

24 β -METHYLCHOLESTA-5,22E,25-TRIEN-3 β -OL AND 24 α -ETHYL-5 α -CHOLEST-22E-EN-3 β -OL FROM *CLERODENDRUM FRAGRANS*

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Abstract—Two minor sterols isolated from *Clerodendrum fragrans* were identified as 24 β -methylcholesta-5,22E,25-trien-3 β -ol and 24 α -ethyl-5 α -cholest-22E-en-3 β -ol of which the former has so far been detected only in a marine sponge. The other sterols identified in the plant were clerosterol, 22E-dehydroclerosterol and several other common sterols.

INTRODUCTION

Plants of the *Clerodendrum* species are known to contain 24 β -ethylcholesta-5,25-dien-3 β -ol (**1f**, clerosterol) and 22E-dehydroclerosterol (**1g**), 24 β -ethylsterols possessing a Δ^{25} -bond, as the major sterols [1–3]. This constitutes the characteristic feature of this species since the great majority of higher plants contain 24 α -alkylsterols (24R if a saturated or Δ^{25} -unsaturated sterol, 24S if the Δ^{22} derivative) which lack a Δ^{25} -bond as the major sterols represented by 24 α -ethylcholest-5-en-3 β -ol (24 α -**1d**, sitosterol) [4]. Thus, we considered it worthwhile to undertake detailed analysis on the sterol constituents of *Clerodendrum* species. This paper describes the thorough investigation on the sterols of the leaves and stems of *C. fragrans* (Vent.) R. Br. which led to the isolation and identification of two very rare sterols, 24 β -methylcholesta-5,22E,25-trien-3 β -ol (**1c**) and 24 α -ethyl-5 α -cholest-22E-en-3 β -ol (24 α -**2e**), in addition to 24 α -**1d**, **1f**, **1g** and several other sterols.

RESULTS AND DISCUSSION

Two sterols **1c** and **2e** were isolated as the acetyl derivatives from *C. fragrans* by virtue of the procedure described in the Experimental section. The mass spectrum of **1c**-acetate showed $[M - \text{HOAc}]^+$, the highest mass ion, at m/z 378 ($\text{C}_{28}\text{H}_{42}$) and other prominent fragmentation ions at m/z 363 $[M - \text{HOAc} - \text{Me}]^+$, 282 $[M - \text{C}_7\text{H}_{12}$ (C-20, 22 vinylic cleavage with 1H transfer) - HOAc] $^+$, 255 $[M - \text{C}_9\text{H}_{15}$ (side chain) - HOAc] $^+$, 253 (255 - 2H) and 213 $[M - \text{C}_9\text{H}_{15} - \text{C}_3\text{H}_6$ (part of ring D) - HOAc] $^+$ indicating that it was an acetate of a C_{28} -sterol with three double bonds, one of which was in the skeleton and the other two were in the C_9 side chain [5–7]. Lack of an ion corresponding to the molecular ion suggests that the skeletal double bond was located at C-5 [5], whereas the ion at m/z 282 indicates one of the two side chain double bonds was located at C-22 [7]. The ^1H NMR spectrum (Table 1) of **1c**-acetate displayed the signals at δ 0.690 (3H, s, 18-H₃), 1.020 (3H, s, 19-H₃), 2.032 (3H, s, 3 β -OAc), 1.012 (3H, d, 21-H₃), 4.60 (1H, m, 3 α -H)

and 5.37 (1H, m, 6-H), which are characteristic for Δ^5 -3 β -yl acetate [2, 7, 8]. Other signals observed were a multiplet around δ 5.24 (2H), which can be assigned to the 22-H and 23-H olefinic protons, and two broad singlets at δ 4.692 and 4.707 (each 1H), typical for a terminal methylene group [2, 7, 8]. A doublet at δ 1.080, highly deshielded by its surrounding two double bonds, is due to the methyl protons at C-28. An isolated multiplet at δ 2.71 (1H), shifted considerably downfield, is attributed to the proton at C-24 which is located in a bis-allylic position. Irradiation of this multiplet collapsed the doublet at δ 1.080 (28-H₃) into singlet, and also simplified the multiplet at δ 5.24 (22-H, 23-H). Finally, a methyl singlet at δ 1.677 is ascribed to the allylic methyl protons at C-27 [2, 7, 8]. The stereochemistry at C-22 was established to be *E* (*trans*) since the 21-H₃ doublet is displayed at δ 1.00–1.01 when it occurs together with a *E*- Δ^{22} double bond, whereas a *Z* (*cis*) double bond shifts it to δ 0.94–0.95 [9]. From the foregoing, **1c**-acetate was regarded as 24-methylcholesta-5,22E,25-trien-3 β -yl acetate. Upon hydrolysis, the acetate gave free sterol **1c** ($[M]^+$, m/z 396; $\text{C}_{28}\text{H}_{44}\text{O}$). The ^1H NMR (Table 1) and MS data of **1c** agreed well with those of 24 β -methylcholesta-5,22E,25-trien-3 β -ol cited in the literature [7], and hence **1c** was confirmed to be 24 β -methylcholesta-5,22E,25-trien-3 β -ol. The 24 β -configuration was supported from the ^1H NMR comparison (Table 1) of the hydrogenated **1c**-acetate, i.e., 24 β -methylcholest-5-en-3 β -ol (24 β -**1b**, 24 β -methylcholesterol) acetate ($[M - \text{HOAc}]^+$, m/z 382), with both of the C-24 epimers of **1b**-acetate.

The mass spectrum of **2e**-acetate showed $[M]^+$ at m/z 456, corresponding to $\text{C}_{31}\text{H}_{52}\text{O}_2$, accompanied with fragmentation ions at m/z 441 $[M - \text{Me}]^+$, 396 $[M - \text{HOAc}]^+$ and 315 $[M - \text{C}_{10}\text{H}_{19}$ (side chain) - 2H] $^+$ indicating that it was an acetate of a C_{29} -sterol with one double bond in the C_{10} side chain [5, 6]. Other ions at m/z 413 $[M - \text{C}_3\text{H}_7$ (allylic cleavage of the terminal isopropyl group)] $^+$, 353 (413 - HOAc) and 344 $[M - \text{C}_8\text{H}_{16}$ (C-20, 22 vinylic cleavage with 1H transfer)] $^+$ suggest that the side chain double bond was located at C-22 [5, 10]. The ^1H NMR spectrum (Table 1) of **2e**-acetate showed signals

Table 1. ¹H NMR data of some sterols isolated from *Clerodendrum fragrans* (400 MHz, CDCl₃, TMS as int. standard)

Acetate	18-H ₃ (s)	19-H ₃ (s)	21-H ₃ (d)	26-H ₃ (d)	27-H ₃ (d)	28-H ₃ (d)	29-H ₃ (t)	3β-OAc (s)	3α-H (m)	6-H (m)	22-H (dd)	23-H (dd)	24-H (m)
24α			0.911(6.6)†	0.851(7.2)	0.803(6.6)	0.774(6.6)			4.60	5.37			
1b*	0.679	1.018						2.030					
24β			0.919(6.5)	0.855(6.6)	0.785(7.7)	0.775(7.7)			4.60	5.37			
24β-1b†	0.675	1.018	0.920(6.6)	0.855(6.6)	0.784(6.6)	0.777(6.6)		2.031					
1c(24β)	0.690	1.020	1.012(6.6)	1.677(s)	4.692(1H, s) 4.707(1H, s)	1.080(6.6)		2.032	4.60	5.37	5.24(2H, m) 5.24(2H, m)		2.71
1c(24β)	0.693	1.010	1.012(6.6)	1.675(s)	4.690(1H, s)	1.080(7.1)			3.52	5.35	5.24(2H, m)		2.71
(3β-OH)					4.705(1H, s)								
1c(24β)§	0.692	1.010	1.012(6.4)	1.676(s)	4.692(1H, s)	1.080(6.9)			3.52	5.35	5.25(2H, m)		2.71
(3β-OH)					4.705(1H, 1.3)								
24α-1d	0.679	1.020	0.922(6.6)	0.837(7.7)	0.815(6.6)		0.847(7.7)	2.032	4.60	5.37			
24α-1e	0.696	1.020	1.021(6.4)	0.846(6.4)	0.796(7.1)		0.804(7.2)	2.035	4.60	5.37	5.013 (8.8, 15.1)	5.154 (8.8, 15.1)	
1f(24β)	0.669	1.016	0.904(6.6)	1.565(s)	4.640(1H, 2.8) 4.726(1H, s)		0.801(7.4)	2.031	4.60	5.37			
1g(24β)	0.690	1.019	1.008(5.5)	1.646(s)	4.695(2H, 0.9)		0.831(7.3)	2.023	4.60	5.37	5.18(2H, m)		
24α-2d	0.647	0.818	0.905(6.6)	0.832(7.1)	0.811(6.0)		0.843(7.3)	2.017	4.68				
24α-2e	0.665	0.819	1.005(6.6)	0.844(6.0)	0.792(7.1)		0.801(7.1)	2.019	4.68		5.005 (8.8, 15.4)	5.145 (8.2, 14.8)	

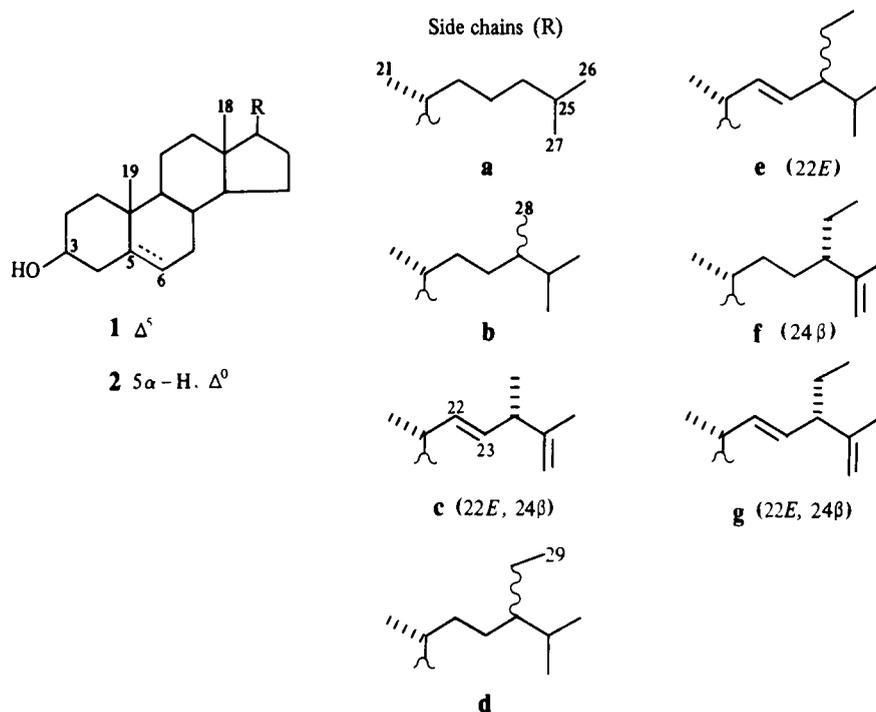
* Mixture of 24α- and 24β-epimers.

† Figures in parentheses denote *J* values (Hz) for doublet and triplet signals.

‡ Prepared from 24β-1c-acetate by hydrogenation.

§ Isolated from *Pseudoaxinella lunacharia* [7] (360 MHz).

|| Prepared from 24α-2e-acetate by hydrogenation.



arising from 3β -acetoxy- 5α -skeleton [δ 0.665 (3H, s, 18-H₃), 0.819 (3H, s, 19-H₃), 2.019 (3H, s, 3 β -OAc) and 4.68 (1H, m, 3 α -H)] [11] and those from 24-ethyl- Δ^{22} side chain [δ 1.005 (3H, d, 21-H₃), 0.844 (3H, d, 26-H₃), 0.792 (3H, d, 27-H₃), 0.801 (3H, t, 29-H₃), 5.005 (1H, dd, 22-H) and 5.145 (1H, dd, 23-H)] [8, 12]. The side chain proton signals were closely correlated with those of 24 α -ethyl-cholesta-5,22E-dien-3 β -ol (24 α -1e, stigmasterol) acetate (Table 1), but differed from those of its 24 β -epimer (24 β -1e-acetate), especially in the chemical shift of 29-H₃ signal [8], which made it possible to determine the configuration at C-24 of 2e-acetate to be α . Thus, 2e was 24 α -ethyl-5 α -cholest-22E-en-3 β -ol (24 α -2e). The 24 α stereochemistry was supported from the following evidence. Hydrogenation of 2e-acetate afforded 24 α -ethyl-5 α -cholestan-3 β -ol (24 α -2d) acetate ([M]⁺, *m/z* 458). The ¹H NMR data of the side chain proton signals of 24 α -2d-acetate were closely correlated with those of 24 α -1d-acetate (Table 1) but differed enough for differentiation from those of its 24 β -epimer [8].

In addition to the above, six sterols were isolated as the acetyl derivatives from *C. fragrans* which were identified as the acetates of cholesterol (1a), a mixture of 24 α - and 24 β -1b (24 α :24 β = 7:3), 1d, 1e, 1f and 1g. Identification of these sterols was performed based on the GC, ¹H NMR [8] and MS data with the exception of 1a-acetate which was identified by GC and MS. Composition of *C. fragrans* sterols was determined on the basis of argentation TLC, GC and ¹H NMR data as the acetyl derivatives as follows: 1a (0.7%), 24 α -1b (1.1%), 24 β -1b (0.4%), 1c (24 β) (2.3%), 24 α -1d (1.8%), 24 α -1e (6.4%), 1f (24 β) (17.3%), 1g (24 β) (67.6%) and 24 α -2e (2.4%).

This study has, thus, demonstrated the occurrence of two very rare sterols, 1c and 24 α -2e, in addition to several other sterols in *C. fragrans*. Sterol 1c has heretofore been reported to occur only in a marine sponge, *Pseudoaxinella lunachatra* [7], and this study seems to be the first instance

for its detection in a plant. Although sterol 2e (24 ξ) has previously been detected in three plants: *Bupleurum falcatum* [13], *Dictyostelium discoideum* [14] and *Lycopersicon esculentum* [15], this study is considered to be the first case of its unambiguous characterization including the determination of the stereochemistry at C-24.

EXPERIMENTAL

Mp: uncorr. Argentation TLC: silica gel-AgNO₃ (4:1) developed \times 3 with CCl₄-CH₂Cl₂ (5:1); HPLC: I: Partisil 5 ODS-2 column (Whatman; 25 cm \times 10 mm i.d.) or II: Altex Ultrasphere ODS column (Beckman; 5 μ m; 25 cm \times 10 mm i.d.), MeOH as mobile phase (flow rate, 4 ml/min) in both systems; GC: OV-17 SCOT glass capillary column (30 m \times 0.3 mm i.d.), column temp. 260°. RR, on HPLC and GC expressed relative to cholesterol (1a) acetate. EIMS (70 eV); probe; ¹H NMR: 400 MHz, CDCl₃, TMS as int. standard; Acetylation: Ac₂O-pyridine at room temp. overnight; Hydrolysis: 5% KOH in EtOH at room temp. overnight; Hydrogenation: EtOH over pre-reduced PtO₂ at atoms. pressure and temp. overnight. The acetates of following sterols: 1a, a mixture of 24 α - and 24 β -1b, 24 α - and 24 β -1d, 24 α - and 24 β -1e, 1f, 1g and 2d, were used as the reference specimens [8, 12]. Leaves and stems of *C. fragrans* were collected locally in India.

Isolation of sterols. Air-dried and powdered leaves and stems (950 g) of *C. fragrans* were extracted with MeOH in a Soxhlet extractor for 48 hr. After removal of the solvent, extracted lipid (70 g) was treated with cold Me₂CO. The Me₂CO soluble part (37 g) was refluxed with 5% KOH in EtOH for 2 hr and then extracted with Et₂O which gave unsaponifiable lipid (11.0 g). CC of the unsaponifiable lipid on silica gel (200 g) [hexane (1.3 l), hexane-Et₂O (9:1, 1 l), hexane-Et₂O (6:1, 1 l), hexane-EtOAc (4:1, 1 l) and then MeOH (1.5 l) as eluants] gave the sterol mixture (810 mg) [eluted with hexane-EtOAc (6:1)]. (The elution was monitored by TLC on precoated silica gel.) This was

acetylated, and the acetate mixture (843 mg) was subjected to argentation TLC to give four bands (referred to as bands 1–4 in the order of polarity, beginning with the least polar). The least polar fraction (12.5 mg) from band 1 (R_f 0.70–0.78) was a mixture which on further argentation TLC afforded two fractions: fraction 1A (2.5 mg) from the less polar band and fraction 1B (4.3 mg) from the more polar band. Fraction 1A was 24 α -2e-acetate. Fraction 1B was a mixture of three components from which was isolated the acetates of **1a**, **1b** (C-24 epimeric mixture) and 24 α -1d by HPLC. The fractions from band 2 (R_f 0.63–0.70) and 3 (R_f 0.41–0.5), on further argentation TLC, gave 24 α -1e-acetate (30.1 mg) and **1f**-acetate (89.8 mg), respectively. A fraction (215.4 mg) from the most polar band 4 (R_f 0.14–0.41) was a mixture of two steryl acetates, and a portion (90 mg) of this fraction was subjected to HPLC giving **1c**-acetate (1.0 mg) and **1g**-acetate (48.8 mg).

24 β -Methylcholesta-5,22E,25-trien-3 β -ol (**1c**) acetate Mp 149.1–151.2°; RR_t = 1.20 (GC), 0.71 (HPLC-I); MS: m/z 378.3241 ($[M - HOAc]^+$, C₂₈H₄₂, rel. int. 100%, requires 378.3284), 363.3034 (C₂₇H₃₉, 7%), 282.2371 (C₂₁H₃₀, 7%), 255.2124 (C₁₉H₂₇, 26%), 253.1941 (C₁₉H₂₅, 13%), 213.1656 (C₁₆H₂₁, 7%).

24 β -Methylcholesta-5,22E,25-trien-3 β -ol (**1c**). Mp 133.0–134.0°; MS: m/z 396.3401 ($[M]^+$, C₂₈H₄₄O, rel. int. 20%, requires 396.3390), 381.3168 (C₂₇H₄₁O, 3%), 378.3266 (C₂₈H₄₂, 5%), 363.3056 (C₂₇H₃₉, 4%), 326.2664 (C₂₃H₃₄O, 3%), 314.2574 (C₂₂H₃₄O, 5%), 309.2598 (C₂₃H₃₃, 6%), 300.2431 (C₂₁H₃₂O, 30%), 285.2327 (C₂₀H₂₉O, 12%), 283.2407 (C₂₁H₃₁, 9%), 271.2059 (C₁₉H₂₇O, 76%), 255.2091 (C₁₉H₂₇, 48%), 253.1950 (C₁₉H₂₅, 8%), 241.1988 (C₁₈H₂₅, 4%), 239.1910 (C₁₈H₂₃, 5%), 229.1998 (C₁₇H₂₅, 6%), 215.1791 (C₁₆H₂₃, 13%), 213.1659 (C₁₆H₂₁, 18%), 81.0701 (C₆H₉, 100%).

24 α -Ethyl-5 α -cholest-22E-en-3 β -ol (24 α -2e) acetate. Mp 139.5–142.0°; RR_t = 1.44 (GC); MS m/z 456.3948 ($[M]^+$, C₃₁H₅₂O₂, rel. int. 42%, requires 456.3965), 441.3742 (C₃₀H₄₉O₂, 4%), 413.3386 (C₂₈H₄₅O₂, 4%), 396.3766 (C₂₉H₄₈, 21%), 381.3586 (C₂₈H₄₅, 3%), 358.2824 (C₂₄H₃₈O₂, 7%), 353.3209 (C₂₆H₄₁, 31%), 344.2743 (C₂₃H₃₆O₂, 32%), 329.2449 (C₂₂H₃₃O₂, 8%), 316.2414 (C₂₁H₃₂O₂, 27%), 315.2289 (C₂₁H₃₁O₂, 37%), 302.2288 (C₂₀H₃₀O₂, 10%), 257.2264 (C₁₉H₂₉, 58%), 255.2086 (C₁₉H₂₇, 18%), 241.1950 (C₁₈H₂₅, 7%), 229.1991 (C₁₇H₂₅, 10%), 215.1797 (C₁₆H₂₃, 16%), 43.0540 (C₃H₇, 100%).

Physical properties of the other *C. fragrans* sterols. Acetyl derivatives. **1a**: Mp 117.0–119.0°, RR_t = 1.00 (GC), 1.00 (HPLC-I)

and -II); mixture of 24 α - and 24 β -**1b**: mp 139.1–140.2°; RR_t = 1.31 (GC), 1.14 (HPLC-II); 24 β -**1b** (prepared from 24 β -**1c**-acetate by hydrogenation): mp 144.5–148.0°; RR_t = 1.31 (GC); 24 α -**1d**: mp 122.4–124.0°; RR_t = 1.63 (GC), 1.26 (HPLC-II); 24 α -**1e**: mp 144.0–146.0°; RR_t = 1.43 (GC), 1.06 (HPLC-II); **1f** (24 β): mp 127.4–128.5°; RR_t = 1.64 (GC), 0.86 (HPLC-I); **1g** (24 β): mp 151.2–153.0°; RR_t = 1.49 (GC), 0.80 (HPLC-I); 24 α -**2d** (prepared from 24 α -2e-acetate by hydrogenation): mp 131.0–132.0°; RR_t = 1.66 (GC). For the ¹HNMR data of the *C. fragrans* sterols, see Table 1.

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