hydrazoic acid may take any role since no trace of III was produced without sodium azide, only recovered I.

Acknowledgment. We thank Professor H. Wynberg for his kindly providing us the infrared data of compound IV and Dr. T. Nishida and Mr. I. Miura of Nippon Electric Varian, Ltd., and Takeda Chemical Industries, Ltd., for nmr measurements.

Tadashi Sasaki, Shoji Eguchi, Takeshi Toru Institute of Applied Organic Chemistry, Faculty of Engineering Nagoya University, Chikusa-ku, Nagoya, Japan Received March 5, 1969

The Synthesis of Adenine 5'-O-Sulfamoyl Nucleosides Related to Nucleocidin¹

Sir:

Continued interest in the potent antitrypanosomal² antibiotic nucleocidin³ has been prompted by the demonstration of its activity as an inhibitor of protein biosynthesis.⁴ A recent communication⁵ has redefined the structure of nucleocidin as 5'-O-sulfamoyl-4'-Cfluoroadenosine (1) (or a stereoisomer) based on ¹H and ¹⁹F nmr data and mass spectroscopy. Nucleocidin is the only known antibiotic containing the sul-



famate ester group and is one of the very few naturally occurring fluoro compounds.⁶ Indeed, sulfamate esters with an unsubstituted nitrogen are rare; only a few simple aliphatic examples have been described.⁷

We wish to report the preparation of 5'-O-sulfamoyladenosine (2) and 5'-O-sulfamoyl-2'-deoxyadenosine (3), which are the first synthetic sulfamate esters of nucleosides and are direct structural models of nucleocidin. A solution of 2',3'-O-ethoxymethylideneadenosine⁸ in 1,2-dimethoxyethane (glyme) was treated with 1 equiv of sodium hydride. After reaction ceased, a solution of 1 equiv of sulfamoyl chloride in glyme was added to the stirred suspension. Purification of 5'-O-sulfamoyl-2',3'-O-ethoxymethylideneadenosine bv

chromatography on silca gel followed by deblocking with 5% formic acid and then aqueous ammonia gave 5'-O-sulfamoyladenosine (2) monohydrate⁹ in 33%over-all yield. The product 2 softens at 153-155° decomposes at 165°; $[\alpha]^{33}D - 33.6^{\circ}$ (c 1.0, DMF); $[\alpha]^{27}D - 33^{\circ}$ (c 1.0, EtOH-0.1 N HCl, 1:1); uv max (pH 1) 257 m μ (ϵ 14,800), (pH 11) 259 m μ (ϵ 15,400); ir (KBr) 1180 cm⁻¹ (5'-OSO₂NH₂); nmr (DMSO-d₆) δ 4.31 (broad s, 3, 5'-H (2) plus 4'-H (1)), 3.50 (s, 2, H_2O), 7.33 (s, 2, 6- NH_2), 7.63 (s, 2, 5'- OSO_2NH_2); addition of D_2O caused the peaks at δ 3.50, 7.33, and 7.63 to disappear with a corresponding increase at δ 3.70 (HDO).

Two equivalents of sulfamovl chloride was added slowly to a solution of 3'-O-acetyl-2'-deoxyadenosine¹⁰ in pyridine-glyme at 0°. The residue obtained after addition of sodium carbonate and evaporation of the solvent was partitioned between ethyl acetate and water and the solid remaining after evaporation of the organic phase was recrystallized from ethanol to give a 39% yield of 5'-O-sulfamoyl-3'-O-acetyl-2'-deoxyadenosine,⁹ mp 157–159°; uv max (pH 1) 257 m μ (ϵ 14,000), (pH 11) 259 m μ (ϵ 16,300); ir (KBr) 1180 cm⁻¹ (5'-OSO₂NH₂), 1740 cm⁻¹ (3'-OAc); nmr (DMSO- d_{6^-} CDCl₃, 30:70) δ 4.43 (broad s, 3, 5'-H (2) plus 4'-H (1)), 2.15 (s, 3, 3'-OAc), 6.92 (s, 2, $6-NH_2$), 7.50 (s, 2, $5'-OSO_2NH_2$).

This product was deblocked with methanolic ammonia and recrystallized from aqueous ethanol, dissolved in water, and then lyophilized to give a 76%yield of amorphous 5'-O-sulfamoyl-2'-deoxyadenosine monohydrate⁹ (3), softens at 114°, decomposes at 170°; $[\alpha]^{30}D - 23.5^{\circ}$ (c 1, H₂O); uv max (pH 1) 258 mµ (ε 15,600), (pH 11) 259 mµ (ε 16,800); ir (KBr) 1180 cm⁻¹ (5'-OSO₂NH₂); nmr (DMSO- d_6) δ 4.16 (broad s, 3, 5'-H (2) plus 4'-H (1)), 7.25 (s, 2, 6-NH₂), 7.57 (s, 2, 5'-OSO₂NH₂), 5.53 (broad s, 1, 3'-OH), 3.35 (s, 2, H_2O); addition of D_2O caused the peaks at δ 7.25, 7.57, 5.53, and 3.35 to disappear with a corresponding increase at δ 3.55 (HDO).

The ir spectra of 2 and 3 are similar to that of nucleocidin^{3a} and show the same band at 1180 cm⁻¹ (covalent $ROSO_2NH_2$). The nmr peak assigned to the 5' protons in 2 or 3 is shifted downfield approximately 0.6 δ in DMSO-d₆ (relative to that of adenosine or 2'deoxyadenosine), which corresponds to the position assigned⁵ to the 5' protons of nucleocidin. The uv spectra of nucleocidin^{3a} are identical with those of 2 and 3. Optical rotations of nucleocidin^{3a} (1) and 2 are essentially identical.

A solution of 5'-O-sulfamoyl-2',3'-O-isopropylideneadenosine⁹ in 0.5% MeOH in MeCN was heated at reflux for 28 hr. The starting material was absent and one new chromatographically homogeneous product (tlc) was present which exhibited identical mobility with 2', 3'-O-isopropylideneadenosine-N³ \rightarrow C⁵'-cyclonucleoside11 in 5% aqueous ammonium chloride on SilicAR-7GF¹² and had uv max (H₂O) 272.5 mµ.¹³ Compound 2 was heated in absolute DMF for 24 hr at

(9) Analysis for C, H, and N agreed within $\pm 0.3\%$ of calculated values.

- (11) V. M. Clark, A. R. Todd, and J. Zussman, J. Chem. Soc., 2952 (1951).
- (12) Mallinckrodt Chemical Works.
- (13) A. Hampton and A. W. Nichol, J. Org. Chem., 32, 1688 (1967).

⁽¹⁾ This work was supported by Grant CA-08109 from the National Cancer Institute of the National Institutes of Health.

⁽²⁾ R. I. Hewitt, A. R. Gumble, L. H. Taylor, and W. S. Wallace, Antibiot. Ann., 722 (1956-1957).

^{(3) (}a) S. O. Thomas, V. L. Singleton, J. A. Lowery, R. W. Sharpe, L. M. Pruess, J. N. Porter, J. H. Mowat, and N. Bohonos, *ibid.*, 716 (1956-1957); (b) C. W. Waller, J. B. Patrick, W. Fulmor, and W. E. Meyer, J. Am. Chem. Soc., 79, 1011 (1957); (c) J. B. Patrick and W. E. Meyer, Abstracts, 156th National Meeting of the American Chemical Society, Atlantic City, N. J., Sept 1968, MEDI-024. (4) J. R. Florini, H. H. Bird, and P. H. Bell, J. Biol. Chem., 241,

^{1091 (1966)}

⁽⁵⁾ G. O. Morton, J. E. Lancaster, G. E. Van Lear, W. Fulmor, and W. E. Meyer, J. Am. Chem. Soc., 91, 1535 (1969). We thank Dr. J. S. Webb, Lederle Laboratories, for a preprint of this work.

⁽⁶⁾ L. Fowden, Proc. Roy. Soc. (London), B171, 5 (1968).
(7) See R. Appel and W. Senkpiel, Z. Anorg. Allg. Chem., 310, 94 (1961), and references therein.

⁽⁸⁾ F. Eckstein and F. Cramer, Chem. Ber., 98, 995 (1965).

⁽¹⁰⁾ M. J. Robins, J. R. McCarthy, Jr., and R. K. Robins, Biochemistry, 5, 224 (1966).

100° with exclusion of moisture. The reaction mixture was chromatographed on SilicAR-7GF and the saltlike product, uv max (MeOH) 269 m μ , was eluted with methanol. Identical treatment of nucleocidin¹⁴ gave similar material, uv max (MeOH) 272 mµ, and 5'-Otosyladenosine¹⁵ (4) gave the $N^3 \rightarrow 5'$ -cyclonucleoside, uv max (MeOH) 271 m μ . This parallel behavior of 1, 2, and 4 defines the stereochemistry of the 1'-adenine and 4'-sulfamoyloxymethyl substituents of nucleocidin as cis.11

The circular dichroism curves of nucleocidin¹⁴ and 2 in water are identical within experimental error throughout the range 230–290 m μ and are similar to that of adenosine.¹⁶ This indicates that the adenine ring is above the sugar plane.^{16,17} Thus, structure 1 for nucleocidin⁵ is consistent with the present data.

It is of interest that 5'-O-sulfamoyladenosine (2) produces 50% inhibition of S. faecalis at 4×10^{-6} M compared to a similar inhibition by nucleocidin at $5 \times 10^{-7} M.^{18}$ Compound 2 also has been found to exhibit pronounced in vitro inhibition of Trypanosoma rhodesiense at 10⁻⁹ M.¹⁹ 5'-O-Sulfamoyladenosine (2) may be viewed²⁰ as an analog of adenosine 5'-phosphate (AMP) which, however, should readily cross cellular membranes due to the nonionic character of the sulfamoyl ester group.

(14) We thank Dr. J. S. Webb, Lederle Laboratories, for a sample of nucleocidin.

(15) R. Kuhn and W. Jahn, Chem. Ber., 98, 1699 (1965).

(16) D. W. Miles, M. J. Robins, R. K. Robins, and H. Eyring, Proc. Natl. Acad. Sci. U. S., 62, 22 (1969).

(17) D. W. Miles, M. J. Robins, R. K. Robins, M. W. Winkley, and H. Eyring, J. Am. Chem. Soc., 91, 831 (1969).

(18) Dr. A. Bloch, Roswell Park Memorial Institute, Buffalo, N. Y., private communication.

(19) Dr. J. Jaffe and Dr. J. J. McCormick, University of Vermont, private communication.

(20) M. G. Stout, M. J. Robins, R. K. Olsen, and R. K. Robins, J. Med. Chem., in press.

> Dennis A. Shuman, Roland K. Robins, Morris J. Robins Department of Chemistry, University of Utah Salt Lake City, Utah 84112 Received March 18, 1969

Generation of Aryl Nitrenes in the Presence of Acetic Acid by Deoxygenation of Aromatic Nitro and Nitroso Compounds¹

Sir:

We wish to report that the presence of acetic acid (5% by volume) in triethyl phosphite solutions employed for photochemical deoxygenation of aromatic nitro compounds² or deoxygenation of aromatic nitroso compounds³ profoundly affects the nature of the reaction products. Our results suggest that a substantial fraction of the aryl nitrenes generated under these conditions are converted to aryl nitrenium ions. This result implies that aryl nitrenes are relatively basic, a conclusion which is of significance to the interpretation of the chemistry of aryl nitrenes in general.

The product mixtures from photochemical deoxygenation of aromatic nitro compounds in the presence of acetic acid include significant amounts of o-hydroxyacetanilides (2) and diethyl aminophenylphosphonates

(1) Supported by National Institutes of Health Grant 14344.

(2) R. J. Sundberg, B. P. Das, and R. H. Smith, Jr., J. Am. Chem. Soc., 91, 658 (1969).

(3) R. J. Sundberg, ibid., 88, 3781 (1966).



(3). Diethyl N-arylphosphoramidates (4) are also formed but in yields generally lower than those observed in the absence of acetic acid. Anilines are also found. The formation of N-aryl 2-acetimidylpyridines from o-methylnitrobenzenes, which is an important process in the absence of acetic acid,² is completely suppressed. Diethyl aminophenylphosphonates are not formed in the absence of acetic acid. Nonphotochemical deoxygenation of aromatic nitroso compounds is also believed⁴ to generate aryl nitrenes, and the product composition from deoxygenation of nitroso compounds is affected in a similar way when acetic acid is present. Table I summarizes the data. Structural assignments for the new compounds o-3a, o-3b, p-3b, o-3c, o-3d, and m-3d rest on correct elemental analyses and definitive infrared and nmr spectral data. The known compounds 2a-d, p-3a, 4b-d, and 5a-d show spectral data in accord with expectation and physical constants in agreement with literature data.

Table I. Product Distribution from Deoxygenations

Reactant	% yields ^a						
	2	0	 	P	4	5	
1a	6	8	с	8	с	4	
ба	11	3	с	6	3	С	
1b	с	2	с	10	7	2	
6b	c	2	с	9	12	2	
1c	27	11	с		2	10	
6c	23	9	с		3	2	
ld	14	6	6		9	4	

" Yields are isolated yields, normally after column chromatography. ^b The designations o, m, and p denote the relationship between the amino and phosphoryl substituents. . Not definitively characterized; yield is less than 3%.

Aryl nitrenium ions are expected to undergo nucleophilic attack principally at the ortho and para carbon atoms⁵ and can be invoked as intermediates in the

(4) J. I. G. Cadogan Quart. Rev. (London), 22, 222 (1968).
(5) P. G. Gassman, G. Campbell, and R. Frederick, J. Am. Chem.
(6) 7377 (1969), P. J. S. Strikt, "On the Strikt Control of the Strikt Soc., 90, 7377 (1968); P. A. S. Smith, "Open Chain Nitrogen Com-