

PII: S0957-4166(97)00162-6

Enantioselective synthesis of both enantiomers of 2-amino-6-phosphonohexanoic acid [(R)- and (S)-AP6], a potent and specific agonist of AMPA receptor subtype

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Abstract: The preparation of both enantiomers of 2-amino-6-phosphonohexanoic acid [(R)- and (S)-AP6] is described. The highly diastereoselective alkylation of imidazolidinones 4 and hydrolysis of the alkylated products [(2R,5R,1'S)-6] and (2S,5S,1'S)-6of proceeds under relatively mild conditions to give the physiologically important, enantiopure aminophosphonic acids (R)-AP6 and (S)-AP6. $(\bigcirc$ 1997 Elsevier Science Ltd

Introduction

Excitatory amino acids (EAA) are the most prevalent neurotransmitters in the mammalian central nervous system (CNS).¹ EAA receptors are thought to offer an abundant and varied opportunity to identify compounds useful to explore normal CNS function as well as to develop new therapeutics for the treatment of several pathological conditions affecting the brain, such as Alzheimer's Disease, Parkinsonism, and Huntington's Disease, as well as neuronal damage resulting from cerebral ischemia and epilepsy.²

Several studies have shown that phosphonate analogues of glutamic acid with side chain lengths of four to six carbon atoms are activators for the N-methyl-D-aspartate (NMDA) receptor site; for instance, (R)- and (S)-AP4 (1), (R)- and (S)-AP5 (2) and (RS)-AP6 (3), AP5 being the most potent.



Although a truly specific agonist for the AMPA-sensitized site has not been identified, data from previous studies suggest that the (S)-isomer of AP6 might be a particularly selective and potent agonist. Unlike NMDA receptors, however, the AMPA-sensitized site displays a preference for (S)- over (R)-isomers; thus, it seemed likely that (S)-AP6 might be uniquely selective for this site.³ As a part of our program on the asymmetric synthesis of amino acids, we have recently described the preparation of biologically active α -amino- ω -phosphonocarboxylic acids.⁴ In the present paper we report the synthesis of (R)- and (S)-AP6 in enantiopure form, using imidazolidinones (2R,1'S)-4 and (2S,1'S)-4.

Results and discussion

The modified Seebach imidazolidinones 4 were prepared according to the procedure described by Juaristi *et al.*⁴ In the present work, we used the diastereomeric pair (2R,1'S)-4 and (2S,1'S)-4, resulting from the conversion of (S)- α -methylbenzylamine. Imidazolidinones (2R,1'S)-4 and (2S,1'S)-4 were then treated separately with LDA, and the resulting enolates were added to diethyl 4-bromobutylphosphonate (5). The desired alkylated products **6** were obtained with very high diastereoselectivity (>98%) as determined by ¹H NMR and ¹³C NMR (Scheme 1). The trans relative configuration in the main product was established in analogy to previous work.⁵

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Adducts (2R,5R,1'S)-6 and (2S,5S,1'S)-6 were purified by flash chromatography to afford the pure derivatives in 62.1% and 61.4% yield, respectively. Hydrolysis to (R)-3 and (S)-3 proceeded conveniently under relatively mild conditions with 6N HCl, at 115°C during 16 h. Following extraction with CH₂Cl₂ and treatment with propylene oxide, we obtained enantiopure⁶ (R)-(-)-AP6 and (S)-(+)-AP6, in 96.2% and 91.3% yields, respectively (Scheme 1).

Experimental⁷

Diethyl 4-bromobutylphosphonate (5)

In a 50 mL round-bottom Schlenk flask provided with magnetic stirrer and condenser, was placed 14.4 mL (25.9 g, 0.12 mol) of 1,4-dibromobutane and heated to 80°C before the addition of 5.2 mL (5.0 g, 30.1 mmol) of triethylphosphite. The reaction mixture was stirred at 80°C during 1 h, and then heated to 160°C for 5 h. The crude product was allowed to cool to room temperature and was purified by distillation in a Kugelrohr apparatus, bp 128°C/0.5 mm, as a viscous, colorless liquid in 78.2% yield. ¹H NMR (270 MHz, CDCl₃) δ 1.30 (t, J=7.3 Hz, 6H, CH₂CH₃), 1.66–1.86 (m, 4H, CH₂CH₂P), 1.97 (p, J=6.6 Hz, 2H, BrCH₂CH₂), 3.42 (t, J=6.6 Hz, 2H, BrCH₂), 4.12 (p, J=7.3 Hz, 4H, CH₂CH₃). ¹³C NMR (67.80 MHz, CDCl₃) δ 15.9 (d, ³J_{C/P}=6.1 Hz, CH₂CH₃), 20.6 (d, ²J_{C/P}=4.89 Hz, CH₂CH₂P), 24.12 (d, ¹J_{C/P}=141.6 Hz, CH₂P), 32.50 (d, ³J_{C/P}=14.5, BrCH₂CH₂), 32.85 (s, BrCH₂), 60.9 (d, ²J_{C/P}=6.1 Hz, CH₂CH₃). ³¹P NMR (36.23 MHz, CDCl₃) δ 31.28.

(2R, 5R, 1'S)-1-Carbobenzyloxy-2-tert-butyl-3- $(\alpha$ -methylbenzyl)-5-(diethyl 4-butylphosphonate)-1,3-imidazolidin-4-one [(2R, 5R, 1'S)-6]

In a Schlenk flask was placed 15 mL of dry THF under nitrogen. The flask was inmersed in a dry ice-acetone bath at -78° C and then 0.2 mL (1.45 mmol) of diisopropylamine followed by 0.7 mL (1.45 mmol) of 2.0 M *n*-BuLi was added. The resulting solutions was stirred for 30 min before the addition of 0.5 g (1.32 mmol) of (2*R*,1'*S*)-4 in 15 mL of THF. The resulting enolate solution was stirred 1 h and then 0.4 g (1.45 mmol) of bromide **5** was added. The reaction mixture was stirred for 1 h, quenched with 5 mL of saturated aqueous ammonium chloride, extracted with two 20 mL portions of CH₂Cl₂, dried with Na₂SO₄, and concentrated. The crude product was purified by flash chromatography (Hex-iPrOH, 95:5) to give 0.47 g (62.1% yield) of a colorless semisolid, $[\alpha]_D^{28}$ =+57.0 (c=1, CH₂Cl₂). ¹H NMR (270 MHz, CDCl₃) δ 0.83 (s, 9H, C(CH₃)₃), 1.31 (t, J=7.3 Hz, 6H, CH₂CH₃), 1.72 (d, J=7.3 Hz, CH₃CH), 0.95-2.10 (broad, 8H, CH₂CH₂CH₂CH₂CH₂P), 4.07 (p, J=7.3 Hz, 4H, CH₂CH₃), 4.16 (m, C(5)–H), 4.62 (q, J=7.3 Hz, CH₃CH), 5.08 (s, 2H, CH₂Ph), 5.11 (s, C(2)–H), 7.21–7.45 (m, 10H, H_{arom}). ¹³C NMR (67.80 MHz, CDCl₃) δ 16.50 (d, ³J_C/P=5.6 Hz, CH₂CH₃), 20.53 (s, CH₃CH), 22.30 (d, ²J_C/P=4.4 Hz, CH₂CH₂P), 23.94 (d, ³J_C/P=17.6 Hz, BrCH₂CH₂), 25.62 (d, ¹J_C/P=141.0 Hz, CH₂P), 26.08 (s, C(CH₃)₃), 29.71 (s, BrCH₂), 40.53 (s, C(CH₃)₃), 59.66 (s, CH₃CH), 60.35 (s, C(5)),

61.42 (d, ${}^{2}J_{C/P}$ =6.6 Hz, CH₂CH₃), 67.46 (s, CH₂Ph), 82.21 (s, C(2)), 127.10, 127.37, 128.46, 128.56, 128.67, 128.78, 135.77, 141.22 (C_{arom}), 153.80 (broad, NCOO), 172.86 (s, CO). ³¹P NMR (36.23 MHz, CDCl₃) δ 31.91. Anal. Calcd. for C₃₁H₄₅N₂O₆P: C, 65.02; H, 7.92. Found: C, 64.87; H, 8.27.

(2S,5S,1'S)-1-Carbobenzyloxy-2-tert-butyl-3- $(\alpha$ -methylbenzyl)-5-(diethyl 4-butylphosphonate)-1,3-imidazolidin-4-one [(2S,5S,1'S)-6]

The same procedure described for the preparation of (2R,5R,1'S)-6 was followed with 0.5 g (1.32 mmol) of (2S,1'S)-4, to afford 0.46 g (61.4% yield) of the desired product as a white solid, mp 64–65°C, $[\alpha]_D^{28}=-22.7$ (c=1.67, CH₂Cl₂). ¹H NMR (270 MHz, CDCl₃) δ 1.02 (s, 9H, C(CH₃)₃), 1.30 (t, J=6.6 Hz, 6H, CH₂CH₃), 2.02 (d, J=7.3 Hz, 6H, CH₃CH), 1.08–2.0 (broad, 8H, CH₂CH₂CH₂CH₂CH₂P), 4.03 (p, J=7.3 Hz, 4H, CH₂CH₃), 4.10 (m, C(5)–H), 4.53 (q, J=7.3 Hz, CH₃CH), 5.25 (s, C(2)–H), 5.28 (s, 2H, CH₂Ph), 7.15–7.55 (m, 10H, H_{arom}). ¹³C NMR (67.80 MHz, CDCl₃) δ 16.48 (d, ³J_{C/P}=5.6 Hz, CH₂CH₃), 16.80 (s, CH₃CH), 22.10 (d, ²J_{C/P}=4.4 Hz, CH₂CH₂P), 23.09 (d, ³J_{C/P}=19.9 Hz, BrCH₂CH₂), 25.36 (d, ¹J_{C/P}=141.0 Hz, CH₂P), 26.26 (s, C(CH₃)₃), 32.71 (s, BrCH₂), 41.21 (s, C(CH₃)₃), 57.57 (s, CH₃CH), 60.26 (s, C(5)), 61.24 (d, ²J_{C/P}=5.5 Hz, CH₂CH₃), 67.34 (s, CH₂Ph), 81.10 (s, C(2)), 127.65, 127.97, 128.31, 128.46, 128.59, 135.77, 140.48 (C_{arom}), 154.0 (broad, NCOO), 172.57 (s, CO). ³¹P NMR (36.23 MHz, CDCl₃) δ 31.95.

(R)-(-)-2-Amino-6-phosphonohexanoic acid [(R)-3]

In a glass ampoule provided with magnetic stirrer was placed 0.35 g (0.61 mmol) of (2R,5R,1'S)-6 and 5 mL of 6N HCl. The ampoule was sealed and heated for 16 h in a oil bath at 115°C. The reaction mixture was then allowed to cool to room temperature, extracted with two 10 mL portions of CH₂Cl₂, the aqueous phase was concentrated and the residue suspended in 10 mL of anhydrous hot ethanol, allowed to cool to room temperature and treated dropwise with propylene oxide until the solution became turbid. At this point, the precipitated solid was filtered under vacuum and recrystallized from EtOH/H₂O (1:1) to afford 0.12 g (96.2% yield) of the corresponding (*R*)-AP6 as a white solid, mp 235°C (foam), $[\alpha]_D^{28}$ =-18.0 (c=1, 6N HCl). ¹H NMR (270 MHz, D₂O) δ 1.40–1.85 (m, 6H, CH₂CH₂CH₂P), 1.85–2.15 (m, 2H, CHCH₂), 4.02 (t, J=5.9 Hz, CH). ¹³C NMR (67.80 MHz, D₂O) δ 22.27 (d, ²J_C/P=4.4 Hz, CH₂CH₂P), 25.29 (d, ³J_C/P=16.5 Hz, CH₂CH₂CH₂P), 26.72 (d, ¹J_C/P=133.3 Hz, CH₂P), 29.55 (s, CHCH₂), 53.47 (s, CH), 173.24 (s, CO). ³¹P NMR (36.23 MHz, D₂O) δ 30.74.

(S)-(+)-2-Amino-6-phosphonohexanoic acid [(S)-3]

The same procedure described for the hydrolysis of (2R,5R,1'S)-6 was carried out with 0.3 g (0.52 mmol) of (2S,5S,1'S)-6 and 5 mL of 6N HCl to give 0.1 g (91.3% yield) of (S)-AP6 as a white solid, mp 235°C (foam), $[\alpha]_D^{28}$ =+17.7 (c=1.12, 6N HCl). The ¹H, ¹³C, and ³¹P NMR spectra were similar to those for (R)-3.

Acknowledgements

We are grateful to V. M. González-Díaz for assistance in recording the ³¹P NMR spectra, and to CONACyT for financial support via grant L006-E9607.

References

- 1. Ferkany, J. W.; Willets, J.; Borosky, S. A.; Clissold, D. B.; Karbon, E. W.; Hamilton, G. S. Bioor. and Med. Chem. Lett., 1993, 3, 33.
- a) Collingridge, G. L.; Sawyer, W. *TIPS*, **1990**, *11*, 290. b) Klockgether, T.; Turski, L. Ann. Neurology, **1990**, 28, 529. c) Young, H. B.; Greenamyre, J. T.; Hollingsworth, Z.; Albin, R. I.; D'Amato, C.; Shoulson, I.; Penny, J. B. Science, **1988**, 241, 981. d) Zivin, J. A.; Choi, D. W. Sci. Amer., **1991**, 265, 36. e) Dingledine, R.; McBain, C. J.; McNamara, J. O. *TIPS*, **1990**, *11*, 334.
- 3. Harris, E. W.; Cotman, C. W. Exp. Brain Res., 1983, 52, 455.
- 4. García-Barradas, O.; Juaristi, E. Tetrahedron, 1995, 51, 3423.

- 5. Juaristi, E.; Anzorena, J. L.; Boog, A.; Madrigal, D.; Seebach, D.; García-Baez, E. V.; García-Barradas, O.; Gordillo, B.; Kramer, A.; Steiner, I.; Zurcher, S. J. Org. Chem., 1995, 60, 6408.
- 6. Under similar hydrolytic conditions, the lower homologues (R)- and (S)-AP5 were prepared in enantiopure form.⁴
- 7. For a description of general experimental data, see ref. 4.

(Received in USA 13 March 1997)