

STEREOSELECTIVE SYNTHESIS OF (S)-PROPANOL AMINES: LIVER MICROSOMES
MEDIATED OPENING OF EPOXIDES WITH ARYLAMINES

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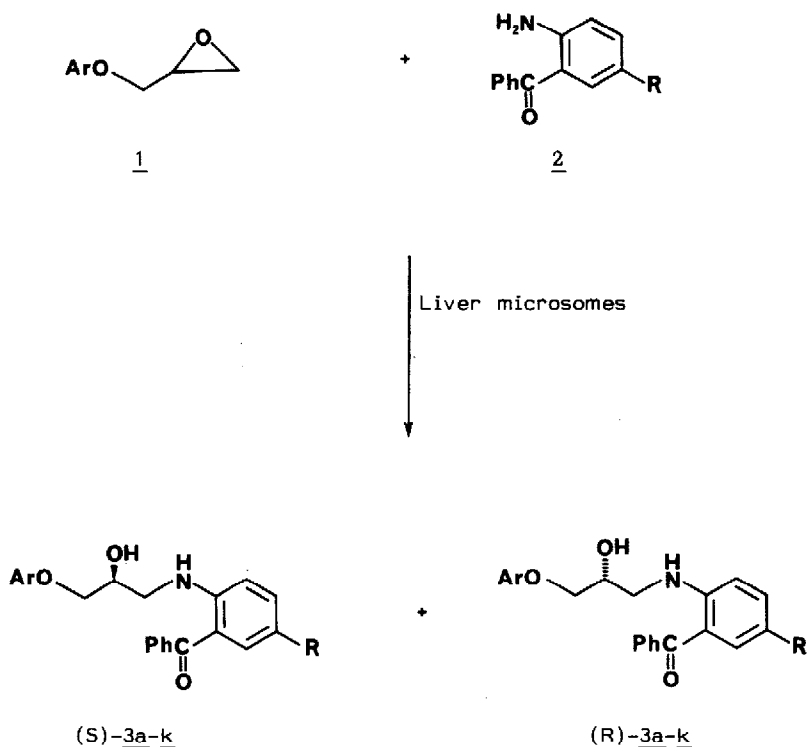
Key words: (S)-propanol amines; liver microsomes; stereoselective synthesis

Abstract: Stereoselective synthesis of 2-(S)-propanol amines by the ring-opening of racemic epoxides with arylamines in presence of rat liver microsomes.

The ring-opening reaction of epoxides with amines is of significance particularly from the stereoselectivity point of view. Most of the biologically important compounds such as in the beta-adrenergic blocking agents^{1,2} the activity usually resides mainly in the (S)-enantiomer³ and are often synthesized by the opening of epoxides with corresponding amines. It is well established that liver microsomes produce biotransformations with remarkable selectivity.⁴ This feature can also be correlated to the *in vitro* metabolism studies of various compounds, wherein regio- and stereoselective phenomena are often experienced.⁵

The stereoselective opening of the epoxides to diols by liver microsomes has been extensively studied.⁶ To our knowledge, there are no reports on the opening of epoxides with amines in presence of liver microsomes. In continuation of our efforts on the application of enzymes as biocatalysts^{7,8} in stereoselective synthesis, the present investigation describes the enantioselective ring-opening reaction of 1-aryloxy-2,3-epoxypropanes with arylamines in presence of rat liver microsomes.

In a typical reaction, to 1a (150 mg) and half a molar equivalent amount of 2a in ethanol (3 ml) as well as 0.1 M phosphate buffer (30 ml, pH 9.5) solution was added freshly prepared rat liver microsomal preparation⁷ (4 ml) of protein content 4.8 mg/ml. The mixture was incubated at 37°C for 24 h and was extracted thrice with chloroform (occasionally appearing emulsions were broken by centrifugation at 2000-4000 g). The organic phase was dried over Na₂SO₄ and evaporated to dryness under reduced pressure. The residue was subjected to column chromatography, chloroform-methanol (97:3) to give 3a, m.p. 93-95°C.



Besides epoxide **1a**, the selective ring-opening reaction for several epoxides with arylamines was carried out successfully employing the present method. The results are illustrated in Table 1. These reactions were monitored by HPLC.⁹ Analytical and spectroscopic data were satisfactory.¹⁰ A control incubation with boiled microsomal fraction did not produce **3**.

The absolute configuration of **3k** obtained from **1k** was established by recovering unreacted **1k** and further reacting with isopropylamine to give propranolol which was then compared with chirally pure (R)-propranolol.¹¹ The unambiguous assignment of the predominant configuration of **3** as (S) and recovered **1** as (R) was thus possible.

Earlier, we synthesized compounds **3** in the racemic form with an interest to explore their biological potential by the well established method of the ring-opening of epoxides with arylamines in protic solvents under reflux conditions.¹² As such, this study enables now to derive a correlation between the stereochemistry and biological activity.

Table. Liver microsomes mediated ring-opening of racemic epoxides with arylamines

	R	Ar	% Yield	% e.e. ^a
<u>a</u>	Cl	Ph	40	86
<u>b</u>	H	4-AcNHC ₆ H ₄	43	77
<u>c</u>	Cl	4-AcNHC ₆ H ₄	37	51
<u>d</u>	H	4-ClC ₆ H ₄	42	57
<u>e</u>	Cl	4-MeOC ₇ H ₄	38	61
<u>f</u>	H	2-MeCOC ₆ H ₄	46	52
<u>g</u>	Cl	2-MeCOC ₆ H ₄	48	88
<u>h</u>	H	2-PhCOC ₆ H ₄	27	65
<u>i</u>	Cl	2-PhCOC ₆ H ₄	33	68
<u>j</u>	H	α -naphthyl	46	75
<u>k</u>	Cl	α -naphthyl	41	85

^a Compounds **3** were derivatized to diacetates and e.e. was determined by HPLC employing LKB enantiopac, α -AGP 10 μ m column (4.0x100 mm), 0.01 M phosphate buffer containing 0.50% (v/v) of acetonitrile at 0.7 ml/min and 220 nm wavelength and were calibrated by the authentic racemic forms of **3**.

In conclusion, the present investigation employing liver microsomes provides a mild and stereoselective procedure for the synthesis of 2-(S)-propanol amines. Further studies to determine the full scope of this methodology by the addition of cosolvents and cofactors are in progress.

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- 9 Employing TSK Si-150, 5 μ m column (4.6x250 mm), chloroform-butanol (90:10) at 0.7 ml/min flow rate and 254 nm wavelength.
- 10 Selected spectral data for **3a**: I.R. ν_{max} (KBr) 3430, 3210, 1600, 1580 cm^{-1} ; ^1H NMR (80 MHz, $\text{CDCl}_3 + \text{D}_6\text{-DMSO}$) δ 8.7 (1H, br s), 6.7-7.1 (13H, m), 4.0-4.4 (3H, m), 3.6 (2H, m), 2.9 (1H, br d, $J=6.5$ Hz).
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IICT Communication No. 2701

(Received in UK 22 April 1992)