46), which were separated on the initial Chiralpak[®] OD column from the mixture of **18a/18b**.

^k See Ref. 4a

'evolved' carba-analogs (e.g., 4) that lack the amide group beta to the carboxylic acid. These derivatives had improved selectivity relative to $\alpha_{IIb}\beta_3$ and in the case of some β -aryl substituted carba-analogs, such as 4, significant improvements in oral bioavailability were observed. However, no β -heterocyclic groups were found to exhibit good oral bioavailability (e.g., pyridyl) initially. This paper focuses on quinoline-containing structures of general formula 5. The structure was modified by varying the spacer units and taking advantage of the selective reduction of the quinoline ring that occurred during hydrogenation.



The syntheses of targets 5 are described in Scheme 1. A strategy was required that would allow variation of the spacer between the piperidine and the quinoline substituent so that the optimum arrangement could be determined.¹⁰ Boc-protected piperidine carboxylic acids 6 and 7 were converted to their corresponding Weinreb amides 8 and 9, which were reacted with the lithium reagent derived from 3-bromoquinoline to yield the ketones 10 and 11. The latter reaction was best performed by premixing the Weinreb amide and aryl bromide in THF, followed by the addition of butyl lithium.¹¹ The resultant ketones were elaborated to the unsaturated esters 12 and 13 by reaction with the sodium salt of trimethyl phosphonoacetate. The mixture of E- and Z-isomers of 12 was hydrogenated in the presence of 10% palladium on carbon to give three different products, depending on the reduction conditions. One could produce quinoline 14 by conducting the hydrogenation at low pressure (8 psig). Over-reduced products 15 and 16 resulted from using intermediate pressure (30 psig) for two days. The optimal conditions for producing the 1,2,3,4-tetrahydroquinoline 15 were to hydrogenate in toluene with triethylamine present at higher pressure and temperature.¹² The stereoisomers of 15 (four isomers) were separated at this stage by chiral HPLC.¹³ The isomer lettering order is based on the elution order



Scheme 1. (a) MeONHMeHCl, NMM (6molequiv), HOBt (0.5 mol equiv), HBTU (1.2 molequiv), MeCN, 6–8 h, 85–95%. (b) 3-Bromoquinoline (2molequiv), THF, BuLi (2molequiv), $-90 \,^{\circ}$ C (10 or 11, 62%). (c) (MeO)₂P(O)CH₂–CO₂Me (4molequiv), NaHMDS (3 mol equiv), THF, $-50 \,^{\circ}$ C, reflux after ketone addn., 4h (12, 100%; 13, 65%). (d) Equal wt of 10% Pd/C, MeOH, H₂, at 0–8 psig for 24h (14, 50%; 17, 68%); at 30 psig for 48h (15, 37%, 16, 28%; 18, 49%, 19, 18%); or half wt of 10% Pd/C, toluene, Et₃N, 50 $\,^{\circ}$ C, 50 psig, 28h (15, 73%, 16, 3%). (e) 2N HCl in dioxane, anisole (cat.), 2h, 100%. (f) For 20–23, NMM (5molequiv), HOBT (0.5molequiv), HBTU (1.2molequiv), MeCN, DMF, 8–12h (87–93%); for 24, PyBroP, DMF, CH₂Cl₂, *i*-Pr₂NEt, 0 $\,^{\circ}$ C, 1.5h (80–94%). (g) 3N NaOH, MeOH, 24h (79–97%) or 4N HCl, 48h (75–95%).

(isomer-a is faster eluting) of intermediate 15. The related intermediates 17–19 were prepared similarly from 13. Intermediates 14–19 were converted to targets 5 by similar methods. The Boc-group was removed with HCl to give the piperidines, which were individually coupled with tetrahydronaphthyridine acids 25^{14} or $26.^{10}$ Conditions using *O*-benzotriazol-1-yl-*N*,*N*,*N'*,*N'*-tetramethyluronium hexafluorophosphate (HBTU), 1-hydroxybenzotriazole (HOBT), and *N*-methylmorphol-

ine (NMM) were effective with **20–23**. For **24**, the usual coupling conditions were unacceptable because of oxidation to the quinoline. The coupling reagent, bromotrispyrrolidinophosphonium hexafluorophosphate (PyBroP), was found to be suitable, without accompanying oxidation. Hydrolysis of the methyl ester was performed under acidic or basic conditions to afford targets of type **5** (**27–31**, Table 1).

The N–Me derivative **33** was produced by alkylation of the intermediate 1,2,3,4-tetrahydroquinoline **15** (2,6-di-(*t*-butyl)pyridine (1.2 molequiv), MeI (1.2 molequiv), DMF, 20h, 31%) to give the *N*-methyl intermediate **32**, which was converted to **33** as described above. Compound **34** (Table 1) was prepared by the same method as described in Scheme 1, beginning with 5-bromo-2-methoxypyridine. The isonipecotamide structures (**2**, **3**) were prepared as described earlier.⁸ Full experimental methods for the preparation and chiral HPLC separation of a related series are discussed elsewhere.¹⁵

The target compounds were evaluated as antagonists of vitronectin binding to $\alpha_V \beta_3$ and $\alpha_V \beta_5$, and fibrinogen binding to $\alpha_{IIb}\beta_3$ (Table 1). SB 265123, included as a reference compound, is reported to be a potent antagonist of both α_V integrins and to be selective relative to α_{IIb} (300-fold).^{4a} The original quinoline-substituted isonipecotamide lead 2 was a potent antagonist of $\alpha_V \beta_3$, but was weakly active against $\alpha_V \beta_5$ and not selective relative to $\alpha_{IIb}\beta_3$. It was thought that dual antagonism of $\alpha_V\beta_3$ and $\alpha_V \beta_5$ might be an advantage in certain disease states.² Replacement of the cyclic guanidine group with the tetrahydronaphthyridine group in 3 led to a great improvement in selectivity, which was first observed by Coleman et al.⁹ and noted by us in other series.^{15,17} When the amide of 3 was replaced by a carbon linker in 27, good dual potency at $\alpha_V \beta_3$ and $\alpha_V \beta_5$ was achieved, along with excellent selectivity (1400-fold). Compound 27 was compared to 2 in a rat pharmacokinetics (PK) study,¹⁸ but there was no improvement in oral bioavailability for **27** (F = 4%, $C_{\text{max}} = 0.5 \,\mu\text{M}$, $t_{1/2} = 3 \,\text{h}$) versus **2** (F = 4%, $C_{\text{max}} = 0.8 \,\mu\text{M}$, $t_{1/2} = 7 \,\text{h}$, dosed 30 mg/kg po). This result was surprising since a related $\alpha_V \beta_3$ antagonist containing the tetrahydronaphthyridine and quinoline groups was orally bioavailable in dogs.⁹ Analogs of 27 with the quinoline reduced to 5,6,7,8- and 1,2,3,4-tetrahydroquinolines (THQ), 28 and 29, were obtained as byproducts. Compounds 28 and 29 have potency and selectivity comparable to 27, so it was of interest to test their PK properties. Although 5,6,7,8-THQ 28 was not detectable following oral administration, 1,2,3,4-THQ 29 was observed at high levels in the plasma in a pilot rat PK experiment (21µM at 2h; 15mg/kg, po). Therefore, the four isomers of **29** (two *dl* pairs) were prepared and evaluated. Two of the isomers, 29b and 29d had good potency in binding assays, so they were examined in rat PK experiments.¹⁹ A dramatic improvement in oral absorption was noted with 29d (F = 22%, $C_{\text{max}} = 3 \,\mu\text{M}, t_{1/2} = 2 \,\text{h}$) versus quinoline 27. Although the quinolinyl group has been used extensively in integrin antagonists, the THQ-group has not yet been described. The N-Me analog 33 showed decreased potency relative to 29.

It was of interest to compare the 2-methoxypyridine analog **34**, as this group has displayed good PK properties.^{4b} In our series, neither the racemate **34** (F = 8%, $C_{\text{max}} = 0.9 \,\mu\text{M}$, $t_{1/2} = \text{ND}$) nor the active isomer **34a** (F = 2%, $C_{\text{max}} = 0.1 \,\mu\text{M}$, $t_{1/2} = 2$ h) had the oral bioavailability seen for **29d**. However, the $\alpha_V\beta_3$ potency of **34a** was noteworthy.

The quinoline and THQ compounds **30** and **31** (m = 2, n = 0, Table 1), which lack a carbon atom between the piperidine and β -position of the carboxylic acid, were evaluated; they have one additional linker carbon attached to the tetrahydronaphthyridine group. Racemic **30** and **31** had good potency against $\alpha_V\beta_3$, but lesser potency against $\alpha_V\beta_5$ as compared to **27** and **29**. The most active THQ isomer, **31a**, was 30-fold more potent against $\alpha_V\beta_3$ than $\alpha_V\beta_5$. Both of the active THQ isomers, **31a** (F = 46%, $C_{max} = 8\,\mu$ M, $t_{1/2} = 2h$) and **31d** (F = 52%, $C_{max} = 58\,\mu$ M, $t_{1/2} = 1h$), showed even better oral bioavailability than what was seen above. The corresponding quinoline, **30**, was not well absorbed (F = 1%).

The relationship of stereochemistry to biological activity is different in the THQ analogs 29 and 31. The enantiomeric pairing of isomers was determined by measuring optical rotations of the intermediates (15, 18) and final products (29, 31; Table 1). The absolute values of the four optical rotations were not that different for the four isomers of 31, making it more difficult to pair the finalproduct isomers. For this reason, ¹H and ¹³C NMR studies were done (D₂O, 600 MHz) on the four isomers of 31. The spectra of 31a and 31d were identical to each other and distinct from the spectra of **31b** and **31c**; the spectra of 31b and 31c were also identical. From this analysis, the two more potent isomers of **29** (**b** and **d**) have a diastereomeric relationship, whereas the two more potent isomers of 31 (a and d) are enantiomers of each other. Therefore, the absolute stereochemistry at the β -position is different for the more potent isomers of 31, which is not commonly the case with aryl substituents.

Conclusion

The quinoline containing carba-analog **27** was modified by hydrogenation, which improved its oral bioavailability. Conditions were developed to selectively reduce the quinoline to the 1,2,3,4-tetrahydroquinoline unit. These 1,2,3,4-THQ-containing α_V antagonists had good potency and the highest oral bioavailability in this series thus far. The four individual isomers of the THQ-analogs **29** and **31** were obtained by using chiral HPLC. Biological activity resided primarily in two of the isomers in each case.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j. bmcl.2004.08.067.

References and notes

- (a) Mousa, S. A. Med. Res. Rev. 2003, 23, 190; (b) Tucker, G. C. Curr. Opin. Invest. Drugs 2003, 4, 722.
- Trikha, M.; Zhou, Z.; Nemeth, J. A.; Chen, Q.; Sharp, C.; Emmell, E.; Giles-Komar, J.; Nakada, M. T. *Int. J. Cancer* 2004, *110*, 326–335.
- 3. Miller, W. H.; Keenan, R. M.; Willette, R. N.; Lark, M. W. Drug Discovery Today 2000, 5, 397.
- 4. (a) Miller, W. H.; Bondinell, W. E.; Cousins, R. D.; Erhard, K. F.; Jakas, D. R.; Keenan, R. M.; Ku, T. W.; Newlander, K. A.; Ross, S. T.; Haltiwanger, R. C.; Bradbeer, J.; Drake, F. H.; Gowen, M.; Hoffman, S. J.; Hwang, S.-M.; James, I. E.; Lark, M. W.; Lechowska, B.; Rieman, D. J.; Stroup, G. B.; Vasko-Moser, J. A.; Zembryki, D. L.; Azzarano, L. M.; Adams, P. C.; Salyers, K. L.; Smith, B. R.; Ward, K. W.; Johanson, K. O.; Huffman, W. F. Bioorg. Med. Chem. Lett. 1999, 9, 1807; (b) Hutchinson, J. H.; Halczenko, W.; Brashear, K. M.; Breslin, M. J.; Coleman, P. J.; Duong, L. T.; Fernandez-Metzler, C.; Gentile, M. A.; Fisher, J. E.; Hartman, G. D.; Huff, J. R.; Kimmel, D. B.; Leu, C.-T.; Meissner, R. S.; Merkle, K.; Nagy, R.; Pennypacker, B.; Perkins, J. J.; Prueksaritanont, T.; Rodan, G. A.; Varga, S. L.; Wesolowski, G. A.; Zartman, A. E.; Rodan, S. B.; Duggan, M. E. J. Med. Chem. 2003, 46, 4790.
- Badger, A. M.; Blake, S.; Kapadia, R.; Sarkar, S.; Levin, J.; Swift, B. A.; Hoffman, S. J.; Stroup, G. B.; Miller, W. H.; Gowen, M.; Lark, M. W. Arthritis Rheum. 2001, 44, 128.
- Hoekstra, W. J.; Maryanoff, B. E.; Damiano, B. P.; Andrade-Gordon, P.; Cohen, J. H.; Costanzo, M. J.; Haertlein, B. J.; Hecker, L. R.; Hulshizer, B. L.; Kaufman, J. A.; Keane, P.; McComsey, D. F.; Mitchell, J. A.; Scott, L.; Shah, R. D.; Yabut, S. C. J. Med. Chem. 1999, 42, 5254.
- Others have shown that selectivity can be reversed by utilizing similar templates: Duggan, M. E.; Duong, L. T.; Fisher, J. E.; Hamill, T. G.; Hoffman, W. F.; Huff, J. R.; Ihle, N. C.; Leu, C.-T.; Nagy, R. M.; Perkins, J. J.; Rodan, S. B.; Wesolowski, G.; Whitman, D. B.; Zartman, A. E.; Rodan, G. A.; Hartman, G. D. J. Med. Chem. 2000, 43, 3736.
- DeCorte, B. L.; Kinney, W. A.; Liu, L.; Ghosh, S.; Brunner, L.; Hoekstra, W. J.; Santulli, R. J.; Tuman, R. W.; Baker, J.; Burns, C.; Proost, J. C.; Tounge, B. A.; Damiano, B. P.; Maryanoff, B. E.; Johnson, D. L.; Galemmo, R. A., Jr. *Bioorg. Med. Chem. Lett.* 2004, 14, 5227.
- Coleman, P. J.; Askew, B. C.; Hutchinson, J. H.; Whitman, D. B.; Perkins, J. J.; Hartman, G. D.; Rodan, G. A.; Leu, C.-T.; Prueksaritanont, T.; Fernandez-Metzler, C.; Merkle, K. M.; Lynch, R.; Lynch, J. J.; Rodan, S. B.; Duggan, M. E. *Bioorg. Med. Chem. Lett.* 2002, *12*, 2463.
- Procedures were adapted from those described in: Askew,
 B. C.; Coleman, P. J.; Duggan, M. E.; Halczenko, W.; Hartman, G. D.; Hunt, C. A.; Hutchinson, J. H.; Meissner, R. S.; Patane, M. A.; Smith, G. R.; Wang, J. U. S. Patent 6,048,861, 2000; *Chem. Abstr.* 1999, 404930.
- 11. To a rapidly stirred, cold ($-90 \,^{\circ}$ C) solution of the Weinreb amide **8** (51.2 g, 179 mmol) and 3-bromo-quinoline (74.7 g, 358 mmol) in THF (205 mL) was added a 1.6 M solution BuLi in hexanes (225 mL, 358 mmol), maintaining the internal temperature $\leq -84 \,^{\circ}$ C. The mixture was stirred at $-90 \,^{\circ}$ C for 1.5 h and the cold bath was removed. The mixture was stirred for 1 h and placed in an ice/water bath until the internal temperature rose to $-6 \,^{\circ}$ C. Saturated aqueous NH₄Cl (500 mL) was added at such a rated that the internal temperature was $\leq 10 \,^{\circ}$ C and the resulting

mixture was stirred under nitrogen for 18h. The aqueous phase was extracted with EtOAc and the organic extracts were combined, dried over Na₂SO₄, filtered, and evaporated to give 105g of a dark oil, which was purified by column chromatography (1.7kg of flash grade silica gel; eluant: toluene 3L, 15% EtOAc in toluene 14L, 22% EtOAc in toluene 9L, 28% EtOAc in toluene 7L and finally 60% EtOAc in toluene 1.8L). Evaporation of appropriate fractions gave 40.57 g of a solid, which was triturated with 12:1 hexane/EtOAc (130 mL) and allowed to stand for 18h. The resulting solid was collected by filtration and washed with portions of hexane to give 37.8 g of the product as a yellow solid. The mother liquors were concentrated, treated with hexane, EtOAc, and acetone and filtered to give 2.06g of the product as a yellow solid. The two batches were combined to give 39.8 g of 10 (62% yield, mp 114-116°C). ¹H NMR (CDCl₃, 300 MHz): δ 1.17–1.34 (m, 2H), 1.46 (s, 9H), 1.74–1.84 (m, 2H), 2.16-2.33 (m, 1H), 2.69-2.84 (m, 2H), 3.04 (d, J = 7.6 Hz, 2H), 4.03–4.23 (m, 2H), 7.61–7.68 (m, 1H), 7.83–7.89 (m, 1H), 7.97 (dd, J = 1.5, 9.1 Hz, 1H), 8.16 (d, J = 9.1 Hz, 1 H), 8.70 (d, J = 1.2 Hz, 1 H), 9.43 (d,J = 1.2 Hz, 1 H). MS (ES+) m/z 396.1 (M+MeCN+H⁺). Anal. Calcd for C21H26N2O3.0.2 C6H14: C, 71.74; H, 7.84; N, 7.54. Found: C, 71.77; H, 7.55; N, 7.98.

- 12. A solution of **12** (17.1 g, 41.7 mmol) was combined with 10% Pd/C (8.6 g) in toluene (210 mL) with Et₃N (2.1 mL). The reaction mixture was shaken on a Parr apparatus at 50 °C and 50 psig for about 28 h. It was stopped when the hydrogen uptake slowed. The reaction mixture was filtered through Celite[®] and evaporated. After purification by flash chromatography on silica gel (11-cm diameter, gradient elution with 20–30% EtOAc in heptane), **15** was isolated as a gum (12.5 g, 72%). ¹H NMR (CDCl₃, 300 MHz): δ 1.0–1.6 (m, 6H), 1.45 (s, 9H), 2.0–2.7 (m, 8H), 3.00 (m, 1H), 3.26 (m, 1H), 3.67 (s, 3H), 3.83 (m, 1H), 4.11 (m, 2H), 6.49 (d, *J* = 8 Hz, 1H), 6.62 (t, *J* = 7 Hz, 1H), 6.97 (m, 2H). MS (ES+) *m*/*z* 417.1 (M+H⁺). HRMS (FAB⁺) *m*/*z* (M⁺) Calcd for C₂₄H₃₆N₂O₄: 416.2675. Found: 416.2662.
- 13. The four isomers of **15** were separated by sequential chiral chromatography. Isomers (lettered based on elution order,

with isomer-**a** eluting first) **15a** and **15b** were separated from isomers **15c** and **15d** using a Chiralpak[®] OD column [cellulose tris-(3,5-dimethyl-phenylcarbamate) coated on 20-µm silica gel, 41 × 5 cm; using methanol as eluant: 100 vol% at 80 mL/min; 220 nM]. The isomers **15a** $([\alpha]_{D}^{23}$ +30, *c* 0.59, MeOH) and **15b** $([\alpha]_{D}^{23} +62, c$ 0.54, MeOH) were separated using a Chiralpak[®] AD column [amylose tris-(3,5-dimethyl-phenylcarbamate) coated on 20-µm silica gel, using ethanol as eluant under the same conditions as above]. The isomers **15c** $([\alpha]_{D}^{23} -65, c$ 0.68, MeOH) and **15d** $([\alpha]_{D}^{23} -31, c$ 0.55, MeOH) were separated similarly on a Chiralpak[®] AD column. The purity of each isomer was greater than 99% by HPLC.

- Duggan, M. E.; Hartman, G. D.; Meissner, R. S.; Perkins, J. J. U. S. Patent 6,410,526, 2002; *Chem. Abstr.* 2000, 861451.
- Luci, D. K.; Santulli, R. J.; Gauthier, D. A.; Ghosh, S.; Kinney, W. A.; DeCorte, B.; Galemmo, R. A., Jr.; Lewis, J. M.; Proost, J. C.; Tounge, B. A.; Dorsch, W. E.; Wagaman, M. W.; Damiano, B. P.; Maryanoff, B. E. *Heterocycles* 2004, 62, 543.
- Hoekstra, W. J.; Beavers, M. P.; Andrade-Gordon, P.; Evangelisto, M. F.; Keane, P. M.; Press, J. B.; Tomko, K. A.; Fan, F.; Kloczewiak, M.; Mayo, K. H.; Durkin, K. A.; Liotta, D. C. J. Med. Chem. 1995, 38, 1582.
- Lawson, E. C.; Kinney, W. A.; Costanzo, M. J.; Hoekstra, W. J.; Kauffman, J. A.; Luci, D. K.; Santulli, R.; Tounge, B. A.; Yabut, S. C.; Andrade-Gordon, P.; Maryanoff, B. E. Lett. Drug Des. Discovery 2004, 1, 14.
- 18. Rat PK experiments were done in animals dosed po at 10 mg/kg (N = 4) versus iv at 2 mg/kg (N = 4), unless otherwise noted. The drug levels in plasma were compared by LC/MS. Oral dosing was done in D5W (5% Dextrose Injection USP, Baxter) adjusted to pH2 with 0.1 N HCl solution. All compounds were fully soluble in this formulation.
- 19. As a precaution, analytical reference and plasma samples of **29** and **31** were treated with acidified $(0.1\% \text{ CF}_3\text{CO}_2\text{H})$ MeCN during the protein precipitation step, because significant oxidation to the quinoline occurs in neutral MeCN.