

4-[1-(4-Chlorphenyl)-propyl]-1,2-diphenylpyrazolidin-3,5-dion (2c)

Die Darstellung erfolgte analog *Rohkopf*⁸⁾ aus 31.2 g 4-Chlorphenylpropylmalonester und 18.4 g Diphenylhydrazin in 100 ml 0.1 M-Natriumethylat. Farblose Kristalle (Dioxan), Schmp. 213°, Ausb. 55 % d. Th. – C₂₄H₂₁ClN₂O₂ (404.4) Ber. c 71.2 H 5.23 N 6.9 Gef. C 70.9 H 5.17 N 7.0. – IR (KBr): 3440 (br), 3050, 2940, 2860, 1750, 1710, 1590, 1480, 1450, 1430, 1295, 1190, 1150, 1095, 1070, 1020, 900, 800, 750, 690, 640, 600 cm⁻¹. – ¹H-NMR (CDCl₃, 270 MHz): δ (ppm) = 7.30–7.06 (m, 10H, arom.), 6.94 ("d", J = 8/2 Hz, 2H, arom.), 6.80 ("d", J = 8/2 Hz, 2H, arom.), 3.56 (d, 1H, H-4), 3.40 (m, 1H, aliph., C*-H), 2.22 (m, 2H, aliph., C-2'), 0.94 (t, J = 7 Hz, 3H, aliph., CH₃). – MS (80 eV): m/z = 404/406 (58/18 %, M⁺), 301 (4), 284 (4), 252 (87), 212 (56), 183 (82), 153 (91), 131 (100), 125 (91), 103 (27), 91 (11), 77 (78), 51 (16).

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[Ph 28]

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Syntheses of Leukotriene Analogs

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also unsaturated fatty acids other than arachidonic acid can be metabolized to leukotrienes⁵⁾. For example the docosaheptaenoic acid can be transformed to only weakly active leukotriene analogs by the 5-lipoxygenase enzyme. On the other hand the C₂₂ unsaturated fatty acid is reported to be a competitive inhibitor of the cyclooxygenase enzyme which produces prostaglandins. Thus, this C₂₂ fatty acid may have cardiovascular protecting properties⁶⁾. This paper describes the synthesis of C₂₂ leukotriene analogs as well as C₂₀ analogs to make these compounds accessible for biological screening.

The epoxyaldehyde **1**⁷⁻⁹⁾ reacted with two equivalents of formylmethylene – triphenylphosphorane in refluxing benzene to give a mixture of the dienealdehyde **2a** and the α , β -unsaturated aldehyde **2b** in the ratio of 70 : 30 (¹H-nmr). The mixture was coupled with undecyltriphenylphosphorane to give the C₂₂ leukotriene-A-analog **3** (70 %) and the C₂₀ tetrahydro-analog **4**¹⁰⁾ (30 %). **3** and **4** were separated with hplc on μ -porasil. Structure and purity of **3** and **4** were corroborated by UV and ¹H-nmr. The triene system in **3** exhibits the characteristic absorptions at 270, 280 and 290 nm, whereas the diene **4** absorbs at 237 nm. Triene **3** and diene **4** are conveniently distinguished by silicagel-tlc (ether:pentane/triethylamine = 50 : 50 : 1). When sprayed with 50 % sulfuric acid, **3** is stained deep-blue while **4** is stained red.

The reaction of **3** and **4** with glutathione, cysteinylglycine, and cysteine according to our earlier described procedure leads to the peptide – conjugates **5a-c** and **6a-c**, respectively, in 60 % isolated yield. This technique is superior to all variants working with protected peptides, because no isomerization is observed, which is an obstacle in protecting group removal¹¹⁻¹³⁾.

The biological activity of the leukotriene analogs was tested on isolated guinea pig ileum. The C₂₂ analogs of the natural leukotrienes had an activity comparable with the leukotrienes of the 3-series¹⁴⁾.

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Experimental Part

¹H-nmr (CDCl₃, int. stand. TMS): Bruker HX-90R; UV: Zeiss DMR-21; HPLC: Waters M-45, μ -Porasil, 30 cm $\times$ 7.9 mm and RP-C₁₈ 30 cm $\times$ 7.9 mm, respectively.

Methyl 10-formyl-5(S),6(S)-epoxy-7E,9E-decadienoate (**2a**)

Methyl 8-formyl-5(S),6(S)-epoxy-7E-octenoate (**2b**)

610 mg (20 mmol) of formylmethylenetriphenylphosphorane is added under argon to a solution of 172 mg (10 mmol) **1** in 10 ml benzene and the reaction mixture is heated 24 h under reflux. Subsequent to flash chromatography (ether) the 70 : 30 mixture of **2a**, **b** is used immediately. Yield: 125 mg (82 %).

C₂₂-Leukotriene-A-methylester (**3**) and C₂₀-Tetrahydroleukotriene-A-methylester (**4**)

At -78°C 175 mg (8 mmol) **2a**, **b** is added to the solution of 8 mmol of undecyltriphenylphosphorane in 5 ml THF/1 ml HMPA. The mixture is stirred for 5 min. 30 ml Ether and 1 ml phosphate buffer (pH 7) are added. The ether phase is separated and dried (Na₂SO₄). After flash chromatography over

silicagel (ether/1% triethylamine) the eluate is purified by hplc (μ -Porasil; hexane/ethylacetate/triethylamine = 100:1:1). Yield **3**: 120 mg (65%), **4**: 36 mg (29%). ^1H -nmr of **3**: δ (ppm) = 2.39 (H-2), 1.52–2.06 (H-3,4), 2.87 (H-5, $J_{5,6}$ = 2.2 Hz), 3.14 (H-6, $J_{6,7}$ = 8.0 Hz), 5.38 (H-7, $J_{7,8}$ = 14.9 Hz), 6.50 (H-8, $J_{8,9}$ = 10.3 Hz), 6.15 (H-9, $J_{9,10}$ = 14 Hz), 6.56 (H-10, $J_{10,11}$ = 11 Hz), 6.01 (H-11, $J_{11,12}$ = 10.1 Hz), 5.51 (H-12, $J_{12,13}$ = 7.5 Hz), 2.16 (H-13), 1.27 (H-14–21), 0.89 (H-22), 3.68 (OCH₃). UV (hexane) λ_{max} = 270 sh, 280, 290 sh nm. Spectral data of **4** see lit.¹⁰⁾.

Synthesis of the peptide conjugates **5a–c** and **6a–c** according to lit.^{11–13)}.

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- 14 The results of the full biological studies will be reported separately.

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Charge-Transfer Complexes of Sulfonamide Drugs with Tetracyanoethylene

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Formation of 1:1 complexes occurs between the sulfonamide drugs and tetracyanoethylene. Values of the formation constant (K_{ct}), the molar absorptivity (ϵ), the transition energy (E) and of the product ($K_{\text{ct}} \times \epsilon$) have been obtained from optical data of the complexes. From the energies of charge-transfer transitions, the ionization potentials of the donors have been obtained. The spectroscopic data indicate that the complexes are weak in nature.