# TWO NEW DITERPENE-BASED ALKALOIDS FROM ICACINA GUESFELDTII

PENGE ON'OKOKO and MAURICE VANHAELEN\*

Institut de Pharmacie, Campus Plaine B205-4, Université Libre de Bruxelles, 1050 Brussels, Belgium

(Received 17 May 1979)

Key Word Index-Icacina guesfeldtii; Icacinaceae; leaf; root; lactonic alkaloids; pimarane skeleton.

Abstract—From Icacina guesfeldtii (leaves and roots), two new diterpene-based alkaloids have been isolated and identified as icaceine (2) and De-N-methylicaceine (3). Icacine (1) occurred both in the leaves and roots. Structure determination was performed by spectroscopic and chemical methods. As icacine (1), these two bases are the first alkaloids with a pimarane skeleton isolated from plants.

### INTRODUCTION

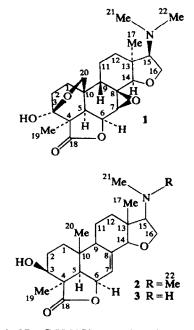
Icacina guesfeldtii Ascher is a shrub endemic to different regions of tropical Africa. Around Lodja (Zaire), the root decoction is used in popular medicine as an anticonvulsant. From a search for the active principle(s), alkaloids were detected in the tuberous roots and in the leaves. A previous study has revealed the presence of icacine (1) in the root; the structure of this new diterpenic alkaloid was established by X-ray analysis [1]. Further investigations, herein reported, led to the isolation and the identification of two additional alkaloids related to the same pimarane skeleton. These compounds, icaceine (2) and De-Nmethylicaceine (3) occur almost exclusively in the leaves. Icacine was detected in larger quantities in the leaves than in the roots.

# **RESULTS AND DISCUSSION**

A chloroform extract of the leaves obtained by a standard extraction procedure afforded a mixture of alkaloids; they were further purified by PLC on alumina to yield three bases.

1 was identical to icacine previously isolated from the root and identified by X-ray analysis [1]. Direct comparison with an authentic sample was carried out by GLC, TLC and by IR, MS and <sup>1</sup>H NMR spectroscopy.

2, icaceine, which afforded by Se dehydrogenation a mixture from which 1,7-dimethylphenanthrene (pimanthrene) was isolated and identified by UV spectroscopy [2], GLC, HPTLC and MS. As 1 and 2 afforded the same dehydrogenation product, a pimarane skeleton was assigned to 2. High resolution MS established the formula as  $C_{22}H_{33}N_1O_4$ ; the base



peak (m/e 87, C<sub>4</sub>H<sub>0</sub>NO) was identical to that of icacine (1) and thus related to the fragmentation of an identical ring D. As in icacine (1), absorption at 1735 cm<sup>-1</sup> suggested the presence of a  $\gamma$ -lactone. Unlike 1, however, the formation of a monoacetate of 2 upon acetylation at room temperature indicated the presence of one secondary alcohol; furthermore, the IR spectrum of 2 showed absorption at 1655 cm<sup>-1</sup> which suggested the presence of a double bond. The absorption at 3020 cm<sup>-1</sup> observed in 1 (epoxide) did not appear in the IR of 2. The <sup>1</sup>H NMR spectrum of 2 confirmed these assignments: it exhibited one olefinic proton at  $\delta$  5.89 (*dd*), one proton (-OH) at 3.64 and one proton on a carbon bearing a hydroxyl at 2.05

<sup>\*</sup> To whom correspondence should be addressed.

(m), which moved downfield to 4.95 as a dd upon acetylation. The C-20 methylene protons shown by 1 ( $\delta$  4.76 and 3.56) were not present, but 2 exhibited a singlet at  $\delta$  1.01 (3H) which can be related to an additional angular methyl. Other signals from ring D were identical to those of icacine. The chemical shifts and couplings of H-14, H-7, H-6 and H-5 were in agreement with the structure proposed for 2; they were similar to those observed for annonalide [3] and nomilactone [4] and further checked by double resonance experiments. Epoxidation of 2 or deoxidation of 1 to correlate both structures were unsuccessful.

However, the stereochemistry of (2) was not completely elucidated, but it was assumed that ring A presented a chair configuration and that the 3-OH was in the  $\beta$ -position: hydrogen bonding between the 3-OH and 18C=O was observed for 2 but not for 1; molecular rotation differencies  $\Delta$  (OAc) –  $\Delta$  (OH) value (-19°) [5], as well as the comparison of the chemical shifts of H-19 exhibited by 1 ( $\delta$  1.38), icacine acetate ( $\delta$  1.42), icaceine (2) ( $\delta$  1.54) and icaceine acetate ( $\delta$  1.38) suggested a 3 $\beta$ -OH, 5 $\alpha$ -H configuration. Furthermore, values of  $J_{5,6}$  and  $J_{6,7}$  identical to earlier published data [3, 4, 6] also supported the proposed stereochemistry of rings A and B.

3, De-N-methylicaceine, on methylation with MeI afforded 2 (MS, <sup>1</sup>H NMR, IR and TLC comparison). On treatment with CD<sub>3</sub>I, 3 afforded a trideuterio derivative whose MS displayed a base peak at m/e 90: this result unequivocally confirmed the origin of the base peak m/e 87 shown by 1 and 2, which is derived from the ring D fragmentation.

The largest accumulation of the alkaloids was found in the leaves, as shown in Table 1; it is reasonable to assume that they were synthetized in the aerial parts of the plant. The occurrence of annonalide in Annona coriacea [3] and momilactone in Oryza sativa [4] indicates that diterpenoids could be precursors of these lactonic alkaloids: further studies on their biogenesis and the isolation of diterpenoid precursors are in progress.

Table 1. Determination of alkaloids in Icacina guesfeldtii (as % dry out)

	Root	Leaf
Icacine (GLC)	0.070	0.180
Icaceine (GLC)	0.002	0.014
De-N-methylicaceine (GLC)	0.003	0.037
Total (titrimetry)	0.091	0.218

#### EXPERIMENTAL

Mps are uncorr. IR spectra were measured in KBr discs. NMR spectra were recorded at 270 MHz in CDCl<sub>3</sub>, using TMS as internal reference; chemical shift values are reported in  $\delta$  (ppm) units. MS were obtained by direct inlet, 70 eV.

Plant material. Roots and leaves of *I. guesfeldtii* were collected around Lodja (Zaire) in September 1977. Plants were identified by Dr. C. Evrard (Université Catholique de Louvain, Belgium). A voucher specimen has been deposited in the Botanical Laboratory of the National University of Zaire (Kinshasa).

Extraction and separation. The air-dried ground leaves or roots were extracted with EtOH. The EtOH-soluble residue

was taken up by CHCl<sub>3</sub> and extracted with 2 N HCl. The combined aq. solns were basified by addition of NH<sub>4</sub>OH and the liberated bases extracted with CHCl<sub>3</sub>. The crude chloroform residue was stirred twice with Me<sub>2</sub>CO at 4° and the yellow solvent discarded after decai. ation of the solid. Further purification was obtained by PLC on neutral Al<sub>2</sub>O<sub>3</sub> (toluene-MeCO-EtOH-NH<sub>4</sub>OH, 40:40:8:3; Dragendorff reagent was used for detection of spots. Three alkaloids were recovered from the leaves (1 kg):  $R_f$  0.60 icacine (1) (1.5 g),  $R_f$  0.79 icaceine (2) (95 mg) and  $R_f$  0.66 De-N-methylicaceine (3) (50 mg).

Alkaloid assays were performed on the crude CHCl<sub>3</sub> extract by a standard volumetric method and GLC. GLC analysis was performed on 3% SE-30 on Chromosorb W (1 m×2 mm column), using programmed temp. from 215 to 265° and a flow rate of 60 ml N<sub>2</sub>/min; n-tetracosane was used as int. standard.

Icacine (1). The alkaloid crystallized from MeOH-CHCl<sub>3</sub> (4:1) as white needles, mp (decomp.) 250-280°, IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup>: 3520, 3020 and 1758,  $[\alpha]_{D}^{D}-51.3^{\circ}$  (c 0.305, CHCl<sub>3</sub>) <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.16 (3H, s, H-17), 1.38 (3H, s, H-19), 2.11 (1H, dd,  $J_1 = 7.5$ ,  $J_2 = 2.2$  Hz, H-5), 2.21 (6H, s, H-21/22), 2.42 (1H, t, J = 8.6 Hz, H-15), 3.01 (1H, s, H-14), 3.34 (1H, d, J = 4.8 Hz, H-7), 3.56 (1H, dd,  $J_1 = 9.8$ ,  $J_2 =$ 2.0 Hz, H-20), 3.86 (1H, dd,  $J_1 = 8.6$ ,  $J_2 = 8.1$  Hz, H-16), 3.88 (1H, s, -OH), 4.04 (IH, dd,  $J_1 = 8.1$ ,  $J_2 = 8.1$  Hz, H-16'), 4.76 (1H, dd,  $J_1 = 9.9$ ,  $J_2 = 2$  Hz, H-20'), 4.96 (1H, dd,  $J_1 = 7.5$ ,  $J_2 = 4.5$  Hz, H-6), MS: m/e 405.2143 (M<sup>+</sup> C<sub>22</sub>H<sub>31</sub>NO<sub>6</sub> requires 405.2150), 308.1854 (C<sub>17</sub>H<sub>26</sub>NO<sub>4</sub> requires 308.1861) and 87.0676 (base peak, C<sub>4</sub>H<sub>9</sub>NO requires 87.0684).

*Icaceine* (2). The alkaloid crystallized from MeOH-CHCl<sub>3</sub> (4:1) as white needles, mp (decomp. sublimation) 220-250°. IR  $\nu_{max}^{RBT}$  cm<sup>-1</sup>: 3490, 1785, 1737 and 1655  $[\alpha]_D^{20}$  - 196.68° (c 0.995, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.01 (3H, s, H-17), 1.06 (3H, s, H-20), 1.54 (3H, s, H-19), 1.85 (1H, d, J = 4.4 Hz, H-5), 2.05 (1H, m, H-3), 2.22 (6H, s, H-21/22), 2.42 (1H, t, J = 8.4 Hz, H-15), 3.64 (1H, s, OH), 3.84 (1H, s, H-14), 3.89 (1H, dd,  $J_1 = 8.8$ ,  $J_2 = 8.1$  Hz, H-16), 4.07 (1H, dd,  $J_1 = 8.1$ ,  $J_2 = 8.1$  Hz, H-16'), 4.9 (1H, dd,  $J_1 = 5.1$ ,  $J_2 = 4.4$  Hz, H-6), 5.9 (1H, dd,  $J_1 = 5.1$ ,  $J_2 = 1.5$  Hz, H-7), MS: m/e 375.2404 (M<sup>+</sup>C<sub>22</sub>H<sub>33</sub>N<sub>1</sub>O<sub>4</sub> requires 375.2409), 346.2371 (C<sub>21</sub>H<sub>32</sub>N<sub>1</sub>O<sub>3</sub> requires 346.2381), 316.2271 (C<sub>20</sub>H<sub>30</sub>N<sub>1</sub>O<sub>2</sub> requires 316.2276), 289.1806 (C<sub>18</sub>H<sub>25</sub>O<sub>3</sub> requires 289.1803), 271.1693 (C<sub>18</sub>H<sub>23</sub>O<sub>2</sub> requires 271.1697), 87 (base peak).

De-N-methylicaceine (3). The alkaloid crystallized from MeOH-CHCl<sub>3</sub> (4:1) as white needles, mp (decomp., sublimation 210-235°. IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup>: 3220, 1770, 1737, 1655. [ $\alpha$ ]<sub>D</sub><sup>20</sup> -165.79° (c 0.535, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  0.96 (3H, s, H-17), 1.06 (3H, s, H-20), 1.53 (3H, s, H-19), 1.85 (1H, d, J = 4.4 Hz, H-5), 2.06 (1H, m, H-3), 2.46 (3H, s, H-21), 3.04 (1H, t, J = 8.4 Hz, H-15), 3.66 (1H, s, -OH), 3.65 (1H, dd, J<sub>1</sub> = 8.8, J<sub>2</sub> = 8.1 Hz, H-16), 3.84 (1H, s, H-14), 4.21 (1H, dd, J<sub>1</sub> = 8.4, J<sub>2</sub> = 8.1 Hz, H-16), 3.84 (1H, s, H-14), 4.21 (1H, dd, J<sub>1</sub> = 8.4, J<sub>2</sub> = 8.1 Hz, H-16), 4.89 (1H, dd, J<sub>1</sub> = 5.5, J<sub>2</sub> = 4.4 Hz, H-6), 5.9 (1H, dd, J<sub>1</sub> = 5.1, J<sub>2</sub> = 1.4 Hz, H-7); MS: m/e 361.2251 (M<sup>+</sup> C<sub>21</sub>H<sub>31</sub>N<sub>1</sub>O<sub>4</sub> requires 361.2252), 331.2141 (C<sub>20</sub>H<sub>29</sub>N<sub>1</sub>O<sub>3</sub> requires 316.1911), 288.1716 (C<sub>18</sub>H<sub>24</sub>O<sub>3</sub> requires 288.1724), 260.1763 (C<sub>17</sub>H<sub>24</sub>O<sub>2</sub> requires 260.1775), 74 (base peak).

3-Acetylicaceine. A soln of 2 (20 mg) in 0.5 ml Py and 0.5 ml Ac<sub>2</sub>O was allowed to stand overnight at room temp. After purification on neutral Al<sub>2</sub>O<sub>3</sub> (toluene-Me<sub>2</sub>CO-EtOH-NH<sub>4</sub>OH, 40:40:8:3,  $R_f$  0.92), the derivative crystallized from MeOH-CHCl<sub>3</sub> (4:1) as white needles, mp

(decomp., sublimation) 190-215°, IR  $\nu_{\max}^{\text{KBr}}$  cm<sup>-1</sup>: 1770, 1735, 1660,  $[\alpha]_D^{20} - 180.34^\circ$  (c 0.295, CHCl<sub>3</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>): 8 1.01 (3H, s, H-17), 1.11 (3H, s, H-20), 1.38 (3H, s, H-19), 1.9 (1H, d, J = 4 Hz, H-5), 2.19 (3H, s, CH<sub>3</sub>COO-), 2.22 (6H, s, H-21/22), 2.42 (1H, t, J = 8.4 Hz, H-15), 3.83 (1H, s, H-14), 3.88 (1H, dd, J<sub>1</sub> = 8.5, J<sub>2</sub> = 8.1 Hz, H-16), 4.07 (1H, dd, J<sub>1</sub> = 8.1, J<sub>2</sub> = 8.1 Hz, H-16), 4.84 (1H, dd, J<sub>1</sub> = 4.8, J<sub>2</sub> = 4.4 Hz, H-6, 4.95 (1H, dd, J<sub>1</sub> = 11.0, J<sub>2</sub> = 7.0 Hz, H-3), 5.89 (1H, dd, J<sub>1</sub> = 5.1, J<sub>2</sub> = 1.5 Hz, H-7), MS: m/e 417 (M<sup>+</sup>), 331, 271, 87 (base peak).

Methylation of De-N-methylicaceine (3). A soln of 3 (20 mg) in MeOH (1.1 ml) was treated at room temp. by 180  $\mu$ l (30  $\mu$ l every 30 min) of MeI or CD<sub>3</sub>I and then left for 12 h at room temp. Unreacted 3 was removed by chromatography on neutral Al<sub>2</sub>O<sub>3</sub> (toluene-Me<sub>2</sub>CO-EtOH-NH<sub>4</sub>OH, 40:40:8:3 (yield: 10 mg).

Se dehydrogenation. 1, 2, or 3 (5 mg) mixed with Se (25 mg) was heated at 300° for 7 hr in a capillary tube. The residue was taken up in *n*-hexane and purified by TLC on Si gel. Further comparison with 1,7-dimethylphenanthrene was achieved by UV spectroscopy, TLC on Si gel GF<sub>254</sub> (HPTLC plates for the nano-TLC Merck, *n*-hexane,  $R_f$  0.2), by GLC on a SE-52 column (50 m×0.5 mm, carrier gas 6 ml He/min, oven temp. 220°,  $R_t$  phenanthrene 5.3 min,  $R_t$  1,7-dimethylphenanthrene 10 min) and by MS. Authentic 1,7-

dimethylphenanthrene was obtained by Se dehydrogenation of rimuene [7].

Acknowledgements—The authors are grateful to Dr. J. D. Connolly for an authentic sample of rimuene.

## REFERENCES

- On'okoko, Penge, Hans, M., Colau, B., Hootele, C., Declerco, J. P., Germain, G. and Van Meersche, M. (1977) Bull. Soc. Chim. Belg. 86, 655.
- Jacobs, W. A. and Huebner, C. F. (1947) J. Biol. Chem. 170, 189.
- 3. Mussini, P., Orsini, F., Pellisoni, F. and Ferrari, G. (1973) J. Chem. Soc. Perkin Trans 1, 2551.
- 4. Orsini, F., Pellizoni, F., McPhail, A. T., Onan, K. D. and Wenkert, E. (1977) Tetrahedron Letters 1085.
- Braude, E. A., Nachod, F. C. and Phillip, W. D. (1955) Determination of Organic Structure by Physical Methods, p. 112. Academic Press, New York.
- Ellestad, G. A., Evans, R. H. and Kunstmann, M. P. (1969) J. Am. Chem. Soc. 91, 2134.
- Connolly, J. D., McCrindle, R., Murray, R. D. H. and Overton, K. H. (1966) J. Chem. Soc. C 273.