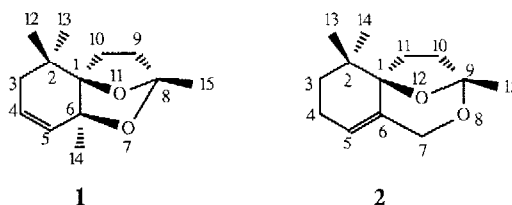


## STRUCTURE AND SYNTHESIS OF TWO NOVEL IONONE-TYPE COMPOUNDS IDENTIFIED IN QUINCE BRANDY (*CYDONIA OBLONGA* MIL.)

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**Summary** Two new ionone-type compounds (**1** and **2**) were isolated from quince brandy (*Cydonia oblonga* Mil.) and their structures deduced from spectroscopic data and confirmed by synthesis.  $^{13}\text{C}$ -,  $^1\text{H}$ -NMR and MS data as well as olfactive properties of **1** and **2** are given.

In the analysis of the brandy of quince fruit [1] [2] (*Cydonia oblonga* Mil.), the two novel acetals **1** [3] [4] and **2** [4] with an ionone skeleton were identified.



The extract, prepared by continuous extraction of the diluted brandy with ether, was separated on silica gel (60, Merck, 0.040-0.063, 230-400 mesh) with pentane/ether = 9/1. The two new acetals were in a fraction with an  $R_f$  = 0.17 (pentane/ether = 9/1). On GC (capillary column, Supelcowax, 60 m, 0.25 mm ID, 80-220°C, 5°/min, 1.5 kg He) **1** eluted after 18'20" and **2** after 23'40". The  $^1\text{H}$ -NMR spectrum (Bruker WH 360,  $\text{CDCl}_3$ , ppm, J in Hz) of compound **1** [5] showed four tertiary methyl groups ( $\delta$  = 0.97, 1.13, 1.37, 1.52), two of which seemed to be adjacent to oxygen, two olefinic protons of one double bond ( $\delta$  = 5.36, 5.53) attached to a quaternary centre on one side and a methylene group on the other side, ( $\delta_A$  = 1.74,  $\delta_B$  = 2.28). The number of hydrogen atoms and the mass spectral data (Finnigan 4021 C, 70 eV, m/e, % of base peak) suggested a  $\text{C}_{13}\text{H}_{20}\text{O}_2$  compound (four parameters). The  $R_f$  was in accord with a diether or an acetal, and in the NMR no hydrogen atoms adjacent to oxygen were visible. Thus the structure shown was in good agreement with spectral data and was confirmed by synthesis. The relative stereochemistry shown with the oxygen atoms in *cis*-configuration was the more stable one and was corroborated by nOe (irrad.  $\text{H}_3\text{-C}(14) \rightarrow$  increases  $\text{H}^1\text{-C}(9)$ ,  $\text{H}^1\text{-C}(10)$  and  $\text{H}_3\text{-C}(13)$ , irrad.  $\text{H}_3\text{-C}(13) \rightarrow$  increases  $\text{H}^1\text{-C}(10)$  and  $\text{H}_3\text{-C}(14)$ ).

The  $^1\text{H}$ -NMR spectrum of **2** [6] revealed three methyl groups as singlets ( $\delta=0.93, 1.01, 1.48$ ), an AB system at  $\delta_{\text{A}}=4.07$  ( $J=13$ ) and  $\delta_{\text{B}}=4.45$  ( $J=13$  with fine splitting), suggesting a cyclic  $\text{CH}_2$ -group attached to oxygen, and a vinyl proton at  $\delta=5.48$ . The number of hydrogen atoms together with the mass spectrum ( $m/e = 208$ ) suggested a  $\text{C}_{13}\text{H}_{20}\text{O}_2$  compound having four parameters. The  $R_f$  on TLC was in agreement with the acetal structure. The much larger synthetic sample also allowed measurement of a  $^{13}\text{C}$  spectrum.

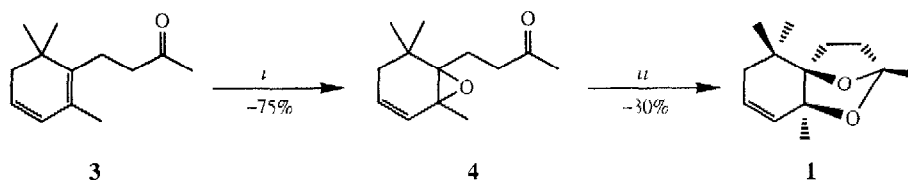
Acetal **1** was synthesized from the known ionone **3** [7] in two steps (*Scheme 1*). Peracetic acid epoxidation with a NaOAc buffer followed by careful neutral work-up allowed isolation of epoxyketone **4** [8] (75% yield). Winterhalter et al. [3], under slightly different conditions, did not obtain this epoxide which is extremely acide-labile and is unstable even with silica gel at room temperature. Dilute sulfuric acid in acetone transformed epoxide **4** directly into acetal **1** (30% yield, remainder polymers) [9][5].

Acetal **2** was prepared in three steps (*Scheme 2*). 2,6,6-Trimethyl-2-cyclohexenone (**5**) [10] was oxidized with  $\text{SeO}_2$  to yield, in agreement with Sharpless' mechanism [11], acetoxyketone **6** as the main product which was isolated in pure form by column chromatography. Reaction of **6** with an organolithium reagent (prepared from 2-(2-bromoethyl)-1,3-dioxalan **7** [13] by metal-halogen exchange with *tert*-butyllithium, see also [14]) gave the key intermediate **8** [15] possessing the correct carbon skeleton with all oxygen functions at the right place. It is not stable in the GC and was purified by column chromatography (49% yield). Mild acid treatment of **8** cleaved the ethylene glycol acetal and formed the internal acetal **2** which was isolated by bulb-to-bulb distillation at  $100\text{--}110^\circ/14$  Torr (50% yield) [9][6].

The odour of our repetitively purified synthetic sample of **1** is rather weak, vaguely woody and camphoraceous, as opposed to Winterhalter's sample which he describes as "highly fruity with an ionone-like aroma" [3]. Acetal **2** exhibits a camphoraceous, herbal, slightly floral note.

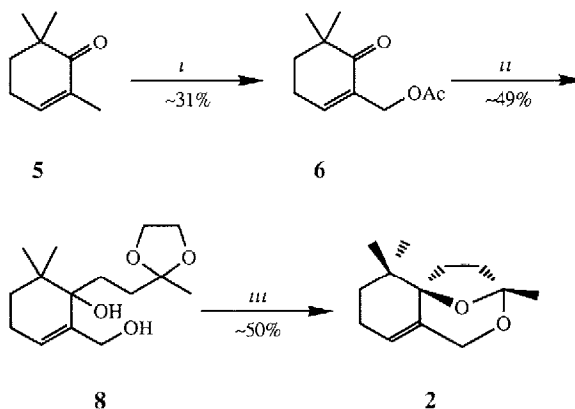
**Acknowledgement** We wish to thank Mr. W. Thommen and Mr. R. Brauchli for the measurements of the NMR spectra, Dr. P. A. Blanc, Firmenich SA, for the perfumistic evaluations, and Drs. B. Maurer and A. F. Thomas, Firmenich SA, for helpful discussions.

*Scheme 1*



Reagents: *i*) 40%  $\text{AcOOH}$  in  $\text{AcOH}/\text{CH}_2\text{Cl}_2/\text{NaOAc}/20^\circ/4\text{h}$ , *ii*) aq. 1N  $\text{H}_2\text{SO}_4$ -acetone 1/1/2h RT

Scheme 2



Reagents i)  $\text{SeO}_2/\text{AcOH}-\text{Ac}_2\text{O}$  3:1/7h reflux ii)  $\text{Br}-\text{CH}_2-\text{CH}_2-\text{O}-\text{CH}_2-\text{CH}_2-\text{Br}$  /  $\text{Li}^+$  / THF-Et<sub>2</sub>O-pentane 2:5:1/-95°/1h, then addition of **6** /-90°/1h→ RT, iii) aq 70%  $\text{HClO}_4$ -H<sub>2</sub>O-THF 3:3:5/R1/3h

### REFERENCES AND NOTES

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- [3] Compound **1** was independently found in Riesling wine by P Winterhalter, Institut für Lebensmittel-Chemie, Universität Würzburg, Germany, P Winterhalter, M A Sefton and P J Williams, *Chem Ind (London)* **1990**, 463
- [4] Chemical Abstract nomenclature  
2,2,6,8-tetramethyl-7,11-dioxatricyclo[6.2.1.0<sup>1,6</sup>]undec-4-ene (**1**)[3] and 2,2,9-trimethyl-8,12-dioxatricyclo[7.2.1.0<sup>1,6</sup>]dodec-5-ene (**2**)
- [5] Spectral data of **1**  
MS 208 (0.5, M<sup>+</sup>), 193 (0.5), 180 (0.5), 175 (0.5), 165 (2), 148 (17), 138 (47), 125 (37), 123 (34), 119 (13), 109 (11), 95 (10), 43 (100) <sup>1</sup>H-NMR (360 MHz) 0.97 (s, 3H), 1.13 (s, 3H), 1.37 (s, 3H), 1.52 (s, 3H), 1.74 (dd, J<sub>1</sub>=17.5, J<sub>2</sub>=5.5, 1H), 1.76-2.00 (m, 4H), 2.28 (br ddd, J<sub>1</sub>=17.5, J<sub>2</sub>=3.0, J<sub>3</sub>=2.1, 1H), 5.37 (dd, J<sub>1</sub>=10.2, J<sub>2</sub>=3.0, 1H), 5.54 (ddd, J<sub>1</sub>=10.2, J<sub>2</sub>=5.5, J<sub>3</sub>=2.1, 1H) nOe, F<sub>2</sub> H<sub>3</sub>-C(12) → H', H"-C(3), H"-C(10) and H<sub>3</sub>-C(13), H<sub>3</sub>-C(13) → H'-C(10), H"-C(3), H<sub>3</sub>-C(12) and H<sub>3</sub>-

- C(14), H<sub>3</sub>-C(14) → H-C(5), H'-C(9), H'-C(10) and H<sub>3</sub>-C(13) <sup>13</sup>C-NMR (90 MHz) (of synth sample) 131.4 (d, C(5)), 123.7 (d, C(4)), 106.9 (s, C(8)), 91.7 (s, C(1)), 79.4 (s, C(6)), 37.9 (t, C(3)), 37.5 (t, C(9)), 33.3 (s, C(2)), 25.4 (q, C(12)), 24.3 (2q, C(13), C(14)), 24.1 (t, C(10)), 19.5 (q, C(15))
- [6] Spectral data of **2**  
 MS 208 (5, M<sup>+</sup>), 193 (3), 190 (1), 180 (3), 178 (12), 165 (8), 151 (5), 133 (17), 124 (12), 105 (21), 95 (29), 81 (15), 67 (18), 43 (100) <sup>1</sup>H-NMR (360 MHz) 0.93 (s, 3H), 1.01 (s, 3H), 1.36 (ddd, J<sub>1</sub>=14, J<sub>2</sub>=6, J<sub>3</sub>=5, 1H), 1.48 (s, 3H), 1.62 (ddd, J<sub>1</sub>=14, J<sub>2</sub>=7, J<sub>3</sub>=6, 1H), 1.73 (ddd, J<sub>1</sub>=13, J<sub>2</sub>=9, J<sub>3</sub>=5, 1H), 1.91 (ddd, J<sub>1</sub>=13, J<sub>2</sub>=13, J<sub>3</sub>=5, 1H), 2.03 (m, 1H), 2.10 (ddd, J<sub>1</sub>=13, J<sub>2</sub>=13, J<sub>3</sub>=5, 1H), 2.21 (ddd, J<sub>1</sub>=2, J<sub>2</sub>=2, J<sub>3</sub>=2, 1H), 4.07 (d, J=13, 1H), 4.45 (br d, J=13, 1H), 5.48 (br s, 1H) <sup>13</sup>C-NMR (90 MHz) of synth sample 135.3 (s, C(6)), 119.6 (d, C(5)), 106 (s, C(5)), 85.7 (s, C(1)), 65.6 (t, C(7)), 34.9 (t, C(10)), 34.7 (t, C(3)), 33.8 (s, C(2)), 29.6 (t, C(11)), 24.3 (q, C(15)), 23.7 (q, C(14)), 22.8 (q, C(13)), 21.9 (t, C(4))
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- [8] Spectral data of **4**  
 MS 208 (1, M<sup>+</sup>), 190 (4), 172 (9), 157 (13), 147 (10), 138 (16), 131 (32), 119 (24), 109 (30), 91 (24), 77 (18), 67 (13), 55 (15), 43 (100) <sup>1</sup>H NMR (360 MHz) 0.93 (s, 3H), 1.09 (s, 3H), 1.35 (s, 3H), 1.59 (dd, J<sub>1</sub>=7, J<sub>2</sub>=17, 1H), 1.96 (ddd, J<sub>1</sub>=7, J<sub>2</sub>=9, J<sub>3</sub>=15, 1H), 2.05-2.15 (m, 2H), 2.16 (s, 3H), 2.56 (dd, J<sub>1</sub>=7, J<sub>2</sub>=9, 2H), 5.68 (dd, J<sub>1</sub>=10, J<sub>2</sub>=3, 1H), 5.76 (ddd, J<sub>1</sub>=2, J<sub>2</sub>=7, J<sub>3</sub>=10, 1H)
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- [12] Spectral data of **6**  
 MS 196 (1, M<sup>+</sup>), 153 (37), 140 (27), 121 (3), 112 (12), 98 (100), 93 (15), 84 (8), 70 (15), 53 (13), 43 (83) <sup>1</sup>H-NMR (360 MHz) 1.13 (2s, 6H), 1.85 (t, J=6, 2H), 2.08 (s, 3H), 2.44 (dt with fs, J<sub>1</sub>=6, J<sub>2</sub>=4, 2H), 4.73 (s with fs, 2H), 6.89 (t, J=4, 1H)
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- [15] Spectral data of **8**  
 MS 252 (1, M<sup>+</sup>), 235 (0.5), 214 (5), 190 (8), 175 (7), 157 (9), 147 (11), 131 (17), 119 (18), 105 (26), 87 (100), 67 (14), 55 (16), 43 (72) <sup>1</sup>H-NMR (360 MHz) 0.98 (2s, 6H), 1.31 (s, 3H), 1.39 (dd with fs, J<sub>1</sub>=7, J<sub>2</sub>=11, 1H), 1.64-2.19 (M, 7H), 3.96 (M, 5H), 4.36 (d, J=11, 1H), 5.71 (dd, J<sub>1</sub>=3, J<sub>2</sub>=3, 1H)

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