A convenient method for the synthesis of methyl 2-acetamido-2-deoxy-3-0-(2,3,4,6-tetra-0-acetyl-β-D-galactopyranosyl)-α-D-glucopyranoside

CHRISTOPHER J. GRIFFITHS AND HELMUT WEIGEL

The Bourne Laboratory, Royal Holloway College (University of London), Egham, Surrey TW20 OEX (Great Britain)

(Received March 16th, 1977; accepted for publication April 24th, 1977)

The disaccharide 2-acetamido-2-deoxy-3- $O-\beta$ -D-galactopyranosyl-D-glucopyranose (1) is a constituent part of "lacto-N-tetraose", which is present in human milk¹, and of the branched trisaccharide 2-acetamido-2-deoxy-4-O- α -L-fucopyranosyl- $3-O-\beta$ -D-galactopyranosyl- β -D-glucopyranose, which is the Le^a blood-group antigenic determinant². Its synthesis involved a Koenigs-Knorr reaction of 2,3,4,6-tetra-Oacetyl- α -D-galactopyranosyl bromide with the 4,6-O-benzylidene derivative of either benzyl 2-acetamido-2-deoxy-α-D-glucopyranoside³ or 2,2,2-trichloroethyl 2-acetamido-2-deoxy- β -D-glucopyranoside⁴, and subsequent removal of the benzylidene group by catalytic hydrogenolysis. Recent syntheses of blood-group antigenic determinants⁵ prompt us to report a convenient method for the synthesis of 3-Osubstituted derivatives of methyl 2-acetamido-2-deoxy-a-D-glucopyranoside. The 4.6-benzeneboronate (2) of methyl 2-acetamido-2-deoxy- α -D-glucopyranoside was chosen as the intermediate for the following reasons: (a) it is readily obtainable in nearly quantitative yield; (b) benzeneboronates are useful intermediates in the synthesis of alkyl derivatives⁶, esters⁷, and glycosides⁸; and (c) chromatographic work-up of the reaction mixtures using lyotropic solvents effects complete hydrolysis of benzeneboronates⁶.



The evidence upon which the structure of 2 was assigned is as follows. Electronimpact mass spectrometry produced, by the pathway shown in Fig. 1, an ion with m/e 159.0626 (C₉H₈BO₂; $\% \sum_{40}$, 3.2), the formation of which, together with the ion m/e 160 (C₉H₉BO₂; $\% \sum_{40}$, 1.8), is characteristic of the 2-phenyl-1,3,2-dioxaborinane ring system^{9, 10}. Methylation of the benzeneboronate with the BF₃-diazomethane reagent^{11,12}, followed by hydrolysis of the benzeneboronate ring and acetylation, gave a single carbohydrate product (g.l.c.), the e.i.-mass spectrum of which was virtually identical with that of authentic methyl 2-acetamido-4,6-di-O-acetyl-2-deoxy-3-O-methyl- α -D-galactopyranoside¹³.

The product of the Koenigs-Knorr reaction, namely, methyl 2-acetamido-2deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-glucopyranoside 4,6benzeneboronate (3) was not isolated, as the chromatographic work-up of the reaction mixture effected complete hydrolysis of the benzeneboronate ring, affording, directly, methyl 2-acetamido-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-glucopyranoside (4) in 91% yield. The assignment of structure to 4 is based on (a) the method of synthesis; (b) the e.i.-induced formation of the ion m/e 534 through loss of MeO from the molecular ion; (c) optical rotatory power (application of isorotation rules with methyl 2-acetamido-2-deoxy- α -D-glucopyranoside and methyl 2,3,4,6-tetra-O-acetyl- β -D-galactopyranoside); and (d) the splitting (8.5 Hz) of the n.m.r. signal for H-1 of the D-galactopyranosyl unit.

As methyl 2-acetamido-2-deoxy- α -D-glucopyranoside can be obtained from 2-acetamido-2-deoxy-D-glucose in yields of >90%, the method described here



Fig. 1. E.i.-induced fragmentation of methyl 2-acetamido-2-deoxy- α -D-glucopyranoside 4,6-benzeneboronate (2); m^*/e = mass number of metastable ions.



represents a >80% overall yield of 4 from 2-acetamido-2-deoxy-D-glucose in only three operations.

EXPERIMENTAL

General methods. — T.I.c. was performed on silica gel (Polygram Sil G) with ethyl acetate-dichloromethane (1:1). Compounds were detected with iodine vapour, followed by spraying with sulphuric acid (5% in ethanol) and heating at 120°.

G.l.c.-mass spectrometry was performed with a Perkin-Elmer F11 gas chromatograph, operating at 200° and containing a glass column (6 ft \times 0.25 in.) packed with 3% of OV-225 on Chromosorb Q (100-120 mesh). The carrier gas, helium, was removed from the effluent by passage through a Biemann separator. The effluent was then passed into a Hitachi RMS-4 mass spectrometer operating at 80 eV and 50 μ A target current. Low- and high-resolution mass spectra were also recorded on an A.E.I. MS-902 spectrometer operating at 70 eV using a direct insertion method.

N.m.r. spectra were recorded (internal Me_4Si) with a Varian E.M. 360 spectrometer.

Methyl 2-acetamido-2-deoxy- α -D-glucopyranoside 4,6-benzeneboronate (2). — Methyl 2-acetamido-2-deoxy- α -D-glucopyranoside (0.15 g) and benzeneboronic anhydride (0.07 g) were heated in toluene under reflux for 4 h, using a Dean and Stark head to remove the water produced. The resulting solution was evaporated, and the residue recrystallised from 2-methoxyethanol to give 2 (0.2 g), m.p. 287–288°, $[\alpha]_D^{25} + 132^\circ$ (c 1.0, pyridine); the highest mass number recorded in e.i.-m.s. was m/e 322.1462, corresponding to the $[M + 1]^+$ ion (Found: C, 55.81; H, 6.24; B, 3.30; N, 4.52%, C₁₅H₂₀BNO₆ calc.: C, 56.07; H, 6.23; B, 3.43; N, 4.66%).

Methyl 2-acetamido-2-deoxy-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)- α -D-glucopyranoside (4). — Compound 2 (0.93 g) was shaken with 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide (1.38 g) and mercuric cyanide (0.37 g) in nitromethane (120 cm³) at room temperature for 48 h; complete dissolution had then occurred. T.l.c. showed that the reaction was complete and that one major product ($R_{\rm F}$ 0.04) had been formed. The solution was evaporated, and the residue passed through a column (3 × 30 cm) of silicic acid (SIL-R 100–300 mesh, Sigma Chemical Co.) with a lyotropic series of solvents. A chromatographically pure product (1.40 g) emerged with ethyl acetate–ethanol (10:1). Recrystallisation from ethyl acetate afforded 4, m.p. 189–190°, $[\alpha]_{\rm D}^{25}$ + 48.5° (c 1.0, chloroform); p.m.r. data: δ 6.06 (bd, 1 H, NH), 4.76 (d, 1 H, $J_{1,2}$ 3.5 Hz, H-1), 4.65 (d, 1 H, $J_{1',2'}$ 8.5 Hz, H-1'), 3.45 (s, ~ H, CH₃O), 2.02–2.18 (m, 15 H, AcN and 4 AcO) (Found: C, 48.58; H, 6.39; N, 2.68%. C₂₃H₃₅NO₁₅ calc.: C, 48.84; H, 6.24; N, 2.48%).

ACKNOWLEDGMENTS

The authors thank the Royal Holloway College for financial support of this investigation, and the S.R.C. for the award of a studentship (C.J.G.).

REFERENCES

- 1 R. KUHN, A. GAUHE, AND H. H. BAER, Chem. Ber., 87 (1954) 289-300.
- 2 V. P. REGE, T. J. PAINTER, W. M. WATKINS, AND W. T. J. MORGEN, Nature (London), 204 (1964) 740-742.
- 3 H. M. FLOWERS AND R. W. JEANLOZ, J. Org. Chem , 28 (1963) 1377-1379.
- 4 R. U. LEMIEUX AND H. DRIGUEZ, J. Am. Chem. Soc., 97 (1975) 4063-4069.
- 5 R. U. LEMIEUX, K. B. HENDRIKS, R. V. STICK, AND K. JAMES, J. Am. Chem. Soc., 97 (1975) 4056-4062; R. U. LEMIEUX AND H. DRIGUEZ, *ibid.*, 97 (1975) 4069-4075; R. U. LEMIEUX, D. R. BUNDLE, AND D. A. BAKER, *ibid.*, 97 (1975) 4076-4083.
- 6 E. J. BOURNE, I. R. MCKINLEY, AND H. WEIGEL, Carbohydr. Res., 25 (1972) 516-517.
- 7 A. M. YURKEVICH, S. G. VERENIKINA, E. G. CHAUSER, AND N. A. PREOBRAZHENSKII, Zh. Obshch. Khim., 36 (1966) 1746–1749.
- 8 R. J. FERRIER AND D. PRASAD, J. Chem Soc., (1965) 7429-7432.
- 9 I. R. MCKINLEY AND H. WEIGEL, Chem. Commun., (1972) 1051-1052.
- 10 D. S. ROBINSON, J. EAGLES, AND R. SELF, Carbohydr. Res., 26 (1973) 204-207.
- 11 I. O. MASTRONARDI, S. M. FLEMATTI, J. O. DEFERRARI, AND E. G. GROS, Carbohydr. Res., 3 (1966) 177-183.
- 12 I. R. MCKINLEY AND H. WEIGEL, Carbohydr. Res., 31 (1973) 17-26.
- 13 K. HEYNS, G. KIESSLING, AND D. MÜLLER, Carbohydr. Res., 4 (1967) 452-464.