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# A CHALCONE GLYCOSIDE FROM CLERODENDRON PHLOMIDIS

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Key Word Index—*Clerodendron phlomidis*; Verbenaceae; flowers; leaves; pectolinarigenin; chalcone glycoside; 7-hydroxyflavone; 7-hydroxyflavanone 7-O-glucoside.

Abstract—A new chalcone glycoside, together with pectolinarigenin, 7-hydroxyflavone and 7-hydroxyflavanone 7-O-glucoside have been isolated from the flowers and leaves of *Clerodendron phlomidis*. The structure of the chalcone glycoside has been established as 4,2',4'-trihydroxy-6'-methoxychalcone 4,4'-D-diglucoside by spectroscopic and degradative methods.

## INTRODUCTION

Clerodendron phlomidis L. (Verbenaceae) is a small tree distributed throughout India and various medicinal properties are attributed to it in the Indian System of Medicine [1]. A number of compounds, namely clerodin,  $\beta$ -sitosterol, lupeol acetate, scutellarein, (24S)-ethyl cholesta, 5,22,25'-triene-3- $\beta$ -ol, D-mannitol, ceryl alcohol, pectolinarigenin, palmitic acid, cerolic acid, apigenin, luteolin and hispidulin [2–7] have earlier been reported from C. phlomidis. We report here the isolation of flavonoids pectolinarigenin and a new chalcone glycoside from flowers and 7-hydroxyflavone and 7-hydroxyflavanone-7-O-glucoside from the leaves of the plant.

### **RESULTS AND DISCUSSION**

The methanolic extract of the flowers of *C. phlomidis* yielded the chalcone glycoside (1), mp 186–188°,  $C_{28}H_{34}O_{15}$ . With Mg/HCl it developed a magenta colour. The UV spectrum showed absorption bands like that of chalcone [8]. It showed peaks in its IR spectrum at 3400 cm<sup>-1</sup> (*br*) for a polyhydroxy system, at 2775 cm<sup>-1</sup> for a methoxyl and at 1645 cm<sup>-1</sup> for a conjugated carbonyl group. On acid hydrolysis it gave glucose and an aglycone (2).

The aglycone (2)  $C_{16}H_{14}O_5$ , mp 257–259°, furnished a diacetate (3), mp 171–173°,  $C_{20}H_{18}O_7$ . The <sup>1</sup>H NMR spectrum of (2) showed a methoxyl group signal ( $\delta$ 3.72), a typical four peak pattern doublet for 4'-oxygenated B ring ( $\delta$ 7.25, 6.80, J = 9 Hz each), two meta coupled doublets ( $\delta$ 5.83, 6.02, J = 2.2 Hz each) and ABX pattern of protons ( $\delta$ 2.70, m and 5.25 1H, dd, J = 5 Hz) like that of naringenin-5-methyl ether [9] (lit. mp 258–261°, diacetate, mp 171.5–173°). The mass spectrum showed

molecular ion peak at m/z 286.0946 and had characteristic ion peaks (m/z 166 and m/z 120) due to retro-Diels-Alder type fragmentation indicating the presence of methoxyl group in ring A. These data suggest that 2 is naringenin 5-methyl ether.

The <sup>13</sup>C NMR of 1 showed signals at  $\delta$ 101.1 and 101.7 due to anomeric carbons of two D-glucose units. Compound 1 on acetylation (Ac<sub>2</sub>O/Py) gave a nona-acetate (4) ( $\delta$ 1.95–2.42). The <sup>1</sup>H NMR spectrum of 1 showed two anomeric protons at  $\delta$ 5.1 and 5.3 which each appeared as doublets (J = 3 Hz). These data show that 1 contains two D-glucose units which have  $\alpha$ -linkages with the aglycone. The formation of naringenin 5-methyl ether supports the structure of the aglycone as isosalipurpol 6'-methyl ether (5).

The attachment of sugar units at C-4 and C-4', respectively, was apparent from the UV spectrum of 1 which showed a bathochromic shift of 50 nm in presence of NaOMe without increase in intensity and no bathochromic shift with NaOAc. The <sup>1</sup>H NMR spectrum of 1 showed signals for the presence of twelve glucosyl protons ( $\delta$ 3.25–4.0, m), one methoxyl ( $\delta$ 3.80, br s), two anomeric protons, two broad singlets ( $\delta 6.35$ , 6.50), for C-3' and C-5' protons, two broad singlets each ( $\delta 6.50$ , 7.00) for C-3, C-5 and C-2, C-6 protons. It also showed two signals for one proton doublet each (J = 16 Hz) for C- $\alpha$ and C- $\beta$  protons of chalcone skeleton. FAB-MS showed a molecular ion peak at m/z 610 and significant peaks at m/z 287 due to the loss of two glucose units and m/z 168 and 120 due to cleavage of the isosalipurpol 6'-methyl ether unit into two halves. The <sup>13</sup>C NMR data are shown on structure 1. Thus 1 is 4,2',4'-trihydroxy-6'-methoxychalcone  $4.4'\alpha$ -D-diglucoside.

### EXPERIMENTAL

Leaves and flowers of *C. phlomidis* were collected from Varanasi district and verified by the Department of

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Botany, Banaras Hindu University and a specimen sample is kept in the department.

Flowers (250 g) were dried, powdered and successively extracted with EtOAc and MeOH in a Soxhlet extractor. CC resolution on SiO<sub>2</sub> gel of the EtOAc and MeOH extracts furnished pectolinarigenin (31 mg), mp 212–214° (lit. mp 213–215°) [10] and chalcone glycoside (1) (75 mg), respectively. The dried leaves (500 g) similarly extracted with MeOH and chromatographed over SiO<sub>2</sub> gel gave, respectively, 7-hydroxyflavone (42 mg), mp 142–144° [11] and 7-hydroxy-flavanone 7-O-glucoside (53 mg) [11]. The structure of the known compounds were established by a comparison with spectral data with reported data and also by authentic samples.

Chalcone glycoside (1) was crystallized from MeOH as yellow granules (75 mg), mp 186–188°. UV  $\lambda_{max}^{MeOH}$  (nm): 245 sh (log  $\varepsilon$  4.25) and 364 (log  $\varepsilon$  4.40), <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta$ 7.80 (1H, d, J = 16 Hz), 7.75 (2H, m), 7.60 (1H, d, J = 16 Hz), 7.00 (2H, m), 6.50 (1H, br s), 6.35 (1H, br s), 3.25–4.00 (12H, m). The acid hydrolysate showed a single spot on PC which corresponded to D-glucose (Co-PC with authentic samples). Diacetate of aglycone 3, mp 171–173°. UV  $\lambda_{max}$  273 and 320 nm, IR  $\nu_{max}$  1743 and 1682 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$ 2.25 (s, 6H), 2.70 (dd, J = 16, 5 Hz, 1H), 6.31 (d, J = 8 Hz, 1H), 7.44 (d, J = 8 Hz, 1H); (Found C, 64.66, H, 5.28, Calculated for C<sub>20</sub>H<sub>18</sub>O<sub>7</sub>: C, 64.86, H, 4.90%).

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#### REFERENCES

- Kirtikar, K. R. and Basu, B. D. (1975) Indian Medicinal Plants (Blatter, E., Casisus, J. F. and Mhasker, K. S., eds), Vol. III, p. 1947.
- Joshi, K. C., Singh, P. and Mehra, A. (1979) Planta Medica 37, 64.
- Gupta, R. K., Chandra, S. and Mahadevan, V. (1967) Indian J. Pharm. 29, 102.
- Subramanian, S. S. and Nair, A. G. R. (1972) J. Indian Chem. Soc. 49, 1069.
- Subramanian, S. S., Nair, A. G. R. and Vedantham, T. N. C. (1973) Phytochemistry 12, 2078.
- 6. Seth, K. K., Pandey, V. B. and Dasgupta, B. (1982) *Pharmazie* 37, 74.
- Bhakuni, D. S., Srivastava, S. N., Sehgal, S. L. and Kaul, K. N. (1962) J. Sci. Ind. Res. (India) 21, 48.
- 8. Imperato, F. (1978) Phytochemistry 17, 822.
- Maruyama, M., Hayasaka, K. and Sasaki, Shin-ichi (1974) Phytochemistry 13, 286.
- Subramanian, S. S. and Nair, A. G. R. (1972) Phytochemistry 11, 3095.
- Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) The Systematic Identifications of Flavonoids, pp. 64 and 169. Springer, New York.