

ISOLATION AND STRUCTURE OF AXISONITRILE-1 AND AXISOTHIOCYANATE-1 TWO UNUSUAL SESQUITERPENOIDS FROM THE MARINE SPONGE *AXINELLA CANNABINA*^a

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Abstract—Two sesquiterpenoids, axisonitrile-1 and axisothiocyante-1, have been isolated from the sponge *Axinella cannabina*. On the basis of chemical and physico-chemical evidence structure 1 is suggested for axisonitrile-1 and structure 2 for axisothiocyante-1.

The only known naturally occurring isonitrile is the antibiotic xanthocillin discovered by Rothe¹ in cultures of *Penicillium notatum* Westling and *Penicillium chrysogenum*. During our studies on the metabolites of Porifera² we isolated a sesquiterpenoid isonitrile, axisonitrile-1, from the marine sponge *Axinella cannabina* and describe the assignment of structure 1 to this compound. In addition, we isolated from the same sponge axisothiocyante-1 (2) and the structure determination proved it to be an isothiocyante strictly related to 1.

Axisonitrile-1 (1). Fresh material was extracted with acetone and the ether soluble fraction, after chromatography on silica gel, afforded 1, C₁₆H₂₅N (elemental analyses and mass spectrum), m.p. 43–45°, [α]_D + 22.6.

The presence of the isonitrile group was deduced from the IR (ν_{max} 2130 cm⁻¹) and mass spectra [intense ion at *m/e* 204 (M⁺ —HCN)]. The NMR spectrum suggests that the isonitrile function is linked to a methine group (δ 3.13, 1H, bm).

Both IR (ν_{max} 3050, 1640 and 895 cm⁻¹) and NMR spectra (δ 4.75, 2H, s) clearly indicate that a >C=CH_2 group is present in 1. A further structural feature, revealed by NMR, is the presence of three Me groups, one tertiary (δ 0.99, s) and the other two secondary (δ 0.85 and 1.03, d, J = 6 Hz). The IR data (ν_{max} 1385 and 1375 cm⁻¹) indicates that the two secondary methyls are part of an isopropyl group. These facts strongly suggest that 1 is a bicyclic sesquiterpenoid isonitrile.

Further information consistent with the presence

of the unit $\begin{array}{c} \text{N}^+ \equiv \text{C}^- \\ | \\ \text{>CH}-\text{CH}-\text{CH}=\text{Me}_2 \end{array}$ (A) was obtained by conversion of axisonitrile-1 into axisothiocyante-1 by treatment with sulphur at

120°. Since an accurate analysis of NMR spectrum of 2, as described below, suggests the presence of

the unit $\begin{array}{c} \text{N}=\text{C}=\text{S} \\ | \\ \text{>CH}-\text{CH}-\text{CH}=\text{Me}_2 \end{array}$ it follows that unit

A must be present in 1. This was confirmed by the following experiments: LAH reduction of 1, afforded the amine 3, *n*_D²⁵ 1.4940, [α]_D + 15.5, M⁺ 235 *m/e*; 3 by methylation and subsequent treatment with AgOH gave the corresponding quaternary base which, by thermal decomposition, afforded 4 in high yield, *n*_D³⁰ 1.5014; [α]_D - 91.7; M⁺ 204 *m/e*.

Compound 4 contains the unit $\text{>CH}-\text{CH}=\text{C}=\text{Me}_2$ as shown by its NMR spectrum: δ 4.86 (1H, doublet broadened by long range coupling, J = 9 Hz, H—C₁₀) and 1.64 and 1.52 (6H, singlets broadened by long range coupling, H₃—C₁₂ and H₃—C₁₃).

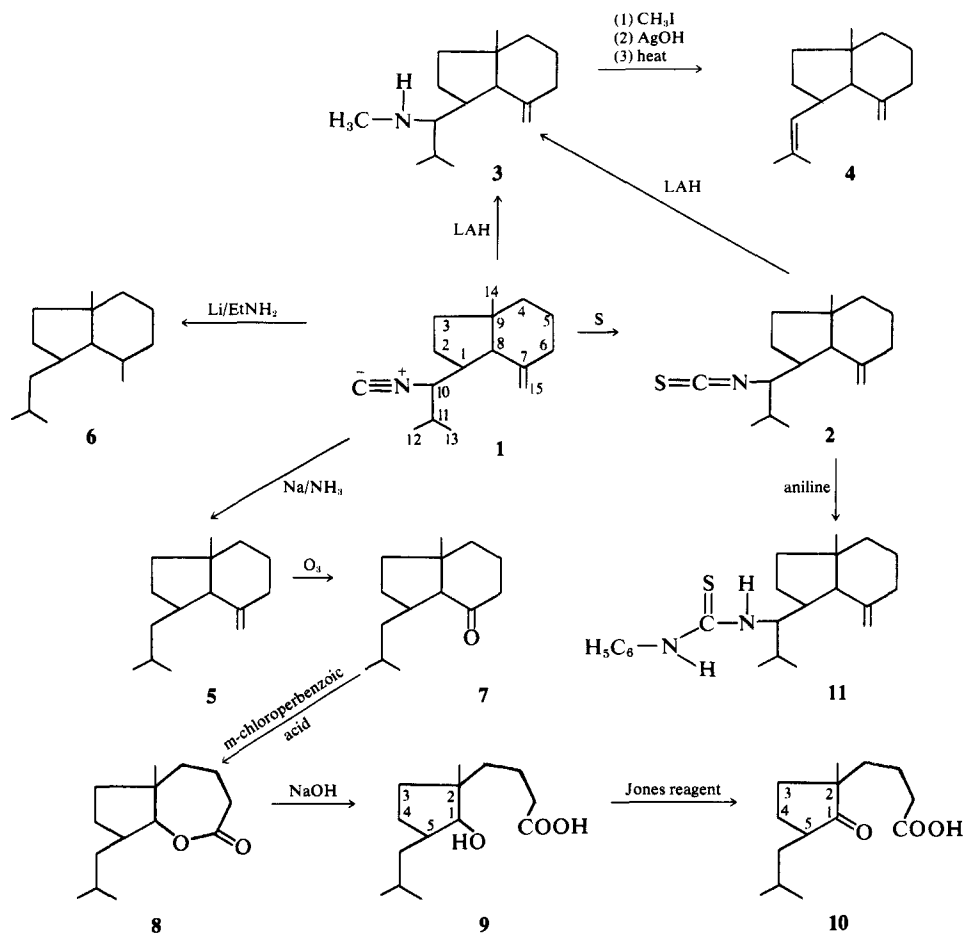
Additional proof for the structure of axisonitrile-1 was provided as follows: Compound 1 afforded 5 [*n*_D²⁵ 1.4802; [α]_D + 8.5; M⁺ 206 *m/e*; δ 4.65 (2H, m, H₂—C₁₅)] by reduction with sodium in liquid ammonia and 6, axane, (*n*_D²⁵ 1.4753; [α]_D - 5.5; M⁺ 208 *m/e*) by treatment with lithium in ethylamine.

Ozonization of 5, followed by decomposition of the ozonide with Na₂SO₃aq, gave ketone 7, *n*_D²⁵ 1.4773; [α]_D + 39.1, M⁺ 208 *m/e*. The IR spectrum of 7 (ν_{max} 1707 cm⁻¹) suggests that the keto group is in a 6-membered ring.

Evidence for the position of the keto group in 7 and consequently of the *exo*-methylene in 1 was secured by deuterium exchange of enolisable hydrogens of 7. The mass spectrum of deuterated compound revealed that three deuterium atoms are incorporated and as a consequence the unit

$\begin{array}{c} | \\ -\text{HC}-\text{C}-\text{CH}_2- \\ || \\ \text{CH}_2 \end{array}$ must be present in the 6-membered ring of axisonitrile-1.

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The ketone **7**, by Baeyer-Villiger oxidation, afforded lactone **8**, n_D^{20} 1.4895; $[\alpha]_D -33.6$; M^+ 224 m/e ; $\nu_{\max}$ 1745 cm^{-1} (CO lactone). In the NMR spectrum a doublet at δ 3.72 (1H, $J = 5$ Hz,

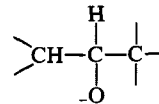
$\text{H}-\text{C}-\text{O}-$) is present.

Alkaline hydrolysis of **8** gave the hydroxy acid **9**, m.p. 92–94°; $[\alpha]_D -36.8$; M^+ 242 m/e ; $\nu_{\max}$ 1705 cm^{-1} , δ 3.25 (1H, d, $J = 6.5$ Hz, $\text{H}-\text{C}-\text{OH}$).

Finally **9**, by treatment with Jones reagent afforded **10**, n_D^{25} 1.4690; $[\alpha]_D -107.0$; M^+ 240 m/e . The IR spectrum shows a band at 1738 cm^{-1} consistent with the presence of a cyclopentanone system.

Only one hydrogen must be present on the C atoms α to $\text{C}=\text{O}$ (C_2 and C_5): in fact, the NMR spectrum of **8** (δ 3.72, 1H, d, $\text{H}-\text{C}-\text{O}-$) and **9** (δ

3.25, 1H, d, $\text{CH}-\text{OH}$) indicates that in the cyclopentane ring the part structure



present.

In the light of these results the three substituents present on C_2 and C_5 in **8** and **9** and, consequently, in **10** must be a tertiary Me group, an isobutyl group and the acidic residue arising from the oxidative degradation of the 6-membered ring.

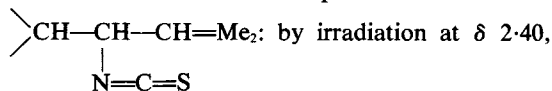
Since the isobutyl group, as reported above, is linked to a methyne group, the Me and $-(\text{CH}_2)_3\text{COOH}$ groups are located on the same C atom.

These considerations, together with all the other results, led us to propose structure **1** for axisonitrile-1.

Axisothiocyanate-1 (2). The second compound present in *Axinella cannabina* in smaller amounts was also isolated from the ether soluble fraction of the acetone extract by chromatography on silica

gel. Compound 2 is an oily substance, $C_{16}H_{25}NS$ (elemental analyses and mass spectrum) $n_D^{25} 1.5394$; $[\alpha]_D + 5.9$. Spectral data [$\nu_{\max}$ 3050, 1650 and 895 cm^{-1} ; δ 4.78 (2H, bm)] point to the presence of a >C=CH_2 group. Axisothiocyanate-1 possess three Me groups, one tertiary (δ 0.98, 3H, s) and two secondary (δ 0.89, 3H, d, $J = 7\text{ Hz}$ and δ 1.00, 3H, d, $J = 7\text{ Hz}$). The presence of an isothiocyante function was deduced from the IR ($\nu_{\max}$ 2120 cm^{-1}), UV ($\lambda_{\max}$ 243 nm, ϵ 2500) and mass spectra [ions at m/e 230 ($M^+ - \text{HS}$) and 204 ($M^+ - \text{HNCS}$)]. This was confirmed by treatment of 2 with aniline which gave in high yields thiourea 11 m.p. 63–66°; $[\alpha]_D - 33.5$; M^+ 356 m/e . Inspection of the NMR spectrum of 2 also indicated that the isothiocyante group is linked to a methyne group (δ 3.27, 1H, t, $J = 5.5\text{ Hz}$).

Further analysis of the NMR spectrum of 2 and spin decoupling experiments provided useful information consistent with the presence of the unit



the triplet at δ 3.27 (1H, $\text{H}-\text{C}_{10}$) collapses into a doublet; by irradiation at δ 2.00 the triplet at δ 3.27 is simplified into a doublet, while the two Me doublets at δ 0.89 and 1.00 (H_3-C_{12} and H_3-C_{13}) collapse into two singlets.

All these facts suggest a close relationship of 2 with axisonitrile-1 (1) as proved: Compound 1 by treatment with sulphur at 120° afforded 2; furthermore 2 by reduction with LAH gave the same amine (3) as obtained from 1. Since the structure of 1 has been established it follows that structure 2 can be assigned to axisothiocyanate-1.

Axisothiocyanate-1, as well as axisonitrile-1, are sesquiterpenes with a skeleton which has not been found before in a naturally occurring compound.

Biogenetically they are interesting molecules; their structures suggest, in fact, new biogenetic pathways for the formation *in vivo* of the carbon skeleton as well as the isonitrile and isothiocyante functions.

EXPERIMENTAL

The UV and IR (CCl_4 solns) spectra were recorded on a Perkin-Elmer 402 and 157 spectrophotometer. NMR spectra were determined on a Perkin-Elmer R12A and Varian HA-100 spectrometers in CCl_4 solns using TMS as internal reference with $\delta = 0$; s = singlet, d = doublet, t = triplet, m = multiplet, b = broad. Mass spectra were taken on AEI MS 902 instrument. Optical rotations were measured with a Perkin-Elmer 141 polarimeter. Elemental analyses were performed by Mr. S. De Rosa (Laboratorio per la Chimica e Fisica di Molecole di Interesse Biologico del CNR-Arco Felice-Napoli). TLC and PLC separations were effected using glass packed precoated silica gel F₂₅₄ plates (E. Merck). GLC's were run using a Perkin-Elmer 881 instrument with glass columns $2\text{ m} \times 0.5\text{ cm}$ (flow of nitrogen 30 ml/min).

Sponges (*Axinella cannabina*), collected in the bay of

Taranto, were obtained from Stazione di Biologia marina del Salento-Porto Cesareo (dir. Prof. P. Parenzan).

Isolation of axisonitrile-1 (1) and axisothiocyanate-1 (2) from the sponge *Axinella cannabina*. Fresh sponges (500 g, dry after extraction) were extracted 4 times with acetone at room temp for 2 days. The combined extracts (81) were concentrated under red press and the remaining aqueous residue was extracted with Et_2O (2l in 3 portions). The organic phase was taken to dryness leaving an oily residue (7.6 g), which was chromatographed on a SiO_2 (760 g) column (eluent: $40-70^\circ$ light petroleum– C_6H_6 8:2). Fractions of 450 ml were collected. Fractions 10–14, on evaporation, afforded mg 370 of 1 gas-chromatographically pure (2.5% SE 30 on chromosorb W at 145° , 162° , 185°); m.p. $43-45^\circ$; $[\alpha]_D + 22.6$ (c 1, CHCl_3); M^+ 231 m/e ; $\nu_{\max}$ 3050, 2130, 1640, 1385, 1375, 895 cm^{-1} ; δ 4.75 (2H, s, H_2-C_{15}), 3.13 (1H, bm, $\text{H}-\text{C}_{10}$), 1.03 (3H, d, $J = 6\text{ Hz}$, H_3-C_{12} or H_3-C_{13}), 0.99 (3H, s, H_3-C_{14}), 0.85 (3H, d, $J = 6\text{ Hz}$, H_3-C_{12} or H_3-C_{13}). (Found C, 83.12; H, 10.75; N, 6.09. Calc. for $\text{C}_{16}\text{H}_{25}\text{N}$ C, 83.05; H, 10.89; N, 6.05%).

Fractions 3–4 were evaporated to dryness and the oily residue (130 mg) was further purified on a SiO_2 (13 g) column using $40-70^\circ$ light petroleum as eluent. Fractions of 10 ml were collected. By evaporation of the fractions 10–12, 70 mg of 2 were obtained as an oily product, $n_D^{25} 1.5394$; $[\alpha]_D + 5.9$ (c 2.5, CHCl_3); M^+ 263 m/e ; $\nu_{\max}$ 3050, 2120, 1650, 1385, 1375, 895 cm^{-1} ; δ 4.78 (2H, bm, H_2-C_{15}), 3.27 (1H, t, $J = 5.5\text{ Hz}$, $\text{H}-\text{C}_{10}$), 1.00 (3H, d, $J = 7\text{ Hz}$, H_3-C_{12} or H_3-C_{13}), 0.98 (3H, s, H_3-C_{14}), 0.89 (3H, d, $J = 7\text{ Hz}$, H_3-C_{12} or H_3-C_{13}). (Found C, 72.75; H, 9.83; N, 5.30; S, 12.20. Calc. for $\text{C}_{16}\text{H}_{25}\text{NS}$ C, 72.95; H, 9.57; N, 5.32; S, 12.16%).

LAH reduction of 1 to 3. LAH (600 mg) and 1 (2.4 g) in dry Et_2O (100 ml) were refluxed for 3 h. EtOAc was added to destroy unreacted LAH. After addition of H_2O and extraction with Et_2O , the organic phase was washed, dried and taken to dryness. The residue was purified by column chromatography (SiO_2 , 120 g) using Et_2O as eluent. Fractions of 60 ml were collected. From the fractions 7–10, after evaporation of the solvents, 1.1 g of 3 were obtained, M^+ 235 m/e ; $n_D^{25} 1.4940$; $[\alpha]_D + 15.5$ (c 2.5, CHCl_3); $\nu_{\max}$ 3380, 3040, 1640, 1380, 1370, 895 cm^{-1} ; δ 4.70 (2H, s, H_2-C_{15}), 2.32 (3H, s, $\text{N}-\text{CH}_3$), 0.95 (3H, s, H_3-C_{14}), 0.88 (3H, d, $J = 6\text{ Hz}$, H_3-C_{12} or H_3-C_{13}), 0.80 (3H, d, $J = 6\text{ Hz}$, H_3-C_{13}) (CDCl_3).

Hofmann exhaustive methylation of 3 to 4. A mixture of 3 (2.2 g), MeI (10 ml) K_2CO_3 (2.5 g) in H_2O (40 ml) was refluxed for 7 h. After cooling, excess MeI was removed *in vacuo* and the soln was extracted repeatedly with CHCl_3 . The organic phase, after evaporation of the solvent, afforded 3.3 g of the quaternary salt, which, without further purification, was dissolved in MeOH (40 ml) and water (2 ml). After addition of Ag_2O (4 g) the mixture was stirred for 2 h at room temp; the ppt was removed by filtration and washed with CHCl_3 . The filtrate, taken to dryness, gave 2.97 g of crude quaternary base, which was heated at $160-180^\circ$ for 30 min.

The distillate was dissolved in Et_2O (10 ml), dried over CaSO_4 and evaporated to dryness. The residue (mg 900) was purified by PLC (8 plates) using $40-70^\circ$ light petroleum as eluent. The band R_f 0.7 (UV light), after elution with Et_2O , gave mg 530 of 4, $[\alpha]_D - 91.7$ (c 2.7, CHCl_3); $n_D^{20} 1.5014$; M^+ 204 m/e ; $\nu_{\max}$ 1645 and 890 cm^{-1} ; δ 4.86 (1H, bd, $J = 9\text{ Hz}$, $\text{H}-\text{C}_{10}$), 4.52 (2H, m, H_2-C_{15}), 0.95 (3H, s, H_3-C_{14}) and 1.64 and 1.52 (each 3H, bs, H_3-C_{12} and H_3-C_{13}).

Reduction of 1 with Na/NH₃ to 5. To a soln of 1 (300 mg) in liquid NH₃ (25 ml) and Et₂O (25 ml) at -45° under stirring Na was slowly (2 h) added. Excess Na was destroyed with a little NH₄Cl and the resultant mixture was taken to dryness. The residue, after addition of H₂O, was extracted with 40–70° light petroleum and the organic phase was evaporated to dryness to give 290 mg of an oily product, which was chromatographed on a SiO₂ (10 g) column (eluent 40–70° light petroleum). Fractions 1–3 (75 ml), evaporated to dryness, gave 224 mg of 5 gas-chromatographically pure (2.5% SE 30 on chromosorb W at 118° and 128°); *M*⁺ 206 *m/e*; *n*_D²⁵ 1.4802; [*α*]_D + 8.5 (c 1, CHCl₃); *ν*_{max} 3050, 1640, 1380, 1375, 890 cm⁻¹; δ 4.65 (2H, m, H₂—C₁₅), 0.95 (3H, s, H₃—C₁₄), 0.85 (3H, d, J = 6 Hz, H₃—C₁₂ or H₃—C₁₃), 0.80 (3H, d, J = 6 Hz, H₃—C₁₂ or H₃—C₁₃).

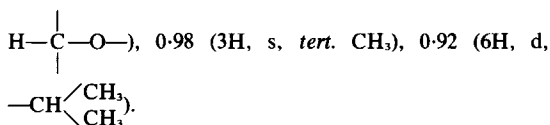
Reduction of 1 with Li/EtNH₂ to axane (6). To a soln of 1 (145 mg) in anhyd EtNH₂ (7 ml), Li (60 mg) was slowly added at 16°. After 90 min a little NH₄Cl was added and the EtNH₂ was evaporated. After addition of H₂O (10 ml) the suspension was extracted with 40–70° light petroleum; the organic phase was washed with H₂O, dried over CaSO₄ and taken to dryness.

The residue (128 mg) was chromatographed on SiO₂/AgNO₃ (7:3; 12 g) column (eluent 40–70° light petroleum—Et₂O 49:1). Fractions of 10 ml were collected. Fractions 11–20, after removal of the solvent, afforded 35 mg of 6, gas-chromatographically pure (2.5% SE 30 on chromosorb W at 128° and 150°); *M*⁺ 208 *m/e*; *n*_D²⁵ 1.4753; [*α*]_D - 5.5 (c 2, CHCl₃); δ 0.97 (3H, s, H₃—C₁₄), 0.90 (3H, d, J = 7 Hz, H₃—C₁₅; irradiation at δ 1.75 collapses this doublet to a singlet), 0.86 (6H, d, J = 6 Hz, H₃—C₁₂ and H₃—C₁₃; irradiation at δ 1.56 collapses this doublet to a singlet).

Ozonolysis of 5 to 7. Ozonized O₂ (2% O₃) was passed through a soln of 5 (2.2 g) in MeOH—EtOAc (1:1, 100 ml) at -40° for 2 h. The ozonide was decomposed with a saturated aqueous soln of Na₂SO₃ (1 g) at 40° for 30 min. After removal of MeOH and EtOAc under red press, the suspension was extracted with 40–70° light petroleum. The organic phase was dried over CaSO₄ and taken to dryness. The residue (1.8 g) was chromatographed on a SiO₂ (50 g) column using the following solvent systems: n-hexane—C₆H₆ 7:3 (360 ml), n-hexane—C₆H₆ 6:4. Fractions of 40 ml were collected. Fractions 23–46, after removal of the solvents, gave 7 (750 mg), *M*⁺ 208 *m/e*; [*α*]_D + 39.1 (c 3, CHCl₃); *n*_D²⁵ 1.4773; *ν*_{max} 1707, 1380, 1375 cm⁻¹; δ 1.07 (3H, s, H₃—C₁₄), 0.84 (6H, d, J = 6 Hz, H₃—C₁₂ and H₃—C₁₃).

Deuteration of enolisable hydrogens of 7. Compound 7 (5 mg), Na (15 mg), D₂O (0.5 ml) and MeOD (0.5 ml) were heated at 70° for 48 h in a sealed tube. After removal of MeOD *in vacuo*, the suspension was diluted with D₂O, acidified with N DCl in D₂O and extracted with Et₂O. The organic phase was taken to dryness to give 4 mg of 7-d, of 91% isotopic purity, *M*⁺ 211 *m/e*.

Baeyer-Villiger oxidation of 7 to 8. A soln of 7 (240 mg) and *m*-chloroperbenzoic acid (300 mg) in CHCl₃ (8 ml) was refluxed for 5 h. After evaporation of the solvent, the residue was dissolved in Et₂O (30 ml). The soln was washed repeatedly with 2N Na₂CO₃ and then with H₂O. After evaporation of Et₂O the residue was chromatographed on a SiO₂ (15 g) column using the following solvents: C₆H₆ (150 ml), C₆H₆—Et₂O 9:1. Fractions of 50 ml were collected. The fractions 23–24, taken to dryness, gave mg 167 of 8, [*α*]_D - 33.6 (c 2, CHCl₃); *n*_D²⁰ (1.4895; *M*⁺ 224 *m/e*; *ν*_{max} 1745 cm⁻¹; δ 3.72 (1H, d, J = 5 Hz,



Alkaline hydrolysis of 8 to 9. To a soln of 8 (120 mg) in dioxane (2 ml) 10% Na₂CO₃aq (8 ml) was added. After refluxing for 2 h the soln was washed with Et₂O, acidified with 2N HCl to pH 4 and extracted with Et₂O. The ethereal extract was washed with H₂O, dried over CaSO₄ and evaporated to dryness to give 94 mg of 9, which was crystallized from 60–80° light petroleum, m.p. 92–94°; [*α*]_D - 36.8 (c 2.5; CHCl₃); *M*⁺ 242 *m/e*; *ν*_{max} 1705 cm⁻¹; δ 3.25

(1H, d, J = 6.5 Hz, H—C—OH, 0.95 (3H, s, tert Me), 0.86 (6H, d, J = 6 Hz —CH=Me₂) (CDCl₃).

Oxidation of 9 to 10. The hydroxyacid 9 (60 mg) in acetone (5 ml) was treated with Jones reagent for 30 min at room temp. Following the usual work-up 10 (40 mg) was obtained and was purified by PLC (eluent C₆H₆—Et₂O 1:1, *R*_f 0.4), *n*_D²⁵ 1.4690; [*α*]_D - 107.0 (c 0.7, CHCl₃); *ν*_{max} 1738 and 1705 cm⁻¹.

Treatment of 2 with aniline to obtain 11. Compound 2 (50 mg) and excess aniline were kept at room temp for 24 h. After dilution with H₂O, the suspension was extracted with Et₂O (50 ml in 3 portions). The combined ethereal extracts, after washing with H₂O, were dried and taken to dryness. The residue (57 mg) was chromatographed on PLC (eluent C₆H₆—Et₂O 9:1). The band *R*_f 0.7 (UV light) was eluted with Et₂O to give 45 mg of 11, m.p. 63–66°; [*α*]_D - 33.5 (c 3, CHCl₃); *M*⁺ 356 *m/e*; *ν*_{max} 3380, 3050, 1635, 895 cm⁻¹; δ 7.22 (5H, m, aromatic protons), 4.57 (2H, m, H₂—C₁₅), 0.95 (3H, d, J = 6 Hz, H₃—C₁₂ or H₃—C₁₃), 0.89 (3H, s, H₃—C₁₄), 0.72 (3H, d, J = 6 Hz, H₃—C₁₂ or H₃—C₁₃).

Treatment of 1 with sulphur to obtain 2. Compound 1 (200 mg) and S (70 mg) were heated at 120° for 16 h; after addition of 40–70° light petroleum (30 ml) and filtration, the soln was taken to dryness and the residue was purified by PLC (2 plates) (eluent: 80–100° light petroleum). The band *R*_f 0.5, eluted with C₆H₆, gave 100 mg of 2.

LAH reduction of 2 to 3. Compound 2 (50 mg) was reduced with the experimental conditions used for 1. Working up as previously described afforded a compound, which was identified as 3 by *n*_D²⁵, [*α*]_D, and chromatographic (TLC in C₆H₆—Et₂O 8:2) and spectral (IR, NMR and MS) properties.

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