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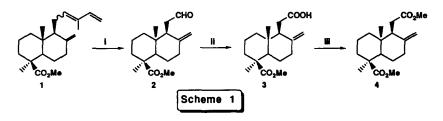
An Efficient Synthesis of the Antifungal Dilactone LL-Z1271a and of other Biologically Active Compounds

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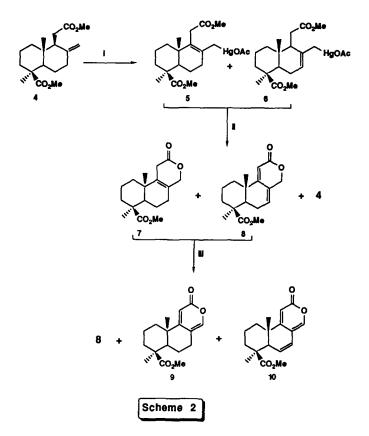
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Abstract: The syntheses of the terpenoid antifungal LL-Z1271 α and other related compounds were achieved starting from the communic acids. The synthetic approach includes a mercuriation-demercuriation reaction of a γ , δ -unsaturated methyl ester in the presence of O₂, giving rise to a lactone ring.

The mold metabolite LL-Z1271 α (14), isolated from the fermentation broths of an Acrostalagmus species, shows antifungal activity *in vitro* against a number of fungi and *in vivo* against some experimental ringworm infections in guinea pigs.¹ Because of this, different procedures for its preparation have been described.² We wish to report herein an improved synthesis of 14 from the readily available carboxilic acid 3,³ using the methyl ester of communic acids 1 as starting material. The esterification of 3 with diazomethane leads to the methyl ester 4 in quantitative yield (scheme 1). In a previous work⁴ we published that the mercuriation reaction in $\Delta^{8(17)}$ of communic acids takes place with the loss of the neighbouring 9-H hydrogen giving rise to the isomerization of the exocyclic double bond. With this in mind, 4 was refluxed for 45 minutes with two equivalents of mercuric acetate in dry toluene (scheme 2), affording a mixture of organomercurials 5 and 6 ⁵ which, after solvent evaporation, was reduced with NaBH4/DMF in the presence of an excess of bubbling O₂, giving lactone 7 (75%), dienolide 8 (15%) and the starting product 4 (5%) .⁶ The dehydrogenation of this mixture was achieved at a 75% yield using DDQ and p-toluenesulfonic acid to give 8, 9 and 10⁷ in the ratio 8:3:1.



i) O3, CH2Cl2, -78°C; S(CH3)2, -78°C to rt, 66.3%; ii) CrO3/H2SO4/H2O, acetone, 90%; iii) CH2N2, Et2O, 100%.

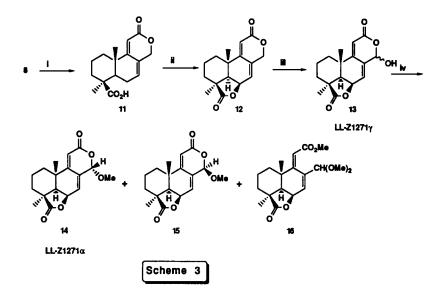


i) Hg(OAc)₂, toluene, reflux, 45min, 100%; ii) NaBH4, O₂, DMF, rt, 90%; iii) DDQ, PTSA, dioxane, reflux, 75%.

The methylester 8 (scheme 3) was transformed by treatment with concentrated sulphuric acid into the free acid 11, and this substance was treated with lead tetraacetate with the aim of closing ring D. The known product 12⁸ was obtained at a 50% yield. The reaction of 12 with three moles of selenium dioxide in refluxing dry dioxane for 1h gave an 85% yield of the lactol 13, identical in properties to LL-Z1271 γ (isolated together with 14 from an *Acrostalagmus* species ^{2b}), and a 15% recovery of 12.

Finally, lactol 13 was treated with methanol/sulphuric acid giving as the main product LL-Z1271 α (14, 50%). In addition, the C-14 epimer 15 (35%) and the dimethyl acetal 16 9 (15%) were isolated from the reaction mixture (PLC).

As outlined in table 1, compounds 12, 13, 14, 15 and 16 were tested against several fungi, 14 and 15 showing a good fungicidal activity. Furthermore, with 14 and 15 protein synthesis blocking assays were performed in an *in vitro* system derived from *S. cerevisiae*, proving that both compounds are good inhibitors of the synthesis of proteins in this assayed system, showing an ED₅₀ 2-6 μ M.



i) H2SO4, rt, 100%; ii) Pb(OAc)4, benzene, hv, 60 h, 50%; iii) SeO2, 3 eq., dioxane, reflux, 1h; iv) CH3OH, H2SO4, rt.

Table 1. Antifungal Activity; MIC (µg/ml):

	Α	В	С	
12	<12.5	<6.25	<3.12	A: Saccharomyces cerevisiae B: Candida albicans C: Cryptococcum neoformans
13	>25	>25	>25	
14	<3.12	<3.12	<3.12	
15	<6.25	<6.25	<3.12	
16	<6.25	>25	<25	

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- 5: ¹H-NMR (400 MHz, CDCl₃): δ 0.70 (s, CH₃-C₁₀), 0.97 (ddd, J = 13.5, 13.5, 4.3 Hz, H₃α), 1.03 (ddd, J = 13.3, 13.5, 4.1 Hz, H₁α), 1.18 (s, CH₃-C₄), 1.34 (dd, J = 12.5, 1.5 Hz, H₅), 1.48 (1H, dm, J = 15 Hz), 1.65 (1H, dm, J = 12.5 Hz), 1.7- 1.8 (2H, m), 1.95- 2.2 (4H, m), 2.03 (s, OAc), 2.32 (d, J = 10.8 Hz, H_{11a}), 2.69 (d, J = 10.8 Hz, H_{11b}), 3.06 (d, J = 16.9 Hz, H_{14a}), 3.17 (d, J = 16.9 Hz, H_{11b}), 3.59 (s, OCH₃-C₁₂), 3.66 (s, OCH₃-C₄). H₇ for 6 can be seen at δ 5.22.
- 6. $7: {}^{1}$ H-NMR (300 MHz, CDCl₃): δ 0.78 (s, CH₃-C₁₀), 1.00 (ddd, J = 13.5, 13.5, 4.3 Hz, H_{3α}), 1.10 (ddd, J = 13, 12.8, 4.2 Hz, H_{1α}), 1.19 (s, CH₃-C₄), 1.33 (dd, J = 12.3, 1.8 Hz, H₅), 1.53 (dm, J = 14.2 Hz, H_{2α}), 1.69 (1H, dm, J = 12.8 Hz), 1.74-1.85 (2H, m), 1.94-1.99 (2H, m), 2.06 (1H, dm, J = 11 Hz), 2.21 (1H, dm, J = 13.5 Hz), 2.87 (dq, J = 20, 2.3 Hz, H_{11a}), 2.95 (dq, J = 20, 2.3 Hz, H_{11b}), 3.61 (s, OCH₃), 4.56 (dm, J = 15 Hz, H_{14α}), 4.63 (dm, J = 15 Hz, H_{14β}); 1³C-NMR (75 MHz, CDCl₃): δ 17.07 (C₂₀), 19.20 (C₂), 20.00 (C₆), 27.75 (C₇), 28.31 (C₁₈), 29.08 (C₁), 35.98 (C₃), 37.49 (C₁₁), 37.60 (C₁₀), 43.72 (C₄), 51.30 (OCH₃), 52.76 (C₅), 71.17 (C₁₄), 124.00 (C₈), 133.72 (C₉), 170.94 (C₁₂), 177.46 (C₁₉).

8 : ¹H-NMR (300 MHz, CDCl₃): δ 0.93 (s, CH₃-C₁₀), 1.08 (ddd, J = 13.5, 13.5, 4 Hz, H_{3α}), 1.22 (s, CH₃-C₄), 1.46 (ddd, J = 13.7, 14.3, 4.2 Hz, H_{1α}), 1.63 (dd, J = 11.7, 4.7 Hz, H₅), 1.65 (dm, J = 13.5 Hz, H_{2α}), 1.92 (ddddd, J = 14.3, 13.5, 13.5, 3.4, 3.4 Hz, H_{2β}), 1.93 (dddd, J = 13.7, 3.5, 1.7, 1 Hz, H_{1β}), 2.24 (dddd, J = 13.5, 3.4, 3.2, 1.7 Hz, H_{3β}), 2.56 (ddd, J = 19.7, 5.7, 5 Hz, H_{6α}), 2.91 (ddm, J = 19.7, 11.7 Hz, H_{6β}), 3.69 (s, OCH₃), 4.78 (dddd, J = 13.2, 2.2, 1.1, 1.1 Hz, H_{14α}), 4.86 (dddd, J = 13.2, 3.7, 2, 2 Hz, H_{14β}), 5.72 (brs, H₁₁), 6.12 (m, H₇); ¹³C-NMR (75 MHz, CDCl₃): δ 19.35 (C₂₀), 19.45 (C₂), 24.88 (C₆), 28.27 (C₁₈), 36.01 (C₁), 37.53 (C₁₀), 37.72 (C₃), 44.22 (C₄), 49.16 (C₅), 51.70 (OCH₃), 69.73 (C₁₄), 109.96 (C₁₁), 125.21 (C₈), 131.11 (C₇), 162.98 (C₉), 165.63 (C₁₂), 177.12 (C₁₉).

7. 9: ¹H-NMR (300 MHz, CDCl₃): δ 1.02 (s, CH₃-C₁₀), 1.08 (ddd, J = 13.5, 13.5, 4.2 Hz, H_{3α}), 1.27 (s, CH₃-C₄), 1.42 (dd, J = 12, 2 Hz, H₅), 1.45 (ddd, J = 13, 13, 4.4 Hz, H_{1α}), 1.66 (dm, J = 14.3 Hz, H_{6α}), 1.80- 2.00 (m, H_{2α}, H_{6β}), 2.07 (dm, J = 13 Hz, H_{1β}), 2.16 (dddd, J = 14.1, 6, 2.3, 2.3 Hz, H_{2β}), 2.28 (dddd, J = 13.5, 3.3, 3.3, 1.5 Hz, H_{3β}), 2.40 (dddd, J = 16, 12.9, 6.1, 2.1 Hz, H_{7α}), 2.70 (ddm, J = 16, 5.3 Hz, H_{7β}), 3.67 (s, OCH₃), 6.20 (s, H₁₁), 7.23 (brs, H₁₄); ¹³C-NMR (75 MHz, CDCl₃): δ 19.62 (C₆), 20.39 (C₂), 22.57 (C₂₀), 25.26 (C₇), 28.53 (C₁₈), 37.32 (C₃), 37.99 (C₁), 39.29 (C₁₀), 44.10 (C₄), 51.32 (C₅), 51.55 (OCH₃), 110.75 (C₁₁), 114.86 (C₈), 147.42 (C₁₄), 163.55 (C₁₂), 167.70 (C₉), 177.20 (C₁₉).

10 : ¹H-NMR (300 MHz, CDCl₃): δ 0.89 (s, CH₃-C₁₀), 1.10 (ddd, J ~ 13.5, 13.5, 4.2 Hz, H₃α), 1.30 (s, CH₃-C₄), 1.50 (ddd, J ~ 14, 13.7, 4.2 Hz, H₁α), 1.72 (dm, J ~ 13.5 Hz, H₂α), 1.90 (ddddd, J ~ 14.3, 13.5, 13.5, 3.4, 3.4 Hz, H₂β), 2.03 (dddd, J ~ 13.7, 3.5, 1.7, 1 Hz, H₁β), 2.21 (dd, J = 3.1, 2.35 Hz, H₅), 2.31 (dddd, J ~ 13.5, 3.4, 3.2, 1.7 Hz, H₃β), 3.67 (s, OCH₃), 6.15 (s, H₁₁), 6.21 (dd, J = 10.0, 3.1 Hz, H₆), 6.44 (dd, J = 10.0, 2.35 Hz, H₇), 7.28 (s, H₁₄).

- 8. This cyclization was assayed by treatment of 11 with the iodosobenzene diacetate-iodine system in cyclohexane, yielding a complex mixture, the major products being those resulting from the decarboxylation of 11 in C-4.
- 9. 16 : ¹H-NMR (300 MHz, CDCl₃): δ 1.13 (s, CH₃-C₁₀), 1.26 (s, CH₃-C₄), 1.86 (d, J = 4.7 Hz, H₅), 2.29 (ddd, J = 14.9, 6.1, 4.1 Hz, H₃β), 3.17 (s, OCH₃), 3.34 (s, OCH₃), 3.73 (s, COOCH₃), 4.99 (ddd, J = 4.7, 4, 1.9 Hz, H₆), 5.34 (dd, J = 2, 1.9 Hz, H₁₄), 5.70 (s, H₁₁), 6.50 (dd, J = 4, 2 Hz, H₇); ¹³C-NMR (75 MHz, CDCl₃): δ 18.24 (C₂), 21.75 (C₂₀), 25.26 (C₁₈), 28.40 (C₃), 32.13 (C₁), 38.59 (C₁₀), 43.02 (C₄), 50.92 (C₅), 51.69 (OCH₃), 52.71 (OCH₃), 53.24 (OCH₃), 72.34 (C₆), 100.56 (C₁₄), 114.65 (C₁₁), 125.86 (C₇), 139.92 (C₈), 153.98 (C₉), 167.81 (C₁₂), 180.59 (C₁₉).

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