

STRUCTURE OF ACARANOIC AND ACARENOIC ACIDS*

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Abstract For acaranoic and acarenoic acids from the lichen *Acarospora chlorophana*, the revised structures of (–)-2S-methyl-3R-carboxypentadecyl-1 \rightarrow 5S-olide and (–)-2-methylene-3R-carboxypentadecyl-1 \rightarrow 5S-olide respectively have been established.

INTRODUCTION

In 1895, Zopf [1] isolated from the lichen *Acarospora chlorophana* a compound, pleopsidic acid, mp 131–132° and $[\alpha]_D^{18} = 55$ to -60° . Later, Santesson [2] recognized pleopsidic acid as a mixture of two components which he named acaranoic and acarenoic acids and for which he put forward the γ -lactone structures 1 and 2. To support 1, Santesson [2] reacted Me β -ketotetradecanoate with di Me bromosuccinate and received, after hydrolysis and subsequent reduction with NaBH₄, a mixture, mp 112–116°, which he assumed to be identical with racemic 1 by comparison of IR and mass spectra. We have reexamined acaranoic and acarenoic acids and now propose the δ -lactone structures 3 and 4 for the two compounds.

RESULTS AND DISCUSSION

Acaranoic acid has two carbonyl bands in the IR spectrum: in KBr, 1678 (CO₂H) and 1716 cm⁻¹ (δ -lactone-CO-), in CHCl₃, 1708 (–CO₂H) and 1720 cm⁻¹ (δ -lactone-CO-). Me acaranoate (5), easily accessible from 3 by reaction with CH₃N₃, shows only one CO band at 1710 cm⁻¹ (in KBr) (1740 cm⁻¹ in CCl₄). Contrary to these data, similar γ -lactones such as roccellaric acid and its Me ester [3], absorb at 1740 and 1782 cm⁻¹ (in KBr) (1713 and 1772 cm⁻¹ in CHCl₃), respectively. The correct carbon skeleton was given from the ¹H NMR spectrum (270 MHz, CDCl₃, δ values in ppm) of 5: 0.88 (3 H, t, 15-Me), 1.26 (14 H, br. s, 7-CH₂–14-CH₂-), 1.32 (3 H, d, J = 7 Hz, 16-Me), 1.55 (2 H, m, 6-CH₂-), 1.76 (1 H, dt, J_{4a,5} = 12, J_{4a,4c} = 14 Hz, C-4H_a), 2.13 (1 H, dt, J_{4c,5} = 3.5, J_{4c,4a} = 14 Hz, C-4H_c), 2.62 (1 H, td, J_{3,4a} = 12, J_{3,4c} = 3.5 Hz, C-3H), 2.78 (1 H, dq, J_{2,16} = 7, J_{2,3} = 11 Hz, C-2H), 3.75 (3 H, s, CO₂Me), 4.29 (1 H, m, C-5H). A key to the δ -lactone structure as well as the relative stereochemistry is the vicinal coupling of four protons at C-2, C-3, C-4 and C-5. With coupling constants of 11 and 12 Hz these protons must occupy coplanar

positions to each other, which can only be achieved in a 6-membered ring with the protons axially arranged as depicted in 3a.

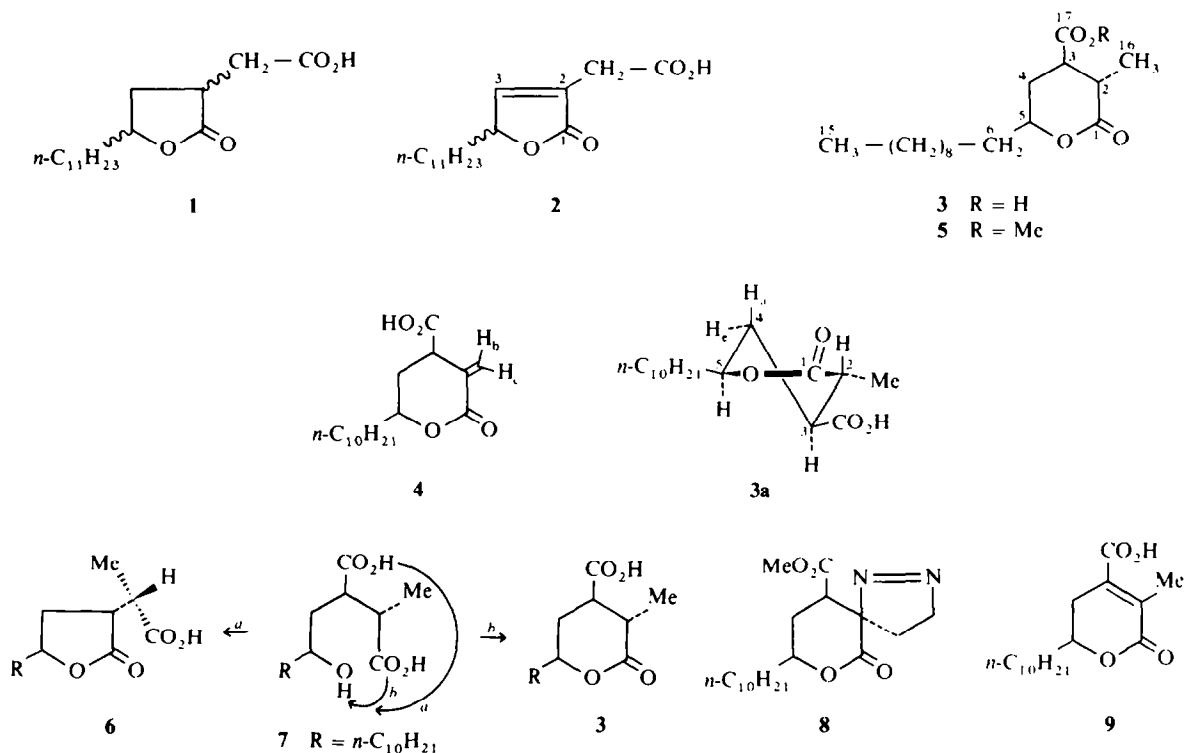
The ¹³C NMR spectrum (25.05 MHz, CDCl₃) of acaranoic acid is in good agreement with structure 3 although the γ -lactone isomer (6) also fits the observed chemical shifts: C-1: 177.0 (s), C-2: 45.3 (d), C-3: 37.7 (d), C-4: 36.1 (t), C-5: 79.6 (d), C-6: 32.7 (t), C-7: 24.8 (t), C-8: C-12: 29.4–29.6 (t), C-13: 32.0 (t), C-14: 22.4 (t), C-15: 14.2 (q), C-16: 16.6 (q), C-17: 172.3 (s).

The sign of the Cotton effect of saturated δ -lactones depends on the chirality of the conformation with a planar lactone group (–C–O–CO–C–) [4]. (–)-Acaranoic acid (3) and its Me ester (5) show positive Cotton effects in the CD curves: 3: $\Delta\epsilon_{222} + 2.23$, 5: $\Delta\epsilon_{222} + 1.83$. This means (–)-acaranoic acid has the chirality shown in 3a and the absolute configuration (–)-2S-methyl-3R-carboxypentadecyl-1 \rightarrow 5S-olide.

The hydroxy acid 7 resulting from hydrolysis of 3 theoretically can relactonize either to γ -lactone acid 6 (path a) or to acaranoic acid (path b). Alkaline hydrolysis of 3 and 5 and subsequent acidification yielded (–)-acaranoic acid: so relactonization follows path b to the more stable δ -lactone acid.

Santesson's structure 2 of acarenoic acid [2] can be eliminated from the ¹H NMR spectrum (270 MHz, CDCl₃), which shows no signal corresponding to a vinylic proton (2, C-3H): 0.88 (3 H, t, J = 7 Hz, 15-Me), 1.26 (14 H, br. s, 7-CH₂–14-CH₂-), 1.3–1.8 (2 H, m, 6-CH₂-), 2.03 (1 H, dt, J_{4a,5} = 12, J_{4a,4c} = 14 Hz, C-4H_a), 2.21 (1 H, ddd, J_{4c,5} = 2, J_{4c,3} = 6, J_{4c,4a} = 14 Hz, C-4H_c), 3.69 (1 H, m, C-3H), 4.23 (1 H, m, C-5H), 5.93 (1 H, d, J_{3,11b} = 2.5 Hz, C-16H_b), 6.64 (1 H, d, J_{3,11c} = 2.5 Hz, C-16H_c). The doublets at 5.93 and 6.64 confirm the presence of an exo-methylene group α to the lactone carbonyl. Structure 4 was finally proved by catalytic hydrogenation of (–)-acarenoic acid which gives as the main product a compound identical with (–)-acaranoic acid (3) in all respects. Hence the absolute configuration of 4 is (–)-2-methylene-3R-carboxypentadecyl-1 \rightarrow 5S-olide. The CD spectrum of 4 shows a positive band at 268 nm ($\Delta\epsilon + 0.20$) and a negative one at 216 nm ($\Delta\epsilon - 5.23$). (–)-Acarenoic

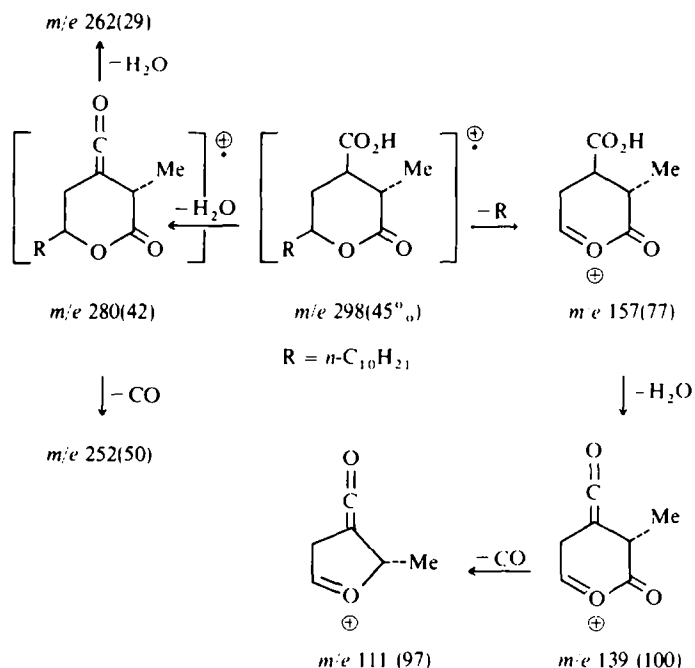
* Part 124 in the series "Lichen Substances". For Part 123 see Huneck, S. (1980) *Lichenologist* (in press).



acid reacts with CH_2N_2 to give the pyrazoline Me ester **8**, whose configuration at C-2 was deduced from the positive CD bands at 327 nm ($\Delta\epsilon +10.0$) and 233 nm ($\Delta\epsilon +4.28$) according to the rule of Sznatzke [5]. This means CH_2N_2 attacks the $\text{C}=\text{C}$ double bond from the re-side. On heating with Ac_2O , **4** is isomerized to *iso*-acarenoic acid (**9**). The main fragments in the mass spectrum of (–)-acarenoic acid are given in Scheme 1.

EXPERIMENTAL

Extraction. *A. chlorophana* (Wahlenb. ex Ach.) Mass. (India, Himalayas, Distr. Pithoragarh, May 1978, voucher specimen in the herbarium of S.H.: 162 g) was extracted with Et_2O (200 ml) for 20 hr and the extract evapd to dryness. The resulting residue was dissolved in C_6H_6 (300 ml) and chromatographed on Si gel (80 g, with 5% H_2O). C_6H_6 (3000 ml) eluted (+)-rhizocarpic acid



Scheme 1. MS fragmentation of (–)-acarenoic acid (**3**) at 40 kV.

(2.05 g, 1.2%), mp 174–175° (CHCl₃–MeOH) and $[\alpha]_D^{24} + 104^\circ$ (CHCl₃, *c* 1.095). Further elution with C₆H₆–Et₂O (1:1) (21.) yielded a mixture of (–)-acaranoic and (–)-acarenoic acids (1.77 g, 1.09%), mp 135–145° (MeOH). Part of this mixture (1 g) was rechromatographed on Si gel (60 g, with 5% H₂O): elution with C₆H₆–Et₂O (9:1) (400 ml) gave pure (–)-acaranoic acid (3, 0.32 g) as plates, mp 150–151° (CHCl₃–MeOH) and $[\alpha]_D^{25} - 38.9^\circ$ (CHCl₃–MeOH, 1:1, *c* 1.36). C₁₇H₃₀O₄ (298.41). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log *ε*): 210 (2.85). IR, $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 688, 722, 748, 814, 850, 920, 960, 990, 1000, 1050, 1102, 1140, 1150, 1178, 1196, 1228, 1250, 1270, 1304, 1330, 1362, 1384, 1398, 1428, 1448, 1470, 1678, 1716, 2800, 2900, 3500. ¹H NMR (270 MHz, CDCl₃–CD₃OD, 4:1, 60°): 0.89 (3 H, *t*, 15-Me), 1.26 (14 H, *br. s*, 7-CH₂–14-CH₂–), 1.35 (3 H, *d*, 16-Me), 1.40–1.60 (2 H, *m*, 6-CH₂–), 1.75 (1 H, *dt*, C-4H₃), 2.20 (1 H, *dt*, C-4H₄), 2.58 (1 H, *td*, C-3H), 2.76 (1 H, *dq*, C-2H), 4.31 (1 H, *m*, C-5H).

The other part of the mixture of 3 and 4 (50 mg) was separated by prep.-TLC (Si gel Merck PF 254 + 366, 20 × 20 × 1 mm, C₆H₆–dioxane–HOAc (90:25:4), 10 mg mixture/plate, detection UV). The band of highest *R_f* value yielded after extraction with Et₂O (–)-acaranoic acid (5 mg), mp 148–150° and the band of lower *R_f* value (–)-acarenoic acid (4, 25 mg), plates, mp 130–132° (C₆H₆) and $[\alpha]_D^{24} - 84.3^\circ$ (CHCl₃, *c* 0.265). C₁₇H₂₈O₄ (296.40). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log *ε*): 219 (3.42). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 684, 718, 810, 928, 954, 1000, 1048, 1080, 1102, 1136, 1178, 1196, 1276, 1310, 1360, 1398, 1422, 1442, 1470, 1614 (C=CH₂), 1670 (–CO₂H), 1720 (lactone-CO–), 2900, 2960, 3500 (–CO₂H).

Me acaranoate (5). Prepared from 3 (0.1 g) in MeOH (10 ml) with CH₃N₃ in Et₂O for 5 min at 20°. Chromatography of the reaction product on Al₂O₃ (6 g, activity II, neutral) in hexane afforded needles, mp 58–59° (pentane or MeOH–H₂O) and $[\alpha]_D^{25} - 31.1^\circ$ (CDCl₃, *c* 1.595). C₁₈H₃₂O₄ (312.44). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 678, 712, 722, 734, 768, 810, 850, 882, 910, 922, 940, 960, 990, 1008, 1020, 1030, 1050, 1064, 1078, 1100, 1114, 1130, 1138, 1160, 1170, 1190, 1210, 1222, 1238, 1270, 1280, 1300, 1328, 1340, 1362, 1380, 1440, 1462, 1472, 1710, 2850, 2940: $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{–1}: 938, 1029, 1072, 1098, 1113, 1172, 1193, 1226, 1270, 1300, 1767, 1380, 1440, 1462, 1740, 2940. MS *m/e* (rel. int.): 312 (M⁺, 30), 294 (19), 281 (21), 267 (24), 253 (30), 239 (38), 171 (73), 155 (26), 146 (77), 143 (42), 142 (73), 139 (49), 128 (42), 127 (46), 114 (48), 102 (54), 101 (100).

Hydrolysis and relactonization of acaranoic acid. (–)-Acaranoic acid (0.1 g) was dissolved in an aq. soln (2 ml) of KOH (50 mg) and kept at room temp. for 20 hr. After acidification with 10% H₂SO₄, extraction with Et₂O and crystallization (CHCl₃–MeOH), plates,

mp 151–152°, were obtained which were identical with (–)-acaranoic acid.

Hydrolysis of Me acaranoate. Me acaranoate (5, 80 mg) was refluxed in a soln of KOH (50 mg) in MeOH (2 ml) and H₂O (1 ml) for 2 hr. After removing the solvents, acidification with 10% H₂SO₄, extraction with Et₂O and crystallization (CHCl₃–MeOH), plates, mp 149–150° and $[\alpha]_D^{25} - 39.6^\circ$ (CHCl₃–MeOH, 1:1, *c* 0.9) were obtained. These were identical with (–)-acaranoic acid.

Hydrogenation of (–)-acarenoic acid. (–)-Acarenoic acid (4, 20 mg) was hydrogenated with Adams catalyst (10 mg) in EtOH under normal conditions for 2 hr. After usual work-up, plates, mp 142–144° (MeOH–H₂O) and $[\alpha]_D^{24} - 39.5^\circ$ (CHCl₃, *c* 0.27) were obtained, identical with (–)-acaranoic acid.

Addition product of CH₃N₃ with (–)-acarenoic acid (8). (–)-Acarenoic acid (0.1 g) was treated with CH₃N₃ in Et₂O for 4 hr. The residue after evapn of Et₂O was chromatographed in C₆H₆ on Si gel (10 g, with 5% H₂O). After elution with C₆H₆ (300 ml), further C₆H₆ (100 ml) eluted the pyrazoline Me ester as plates, mp 48–48.5° (MeOH–H₂O) and $[\alpha]_D^{25} + 238.5^\circ$ (CHCl₃, *c* 0.35). C₂₀H₃₂N₂O₄ (340.46).

Iso-acarenoic acid (9). (–)-Acarenoic acid (10 mg) was heated with Ac₂O (2 ml) to 100° for 2 hr. After usual work-up and crystallization from MeOH–H₂O, plates mp 123–125° were obtained. C₁₇H₂₈O₄ (296.40). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log *ε*): 232 (3.56). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 702, 760, 780, 920, 940, 960, 1000, 1100, 1116, 1156, 1180, 1198, 1232, 1250, 1310, 1370, 1398, 1430, 1470, 1670, 1700, 2960, 3500. MS *m/e* (rel. int.): 296 (M⁺, 75), 278 (62), 260 (29), 252 (47), 233 (64), 225 (48), 207 (48), 193 (38), 179 (53), 165 (53), 155 (100), 139 (98), 126 (100), 111 (93), 97 (93), 87 (96).

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