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## Studies on the Agalwood (Jinkō). VII.<sup>1)</sup> Structures of Phenylethylchromone Derivatives AH<sub>7</sub>, AH<sub>8</sub> and AH<sub>9</sub>

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Three new phenylethylchromone derivatives, tentatively named AH<sub>7</sub>, AH<sub>8</sub> and AH<sub>9</sub>, were isolated from acetone and pyridine extracts of agalwood (jinkō) from Kalimantan. AH<sub>7</sub> from the pyridine extract was characterized as 5,8-dihydroxy-2-(2-phenylethyl)chromone, and AH<sub>8</sub> from the acetone extract was elucidated to be 6,7-dimethoxy-2-[2-(4-methoxyphenyl)ethyl]chromone. AH<sub>9</sub>, acetylated in order to separate it from the mixture, was concluded to be (5*S*,6*S*,7*R*)-5*a*',6*a*',7*a*'-triaceoxy-2-[2-(2-acetoxyphenyl)ethyl]-5,6,7,8,8-pentahydrochromone on the basis of the proton nuclear magnetic resonance (<sup>1</sup>H-NMR) spectrum, dihedral angle and circular dichroism (CD) spectral data.

**Keywords**—2-(2-phenylethyl)chromone; agalwood; Aquilariaceae; polyoxylate; <sup>1</sup>H-NMR; 2D-COSY; CD; dihedral angle

In the preceding papers of this series<sup>1a-c)</sup> it was reported that acetone extract of agalwood (jinkō) contained six 2-(2-phenylethyl)chromone derivatives, tentatively named AH<sub>1</sub> (agarotetrol),<sup>2)</sup> AH<sub>2</sub> (isoagarotetrol), AH<sub>3</sub>, AH<sub>4</sub>, AH<sub>5</sub>, and AH<sub>6</sub> along with other related compounds.

This paper deals with the isolation and characterization of three additional minor constituents from the acetone and pyridine extracts, tentatively named AH<sub>7</sub>, AH<sub>8</sub> and AH<sub>9</sub>. The procedures of isolation are described in the experimental section.

AH<sub>7</sub> (**1**), yellowish needles, mp 198–201 °C, exhibited absorptions due to a trisubstituted γ-pyrone ring, and hydroxyl and phenylethyl groups in the ultraviolet (UV) and infrared (IR) spectra, suggesting it to be a compound closely related to 5,8-dihydroxy-2-(2-phenylethyl)chromone, obtained as a by-product in the acetonide formation of agarotetrol by Yoshii *et al.*,<sup>2)</sup> and the structure was further supported by the proton and carbon-13 nuclear magnetic resonance (<sup>1</sup>H- and <sup>13</sup>C-NMR) spectra (Table II). In the <sup>1</sup>H-NMR spectrum of **1** the doublet signals at δ 6.66 and 7.14 were assigned to 6- and 7-H, respectively, based on the α-effect downfield shifts (*ca.* 0.2–0.3 ppm) following the acetylation at 5- and 8-OH.

Accordingly, AH<sub>7</sub> was concluded to be 5,8-dihydroxy-2-(2-phenylethyl)chromone.

AH<sub>8</sub> (**2**), colorless needles, C<sub>20</sub>H<sub>20</sub>O<sub>5</sub>, mp 139–140 °C, was indicated to be a trimethoxyl derivative of 2-(2-phenylethyl)chromone on the basis of the molecular formula and <sup>1</sup>H-NMR spectrum. The structure of the 6,7-dimethoxylate, as in the case of AH<sub>6</sub>,<sup>1a)</sup> was suggested by the <sup>1</sup>H-NMR spectrum, which showed two singlet signals due to aromatic protons at δ 7.07 and 7.77, and another methoxyl group located at the *para*-position with respect to the 1'-carbon of the phenylethyl group was suggested by the appearance of the proton signals of the A<sup>2</sup>–B<sup>2</sup> system due to the 1,4-disubstituted benzene ring. In the <sup>13</sup>C-NMR spectrum the assignments of carbons at the 5–10 positions were in fairly good accord with the calculated

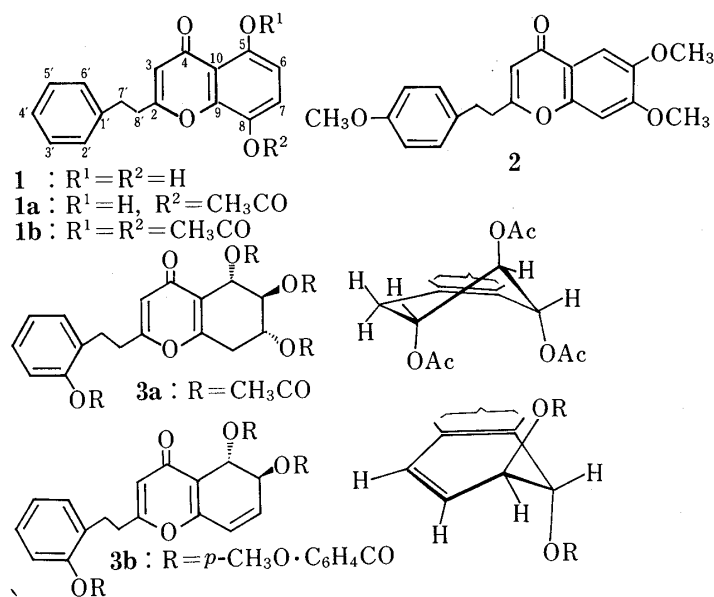
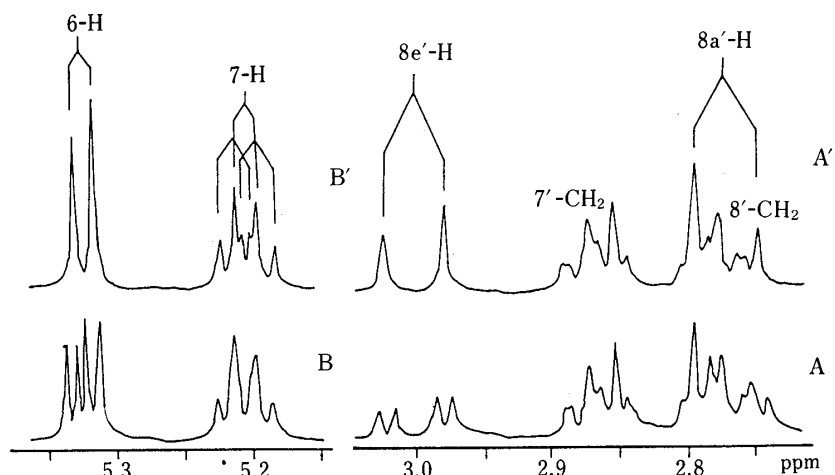


Chart 1

Fig. 1.  $^1H$ -NMR Spectra of  $AH_9$  (**3a**)

A and B are normal spectra, and A' and B' are spectra irradiated at  $\delta$  5.21 and 6.02, respectively.

TABLE I.  $^1H$ -NMR Spectral Data for **3a** and Results of  $^1H$ -Selective Irradiations

H	(400 MHz, in $CDCl_3$ )	
3	6.12 (1H, s)	
5	6.02 (1H, d, $J = 3.5$ Hz)	
6	5.33 (1H, dd, $J = 6.0, 3.5$ Hz)	(d, $J = 6.0$ Hz) <sup>a)</sup> (d, $J = 3.5$ Hz) <sup>b)</sup>
7	5.21 (1H, bq)	(dt, $J = 6.0, 5.0, 5.0$ Hz) <sup>a)</sup>
8a'	2.77 (1H, dd, $J = 18.0, 5.0$ Hz)	(d, $J = 18.0$ Hz) <sup>b)</sup>
8e'	2.99 (1H, dd, $J = 18.0, 5.0$ Hz)	(d, $J = 18.0$ Hz) <sup>b)</sup>
3'	7.06 (1H, dd, $J = 8.0, 1.0$ Hz)	
4'—6'	7.24 (3H, m)	
7'	2.87 (2H, m)	
8'	2.78 (2H, m)	
$CH_3COO$	2.05, 2.07, 2.08, 2.33 (each 3H, s)	

a) Selective irradiation at 5-H. b) Selective irradiation at 7-H.

TABLE II.  $^{13}\text{C}$ -NMR Spectral Data for **1**, **1a**, **1b**, **2** and **3a** (ppm)<sup>a)</sup>

Carbon	<b>1</b>	<b>1a</b>	<b>1b</b>	<b>2</b>	<b>3a</b> <sup>b)</sup>
2	170.14	170.67	169.64	168.82	167.94
3	108.82	109.38	111.51	109.65	113.71
4	183.94	183.32	176.20	178.06	176.70
5	158.71	161.30	159.63	104.70 (104.0) <sup>c)</sup>	65.99
6	110.53	110.68	118.40	148.09 (148.0)	69.12
7	122.74	127.37	126.25	155.08 (154.2)	66.48
8	138.90	130.53	137.19	100.47 (100.1)	27.71
9	152.92	157.86	156.49	152.99 (152.6)	162.26
10	111.51	110.70	118.40	116.96 (118.1)	117.89
1'	140.23	139.79	139.46	132.59 (133.0)	132.46 (133.0) <sup>d)</sup>
2'	128.84	129.00	128.83	129.83 (130.0)	149.74 (151.2)
3'	128.57	128.72	128.73	114.50 (114.4)	123.17 (122.4)
4'	126.75	126.84	126.69	148.09 (158.2)	128.12 (128.0)
5'	128.57	128.72	128.73	114.50 (114.4)	126.53 (126.0)
6'	128.84	129.00	128.83	129.83 (130.0)	130.19 (129.5)
7'	35.69	35.64	35.59	36.02	33.86
8'	32.72	32.54	32.61	32.16	30.11
CH <sub>3</sub> O	<b>2</b> : 55.32, 56.14, 56.58				
CH <sub>3</sub> COO	<b>1a</b> : 20.38, 169.85 <b>1b</b> : 20.45, 21.01, 166.73, 168.14 <b>3a</b> : 20.52 ( $\times 2$ ), 20.62 ( $\times 2$ ), 169.32 ( $\times 2$ ), 169.98 ( $\times 2$ )				

a) The spectrum of **1b** was measured in  $\text{CDCl}_3$ , and others were measured in pyridine- $d_5$ . b) Assignments of  $\text{C}_5$ — $\text{C}_{10}$  were established on the basis of the data for  $\text{AH}_2$  (isoagarotetrol).<sup>1b)</sup> c) Calculated values in parentheses have been described in connection with the data for  $\text{AH}_6$ .<sup>1a,3)</sup> d) Values in parentheses were calculated by application of monomethoxy substituent effects<sup>3)</sup> to the data for  $\text{AH}_1$  (agarotetrol) tetraacetate.<sup>1b)</sup>

TABLE III. "Observed" and Calculated Dihedral Angles in **3a**

	5-H, 6-H		6-H, 7-H	7-H, 8-H	
	e'e	ee	ea	ea'	ee'
"Observed"	58°	42°	—	48°	48°
	( $J=3.5$ Hz)	( $J=6.0$ Hz)		( $J=5.0$ Hz)	( $J=5.0$ Hz)
Conduritols <sup>4)</sup>	62°	—	64°	53°	62°
Büchi	70°	56°	64°	50°	70°

values, as shown in Table II.

Therefore,  $\text{AH}_8$  was concluded to be 6,7-dimethoxy-2-[2-(4-methoxyphenyl)ethyl]chromone, **2**.

$\text{AH}_9$  (**3a**), colorless needles,  $\text{C}_{25}\text{H}_{26}\text{O}_{10}$ , mp 147—148 °C,  $[\alpha]_{\text{D}} -11.1^\circ$ , exhibited some characteristic absorption maxima due to acetoxyl phenylethylchromones in the UV and IR spectra, suggesting it to be a polyoxyl derivative related to agarotetrol and isoagarotetrol.<sup>1b)</sup> The  $^1\text{H}$ -NMR spectrum indicated the presence of four acetoxyl groups ( $\delta$  2.05, 2.07, 2.08 and 2.33) and three methine protons ( $\delta$  5.21, 5.33 and 6.02). However, since the methine proton signal at  $\delta$  5.21 appeared as a broad quartet-like peak and some signals regarded as being due to methylene protons were very complicated, **3a** was subjected to  $^1\text{H}$ -selective decoupling experiments. On  $^1\text{H}$ -selective irradiation at  $\delta$  6.02, a signal at  $\delta$  5.21 was transformed from the broad quartet into a doublet of triplets ( $J=6.0$ , 5.0, 5.0 Hz) and a signal at  $\delta$  5.33 from a doublet of doublets into a doublet ( $J=6.0$  Hz). Similarly, at  $\delta$  5.21 a pair of geminally coupled methylene protons was clearly confirmed to give two doublet signals ( $J=18.0$  Hz) at  $\delta$  2.77

and 2.99 as shown in Fig. 1. Therefore, the hexenyl ring was proved to have three adjacent acetoxyl groups at  $C_5$ ,  $C_6$  and  $C_7$  or  $C_6$ ,  $C_7$  and  $C_8$ . Two protons at  $\delta$  2.77 and 2.99 were ascribable to the methylene protons at  $C_8$  based on the 2D-COSY spectrum, which showed cross peak signals due to the long-range couplings of  $C_8$ -methylene ( $\delta$  2.78) beyond the  $\gamma$ -pyrone ring. The  $^{13}\text{C}$ -NMR spectrum also showed a chemical shift of the  $C_4$  signal analogous with that for isoagarotetrol<sup>1b)</sup> while the chemical shift in the case of  $C_9$  showed no effect ascribable to substitution by an acetoxyl group at  $C_8$ . Since there was also an upfield shift of about 7.0 ppm in the  $C_1$  signal, as shown in Table II, the other acetoxyl group should be attached to the *ortho*-position with respect to 1'-carbon.<sup>3)</sup>

By assuming a half-chair conformation with steric and electrostatic repulsion between 4-CO and 5-OAc<sup>2)</sup> the vicinal methine protons ( $C_5$ — $C_7$ ) of the hexenyl ring can be regarded as being pseudoequatorial ( $e'$ ), equatorial ( $e$ ) and  $e$ , respectively. The dihedral angles calculated by the use of the equation  $J = 11.0 \cos^2 \phi$  and the observed values of  $J_{5,6}$ ,  $J_{6,7}$  and  $J_{7,8}$  are given in Table III.<sup>4)</sup> These values obtained from  $J_{5,6(e',e)}$  and  $J_{7,8(e,e')}$  were in good agreement with the corresponding values of conduritols observed by Abraham *et al.*<sup>4)</sup> Consideration of the difference of  $J_{5,6(e',e)}$  and  $J_{7,8(e,e')}$  values suggested it to be the result of hexenyl ring buckling due to the effects of  $\gamma$ -pyrone ring and three acetoxyl substituents at  $C_5$ ,  $C_6$  and  $C_7$  in the hexenyl ring. Therefore, the stereochemistry of three vicinal methine protons in the cyclohexenyl moiety was supported as being  $5e'$ ,  $6e$  and  $7e$ , respectively.

In order to determine the absolute configuration of **3a**, the *p*-methoxybenzoate (**3b**) was prepared from **3a** followed by hydrolysis of the acetoxyl groups. Compound **3b** showed the three methoxyl group signals at  $\delta$  3.83, 3.85 and 3.87 in the  $^1\text{H}$ -NMR spectrum, suggesting the tri-*p*-methoxybenzoate structure, and further, four methine proton signals at  $\delta$  5.62, 6.22, 6.55 and 6.63 were assigned to the vicinal protons at  $C_5$ — $C_8$  based on the coupling systems arising from the 5,6-dioxy-7,9-cyclohexadienyl ring partial structure which was derived by dehydration between 7-OH and 8-H of **3a**.

Accordingly, **3b** was characterized as  $\Delta^{7,8}$ -5a',6a,2'-tri-*p*-methoxybenzoate. The circular dichroism (CD) spectrum of **3b** in ethanol showed a positive chirality<sup>5)</sup> of the 5,6-dibenzoate groups which was in accordance with the figure of Newman's projection at  $C_5$  and  $C_6$  (Figs. 2 and 3).

Since the conformations of the hexenyl moiety at  $C_5$  and  $C_6$  can be regarded as almost the same in **3a** and **3b**, the drawing shown in Chart 1 represents the absolute structure of **3a**. Consequently, AH<sub>9</sub> was defined as (5*S*,6*S*,7*R*)-2-[2-(2-acetoxyphenyl)ethyl]-5a',6a,7a-tri-acetoxy-5,6,7,8,8-pentahydrochromone, **3a**.

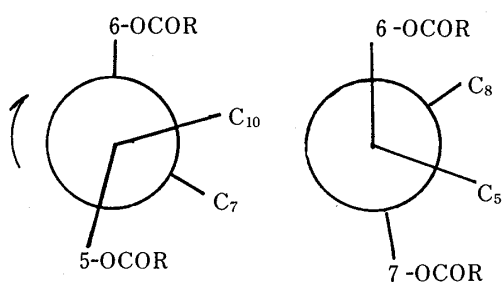


Fig. 2. Newman's Projection Following Büchi's Dreiding Model

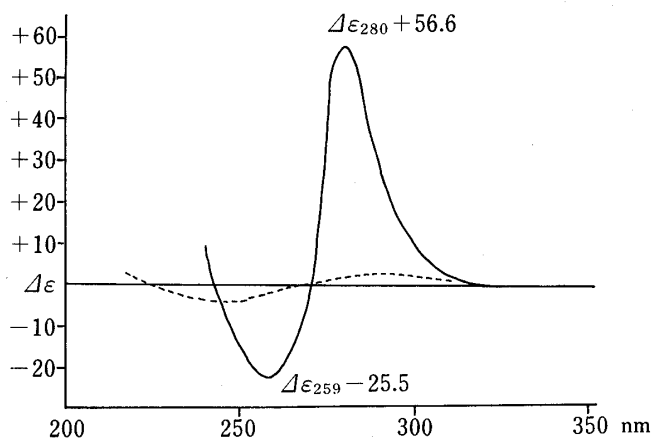


Fig. 3. CD Spectra (MeOH)  
**3a**, ----; **3b**, —

Compounds **1**, **2** and **3a** were elucidated to be polyoxyl derivatives of 2-(2-phenylethyl)chromone. They are the first to be isolated from agalwood, and the hexenyl ring structure of **AH<sub>9</sub>** is of interest in connection with that of isoagarotetrol.

### Experimental

Melting points were determined on a micro melting point apparatus (Yanagimoto) and are uncorrected. The UV and CD spectra were obtained in EtOH (or MeOH) with a Shimadzu UV-200s spectrometer and a JASCO J-500c spectropolarimeter, respectively, and IR spectra (in KBr disks) with a Shimadzu IR 27G spectrometer. The <sup>1</sup>H-NMR spectra were taken on a Varian CFT-20 spectrometer at 79.54 MHz and a JEOL JNM GX-400 spectrometer at 399.65 MHz at 24 °C, and <sup>13</sup>C-NMR spectra on the Varian CFT-20 at 20.0 MHz and the JEOL JNM GX-400 at 100.4 MHz. Chemical shifts are given in δ (ppm) with tetramethylsilane as an internal standard (s, singlet; d, doublet; t, triplet; dd, double doublet; dt, double triplet; ddd, double double doublet; m, multiplet; br, broad). Column chromatographies were performed on Kieselgel 60 (70–230 mesh, Merck), Kiesel 60 silanisiert (70–230 mesh, Merck) and polyamide (Woelm).

**Isolation of AH<sub>7</sub>, AH<sub>8</sub> and AH<sub>9</sub>**—As described previously (refer to Chart 1 in Part I<sup>1a)</sup> of this series) six crystalline compounds, **AH<sub>1</sub>**–**AH<sub>6</sub>** were isolated from the ethereal and acetone extracts of agalwood from Kalimantan. Fr<sub>2</sub> (35.2 g) was further chromatographed on a polyamide column (MeOH–H<sub>2</sub>O, 7:3 v/v) for the isolation of some constituents, with monitoring by thin layer chromatography (TLC) (detection under ultraviolet light). A mixture (20.1 g) of desired constituents was subjected to silica gel chromatography (CHCl<sub>3</sub>–MeOH, 10:1 v/v) and followed by similar chromatography again but using hexane–AcOEt (1:1 v/v) as the eluent. The fractions of **AH<sub>6</sub>** (820 mg) and **AH<sub>8</sub>** (88 mg) were recrystallized from MeOH to give colorless plates (611 mg) and needles (25 mg), respectively. On the other hand, residue 2 (950 g)<sup>11</sup> was extracted 3 times with pyridine (1 l) under reflux for 5 h. The combined filtrate was concentrated to give a dark viscous extract (300 g), and it was extracted with MeOH under reflux. The MeOH extract (75 g) was chromatographed on a silica gel column by using AcOEt and MeOH eluting solutions successively. The mixture (200 mg) of desired constituents from the AcOEt eluate was subjected to silica gel chromatography (hexane–AcOEt, 1:2 v/v) to give a fraction containing **AH<sub>7</sub>** (53 mg). The MeOH eluate (66.6 g) was again column-chromatographed on a silica gel column (CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O, 8.5:1.5:0.15 v/v). The eluate (8.6 g) was acetylated with Ac<sub>2</sub>O–pyridine by standing overnight and the mixture was evaporated to dryness under reduced pressure. The residue (9.2 g) was chromatographed over a column of silica gel (hexane–AcOEt, 1:1 v/v), and the fractionated **AH<sub>9</sub>** (324 mg) was further purified by a silanized silica gel chromatography (MeOH–H<sub>2</sub>O, 7:3 v/v). **AH<sub>7</sub>** (40 mg), yellowish needles, was recrystallized from AcOEt and **AH<sub>9</sub>** (86 mg), colorless needles, recrystallized from an ether solution of the purified **AH<sub>9</sub>** (139 mg).

**AH<sub>7</sub> (1)**—Yellowish needles (from AcOEt), mp 198–201 °C. UV λ<sub>max</sub><sup>EtOH</sup> nm (ε): 240.5 (15125), 300.0 (1681), 358.4 (2581). IR (KBr) cm<sup>−1</sup>: 3120 (OH), 1655, 1615, 1586 (γ-pyrone ring), 852, 822, 749 (phenyl). <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ: 3.02 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 6.05 (1H, s, 3-H), 6.66 (1H, d, *J* = 8.8 Hz, 6-H), 7.14 (1H, d, *J* = 8.8 Hz, 7-H), 7.25 (5H, m, 2'–6' protons), 1.56, 11.64 (each 1H, s, OH). Anal. Calcd for C<sub>17</sub>H<sub>14</sub>O<sub>4</sub>: C, 72.33; H, 5.00. Found: C, 72.30; H, 4.75. <sup>13</sup>C-NMR: Table II.

**Acetylation of 1**—a) The 8-Acetate: An Ac<sub>2</sub>O–pyridine (1:1, v/v, 5 ml) mixture containing **1** (33 mg) was allowed to stand overnight and evaporated to dryness under reduced pressure. The residue was purified by column chromatography (hexane–AcOEt, 1:1 v/v) and recrystallized from hexane–AcOEt (2:1 v/v) to give colorless needles, mp 100–101 °C. UV λ<sub>max</sub><sup>MeOH</sup> nm (ε): 234 (19395), 257 (11741), 334 (4119). IR (KBr) cm<sup>−1</sup>: 3500 (OH), 1770 (ester), 1660, 1630, 1600, 1590 (γ-pyrone), 879, 810, 700 (phenyl). <sup>1</sup>H-NMR (pyridine-*d*<sub>5</sub>) δ: 2.36 (3H, s, CH<sub>3</sub>CO), 2.92 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 6.18 (1H, s, 3-H), 6.80 (1H, d, *J* = 8.9 Hz, 6-H), 7.25 (5H, m, 2'–6' protons), 7.46 (1H, d, *J* = 8.9 Hz, 7-H). Anal. Calcd for C<sub>19</sub>H<sub>16</sub>O<sub>5</sub>: C, 70.36; H, 4.97. Found: C, 69.91; H, 4.92. <sup>13</sup>C-NMR: Table II.

b) The 5,8-Diacetate (**1b**): An Ac<sub>2</sub>O (2 ml)–AcONa (20 mg) solution of **1** (80 mg) was heated on a water bath at 60 °C for 1 h. After addition of ice water, the aqueous mixture was filtered to obtain a precipitate (83 mg). The major product (72 mg) separated from the precipitate by column chromatography (hexane–AcOEt, 2:1 v/v) was recrystallized from acetone to give colorless needles, mp 149–150 °C. UV λ<sub>max</sub><sup>MeOH</sup> nm (ε): 212 (21036), 225 (23570), 256 (4807), 304 (6992). IR (KBr) cm<sup>−1</sup>: 1770, 1718 (ester), 1665, 1620, 1582 (γ-pyrone), 900, 703 (phenyl). <sup>1</sup>H-NMR (CDCl<sub>3</sub>) δ: 2.38, 2.39 (each 3H, s, CH<sub>3</sub>CO), 2.92 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 5.99 (1H, s, 3-H), 6.95 (1H, d, *J* = 8.6, 6-H), 7.22 (5H, m, 2'–6' protons), 7.38 (1H, d, *J* = 8.6 Hz, 7-H). Anal. Calcd for C<sub>21</sub>H<sub>18</sub>O<sub>6</sub>: C, 68.84; H, 4.95. Found: C, 68.68; H, 4.75. <sup>13</sup>C-NMR: Table II.

**AH<sub>8</sub> (2)**—Colorless needles (from hexane–AcOEt, 1:1 v/v), mp 139–140 °C. UV λ<sub>max</sub><sup>MeOH</sup> nm (ε): 230 (31672), 316 (10999). IR (KBr) cm<sup>−1</sup>: 1640, 1600 (γ-pyrone), 831 (phenyl). <sup>1</sup>H-NMR (pyridine-*d*<sub>5</sub>) δ: 2.93 (4H, m, CH<sub>2</sub>CH<sub>2</sub>), 3.66, 3.76, 3.86 (each 3H, s, CH<sub>3</sub>O), 6.35 (1H, s, 3-H), 6.94, 7.25 (each 2H, dt, *J* = 8.7, 3.0, 3.0 Hz, aromatic H), 7.07, 7.77 (each 1H, s, 8- and 5-H). Anal. Calcd for C<sub>20</sub>H<sub>20</sub>O<sub>5</sub>: C, 70.57; H, 5.92. Found: C, 70.60; H, 5.88. <sup>13</sup>C-NMR: Table II.

**AH<sub>9</sub> (3a)**—Colorless needles (from Et<sub>2</sub>O), mp 147–148 °C, [α]<sub>D</sub><sup>19</sup> −11.1° (*c* = 1.08, MeOH). UV λ<sub>max</sub><sup>MeOH</sup> nm (ε):

249 (12973). IR (KBr)  $\text{cm}^{-1}$ : 1754 (ester), 1669, 1633, 1613 ( $\gamma$ -pyrone), 822, 724 (phenyl).  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.05, 2.07, 2.08, 2.33 (each 3H, s,  $\text{CH}_3\text{CO}$ ), 2.78, 2.87 (each 2H, m,  $\text{CH}_2\text{CH}_2$ ), 2.77, 2.99 (each 1H, dd,  $J=18.0, 5.0$  Hz, 8-a'- and 8-e'-H), 5.21 (1H, br q,  $J=10.0, 5.5$  Hz, 7-H), 5.33 (1H, dd,  $J=6.0, 3.5$  Hz, 6-H), 6.02 (1H, d,  $J=3.5$  Hz, 5-H), 6.12 (1H, s, 3-H), 7.06 (1H, dd,  $J=8.0, 1.0$  Hz, 3'-H), 7.24 (3H, m, 4', 5', 6' protons).  $^{13}\text{C-NMR}$ : Table II. High-resolution MS  $m/z$ : Calcd for  $\text{C}_{25}\text{H}_{26}\text{O}_{10}$ : 486.15242. Found: 486.1531.

**Tri-*p*-methoxybenzoate (3b) from 3a**—An absolute MeOH solution (5 ml) of **3a** (23 mg) was heated with anhydrous sodium carbonate (50 mg) at  $50^\circ\text{C}$  for 5 h, and the reaction mixture was filtered. The filtrate was evaporated to dryness *in vacuo*. The residue (16.5 mg) was chromatographed on a silica gel column ( $\text{CHCl}_3$ -MeOH, 20:1 v/v) to afford a main product (7.2 mg). To a pyridine solution (1 ml) of the product (7.2 mg), *p*-methoxybenzoyl chloride (20 mg) was added and the mixture was allowed to stand overnight at  $5^\circ\text{C}$ . The mixture was then evaporated to dryness under reduced pressure. The residue was chromatographed on a silica gel column with *n*-hexane-AcOEt (1:1, v/v) to afford a powder (4.8 mg) from MeOH: (mp  $82\text{--}84^\circ\text{C}$ ),  $[\alpha]_D^{22} +64.4^\circ$  ( $c=1.0$ , MeOH). CD ( $c=1.37 \times 10^{-5}$ , MeOH)  $\Delta\epsilon^{25}$ :  $-25.5$  (259 nm) (negative maximum),  $+56.6$  (280 nm) (positive maximum).  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.85, 2.96 (each 2H, m,  $\text{CH}_2\text{CH}_2$ ), 3.83, 3.85, 3.87 (each 3H, s,  $\text{CH}_3\text{O}$ ), 5.62 (1H, dd,  $J=5.1, 1.7$  Hz, 6-H), 6.14 (1H, s, 3-H), 6.22 (1H, d,  $J=10.1$  Hz, 8-H), 6.55 (1H, dd,  $J=1.7, 1.3$  Hz, 5-H), 6.63 (1H, ddd,  $J=10.1, 5.1, 1.3$  Hz, 7-H), 6.84, 6.88, 6.99 (each 2H, d,  $J=9.0$  Hz, aromatic H), 7.26 (m, aromatic H), 7.91, 7.93, 8.16 (each 2H, d,  $J=9.0$  Hz, aromatic H).

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