9-(1-Fluoro-5-hydroxypentan-2-yl)-9H-guanine: Synthesis and Evaluation of Antiviral Activity

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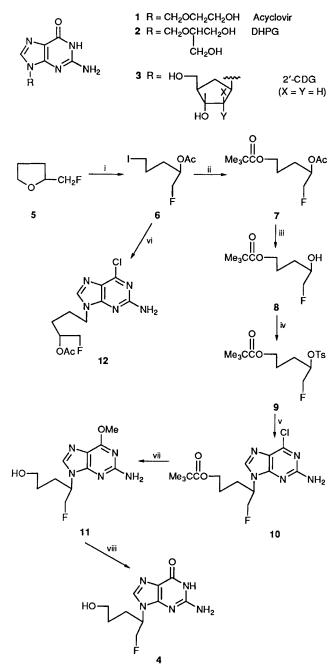
5-Fluoro-4-tosyloxypentyl pivalate has been synthesized in four steps from 2-(fluoromethyl)tetrahydrofuran. Condensation with 2-amino-6-chloropurine gave the N-9 derivative, which was converted via the 6-methoxy analogue into 9-(1-fluoro-5-hydroxypentan-2-yl)-9H-guanine. The latter was evaluated, and found inactive, in a large variety of antiviral assays.

The acyclic guanosine analogues 9-(2-hydroxyethoxymethyl)guanine 1 (acyclovir)¹ and 9-[(1,3-dihydroxypropan-2-yloxy)methyl]guanine 2 (DHPG, ganciclovir)² are potent antiviral agents which are in clinical use. Other antivirally active guanine derivatives have alkyl side-chains at N-9 in which the 2' oxygen atom has been replaced by a methylene group³ or in which the oxygen has been switched to the 3' position as in the phosphonate analogues⁴ or to the 1' position as in the 9-alkoxy series.⁵ Replacement of a hydrogen atom by fluorine does not produce drastic steric changes in a molecule ^{6a} but may enhance the biological activity of the molecule. The much greater electronegativity of fluorine has a strong electronic effect on reactions of neighbouring functional groups and the fluorine atom, being a hydrogen-bond acceptor, is often processed by enzymes acting on the corresponding hydroxy compounds.^{6b} The carbocyclic analogue of 2'-deoxyguanosine, compound 3 (X = Y = H) (2'-CDG), shows potent anti-herpes activity in vitro.⁷ Substitution of 2'-H by 2' β -F, 3 (X = F, Y = H), increased the activity one hundred-fold.⁸ The $2'\alpha$ -F derivative 3 (X = H, Y = F) was less active than 2'-CDG. A few examples of fluoroacyclic guanosine analogues have been reported.⁹⁻¹¹ Of those which showed antiviral activity, all were less potent than acyclovir. In this paper we describe the synthesis of racemic 9-(1-fluoro-5-hydroxypentan-2-yl)-9H-guanine 4, the acyclic version of 2'-CDG, lacking C-3' and with a fluorine substituent at the equivalent of the C-2' position.

Results and Discussion

Synthesis.—A convergent approach to our target molecule 4 requires, as alkylating moiety, a 1-fluoropentane with a masked hydroxy group at C-5 and a reactive leaving group at C-2. One route to this structure involves regioselective ring opening of the known 2-(fluoromethyl)tetrahydrofuran 5¹² (Scheme 1). A variety of reagents have been used to cleave the ether link in tetrahydrofuran.¹³ With unsymmetrically substituted cyclic ethers as substrates, the products depend on the reagents used and on steric factors. Thus, with acetyl toluene-p-sulfonate, 2methyltetrahydrofuran gave 5-acetoxypentan-2-yl-toluene-psulfonate (95%),¹⁴ whereas use of pivaloyl chloride-sodium iodide reagent gave 5-iodopentan-2-yl pivalate (88%).15 Cleavage with acetyl chloride-sodium iodide resulted in a 1:1 mixture of the two possible isomers. In a preliminary investigation the reaction of compound 5 with acetyl toluene-p-sulfonate was unsatisfactory. With acetyl chloride-sodium iodide reagent, attack by the iodide ion on compound 5 occurred exclusively at the less hindered position to give 1-fluoro-5-iodopentan-2-yl acetate 6. Attempts to convert the iodide into the benzyl ether were not successful using the benzyloxy anion, generated by fluorodestannylation of benzyl tributyltin ether.¹⁶ The same

procedure with tributyltin pivalate gave a high yield (94%) of the pivalate ester 7. An equally good yield was later obtained under the conditions described in a more recent publication.¹⁷ The acetate group of the diester 7 was selectively cleaved by methanolic ammonia¹⁸ to yield 5-fluoro-4-hydroxypentyl pivalate 8. Activation of the secondary hydroxy group by conversion into the tributyltin ether, 19 followed by tosylation of the crude product, gave the required pentane 9 in 79% yield. Direct tosylation of 8 gave a 40% yield of 9. The overall yield of compound 9 from compound 5 was 52%. Coupling of the tosyl ester 9 with 2-amino-6-chloropurine in dimethylformamide (DMF) with potassium carbonate, and 18-crown-6 as catalyst,²⁰ gave the N^9 -alkylated purine 10 as the major product (64%). A second, more polar, fraction consisted mainly of the N-7 isomer (8%) with two minor contaminants, as shown by ¹H NMR spectroscopy. The N-9 and N-7 isomers were distinguished by the characteristic differences in their ¹³C and ¹H NMR spectra.²¹ In the case of the N-7 isomer, the 8-H signal and C-8 signal (identified by a ¹³C DEPT spectrum) appeared downfield and the C-5 signal and NH₂ signal were upfield relative to the corresponding signals of the N-9 isomer 10. The chemical-shift and ³J-values in CDCl₃ for the fully coupled ¹³C-¹H spectrum of compound 10 agree with those reported for N⁹-alkylated 2-amino-6-chloropurine.²² The C-5 signal at $\delta_{\rm C}$ 124 was a doublet with ${}^{3}J$ 11.5 Hz; the C-4 signal at $\delta_{\rm C}$ 153.4 was a double doublet with ${}^{3}J$ 6 and 3.5 Hz. The ${}^{1}H$ and ${}^{13}C$ spectra of compound 10 differed from those of its analogue 12, thus precluding the possibility of a rearrangement of the pentyl chain during the reaction sequence. Comparison of the UV spectrum of the major product 10 with the spectra of N^9 - and N^7 -alkylated 2-amino-6-chloropurines²³ confirmed it as the N-9 isomer. Hydrolysis of pivalate 10 to compound 4 in the presence of acid (1 mol dm⁻³ HCl; reflux 4 h); or base (2.5 mol dm⁻³ NaOH; reflux 1 h) yielded foams. In both cases UV and NMR spectra were consistent with structure 4 together with contaminants. Attempts to isolate a pure sample were unsuccessful. Conversion of compound 10, by refluxing it with dry potassium carbonate in methanol, into the 6-methoxy analogue 11 with concomitant removal of the pivaloyloxy protecting group was achieved in 82% yield. The methyl ether was cleaved by treatment of compound 11 with one mole equivalent of bromotrimethylsilane (TMSBr) in DMF to afford a foam with an NMR spectrum consistent with 6-oxo compound 4 and unchanged ether 11. These were separated by chromatography. When chlorotrimethylsilane (TMSCl)-sodium iodide reagent was used for cleavage of the ether, the isolated product was shown to be the hemi-hydrate of the 1:1 sodium chloride complex of the required compound 4. The ¹H NMR spectrum of the complex showed 8-H as a doublet with $J_{8,F}$ 2.5 Hz. This long-range H ••• F coupling has been observed for β-isomers of



Scheme 1 Reagents and conditions: i, NaI, AcCl, MeCN, room temp.; ii, Me₃CCO₂SnBu₃, CsF, DMF, 40 °C; iii, NH₃–MeOH, room temp.; iv, (Bu₃Sn)₂O, toluene; then *p*-TsCl, DMAP, Et₃N, 65 °C; v, 2-Amino-6-chloropurine, K₂CO₃, 18-C-6, DMF, 65 °C; vi, 2-Amino-6-chloropurine, K₂CO₃, DMSO, room temp.; vii, K₂CO₃, MeOH, 65 °C; viii, TMSBr or TMSCl-NaI, DMF, room temp.

2'-fluoroarabinofuranosylpurines 24 and their carbocyclic analogues.⁸ The chemical shifts of ¹NH and NH₂ were downfield relative to those of the non-complexed product as a result of hydrogen bonding to the chloride anion.²⁵ The ¹H NMR spectra of the crude material from the acidic or basic aqueous hydrolyses of compound **10** resembled that of the sodium chloride-complexed product.

Evaluation of Antiviral Activity.—9-(1-Fluoro-5-hydroxypentan-2-yl)-9H-guanine (compound 4) was evaluated and found inactive at the highest concentrations (up to 400 μ g cm⁻³) tested, in the following antiviral assay systems: herpes simplex virus type 1 (strain KOS), herpes simplex virus type 2 (strain G), vaccinia virus, vesicular stomatitis virus and thymidine kinasedeficient (TK⁻) herpes simplex virus type 1 (strain B2006) in human embryonic skin-muscle (ESM) cells; varicella zoster virus (strain OKA and strain YS), cytomegalovirus (strain AD-169 and strain Davis) in human embryonic lung (HEL) cells; poliovirus type 1, Coxsackie B4 virus and respiratory syncytial virus (strain Long) in HeLa cells; influenza A (strain Ishikawa) and influenza B (strain Singapore) in Madin–Darby canine kidney (MDCK) cells; reovirus type 1, parainfluenza virus type 3, Sindbis virus, Semliki forest virus, Junin virus and Tacaribe virus in Vero cells; and human immunodeficiency virus (HIV) type 1 (strain III_B/LAI) and type 2 (strain ROD) in MT-4 cells. Compound 4 did not prove cytotoxic (again, at concentrations up to 400 μ g cm⁻³) to any of the host cells used in these studies.

Experimental

M.p.s were determined on a Gallenkamp capillary apparatus and are uncorrected; UV spectra were obtained on a Unicam SP800A spectrometer. NMR spectra were recorded on a Bruker MSL 300 machine at 300.13 MHz for ¹H and 75.468 MHz for ¹³C. SiMe₄ was the internal standard; *J*-values are given in Hz. IR spectra were recorded on a Perkin-Elmer 883 spectrometer. FAB-MS was measured at the SERC Mass Spectrometry Service Centre, Swansea using fast xenon atoms at 8 kV and 3nitrobenzyl alcohol as matrix. GLC was performed on a GOW MAC 552 with 15% DC 200 on CHROM P 90/100 mesh, $4'/\frac{1}{4}''$. Injector 230 °C, column 175 °C, detector 190 °C. TLC was carried out on Merck silica gel 60F254-coated aluminium sheets and spots were visualised by UV illumination. Column chromatography was carried out on Merck silica gel 60 (230-400 mesh) or 60 (70-230 mesh). Organic extracts were dried over magnesium sulfate and evaporated (rotary evaporator) at 30 °C unless stated otherwise.

1-Fluoro-5-iodopentan-2-yl Acetate 6.-To a stirred mixture of the tetrahydrofuran derivative 5^{12a} (520 mg, 5 mmol) and sodium iodide (1.5 g, 10 mmol) in acetonitrile (5 cm³) at 0 °C was added gradually a solution of acetyl chloride (600 mg, 7.5 mmol) in acetonitrile (10 cm³) over a period of 30 min. The mixture was stirred at ambient temperature for 65 h. Crushed ice and saturated aq. sodium hydrogen carbonate (10 cm³) were then added and the mixture was extracted with diethyl ether $(3 \times 15 \text{ cm}^3)$. The organic layer was washed successively with saturated aq. sodium thiosulfate $(2 \times 15 \text{ cm}^3)$ and brine $(2 \times 15 \text{ cm}^3)$, dried, and evaporated. The product was a pale yellow liquid (1.2 g, 82%). TLC [hexane-chloroform (1:2)] showed a single spot, with $R_f 0.7$. The product rapidly darkened at ambient temperature but was stable for up to at least six months at -20 °C. It was used without further purification to prepare compound 7. A sample was purified for analysis by column chromatography with hexane-chloroform (1:1) as eluent; $v_{max}(film)/cm^{-1}$ 1746, 1237, 1070 and 1032; $\delta_{H^{-1}}$ (CDCl₃) 5.07 (1 H, dm, J21.2, CHCH₂F), 4.47 (1 H, ddd, J47.5, 10.2 and 3.4, CHCH^aF), 4.42 (1 H, ddd, J 47.5, 10.2 and 4.9, CHCH^bF), 3.20 (2 H, t, J 6.7, 5-H₂), 2.10 (3 H, s, AcO) and 1.88 and 1.77 (4 H, 2 × m, 3- and 4-H₂); $\delta_{\rm C}$ (CDCl₃) 170 (C=O), 83.4 (d, J 174, C-1), 70.9 (d, J 19.7, C-2), 30.5 (d, J 5.6, C-3), 28.9 (C-4), 20.9 (Me) and 5.5 (C-5) (Found: C, 31.0; H, 4.0. C₇H₁₂FIO₂ requires C, 30.68; H, 4.41%).

4-Acetoxy-5-fluoropentyl Pivalate 7.—Pivalic acid (510 mg, 5 mmol) and bis(tributyltin) oxide (1.5 g, 2.5 mmol) were refluxed in toluene for 3 h using a water separator. The toluene was evaporated off and the residue was dissolved in dry DMF (5 cm^3). Caesium fluoride (760 mg, 5 mmol) was added, followed by a solution of the iodide 6 (1.37 g, 5 mmol) in DMF (5 cm^3) and the mixture was stirred at 40 °C for 40 h with the exclusion of moisture. The solvent was evaporated off and the residue was

stirred with potassium fluoride (750 mg) and ethyl acetate (25 cm³) for 1 h. The mixture was filtered through a pad of silica gel and eluted with ethyl acetate (100 cm³). The filtrate was washed with saturated aq. sodium hydrogen carbonate $(2 \times 50 \text{ cm}^3)$. dried, and evaporated. The residue was purified by column chromatography with dichloromethane-methanol (100:1) to yield the title compound 7 as a liquid (1.17 g, 94%). GLC showed a single peak, $t_{\rm R}$ 9.3 min; $v_{\rm max}$ (film)/cm⁻¹ 1750, 1734, 1285, 1237, 1163 and 1041; $\delta_{\rm H}({\rm CDCl}_3)$ 5.07 (1 H, dm, J 21.5, CHCH₂F), 4.47 (1 H, ddd, J 47.5, 10.2 and 3.4, CHCH^aF), 4.42 (1 H, ddd, J47.5, 10.2 and 4.8, CHCH^bF), 4.08 (2 H, t, J6, 1-H₂), 2.10 (3 H, s, AcO), 1.73-1.67 (4 H, br m, 2- and 3-H₂) and 1.21 (9 H, s, CMe₃); δ_C(CDCl₃) 178.4 (C=O), 170.4 (C=O), 83.3 (d, J 174, C-5), 71.7 (d, J 19.5, C-4) 63.6 (C-1), 38.7 (CMe₃), 27.1 (CMe₃), 26.2 (d, J 5.7, C-3), 24.5 (C-2) and 20.9 (MeCO) (Found: C, 58.0; H, 8.4. C₁₂H₂₁FO₄ requires C, 58.05; H, 8.53%).

5-Fluoro-4-tosyloxypentyl Pivalate 9.—A solution of the acetate 7 (1.07 g, 4.3 mmol) in methanol saturated with ammonia at 0 °C (40 cm³) was left at room temperature for 17 h. The solvent was evaporated off and the residue was purified by flash chromatography. Elution with dichloromethane-methanol (100:4) gave the alcohol 8 as a liquid (742 mg, 85%). GLC gave a single peak, t_R 5.3 min; v_{max} (film)/cm⁻¹ 3448, 1731, 1289, 1164 and 1037; $\delta_{\rm H}$ (CDCl₃) 4.40 (1 H, ddd, J 47.5, 9.4 and 3.3, CHCH^aF), 4.32 (1 H, ddd, J 47.5, 9.4 and 6.7, CHCH^bF), 4.1 (2 H, t, J 6.6, 1-H₂), 3.9 (1 H, dm, J 16, CHCH₂F), 2.3 (1 H, br s, OH), 1.8 and 1.55 (4 H, 2 × m, 2-and 3-H₂) and 1.2 (9 H, s, CMe₃); $\delta_{\rm C}$ (CDCl₃) 178.6 (C=O), 86.8 (d, J 169, C-5), 70.0 (d, J 19, C-4), 64.0 (C-1), 38.7 (CMe₃), 28.3 (d, J 6.6, C-3), 27.2 (CMe₃) and 24.7 (C-2).

A solution of the alcohol 8 (371 mg, 1.8 mmol) and bis(tributyltin) oxide (596 mg, 1 mmol) in toluene was refluxed for 17 h using a water separator and was then cooled to 65 °C. Toluene-p-sulfonyl chloride (684 mg, 3.6 mmol), triethylamine (364 mg, 3.6 mmol) and 4-(dimethylamino)pyridine (DMAP) (244 mg, 2 mmol) were added and the mixture was stirred at 65 °C for 24 h. It was then stirred for 1 h at room temperature with potassium fluoride (200 mg), filtered, and evaporated. The residual oil was purified by flash chromatography. Elution with hexane-dichloromethane (1:2) gave the *title compound* 9 as an oil (510 mg, 79%), v_{max}(film)/cm⁻¹ 1730, 1600, 1368, 1286, 1178, 1048 and 1036; $\delta_{\rm H}({\rm CDCl}_3)$ 7.80 and 7.34 (4 H, AA'BB'q, J_{AB} 8.5, ArH), 4.72 (1 H, dm, J18.6, CHCH₂F), 4.49 (1 H, ddd, J 47.0, 10.3 and 3.8, CHCH^aF), 4.34 (1 H, ddd, J 47.0, 10.3 and 4.7, CHCH^bF), 4.00 (2 H, t, J 6, 1-H₂), 2.45 (3 H, s, ArMe), 1.8 and 1.6 (4 H, 2 × m, 2- and 3-H₂) and 1.2 (9 H, s, CMe₃); $\delta_{\rm C}({\rm CDCl}_3)$ 178.4 (C=O), 145.0, 133.8 and 127.8 (Ar), 82.9 (d, J 176.2, C-5), 79.5 (d, J 20.3, C-4), 63.2 (C-1), 38.7 (CMe₃), 27.1 (d, J 5.3, C-3), 24.1 (C-2) and 21.6 (CMe₃) (Found: C, 56.7; H, 6.8. C₁₇H₂₅FO₅S requires C, 56.65; H, 6.99%).

2-Amino-6-chloro-9-(1'-fluoro-5'-pivaloyloxypentan-2'-yl)-

9H-purine 10.—A mixture of 2-amino-6-chloropurine (340 mg, 2 mmol), anhydrous potassium carbonate (276 mg, 2 mmol) and 18-crown-6 (528 mg, 2 mmol) in DMF (5 cm³) was stirred for 1 h at 65 °C. A solution of the tosyl compound 9 (721 mg, 2 mmol) in DMF (5 cm³) was added and the mixture was stirred for 65 h at 65 °C. The solvent was evaporated off and the residual oil was co-evaporated with water (2×5 cm³) and ethanol (2×5 cm⁻³) and flash chromatographed. Elution with dichloromethane-methanol (100:1.6) gave an oil (510 mg) which showed two spots, R_f 0.50 and 0.33, on TLC with chloroform-ethyl acetate (1:1). The oil was dissolved in diethyl ether-hexane 1:1 and left at -20 °C for 24 h. A solid ether clathrate separated and was air-dried at room temperature, m.p. 35-40 °C (R_f 0.50). This was vacuum-dried at 100 °C to

2109 give the *title compound* **10** (458 mg, 64%) as an oil, w. (MaOH)/mm 230 (a 7600) 240 (7510) and 212 (2220)

 v_{max} (MeOH)/nm 230 (ε 7600), 249 (7510) and 312 (8320); δ_{H} [(CD₃)₂SO] 8.23 (1 H, s, 8-H), 6.84 (2 H, br s, NH₂), 5.00– 4.66 (3 H, m, CHCH₂F), 3.97 (2 H, t, *J* 6.3, 5'-H₂), 1.97 and 1.45 (4 H, 2 × m, 3'- and 4'-H₂) and 1.1 (9 H, s, CMe₃); δ_{C} [(CD₃)₂SO] 178.6 (C=O), 160.2, 154.6 and 150.4 (Ar), 142.7 (C-8), 124 (C-5), 83.9 (d, *J* 170.8, C-1'), 63.7 (C-5'), 55.5 (d, *J* 18.5, C-2'), 27.4 (CMe₃), 25.5 (d, *J* 5.2, C-3') and 25.2 (C-4'); δ_{C} (CDCl₃) 159.0 (s, C-2), 153.4 (dd, *J* 6 and 3.5, C-4), 151.1 (s, C-6), 141 (ddd, *J* 210, 4.3 and 2.3, C-8) and 124 (d, *J* 11.5, C-5) (Found: C, 50.2; H, 6.0; N, 19.2. C₁₅H₂₁ClFN₅O₂ requires C, 50.35; H, 5.92; N, 19.57%).

Elution of the column with dichloromethane-methanol (100:3.2) gave the impure N-7 isomer of compound 10 as needles (35 mg, 8%), λ_{max} (MeOH)/nm 235 (ε 8950), 250 (4570) and 320 (7550). TLC (ethyl acetate) showed two spots, R_f 0.6 and 0.8; δ_{H} [(CD₃)₂SO] of the major constituent: 8.62 (8-H) and 6.73 (NH₂); δ_{C} [(CD₃)₂SO] 151.1 (C-8) and 115.1 (C-5). A shorter reaction time of 40 h reduced the yield of the N-9 isomer 10 to 45%.

9-(4'-Acetoxy-5'-fluoropentyl)-2-amino-6-chloro-9H-purine 12.---A mixture of the iodide 6 (612 mg, 2.2 mmol), 2-amino-6chloropurine (376 mg, 2.2 mmol) and anhydrous potassium carbonate (304 mg, 2.2 mmol) in dry dimethyl sulfoxide (DMSO) (10 cm³) was stirred at ambient temperature for 5 h. The solvent was removed at 50 °C, and the residue was triturated with dichloromethane (10 cm³) and filtered. Evaporation of the filtrate yielded a yellow oil, which was chromatographed. Elution with chloroform-methanol (100:2) gave the title compound 12 as a solid (410 mg, 65%), m.p. 122-123 °C. A portion was recrystallised from methanol, m.p. 122-123 °C; λ_{max} (MeOH)/nm 229 (ϵ 7340), 247 (5270) and 311 (7630); $\delta_{\rm H}[({\rm CD}_3)_2{\rm SO}]$ 8.15 (1 H, s, 8-H), 6.92 (2 H, s, NH₂), 5.00 (1 H, dm, J 22.5, CHCH₂F), 4.47 (1 H, ddd, J 45.2, 10.4 and 2.9, CHCH^aF), 4.42 (1 H, ddd, J 45.2, 10.4 and 5.2, CHCH^bF), 4.07 (2 H, t, J 7, 1'-H₂), 2.04 (3 H, s, Me) and 1.82 and 1.55 (4 H, $2 \times m$, 2'- and 3'-H₂); $\delta_{C}[(CD_{3})_{2}SO]$ 170 (C=O), 159.7, 154.1 and 149.3 (Ar), 143.2 (d, J 3.6, C-8), 123.3 (C-5), 83.5 (d, J 170, C-5'), 71.3 (d, J18.5, C-4'), 42.7 (C-1'), 26.0 (d, J 6.3, C-3'), 24.7 (C-2') and 20.8 (Me) (Found: C, 45.3; H, 5.0; N, 22.1. $C_{12}H_{15}CIFN_5O_2$ requires C, 45.65; H, 4.79; N, 22.18%). Further elution of the column with the same solvent gave 7-(4'acetoxy-5'-fluoropentyl)-2-amino-6-chloro-7H-purine, m.p. 152-154 °C (100 mg, 16%). A portion recrystallised from methanol had m.p. 153–154 °C (decomp.); λ_{max} (MeOH)/nm 234 (ε 9740), 250 (4570) and 323 (6160); $\delta_{\rm H}[({\rm CD}_3)_2 {\rm SO}]$ 8.38 (1 H, s, 8-H), 6.62 (2 H, s, NH₂), 5.00 (1 H, dm, J 20, CHCH₂F), 4.49 (1 H, ddd, J45.5, 10.4 and 2.9, CHCH^aF), 4.46 (1 H, ddd, J45.5, 10.4 and 5.1, CHCH^bF), 4.30 (2 H, J 7, 1'-H₂), 2.03 (3 H, s, Me) and 1.82 and 1.56 (4 H, 2 × m, 2'- and 3'-H₂); $\delta_{\rm C}[({\rm CD}_3)_2{\rm SO}]$ 170(C=O), 164.3, 159.9 (Ar) 149.4 (d, J3.8, C-8), 142.1 (Ar) 114.7 (C-5), 83.5 (d, J 169.6, C-5'), 71.3 (d, J 18.4, C-4'), 45.8 (C-1'), 26.5 (C-2'), 25.8 (d, J 6.4, C-3') and 20.7 (Me) (Found: C, 45.5; H, 5.0; N, 22.2%).

2-Amino-9-(1'-fluoro-5'-hydroxypentan-2'-yl)-6-methoxy-9H-purine 11.—A mixture of compound 10 (180 mg, 0.5 mmol) and anhydrous potassium carbonate (200 mg, 1.45 mmol) in dry methanol (10 cm³) was stirred and refluxed for 4 h. The solvent was evaporated off and the residue was purified by flash chromatography with dichloromethane-methanol (10:1). The product (112 mg, 82%) was a gum which solidified to give the *title compound* 11 as a solid, m.p. 112–114 °C; λ_{max} (water)/nm pH 7 and pH 12, 217 (5390), 249 (7290) and 281 (ε 8550); $\delta_{\rm H}[(CD_3)_2SO]$ 7.9 (1 H, s, 8-H), 6.3 (2 H, br s, D₂O exch., NH₂), 4.9–4.6 (3 H, 3 × m, CHCH₂F), 4.39 (1 H, dt, D₂O exch., OH), 4.0 (3 H, s, OMe), 3.35 (2 H, dt, D₂O converts to t, J 6.2, 5'-H₂), 1.9 (2 H, m, 4'-H₂) and 1.3 (2 H, dm, 3'-H₂); $\delta_{\rm C}[({\rm CD}_3)_2{\rm SO}]$ 160.6, 159.7 and 154.3 (Ar), 138.5 (C-8), 113.8 (C-5), 83.7 (d, J 170, C-1'), 59.9 (C-5'), 54.2 (d, J 18.7, C-2'), 53.1 (OMe), 28.6 (C-4') and 25.5 (d, J 5, C-3') (Found: C, 48.8; H, 5.9; N, 25.9. C₁₁H₁₆FN₅O₂ requires C, 49.06; H, 5.99; N, 26.01%).

9-(1'-Fluoro-5'-hydroxypentan-2'-yl)-9H-guanine Hvdrate 4.—A solution of TMSBr (0.03 cm³, 0.25 mmol) in DMF (2 cm³) was added to a solution of compound 11 (68 mg, 0.25 mmol) in DMF (2 cm³) under N_2 , and the mixture was stirred for 24 h. The solvent was removed at 40 °C and the residue was co-evaporated with methanol $(3 \times 5 \text{ cm}^3)$. The resulting yellow gum was purified by flash chromatography. Elution with dichloromethane-methanol (10:2) gave unchanged substrate 11 (21 mg). Elution with the same solvents (10:7) gave a foam (42 mg), λ_{max} (water)/nm pH 7, 255 (ϵ 12 020) and 270 (9380); pH 12, 255 (10 010) and 270 (10 510); δ_H[(CD₃)₂SO] 10.65 (1 H, s, D₂O exch., N¹H), 7.79 (1 H, s, 8-H), 6.49 (2 H, s, D_2O exch., NH_2), 4.9-4.6 (3 H, 3 × m, J 50.5 and 9.66, CHCH₂F), 4.44 (1 H, t, J 5.1, D₂O exch., OH), 3.44 after D₂O (2 H, t, J 6.3, 5'-H₂), 1.87 (2 H, m, 4'-H₂) and 1.32 (2 H, dm, 3'-H₂); $\delta_{\rm C}[({\rm CD}_3)_2{\rm SO}]$ 156.8, 153.5 and 151.4 (Ar), 136.1 (C-8), 116.5 (C-5), 83.8 (d, J 170, C-1'), 59.9 (C-5'), 54.1 (d, J 18.7, C-2'), 29.6 (C-4') and 25.7 (d, J 5.0, C-3').

9-(1'-Fluoro-5'-hydroxypentan-2'-yl)-9H-guanine-Sodium

Chloride.—To a mixture of compound 11 (112 mg, 0.4 mmol) and sodium iodide (62.5 mg, 0.42 mmol) in DMF (4 cm³), stirred under nitrogen, was added a solution of TMSCl (0.06 cm³, 0.42 mmol) in DMF (1 cm³). The mixture was stirred for 22 h. The solvent was removed at 40 °C and the residue coevaporated with methanol ($3 \times 5 \text{ cm}^3$). Flash chromatography with dichloromethane-methanol gave the title product as a foam (117 mg, 87%), λ_{max} (water)/nm 251 (ϵ 13 340) and 270 (9930); $\delta_{H}[(CD_{3})_{2}SO]$ 10.93 (1 H, D₂O exch., N¹H), 7.78 (1 H, d, J 2.5, 8-H), 6.76 (2 H, br s, D₂O exch., NH₂), 4.94-4.55 $(3 H, 3 \times m, CHCH_2F), 4.52 (1 H, t, J 5.1, D_2O exch., OH), 3.35$ visible after $D_2O(2H, t, J 6.3, 5'-H_2)$, 1.85 (2 H, m, 4'-H₂) and 1.30 (2 H, dm, 3'-H₂); $\delta_{\rm C}[({\rm CD}_3)_2 {\rm SO}]$ 156.8, 153.7 and 151.3 (Ar), 135.9 (C-8), 116.5 (C-5), 83.8 (d, J 169.9, C-1'), 59.9 (C-5'), 53.0 (d, J 18.7, C-2'), 28.6 (C-4') and 25.7 (d, J 5, C-3') (Found: C, 37.2; H, 4.6; N, 21.6; Cl, 11.1; Na, 7.1. C₁₀H₁₄FN₅O₂·Na-Cl·0.5H₂O requires C, 37.22; H, 4.69; N, 21.70; Cl, 10.99; Na, 7.12%); +ve FAB-MS m/z, 278 (M + Na⁺), 256 (M + H⁺).

Viruses and Antiviral Assays .--- The source of the viruses and methodology of the antiviral assays are described in previous publications: for herpes simplex virus²⁶ and most of the other viruses,27 varicella zoster and cytomegalovirus,28 human immunodeficiency virus²⁹ and the myxoviruses.³⁰

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References

- 1 H. J. Schaeffer, L. Beauchamp, P. de Miranda, G. B. Elion, D. J. Bauer and P. Collins, Nature (London), 1978, 272, 583.
- 2 J. C. Martin, C. A. Dvorak, D. F. Smee, T. R. Matthews and J. P. H. Verheyden, J. Med. Chem., 1983, 26, 759; K. O. Smith, K. S. Galloway, W. L. Kennell, K. K. Ogilvie and B. K. Radatus, Antimicrob. Agents Chemother., 1982, 22, 55; W. T. Ashton, J. D. Karkas, A. K. Field and R. L. Tolman, Biochem. Biophys. Res. Commun., 1982, 108, 1716.
- 3 A. Larsson, B. Oberg, S. Alenius, C. E. Hagberg, N. G. Johansson, B. Lindborg and G. Stening, Antimicrob. Agents Chemother., 1983, 23, 644.
- 4 E. De Clercq, T. Sakuma, M. Baba, R. Pauwels, J. Balzarini, I. Rosenberg and A. Holy, Antiviral Res., 1987, 8, 261; C. U. Kim, B. Y. Luh, P. F. Misco, I. Ghazzouli and J. C. Martin, J. Med. Chem., 1990, 33, 1207, and references cited therein. 5 M. R. Harnden, P. G. Wyatt, M. R. Boyd and D. Sutton, J. Med.
- Chem., 1990, 33, 187; S. Bailey, M. R. Harden, R. L. Jarvest, A. Parkin and M. R. Boyd, J. Med. Chem., 1991, 34, 57.
- 6 (a) J. Mann, Chem. Soc. Rev., 1987, 16, 381; (b) R. H. Abeles and T. A. Alston, J. Biol. Chem., 1990, 265, 16705.
- 7 Y. F. Shealy, C. A. O'Dell, W. M. Shannon and G. Arnett, J. Med. Chem., 1984, 27, 1416.
- 8 A. Borthwick, B. E. Kirk, K. Biggadike, A. M. Exall, S. Butt, S. M. Roberts, D. J. Knight, J. A. V. Coates and D. M. Ryan, J. Med. Chem., 1991, 34, 907.
- 9 P. J. Casara, M. T. Kenny and K. C. Jund, Tetrahedron Lett., 1991, 32. 3823.
- 10 M. R. Harnden, A. Parkin and P. G. Wyatt, J. Chem. Soc., Perkin Trans. 1, 1988, 2757.
- 11 A. Bzowska, E. Kulikowska, D. Shugar, B. Y. Chen, B. Lindborg and N. G. Johansson, Biochem. Pharmacol., 1991, 41, 1791.
- 12 (a) L. Kaulina, L. M. Yagupoli'skii, N. V. Kondratenki, E. P. Vechirko, A. Berzina, E. Silina, M. Lidaks and R. A. Shuk, Khim. Geterotsikl. Soedin., 1982, 246 (English translation: Chemistry of Heterocyclic Compounds, Plenum Publishing Corporation, 1982, 202); (b) J. F. Garst and F. E. Barton, J. Am. Chem. Soc., 1974, 96, 523
- 13 V. K. Yadav and A. G. Fallis, J. Org. Chem., 1986, 51, 3372.
- 14 M. H. Karger and Y. Mazur, J. Org. Chem., 1971, 36, 532.
- 15 A. Oku, T. Harada and K. Kita, Tetrahedron Lett., 1982, 23, 681
- 16 D. N. Harpp and M. Gingras, J. Am. Chem. Soc., 1988, 110, 7737.
- 17 T. Sato, J. Otera and H. Nozaki, J. Org. Chem., 1992, 57, 2166.
- 18 B. E. Griffin, M. Jarman and C. B. Reese, Tetrahedron, 1968, 24, 639.
- 19 I. D. Jenkins, J. P. H. Verheyden and J. G. Moffatt, J. Am. Chem. Soc., 1971, 93, 4323
- 20 J. R. Medich, K. B. Kunnen and C. R. Johnson, Tetrahedron Lett., 1987, 28, 4131
- 21 J. Kjellberg and N. G. Johansson, Tetrahedron, 1986, 42, 6541.
- 22 D. R. Haines, C. K. H. Tseng and V. E. Marquez, J. Med. Chem., 1987, 30, 943
- 23 A. J. H. Nollet, C. M. Hunting and U. K. Pandit, Tetrahedron, 1969, 25, 5971.
- 24 J. A. Wright, N. F. Taylor and J. J. Fox, J. Org. Chem., 1969, 34, 2632.
- 25 J. Bariyanga and T. Theophanides, Inorg. Chim. Acta, 1985, 108, 133.
- 26 E. De Clercq, J. Descamps, G. Verhelst, R. T. Walker, A. S. Jones, P. F. Torrence and D. Shugar, J. Infect. Dis., 1980, 141, 563
- 27 E. De Clercq, Antimicrob. Agents Chemother., 1985, 28, 84.
- 28 E. De Clercq, A. Holy, I. Rosenberg, T. Sakuma, J. Balzarini and P.
- C. Maudgal, Nature (London), 1986, **323**, 464. 29 R. Pauwels, E. De Clercq, J. Desmyter, J. Balzarini, P. Goubau, P. Herdewijn, H. Vanderhaeghe and M. Vandeputte, J. Virol. Methods, 1987, 16, 171.
- 30 M. Hosoya, J. Balzarini, S. Shigeta and E. De Clercq, Antimicrob. Agents Chemother., 1991, 35, 2515.

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