

TRITERPENOID SAPONINS FROM BERNEUXIA THEBETICA

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MING-KUI WANG,* HONG CAI, SHU-LIN PENG, LI-SHENG DING, FENG-E WU and YAO-ZU CHEN†

Laboratory of Natural Materia Medica, Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu, 610041, China; † Department of Chemistry, Zhejiang University, 310027, Hangzhou, P.R. China

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Key Word Index—Berneuxia thibetica; Diapensiaceae; triterpenoid saponins; berneuxia saponins A, B, C; desacyl jegosaponin.

Abstract—Four triterpenoid saponins were isolated from *Berneuxia thibetica*. On the basis of chemical and spectroscopic evidence, three new saponins, berneuxia saponins A, B and C, were elucidated as 21-tigloylbarringtogenol C-3 β -O- α -L-rhamnopyranosyl(1 \rightarrow 2)- β -D-galactopyranosyl(1 \rightarrow 3)[β -D-glucopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranosyl(1 \rightarrow 3)[β -D-glucupyranosyl(1 \rightarrow 2)- β -D-glucuronopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranosyl(1 \rightarrow 2)- β -D-glucupyranosyl(1 \rightarrow 3)[β -D-glucupyranosyl(1 β -D-glucupyranosyl(

INTRODUCTION

Berneuxia thibetica Decne, which is widespread in the southwest of China, is used as a Chinese folk medicine for curing coughs due to pathogenic wind-cold factors, overstrain, asthma and dyspea, and wounds [1].We have reported sterols, triterpenes and flavones from this plant [2–4]. We now report the triterpenoid saponins isolated from the leaves.

RESULTS AND DISCUSSION

The crude saponin fractions were subjected to repeated CC on silica gel and silanised silica gel, affording saponins 1, 2, 3 and 4. The yields were 0.008%, 0.006%, 0.01% and 0.004% of the dry leaves, respectively.

On mineral acid hydrolysis, saponin 1 yielded the aglycone 1a, which was identified as 21-tigloylbarringtogenol C by comparison with an authentic sample. This aglycone was also isolated from the ethyl acetate extract of this plant. Four kinds of sugars, glucuronic acid, glucose, galactose and rhamnose, were detected by PC in the aqueous fraction after the removal of the aglycone. The EI-mass spectrum of its acetate showed fragment ions at m/z 273 [terminal rhamnose(Ac)₃]⁺, 331 [terminal glucose (Ac)₄]⁺ and 561 [rhamnose (Ac)₃ galactose(Ac)₃]⁺. The FAB-mass spectrum showed the molecular ion at m/z 1219

 $[M+H]^+$. The negative FAB-mass spectrum showed the fragment ions at m/z 1217 [M-H]⁻, 1071 [M-rhamnose]⁻, 1055 [M-glucose]⁻ and 909 $[M-rhamnose-galactose]^-$. The ¹³C NMR spectrum indicated the presence of four monosaccharide units. On hydrolysis with acid (1 M HCl), 1 gave prosapogenin 1b. The hydrolysis of 1b gave 21-tigloylbarringtogenol C as the aglycone, and D-glucuronic acid and D-glucose as the sugar components. The EI-mass spectrum of the acetate and Me ester of showed fragment ions at 331 [terminal 1b glucose(Ac)₄]⁺, 605 [glucose(Ac)₄ glucuronic acid-OMe (Ac)₂]⁺. The ¹³C NMR spectrum indicated the presence of two monosaccharide units. The glycosylation shift (82.4 ppm) indicated that the β -Dglucuronic acid was 2-O-glycosylated [5]. Therefore, the structure of 1b was 21-tigloylbarringtogenol C-3- $O-\beta$ -D-glucopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranoside. Comparison of the ¹³C NMR signals of 1 with those reported by Calis et al. [6] showed that the sugar moieties were the same. The downfield shift of C-2 of glucuronic acid in 1b (δ 78.6 of 1 to δ 82.4 of 1b) may be caused by 3-O-glycosylation of β -D-glucuronic acid. On the basis of above, saponin 1 was elucidated 21-tigloylbarringtogenol C-3-O-α-L-rhamas nopyranosyl($1 \rightarrow 2$)- β - D - galactopyranosyl($1 \rightarrow 3$) - $[\beta - D - glucopyranosyl (1 \rightarrow 2) - \beta - D - glucuronopyrano$ side], named berneuxia saponin A.

Saponin 2 yielded the aglycone 2a by mineral acid hydrolysis. Compound 2a was identified as 28-tigloylbarringtogenol C by comparison of the 'H NMR signals with those reported [7]. Four sugars, glu-

^{*} Author to whom correspondence should be addressed.



curonic acid, glucose, galactose and rhamnose, were detected by PC in the aqueons fraction after removal of the aglycone. The FAB-mass spectrum showed the molecular ion at m/z 1219 $[M+H]^+$. The negative FAB-mass spectrum showed the fragment ions at m/z $1217 [M - H]^{-}, 1071 [M - rhamnose]^{-}, 1055 [M - glu \cos^{-1}$ and 909 [M – rhamnose – galactose]⁻. The ¹³C NMR spectrum of 2 resembled that of 1, indicating they were only different in the substitution position of tigloyl group. By comparison of ¹³C NMR signals due to aglycone of 1 with those of 2, the obvious difference of chemical shifts occurred at C-16, C-21, C-22, C-28, C-29, indicating that C-28 was substituted by the tigloyl group. Therefore, saponin 2 were identified as 28-tigloylbarringtogenol C-3-O-α-L-rhamnopyranosyl(1 \rightarrow 2)- β -D-galactopyranosyl(1 \rightarrow 3)-[β -D-glucopyranosyl($1 \rightarrow 2$)- β -D-glucuronopyranoside], named berneuxia saponin B.

On mineral acid hydrolysis, saponin 3 yielded the aglycone 3a. There were 30 signals in ¹³C NMR spectrum of 3a. The EI-mass spectrum showed the molecular ion at m/z 472 [M]⁺. According to the EI-mass spectrum and ¹³C NMR spectrum, 3a was identified as 3β , 16α , 28-trihydroxylolean-12-en-21-one, whose EImass spectrum and ¹H NMR spectrum were identical to those of armillarigenin from Jacquinia armillaris [8]. Four sugars, glucuronic acid, glucose, galactose and rhamnose, were detected by PC in the aqueous fraction after removal of the aglycone. The negative FAB-mass spectrum showed the fragment ions at m/z1117 $[M-H]^-$, 971 $[M-rhamnose]^-$, 955 [M-glucose]⁻ and 809 [M - rhamnose - galactose]⁻. By comparison of the ¹³C NMR signals due to the sugar moieties of 3 with those of 1, the sugar moieties were the same. On the basis of above evidence saponin 3 was elucidated as armillarigenin-3- $O\alpha$ -L-rhamnopyranosyl(1 \rightarrow 2)- β -D-galactopyranosyl(1 \rightarrow 3)[β -D-glucopyranosyl(1 \rightarrow 2)- β -D-glucuronopyranoside], named berneuxia saponin C.

Hydrolysis of 4 yielded 4a as the aglycone and glucuronic acid, glucose, galactose and rhamnose as sugars. The EI-mass spectrum of its acetate showed fragment ions at m/z 273 [terminal rhamnose(Ac)₃]⁺, 331 [terminal glucose (Ac)₄]⁺ and 561 [rhamnose(Ac)₃ $galactose(Ac)_3$ ⁺. The negative FAB-mass spectrum showed the fragment ions at m/z 1135 [M-H]⁻, 989 [M-rhamnose]⁻, 973 [M-glucose]⁻ and 827 $[M-rhamnose-galactose]^-$. Compound 4a had the same R_f as the alkaline hydrolysate of 1a on silica HPTLC with different solvent systems. ¹³C NMR signals due to the aglycone were in accord with barringtogenol C except for the position of C-3 [9]. By comparison, of the ¹³C NMR signals due to sugar moieties of 4 with those of 1, the sugar moieties were the same. On the basis of above, 4 was elucidated as barringtogenol C-3-O- α -L-rhamnopyranosyl(1 \rightarrow 2)- β - D - galactopyranosyl(1 \rightarrow 3) - [β - D - glucopyranosyl $(1 \rightarrow 2)$ - β -D-glucuronopyranoside], which was the same as desacyl jegosaponin from Styrax japonica [10].

EXPERIMENTAL

¹H and ¹³C NMR spectra were recorded at 300 MHz in pyridine- d_5 and CDCl₃ using TMS as int. standard. EI-MS was measured at 40 eV accelerating voltage after acetylation. FAB-MS was measured with VG ZAB mass spectrometer. Optical rotations were measured with PE241 automatic recording spectropolarimeter.

Carbon	1*	1a†	1b*	2*	2a†	3*	3a†	4*
1	38.6	38.4	38.8	39.2	38.6	38.7	38.8	38.4
2	26.0	26.5	26.5	26.7	26.4	27.5	27.2	25.9
3	90.2	78.5	89.2	90.8	79.2	90.2	78.9	90.1
4	39.4	38.4	38.8	40.1	38.8	39.5	38.7	39.2
5	55.4	54.9	55.7	56.1	55.2	55.7	55.3	55.4
6	18.1	18.0	18.3	18.9	18.2	18.1	18.3	18.1
7	32.8	32.4	33.0	33.5	32.8	33.0	32.5	32.7
8	39.7	39.4	39.5	40.4	39.7	39.5	40.3	39.6
9	46.6	46.3	46.9	47.3	47.0	48.1	47.1	46.5
10	36.4	36.6	36.7	37.1	37.9	36.7	36.9	36.3
11	23.5	23.2	23.8	24.3	23.7	23.8	23.5	23.4
12	123.7	123.3	122.7	123.8	124.4	123.9	124.1	122.8
13	143.1	141.5	143.5	143.8	141.1	143.1	140.8	143.4
14	41.4	41.0	41.8	42.2	41.3	42.0	41.5	41.5
15	33.8	32.9	34.2	35.1	33.7	34.3	34.4	33.8
16	67.4	68.8	67.8	68.5	68.3	75.6	74.0	67.8
17	47.7	46.3	47.7	47.2	46.7	46.7	46.5	46.8
18	40.1	40.3	40.3	41.3	40.5	42.0	42.5	40.6
19	47.3	46.7	48.0	48.6	47.1	46.9	47.6	4 7.7
20	36.0	35.4	36.3	36.8	36.9	44.7	44.6	35.9
21	81.4	80.8	81.8	78.6	78.9	216.4	209.8	77.7
22	/5.2	/5.8	/5.3	77.4	75.7	40.5	40.3	75.8
23	27.4	28.7	28.0	28.4	28.9	27.8	28.0	27.5
24	10.2	15.2	10.0	1/.1	15.6	16.6	15.8	16.2
25	15.4	15.2	15.0	10.0	15.2	15.6	15.6	15.2
20	26.0	10.2	10.9	17.5	10.9	17.0	17.3	16.5
27	65.9	20.5	65.9	20.2 67 3	27.2 69 1	20.1	20.4	27.0
20	29.6	27.6	20.8	31.0	28.0	03.2	09.5	07.5
30	20.1	19.3	29.8	10.8	20.0	27.8	27.0	30.1
1'	168.4	169.4	168 5	168.4	167.9	23.9	23.0	19.0
2'	129.4	128.4	129.8	129.5	128.7			
3′	135.9	137.5	136.1	137.7	138.0			
4′	12.0	11.7	12.4	12.7	12.2			
5′	14.1	14.0	14.1	14.7	14.2			
glu-1	104.9		105.7	105.5		104.9		104.6
2	78.6		82.4	79.3		78.9		78.6
3	81.1		77.8	82.1		81.7		81.3
4	71.9		73.0	71.9		72.5		72.1
5	75.2		76.8	76.2		75.7		75.3
6	172.2		170.3 52.0	176.9		175.3		175.1
glc-1	101.4		101.6	102.4		101.7		101.5
2	73.1		72.6	74.1		72.9		73.2
3	77.6		78.1	78.6		78.1		77.7
4	70.8		71.5	71.8		71.2		71.3
5	76.5		77.4	76.6		76.8		75.8
6	62.0		62.6	62.8		62.0		61.7
gal-1	101.9			102.8		102.0		101.9
2	11.8			79.0		78.1		78.1
3	/ J.O 70.9			/6.2		76.1		75.3
+ 5	70.8 76.5			/1.8		71.2		71.3
5	70.3 63 1			//.4		/6.8		76.7
u rha-1	100.1			04.0		03.5		63.1
7	72 1			101.2		100./		100.4
<u>-</u> 3	71.8			72.9		12.3		/2.1
4	73 2			74 7		12.3		72.1
5	69.2			70.1		69.6		693
6	17.6			18.6		18 1		177
				- 5.0		10.1		

* pyridine-*d*₅; † CDCl₃.

Plant material

Leaves of *Berneuxia thibetica* Decne. were collected in Xichang of Sichuan Province, China, and identified by Prof. R. N. Zhao. A specimen is deposited in the Herbarium of the Chengdu Institute of Biology, Chinese Academy of Sciences.

Extraction and isolation of saponins

Dry leaves (1.8 kg) were extracted with 95% EtOH. After removal of solvent by evapn, the combined extracts (180 g) were suspended in H₂O, extracted with petrol, EtOAc and n-BuOH successively. The n-BuOH part (45 g) were dissolved with MeOH, precipitated with Et₂O to obtain crude saponins (40 g). The crude saponin fr. was chromatographed on a silica gel column, eluted with CHCl₃-MeOH-H₂O to yield frs 1-46. Fr. 26 (1.12 g) was purified by silanised silica gel 60 to give saponins 1 (150 mg) and 2 (106 mg). Fr. 43 was purified by silanised silica gel 60 to give saponin 3 (202 mg). Fr. 46 was recrystallized to give saponin **4** (80 mg). Compound **1**, $C_{59}H_{94}O_{26}$, $[\alpha]_D^{13} - 12.2^{\circ}$ (MeOH, c 1.1). EI-MS m/z: 273 [terminal rham $nose(Ac)_{3}]^{+}$, 561[rhamnose(Ac)₃galactose(Ac)₃]⁺, 331[terminal glucose(Ac)₃]⁺. FAB-MS m/z: 1219 $[M+1]^+$ (C₅₉H₉₄O₂₆+H). Negative FAB-MS *m*/*z*: 1217 $[M-1]^-$ (C₅₉H₉₄O₂₆-H), 1071 [M-rham-1055 [M-glucose]⁻, 909 [M-rhamnose]⁻, nose-galactose]⁻. Compound 1a was identified as 21-tigloylbarringtogenol C by comparison with an authentic sample. Compound 2, $C_{59}H_{94}O_{26}$, $[\alpha]_D^{26}$ -15.6° (MeOH, c 1.0). Negative FAB-MS m/z: 1217 $[M-1]^{-}$ (C₅₉H₉₄O₂₆-H), 1071 [M-rhamnose]⁻, 1055 $[M-glucose]^-$, 909 $[M-rhamnose-galactose]^-$. Compound 2a, white powder. ¹H NMR (CDCl₃): δ 6.89 (1H, q, J = 6.4 Hz, 3'-H), 5.35 (1H, br s, 12-H), 4.31 (1H, br s, 16-H), 4.09 (1H, d, J = 12.3 Hz, H-28), $3.84 (1H, d, J = 10.5 Hz, 21\alpha - H), 3.75 (1H, d, J = 12.7)$ Hz, H-28), 3.70 (1H, d, J = 10.5 Hz, 22 β -H), 3.23 (1H, *m*, 3 α -H). Compound 3, C₅₄H₈₆O₂₄, [α]¹³_D - 6.8° (MeOH, c 0.75). FAB-MS m/z: 1141 [M+Na]⁺ $(C_{54}H_{86}O_{24} + H)$. Negative FAB-MS m/z: 1117 [M-1]⁻ $(C_{54}H_{86}O_{24}-H)$, 971 [M-rhamnose]⁻, 955 [M-glucose]⁻, 809 [M-rhamnose-galactose]⁻. Compound 3a, colorless needles, mp 296-298 (MeOH) (lit. 299- 301° [9]). EI-MS m/z: 472 [M]⁺, 454, 436, 424, 264(a), 233, 215, 208(b), 190. [']H NMR (CDCl₃): δ 5.44 (1H, t, J = 3.3 Hz, 12-H), 3.85 (1H, m, 16-H), 3.28 (1H, m, 3-H), 3.25 (1H, d, J = 5.0 Hz, 28-H), 3.21 (1H, d, J = 5 Hz, 28-H), 2.52 (2H, d, J = 6 Hz, 22-H), 1.32, 1.25, 1.14, 1.07, 1.00, 0.93, 0.79 (each 3H, s). Compound 4, $C_{54}H_{88}O_{25}$, white powder. EI-MS m/z: 273 [terminal rhamnose(Ac)₃]⁺, 561 [rhamnose(Ac)₃ galactose(Ac)₃]⁺, 331 [terminal glucose(Ac)₄]⁺. FAB-MS m/z: 1159 [M + Na]⁺ (C₅₄H₈₈O₂₅ + Na). Negative FAB-MS m/z: 1135 [M-1]⁻ (C₅₄H₈₈O₂₅-H), 989 $[M-rhamnose]^-$, 973 $[M-glucose]^-$, 827 $[M-glucose]^$ rhamnose – galactose]⁻.

Acid hydrolysis of the saponins and identification of the resulting monosaccharide

Saponin 1 (100 mg) was dissolved in MeOH and heated with 1M HCl/MeOH for 50 min. The soln was evaporated below 30°. The residue was chromatographed on silica gel to obtain 1b (13 mg). Other frs were further hydrolysed with 5% H₂SO₄. The reaction mixture was diluted in H₂O and extracted with CHCl₃. Compound 1a was isolated from the CHCl₃ layer. The aq. layer was neutralized with Ba(OH)₂ and concentrated, then subjected to PC analysis with authentic samples, Developing solvent BuOH–AcOH–H₂O (4:1:5) (upper layer), detection reagent: aniline – phthalate. Each saponin of 2–4 (20 mg) was heated with 5% H₂SO₄. The sugars were detected as above. The aglycones were obtained from CHCl₃ layer.

Acetylation of saponins

To each saponin (5 mg) was added Ac_2O -pyridine (1:1) (0.5 ml) in a microtube. After standing at room temp for 48 h, the soln was evapd to dryness and then subjected to EIMS analysis.

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REFERENCES

- 1. Jiangsu New Medical College, *The Dictionary of Traditional Chinese Medicines*, 1979.
- Ding, L.-S., Wu, F.-E. and Chen, Y.-Z., Zhongguo Zhongyao Zhazi, 1991, 16, 289.
- Wang, M.-K., Wu, F.-E. and Chen, Y.-Z., Yaoxue Xuebao, 1993, 28, 845.
- Wang, M.-K., Peng, S.-L., Ding, L.-S. and Chen, P.-Q., *Tianran Chanwu Yianjiu Yu Kaifa*, 1997, 9, 43.
- Borel, C., Gupta, M. P. and Hostettmann, K., *Phytochemistry*, 1987, 26, 2685.
- Calis, I., Yuruker, A., Ruegger, H., Wright, A. D. and Sticher O., J. Nat. Prod., 1992, 55(9), 1299.
- Schrutka-Rechtenstamm, R., Robien, W. and Jurenitsch J., *Pharmazie*, 1988, 43(3), 208.
- 8. De Maheas, M.R., Billet, D., Raulais, D. and Chaigreau, M., Bull. Soc. Chim. Fr., 1969, (1), 226.
- Massiot, G., Chen, X., Lavaud, C., Le Men-Olivier, L., Delaude, C., Viari, A., Vigny, P. and Duval, J., *Phytochemistry*, 1992, 31, 3571.
- Kitagawa, I., Yoshikawa, M., Kobayashi, K., Imakura, Y., Im. Kwang S. and Ikenishi Y., *Chem. Pharm. Bull.*, 1980, 28, 296.