

hydrogenase¹⁰ activity, which is absent from the rat liver preparations used here.

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VITAMIN B₁₂. XXVI. DEGRADATION OF FACTOR III TO 5-HYDROXYBENZIMIDAZOLE AND DERIVATIVES AND BIOSYNTHESIS OF FACTOR III

Sir:

Factor III was isolated from fermented sewage.¹ It has been reported to have hematological activity similar to that of vitamin B₁₂, and appeared to differ from vitamin B₁₂ by having an unknown moiety in place of 5,6-dimethylbenzimidazole.² Friederich and Bernhauer³ have now reported 5-hydroxybenzimidazole as a degradation product of Factor III.

Last year, through the generosity of Professor Dr. K. Bernhauer, we received samples of Factor III. We have independently identified 5-hydroxybenzimidazole as a part of the molecule. We have also prepared two crystalline cobalt complexes from 5-hydroxybenzimidazole and Factor B⁴ by biosynthesis. One of these appears to be Factor III from comparison with the substance isolated from sewage.

Factor III was hydrolyzed with 6 *N* hydrochloric acid for 20 hours at room temperature, and the hydrolysate was subjected to paper electrophoresis in 0.5 *N* acetic acid containing a little cyanide. Material showing bright blue-white fluorescence under ultraviolet light separated from the pigments present. On paper chromatography in a butanol-acetic acid-water system,⁵ it separated into two spots (I and II) of unequal intensity with *R_f* values of 0.16 and 0.22. Further hydrolysis of combined I and II with 6 *N* hydrochloric acid at 95° for 24 hours gave a new substance (IV) with an *R_f* value of 0.46. Hydrolysis of Factor III, or of the fluorescent materials I and II, with 6 *N* hydrochloric acid at 150° for 21 hours gave another fluorescent substance (V) with an *R_f* value of 0.65, and phosphate ion was detected in the hydrolysates. These same hydrolysis conditions degrade vitamin B₁₂ to isomers of ribazole phosphate, ribazole and 5,6-dimethylbenzimidazole.⁶

In the case of Factor III, it was assumed that the substances obtained were isomers of a riboside phosphate (I and II), the riboside (IV) and the base (V). The base was isolated as a crystalline picrate, m.p. mainly 220–225°, which was converted to a polymorphic crystalline hydrochloride,

m.p. mainly 185–190°. There was not sufficient material for elemental analysis, so a detailed study of spectra was made.

The absorption spectrum of V seemed to eliminate purines, pyrimidines, pyridines and alkylbenzimidazoles and indicated an hydroxybenzimidazole structure. The previously undescribed 5-hydroxybenzimidazole was prepared by demethylation of 5-methoxybenzimidazole⁷ with hydrobromic acid; m.p. 220–222°; *Anal.* Found: C, 62.26; H, 4.60; N, 20.90. The absorption spectrum ($\lambda_{\text{max}}^{0.1N \text{ NaOH}}$, 2500 (485); 3050 (573); $\lambda_{\text{max}}^{0.1N \text{ HCl}}$, 2400–2500 (shoulder); 2870 (528)) was in good agreement with that of base V. The picrate and hydrochloride of synthetic 5-hydroxybenzimidazole had wide melting point ranges, but mixed melting points with the isolated salts showed no depression. The infrared spectra of the hydrochlorides showed them to be identical.

Factor B and synthetic 5-hydroxybenzimidazole were combined microbiologically using *Escherichia coli* 113-3,⁸ and two red crystalline compounds were isolated. One of these did not separate from Factor III on mixed paper chromatograms. This result confirmed the chemical evidence that 5-hydroxybenzimidazole is present in Factor III and is not an artifact. The structure and synthesis of the other degradation products of Factor III are being studied further.

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ON THE ORIGIN OF THE METHYL GROUPS OF PHOSPHOLIPID CHOLINE IN THE RAT¹

Sir:

We previously reported that the extent of incorporation of the carbon of the methyl group of methionine, labeled with C¹⁴, to phospholipid choline is markedly reduced in the folic acid-deficient rat, and as a tentative explanation of this observation we suggested that in the folic acid-deficient rat the synthesis of the acceptor of the methyl group of methionine for choline formation is inhibited². It is generally assumed that aminoethanol is the acceptor of three methyl groups of methionine *in vivo*, the formation of choline taking place via direct transmethylation from methionine. However, existing evidence does not indicate that folic acid or its biological derivative is a co-factor in the enzymatic reactions involving transmethylation from methionine, neither is folic acid or its derivative involved in the *in vivo* decarboxylation of serine to aminoethanol. It occurred to us, therefore, that the acceptor of the methyl group

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