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# Synthesis of Milbertycins $\beta_9$ and $\beta_{10}$ from Milbertycins $A_3$ and $A_4$ and Their Biological Activities

Takahiro TSUKIYAMA<sup>a</sup>, Ayako KINOSHITA<sup>a</sup>, Reiji ICHINOSE<sup>b</sup> & Kazuo SATO<sup>a</sup>

<sup>a</sup> Agroscience Research Laboratories, Sankyo Co. Ltd. 1041 Yasu, Yasu-cho, Yasu-gun, Shiga 520-2342, Japan

<sup>b</sup> Crop Protection Department, Sankyo Co. Ltd. 7-12 Ginza 2-Chome, Chuo-ku, Tokyo 104-8113, Japan

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#### Note

## **JS**A

## Synthesis of Milbertycins $\beta_9$ and $\beta_{10}$ from Milbertycins $A_3$ and $A_4$ and Their Biological Activities

Takahiro Tsukiyama,<sup>†</sup> Ayako Kinoshita, Reiji Ichinose,<sup>\*</sup> and Kazuo Sato

Agroscience Research Laboratories, Sankyo Co. Ltd., 1041 Yasu, Yasu-cho, Yasu-gun, Shiga 520-2342, Japan \*Crop Protection Department, Sankyo Co. Ltd., 7-12 Ginza 2-Chome, Chuo-ku, Tokyo 104-8113, Japan

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Chemical derivation methods were used to prepare milbemycins  $\beta_9$  and  $\beta_{10}$  from milbemycins  $A_3$  and  $A_4$ . Their acaricidal activities were also assessed against the organophosphorus-sensitive two-spotted spider mite (*Tetranychus urticae*) on primary leaves of cowpea plants (*Vigna sinesis Savi* species) by spraying.

### Key words: milbemycin; acaricide

Milbemycins<sup>1-2)</sup> are sixteen-membered ring macrolides that have been isolated from Streptomyces hygroscopicus subsp. aureolacrimosus. They exhibit notable activities as acaricides, insecticides and anthelmintics. Among them, milbemectin<sup>3)</sup> [a mixture of milberrycins  $A_3$  (1) and  $A_4$  (2) (Fig. 1)] has been developed as an agricultural acaricide. Since the discovery of milbemycins, numerous homologues such as Merck's avermectins<sup>4)</sup> and Cyanamid's LL-F28249 series (nemadectins)<sup>5)</sup> have been isolated and reported. Enormous efforts have been made to study the biosynthetic pathways to milbemycins,6 and many congeners, which have the same sixteen-membered ring moiety, have consequently been isolated. In a recent study on the biosynthetic pathways to milbemycins, Nonaka et al. have reported the discovery of milberrycins  $\beta_9$  (3) and  $\beta_{10}$  (4) (Fig. 1).<sup>6c)</sup> Interest in their biological activities and their necessity as reference materials for further study of the biosynthetic pathways required preparing them by chemical derivation from milberty  $A_3$  (1) and  $A_4$  (2), because of their low biosynthetic productivity.<sup>6c)</sup> We report in this paper the chemical derivations from milbemycins A<sub>3</sub> (1) and A<sub>4</sub> (2) of milberrycins  $\beta_9$  (3) and  $\beta_{10}$  (4), and their acaricidal activities.

The straightforward chemical derivation from milbemycins A<sub>3</sub> (1) and A<sub>4</sub> (2) of milbemycins  $\beta_9$  (3) and  $\beta_{10}$  (4) was achieved as shown in the Figure. Milbemycin A<sub>3</sub> (1) was methylated as reported<sup>7)</sup> to afford 5-*O*methylated derivative **5** (85.9%), together with 5-*O*, 7-*O*-dimethylated derivative **7** (3.8%) as a byproduct. In the case of milbemycin A<sub>4</sub> (2), the yield of 6 decreased to 39.9% due to the increased formation of 5-O, 7-O-dimethylated by-product 8 (22.7%). These results may have been due to the difference in solubility of milberrycins  $A_3(1)$  and  $A_4(2)$  in acetonitrile. In the reaction, milberty  $A_3(1)$  was suspended in acetonitrile, but milbemycin  $A_4$  (2) was completely dissolved. After purification by preparative HPLC, 5-O-methylated derivatives 5 and 6 were oxidized to 27-oxo derivatives 9 and 10 with chromium (VI) oxide  $(CrO_3)^{(8)}$  in 15.6% and 24.4% yields, respectively. Considerable amounts of the over-oxidized products, 5-O-formyl-27-oxo derivative 11 (5.5%) and 12 (6.5%), were formed in this reaction, and residual 5 and 6 were degraded into messy products. Reduction of the 27-lactone moiety of 9 and 10 with diisobutylaluminum hydride (DIBAL-H) provided milberrycins  $\beta_9$  (3) (57.6%) and  $\beta_{10}$  (4) (49.6%). All spectral data for 3 and 4 synthesized here are in agreement with those of the reference materials.6c)

The acaricidal activities of milbemycins  $\beta_9$  (3) and  $\beta_{10}$  (4) were studied by spraying on primary leaves of cowpea plants (*Vigna sinesis Savi* species) infested with the organophosphorus-sensitive two-spotted spider mites (*Tetranychus urticae*). The results are summarized in the Table 1 Parent milbemycins A<sub>3</sub> (1) and A<sub>4</sub> (2) exhibited high acaricidal activities, while milbemycins  $\beta_9$  (3) and  $\beta_{10}$  (4) exhibited only poor acaricidal activities.

### **Experimental**

NMR spectra were measured with a Varian Gemini-200 FT NMR spectrometer (200 MHz) or a Jeol JNM-GX-270 FT NMR spectrometer (270 MHz). Chemical shifts ( $\delta$ ) are expressed in parts per million relative to internal tetramethylsilane. Mass spectra were measured by a Fisions Instruments VG Autospec, and IR spectra were measured by a Perkin Elmer 1600 series FT IR instrument.

<sup>&</sup>lt;sup>†</sup> To whom correspondence should be addressed. Fax: +81-77-588-2538; E-mail: tukiya@yasu.sankyo.co.jp



Fig. 1. Structure of Milbemycins and Reaction Conditions. Milbemycins A<sub>3</sub> (1), A<sub>4</sub> (2),  $\beta_9$  (3) and  $\beta_{10}$  (4); a) MeI, Ag<sub>2</sub>O, acetonitrile; b) CrO<sub>3</sub>, pyridine; c) DIBAL-H, toluene-THF

Methylation of milbertycin  $A_3$  (1) with methyl iodide (MeI) and silver (I) oxide (Åg,O). To a stirred suspension of 1 (1.00 g, 1.89 mmol) in acetonitrile (20 ml) at ambient temperature were added MeI 11.36 mmol) and Ag<sub>2</sub>O (878.3 mg, (0.71 ml, 3.79 mmol). After stirring overnight, the reaction mixture was filtered with Celite,<sup>®</sup> and the resulting filtrate was evaporated in vacuo. The residue was purified by silica gel column chromatography [nhexane (Hex)-ethyl acetate (EtOAc) gradient] and preparative HPLC (YMC-Pack ODS S-365-10, I-10  $\mu$ m 120A, 30 mm I.D.  $\times$  500 mm, acetonitrilewater) to give 881.7 mg (85.9%) of 5 and 3.8 mg (3.8%) of 7 as colorless amorphous solids.

5-O-Methylmilbemycin  $A_3$  (5): IR  $v_{max}$  (film) cm<sup>-1</sup>: 3465, 2960, 2915, 2860, 1730, 1710; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 5.67–5.81(2H, m, H-9, H-10), 5.29–5.44(3H, m, H-3, H-11, H-19), 4.98(1H, m, H-15), 4.71(1H, d, J=14.7 Hz, H-27), 4.61(1H, d, J=14.7 Hz, H-27), 4.15(1H, s, 7-OH), 4.03(1H, d, J=5.5 Hz, H-6), 3.96(1H, m, H-5), 3.40–3.65(1H, m, H-17), 3.51(3H, s, CH<sub>3</sub>O), 3.20–3.35(2H, m, H-2, H-25), 2.35–2.50(1H, m, H-12), 2.10–2.30(3H, m, H-13, H<sub>2</sub>-16), 1.82(3H, br, H<sub>3</sub>-26), 1.62(3H, br,

 Table 1.
 Acaricidal Activity of Milbemycins against Tetranychus urticae

	Mortality (%)	
	10 ppm	1 ppm
Milbemycin $\beta_9$ (3)	10	0
Milbemycin $\beta_{10}$ (4)	0	0
Milbemycin $A_3$ (1)	69	3
Milbemycin $A_4$ (2)	100	32

H<sub>3</sub>-29), 1.14(3H, d, J=6.6Hz, H<sub>3</sub>-28), 1.00(3H, d, J=6.6 Hz, H<sub>3</sub>-31), 0.83(3H, d, J=6.6 Hz, H<sub>3</sub>-30), 0.82–2.05(10H, m, H-13, H<sub>2</sub>-18, H<sub>2</sub>-20, H<sub>2</sub>-22, H<sub>2</sub>-23, H-24); EI-MS (m/z): 542 (M<sup>+</sup>), 492; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>32</sub>H<sub>46</sub>O<sub>7</sub>, 542.3244; found, 542.3243.

5-O, 7-O-Dimethylmilbemycin  $A_3$  (7): IR  $v_{max}$ (film) cm<sup>-1</sup>: 2960, 2915, 2860, 1730; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 5.66–5.90(3H, m, H-3, H-9, H-10), 5.38(1H, dd, J=14.3, 9.9 Hz, H-11), 4.95-5.20(2H, m, H-15, H-19), 4.67(1H, dd, J=13.9, 2.2 Hz, H-27), 4.57(1H, dd, J=13.9, 1.8 Hz, H-27), 4.27(1H, d, J = 5.5 Hz, H-6), 3.94(1H, m, d, H = 5.5 Hz, H-6), 3.94(1H, m, d, H = 5.5 Hz, H = 5.5 Hz, 3.94(1H, m, d, H = 5.5 Hz, H = 5.5 Hz, 3.94(1H, m, d, H = 5.5 Hz, H = 5.5 Hz, 3.94(1H, m, d, H = 5.5 Hz, 3.94(1H,H-5), 3.51-3.55(1H, m, H-17), 3.48(3H, s, CH<sub>3</sub>O), 3.38(1H, dd, J=4.8, 2.2 Hz, H-2), 3.19-3.30(1H, m, H-25), 3.27(3H, s, CH<sub>3</sub>O), 2.40-2.60(1H, m, H-12), 2.10-2.32(3H, m, H-13, H<sub>2</sub>-16), 1.83(3H, br, H<sub>3</sub>-26), 1.55(3H, s,  $H_3$ -29), 1.14(3H, d, J=6.2 Hz,  $H_3$ -28), 1.03(3H, d, J=6.6 Hz, H<sub>3</sub>-31), 0.82(3H, d, J = 6.6 Hz, H<sub>3</sub>-30), 0.73-2.00(10H, m, H-13, H<sub>2</sub>-18,  $H_2$ -20,  $H_2$ -22,  $H_2$ -23, H-24); EI-MS (*m*/*z*): 556 (M<sup>+</sup>), 492; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>33</sub>H<sub>48</sub>O<sub>7</sub>, 556.3400; found, 556.3401.

Methylation of milbemycin  $A_4$  (2) with MeI and  $Ag_2O$ . Using the same procedure as that described for the preparation of 5, milbemycin  $A_4$  (2) (10.00 g, 18.5 mmol) was methylated with MeI (6.9 ml, 110.7 mmol) and  $Ag_2O$  (8.5 g, 36.9 mmol) to give 4.10 g (39.9%) of 6 and 2.39 g (22.7%) of 8 as colorless amorphous solids.

5-O-Methylmilbemycin  $A_4$  (6): IR  $v_{max}$  (film) cm<sup>-1</sup>: 3440, 2960, 2915, 2870, 1730, 1705; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 5.67–5.82(2H, m, H-9, H-10), 5.29–5.43(3H, m, H-3, H-11, H-19), 4.96(1H, m, H-15), 4.71(1H, d, J=14.9 Hz, H-27), 4.62(1H, d, J=14.9 Hz, H-27), 4.16(1H, s, 7-OH), 4.03(1H, d, J=5.7 Hz, H-6), 3.95(1H, m, H-5), 3.45–3.65(1H, m, H-17), 3.51(3H, s, CH<sub>3</sub>O), 3.30(1H, dd, J=4.8, 2.5 Hz, H-2), 3.07(1H, dt,  $J_t$ =9.2 Hz,  $J_d$ =2.6 Hz, H-25), 2.30–2.50(1H, m, H-12), 2.10–2.30(3H, m, H-13, H<sub>2</sub>-16), 2.00(1H, m, H-20), 1.81(3H, br, H<sub>3</sub>-26), 1.53(3H, br, H<sub>3</sub>-29), 1.00(3H, d, J=6.5 Hz, H<sub>3</sub>-28), 0.98(3H, t, J=6.5 Hz, H<sub>3</sub>-32), 0.82(3H, d, J=6.3 Hz, H<sub>3</sub>-30), 0.75–1.90(11H, m, H-13, H<sub>2</sub>-18, H-20, H<sub>2</sub>-22, H<sub>2</sub>-23, H-24, H<sub>2</sub>-31); EI-MS (m/z): 556 (M<sup>+</sup>), 506; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>33</sub>H<sub>48</sub>O<sub>7</sub>, 556.3400; found, 556.3399.

5-O, 7-O-Dimethylmilbemycin  $A_4$  (8): IR  $v_{max}$ (film) cm<sup>-1</sup>: 2960, 2915, 2870, 1730; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>) δ: 5.67-5.91(3H, m, H-3, H-9, H-10), 5.40(1H, dd, J = 13.9, 9.5 Hz, H-11), 4.96-5.13(2H, m, H-15, H-19), 4.65(2H, m, H<sub>2</sub>-27), 4.27(1H, d, *J*=5.1 Hz, H-6), 3.94(1H, d, *J*=5.1 Hz, H-5), 3.60(1H, m, H-17), 3.48(3H, s, CH<sub>3</sub>O), 3.39(1H, m, H-2), 3.28(3H, s, CH<sub>3</sub>O), 3.05(1H, m, H-25), 2.40-2.60(1H, m, H-12), 2.10-2.35(3H, m, H-13, H<sub>2</sub>-16), 1.83(3H, br, H<sub>3</sub>-26), 1.55(3H, br, H<sub>3</sub>-29), 1.03(3H, d, J = 6.6 Hz, H<sub>3</sub>-28), 1.01(3H, t,  $J = 6.2 \text{ Hz}, \text{ H}_3 - 32$ ), 0.81(3H, d,  $J = 6.6 \text{ Hz}, \text{ H}_3 - 30$ ),  $0.79-1.95(12H, m, H-13, H_2-18, H_2-20, H_2-22,$ H<sub>2</sub>-23, H-24, H<sub>2</sub>-31); EI-MS (m/z): 570 (M<sup>+</sup>), 506; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>34</sub>H<sub>50</sub>O<sub>7</sub>, 570.3557; found, 570.3554.

Oxidation of 5 with CrO<sub>3</sub> in pyridine. To stirred pyridine (20 ml) in an ice bath was slowly added CrO<sub>3</sub> (369.0 mg, 3.69 mmol) while maintaining the temperature at under 10°C. To this solution was added a solution of 5-O-methyl-milberrycin A<sub>3</sub> (5; 200.0 mg, 0.37 mmol) in pyridine (2 ml). After stirring for 1 hour, the ice bath was removed, and the mixture stirred at ambient temperature. After stirring for 6 days, the reaction mixture was poured into a mixture of 1 N hydrochloric acid and EtOAc. After stirring for 15 min, the insoluble material was filtered off with Celite, <sup>®</sup> and the filtrate was extracted with EtOAc. The extract was successively washed with 1 N hydrochloric acid, water and brine, dried over magnesium sulfate (MgSO<sub>4</sub>), and evaporated in vacuo. The residue was purified by silica gel column chromatography (Hex-EtOAc gradient) and preparative HPLC (YMC-Pack ODS S-365-10, I-10 µm 120A, 30 mm I.D.  $\times 500 \text{ mm}$ , acetonitrile-water) to give 32.0 mg (15.6%) of 9 and 11.6 mg (5.5%) of 11 as colorless amorphous solids.

5-O-Methyl-27-oxomilbemycin  $A_3$  (9): IR  $v_{max}$ (film) cm<sup>-1</sup>: 3465, 2960, 2915, 2870, 1755, 1730, 1710, 1640; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.30(1H, dd, J=15.0, 11.4 Hz, H-10), 6.53(1H, d, J=11.4 Hz, H-9), 5.85(1H, dd, J=15.0, 9.9 Hz, H-11), 5.70(1H, br, H-3), 5.50(1H, m, H-19), 5.03(1H, m, H-15), 4.69(1H, s, 7-OH), 4.35(1H, d, J=6.5 Hz, H-6), 4.02(1H, m, H-5), 3.65(1H, dd, J=5.5, 2.9 Hz, H-2), 3.50-3.65(1H, m, H-17), 3.43(3H, s, CH<sub>3</sub>O), 3.28(1H, d, J=9.5, 6.2 Hz, H-25), 2.55-2.75(1H, m, H-12), 2.15-2.35(3H, m, H-13, H<sub>2</sub>-16), 1.93(3H, br, H<sub>3</sub>-26), 1.53(3H, br, H<sub>3</sub>-29), 1.15(3H, d, J=6.2 Hz, H<sub>3</sub>-28), 1.03(3H, d, J=6.6 Hz, H<sub>3</sub>-31), 0.84(3H, d, J=6.6 Hz, H<sub>3</sub>-30), 0.82-2.05(10H, m, H-13, H<sub>2</sub>-18, H<sub>2</sub>-20, H<sub>2</sub>-22, H<sub>2</sub>-23, H-24); EI-MS (m/z): 556 (M<sup>+</sup>), 538, 506; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>32</sub>H<sub>44</sub>O<sub>8</sub>, 556.3036; found, 556.3036.

5-O-Formyl-27-oxomilberic A<sub>3</sub> (11): IR  $v_{max}$ (film) cm<sup>-1</sup>: 3450, 2960, 2915, 2870, 1755, 1725, 1640; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.09(1H, s, OCHO), 7.27(1H, dd, J=15.4, 11.4 Hz, H-10), 6.55(1H, d, J=11.4 Hz, H-9), 5.89(1H, dd, J=15.4, 9.9 Hz, H-11), 5.79(1H, br, H-3), 5.70(1H, d, J = 6.2 Hz, H-5), 5.50(1H, m, H-19), 5.03(1H, m, H-15), 4.70(1H, s, 7-OH), 4.45(1H, d, J=6.2 Hz, H-6), 3.55-3.70(1H, m, H-17), 3.50(1H, dd, J=5.1), 2.6 Hz, H-2), 3.28(1H, dd, J=9.5, 6.2 Hz, H-25), 2.55-2.70(1H, m, H-12), 2.15-2.40(3H, m, H-13, H<sub>2</sub>-16), 1.94(3H, br, H<sub>3</sub>-26), 1.54(3H, br, H<sub>3</sub>-29), 1.15(3H, d, J=6.2 Hz, H<sub>3</sub>-31), 1.04(3H, d,  $J = 6.6 \text{ Hz}, H_3 - 28), 0.84(3 \text{ H}, d, J = 6.2 \text{ Hz}, H_3 - 30),$ 0.83-2.05(10H, m, H-13, H<sub>2</sub>-18, H<sub>2</sub>-20, H<sub>2</sub>-22, H<sub>2</sub>-23, H-24); EI-MS (m/z): 570 (M<sup>+</sup>); HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>32</sub>H<sub>42</sub>O<sub>9</sub>, 570.2829; found, 570.2831.

Oxidation of 6 with  $CrO_3$ . Using the same procedure as that described for the preparation of 9, 6 (3.00 g, 5.40 mmol) was oxidized with  $CrO_3$  (4.69 g, 46.94 mmol) in pyridine (330 ml) to give 0.75 g (24.4%) of 10 and 0.20 g (6.5%) of 12 as colorless amorphous solids.

5-O-Methyl-27-oxomilberic A<sub>4</sub> (10): IR  $v_{max}$ (film) cm<sup>-1</sup>: 3475, 2960, 2915, 2870, 1750, 1735, 1710, 1640; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 7.28(1H, J = 15.0,11.2 Hz, H-10), 6.53(1H, dd. d. J=11.2 Hz, H-9), 5.87(1H, dd, J=15.0, 9.6 Hz, H-11), 5.69(1H, br, H-3), 5.40-5.55(1H, m, H-19), 4.99(1H, m, H-15), 4.67(1H, s, 7-OH), 4.34(1H, d, J = 5.5 Hz, H-6), 4.02(1H, d, J = 5.5 Hz, H-5), 3.50-3.65(2H, m, H-2, H-17), 3.42(3H, s, CH<sub>3</sub>O), 3.07(1H, dt,  $J_t = 9.2 \text{ Hz}$ ,  $J_d = 2.3 \text{ Hz}$ , H-25), 2.50-2.70(1H, m, H-12), 2.15-2.30(3H, m, H-13, H<sub>2</sub>-16), 1.92(3H, br, H<sub>3</sub>-26), 1.52(3H, br, H<sub>3</sub>-29), 1.02(3H, d, J=6.9 Hz, H<sub>3</sub>-28), 0.99(3H, t, J=6.9 Hz, H<sub>3</sub>-32), 0.82(3H, d, J=6.4 Hz, H<sub>3</sub>-30),  $0.80-2.05(12H, m, H-13, H_2-18, H_2-20, H_2-22,$ H<sub>2</sub>-23, H-24, H<sub>2</sub>-31); EI-MS (m/z): 570 (M<sup>+</sup>), 552, 520; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>33</sub>H<sub>46</sub>O<sub>8</sub>, 570.3193; found, 570.3192.

5-O-Formyl-27-oxomilbemicin  $A_4$  (12): IR  $v_{max}$ (film) cm<sup>-1</sup>: 3455, 2960, 2915, 2870, 1755, 1730, 1640; <sup>1</sup>H-NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$ : 8.09(1H, s, OCHO), 7.25(1H, dd, J=15.1, 11.4 Hz, H-10), 6.56(1H, d, J=11.4 Hz, H-9), 5.91(1H, dd, J=15.1, 10.1 Hz, H-11), 5.78(1H, br, H-3), 5.70(1H, d, J= 6.3 Hz, H-5), 5.40-5.60(1H, m, H-19), 5.00(1H, m, H-15), 4.70(1H, s, 7-OH), 4.45(1H, d, J=6.3 Hz, H-6), 3.50-3.70(1H, m, H-17), 3.49(1H, m, H-2), Downloaded by [Thammasat University Libraries] at 15:31 07 October 2014

3.08(1H, dt,  $J_t$ =9.1 Hz,  $J_d$ =2.4 Hz, H-25), 2.55-2.80(1H, m, H-12), 2.20-2.35(3H, m, H-13, H<sub>2</sub>-16), 1.93(3H, d, J=1.9 Hz, H<sub>3</sub>-26), 1.53(3H, s, H<sub>3</sub>-29), 1.03(3H, d, J=6.7 Hz, H<sub>3</sub>-28), 1.00(3H, t, J=7.3 Hz, H<sub>3</sub>-32), 0.83(3H, d, J=6.3 Hz, H<sub>3</sub>-30), 0.82-2.05(12H, m, H-13, H<sub>2</sub>-18, H<sub>2</sub>-20, H<sub>2</sub>-22, H<sub>2</sub>-23, H-24, H<sub>2</sub>-31); EI-MS (m/z): 584 (M<sup>+</sup>), 520; HREI-MS (m/z): [M<sup>+</sup>]: calcd. for C<sub>33</sub>H<sub>44</sub>O<sub>9</sub>, 584.2985; found, 584.2987.

Reduction of 9 with DIBAL-H. To a stirred solution of 9 (30 mg, 0.05 mmol) in toluene (1 ml) was added dropwise a 1 M solution of DIBAL-H in tetrahydrofuran (THF; 0.16 ml, 0.16 mmol) while cooling in an ice bath. After stirring for 30 min in the ice bath, an additional 1 M solution of DIBAL-H in THF (0.16 ml, 0.16 mmol) was added dropwise. After stirring for 40 min more in the ice bath, an additional 1 M solution of DIBAL-H in THF (0.32 ml, 0.32 mmol) was again added dropwise. After stirring for a final 40 min in the ice bath, the reaction mixture was poured into 1 N hydrochloric acid and extracted with EtOAc. The extract was successively washed with water and brine, dried over MgSO<sub>4</sub>, and evaporated in vacuo. The residue was purified by preparative TLC (Hex-EtOAc = 1:2) to give 17.4 mg(57.6%) of milberrycin  $\beta_9$  (3) as a colorless amorphous solid.

Reduction of 10 with DIBAL-H. Using the same procedure as that described for the preparation of milbemycin  $\beta_9$  (3), 10 (200.0 mg, 0.35 mmol) was reduced with DIBAL-H in toluene to give 99.9 mg (49.6%) of milbemycin  $\beta_{10}$  (4) as a colorless amorphous solid.

Acaricidal activity against Tetranychus urticae. The primary leaves of cowpea plants (Vigna sinensis Savi species) were infected with the organophosphorus-sensitive two-spotted spider mites (Tetranychus urticae). One day after infection, the infested plants were sprayed by a Mizuho rotary sprayer with 7 ml of a solution containing the test compound at a concentration ranging from 1 to 10 ppm at a rate of 3.5 mg of the test solution per 1 cm<sup>2</sup> of leaf. The plants were assessed after 3 days by examining the adult mites under a binocular microscope to determine the numbers of living and dead individuals. Two plants were used for each concentration and each test compound. The plants were kept during the test in green-house compartments at 25°C.

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### **References and Notes**

- a) Takiguchi, Y., Mishima, H., Okuda, M., Terao, M., Aoki, A., and Fukuda, R., Milbemycins, a new family of macrolide antibiotics: Fermentation, isolation and physico-chemical properties. J. Antibiot., 33, 1120-1127 (1980); b) Okazaki, T., Ono, M., Aoki, A., and Fukuda, R., Milbemycins, a new family of macrolide antibiotics: Producing organism and its mutants. J. Antibiot., 36, 438-441 (1983); c) Mishima, H., Ide, J., Muramatsu, S., and Ono, M., Milbemycins, a new family of macrolide antibiotics: structure determination of milbemycins D, E, F, G, H, J and K. J. Antibiot., 36, 980-990 (1983).
- For reviews of milbemycins and related 16-membered ring macrolides, see: a) Davies, H. G. and Green, R. H., Avermectins and milbemycins. *Natural Product Reports*, 3, 87-121 (1986); b) Davies, H. G. and Green, R. H., Avermectins and milbemycins part 1. *Chem. Soc. Rev.*, 20, 211-269 (1991); c) Ide, J., Okazaki, T., Ono, M., Saito, A., Nakagawa, K., Naito, S., Sato, K., Tanaka, K., Yoshikawa, H., Ando, M., Katsumi, S., Matsumoto, K., Toyama, T., Shibano, M., and Abe, M., Milbemycin: Discovery and development. *Annu. Rep. of Sankyo Res. Lab.*, 45, 1–98 (1993); d) Fisher, M. H., Structure-activity relationships of avermectins and milbemycins. ACS Symp. Ser., 658 (Phytochemicals for pest control), 220-238 (1997).
- Tomlin, C. D. S., "The Pesticide Manual 12th Edition," British Crop Protection Council, 49 Downing Street, Farnham, Surrey GU9 7PH, UK, 2000, P647-648.
- Albers-Schönberg, G., Arison, B. H., Chabala, J. C., Douglas, A. W., Eskola, P., Fisher, M. H., Lusi, A., Mrozik, H., Smith, J. L., and Tolman, R. L., Avermectins. Structure determination. *J. Am. Chem. Soc.*, 103, 4216-4221 (1981).
- 5) Carter, G. T., Nietsche, J. A., and Borders, D. B., Structure determination of LL-F28249 α, β, γ and λ, potent antiparasitic macrolides from *Streptomyces* cyaneogriseus ssp. non cyanogenus. J. Chem. Soc., Chem. Commun., 402-404 (1987).
- a) Ono, M., Mishima, H., Takiguchi, Y., and Terao, M., Milbemycins, a new family of macrolide antibiotics: Fermentation, isolation, physico-chemical properties and bioconversion of milbemycins J and K. J. Antibiot., 36, 509-515 (1983); b) Nonaka, K., Kumasaka, C., Okamoto, Y., Maruyama, F., and Yoshikawa, H., Bioconversion of milbemycin-related compounds: Biosynthetic pathway of milbemycins. J. Antibiot., 52, 109-116 (1999); c) Nonaka, K., Tsukiyama, T., Okamoto, Y., Sato, K., Kumasaka, C., Yamamoto, T., Maruyama, F., and Yoshikawa, H., New milbemycins from Streptomyces hygroscopicus subsp. aureolacrimosus: Fermentation, isolation and structure elucidation. J. Antibiot., 53, 694-704 (2000).
- Naito, S., Nanba, T., Owatari, Y., Nakada, Y., Muramatsu, S., and Ide, J., Milbemycin derivatives: Modification at the C-5 position. J. Antibiot., 47,

 Poos, G. I., Arth, G. E., Beyler, R. E., and Sarett, L. H., Approaches to the total synthesis of adrenal steroids. V. 4b-Methyl-7-ethylenedioxy-1, 2, 3, 4, 4α, 4b, 5, 6, 7, 8, 10,  $10\alpha\beta$ -dodecahydrophenanthrene-4 $\beta$ -ol-1-on and related tricyclic derivatives. *J*, *Am*, *Chem*, *Soc.*, **75**, 422 (1953).