and therefore the neighbouring C-13 and C-8 must be substituted by oxygen functions. (c) The signal at $\delta 2.36$ attributed to the C-12 methyl group was shifted to lower field by 0.65 ppm in comparison with that of cinncassiol D₄ [6-8] indicating the presence of an ether bond between C-12 and C-13. (d) The bond formation between C-13 and C-10 accounted for one isolated methine proton at $\delta 2.61$ attributable to the methine at C-10.

The final structure deduced for cinncassiol E is that shown by 1. Cinncassiol E (1) is noteworthy because it is a novel diterpene with a new skeleton. The stereochemical evidence was not clear, but the stereochemistry of cinncassiol E was tentatively assumed to be as shown in 1 because of its probable biogenetic relationship with the cinncassiol D types of compounds.

EXPERIMENTAL

Isolation of cinncassiol E. The extraction and separation was described in the preceding paper [1], where cinncassiol E (35 mg) is referred to as compound X.

Cinneassiol E (1). FD-MS m/z: 421 [M + Na]⁺, 399 [M + 1]⁺; EI-MS m/z: 380 [M - H₂O]⁺, 352, 290, 194, 169, 149; ¹H NMR (100 MHz, pyridine-d₃): δ 1.07, 1.14 (each 3H, d, J = 6 Hz, 18-Me), 1.73 (3H, d, J = 6 Hz, 1-Me), 1.76 (3H, s, 9-Me), 2.36 (3H, s, 12-Me), 2.13, 2.65 (each 1H, d, J = 13 Hz, 14-H₂), 2.61 (1H, s, 10-H); (CD₃OD): δ 1.00 (6H, d, J = 6 Hz, 18-Me₂), 1.28 (3H, d, J = 7 Hz, 1-Me), 1.32 (3H, s, 9-Me), 1.54, 2.26 (each 1H, d, J = 13 Hz, 14-H₂), 1.60 (3H, s, 12-Me), 1.98 (1H, s, 10-H).

REFERENCES

- Yagi, A., Tokubuchi, N., Nohara, T., Nonaka, G., Nishioka, I. and Koda, A. (1980) Chem. Pharm. Bull. 28, 1432.
- Nohara, T., Nishioka, I., Tokubuchi, N., Miyahara, K. and Kawasaki, T. (1980) Chem. Pharm. Bull. 28, 1969.
- Nohara, T., Tokubuchi, N., Kuroiwa, M. and Nishioka, I. (1980) Chem. Pharm. Bull. 28, 2682.
- Kashiwada, Y., Nohara, T., Tomimatsu, T. and Nishioka, I. (1981) Chem. Pharm. Bull. 29, 2686.
- Nakano, K., Nohara, T., Tomimatsu, T. and Nishioka, I. (1981) Yakugaku Zasshi 101, 1052.
- Nohara, T., Kashiwada, Y., Tomimatsu, T., Kıdo, M., Tokubuchi, N. and Nishioka, I. (1980) *Tetrahedron Letters* 21, 2647.
- Nohara, T., Kashiwada, Y., Murakami, K., Tomimatsu, T., Kido, M., Yagi, A. and Nishioka, I. (1981) Chem. Pharm. Bull. 29, 2451
- Nohara, T., Kashiwada, Y., Tomimatsu, T. and Nishioka, I. (1982) Phytochemistry 21, 2130.
- Koda, A. and Nagai, H. (1974) Proc. Symp. Wakan-Yaku 18, 13.
- 10 Nagai, H., Ichikawa, M., Watanabe, S. and Koda, A. (1978) Proc. Symp. Wakan-Yaku 11, 51.

Phytochemistry, Vol 24, No 8, pp 1850-1852, 1985 Printed in Great Britain 0031-9422/85 \$3.00+0 00 © 1985 Pergamon Press Ltd.

KAURENIC ACID DERIVATIVES FROM STEVIA EUPATORIA

A. ORTEGA, F. J. MORALES and M. SALMÓN

Instituto de Química, Universidad Nacional Autónoma de México Circuito Exterior, Ciudad Universitaria, Coyoacán 04510, México, D F

(Revised received 4 December 1984)

Key Word Index—Stevia eupatoria; Compositae; Eupatoriae; kaurenes; 12β -ethoxy-ent-kaur-9(11),16-dien-19-oic acid, 12α -hydroxy-ent-kaur-16-en-19-oic acid

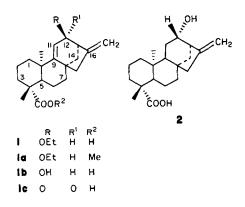
Abstract—Two kaurene type diterpenes were isolated from the aerial part of *Stevia eupatoria*. Their structures and stereochemistry were established by carbon and ¹H NMR, chemical transformation and correlation with known compounds.

INTRODUCTION

Sweet diterpene glycosides isolated from Stevia rebaudiana [1, 2] and S. paniculata [3, 4] produce, by hydrolysis, several hydroxylated kaurenoic acid derivatives. Very few compounds of the latter type have been found in Stevia species which grow in Mexico. In this paper we describe the isolation and structure determination of 12β -ethoxy-ent-kaur-9(11),16-dien-19-oic acid (1) and the known 12α -hydroxy-ent-kaur-16-en-19-oic acid (2) from S. eupatoria.

RESULTS AND DISCUSSION

The less polar fraction of S. eupatoria, afforded the ethoxy diterpenic acid (1) $C_{22}H_{32}O_3$, [M]⁺ at m/z 344. The IR spectrum revealed the presence of a hydroxyl



group of a carboxylic acid $(3300-2700 \text{ cm}^{-1})$, a carbonyl (1699 cm^{-1}) and a terminal methylene group $(1650, 980 \text{ and } 885 \text{ cm}^{-1})$. The ¹H NMR spectrum (Table 1), showed signals for two tertiary methyl groups, an exocyclic methylene and a vinylic proton of a trisubstituted double bond.

Treatment of 1 with diazomethane gave the methyl ester (1a). Its ¹H NMR spectrum exhibited the signal of a methyl of a carbomethoxy group confirming the presence of a carboxylic acid.

Compound 1 losses C_2H_4 upon treatment with acid yielding 1b, $C_{20}H_{28}O_3$, $[M]^+$ at m/z 316, which exhibited in the IR spectrum a strong hydroxyl absorption band at 3440 cm⁻¹. The ¹H NMR spectrum of 1b did not show signals for the ethoxy group and a new signal was observed, which disappeared upon D_2O addition, which was attributable to a hydroxy group.

Oxidation of the hydroxy acid 1b with Jones reagent gave an α,β -unsaturated ketone (1c), $C_{20}H_{26}O_3$, IR 1650 cm⁻¹; λ_{max}^{EiOH} 250 nm, log ϵ 3.91. The ¹H NMR spectrum (Table 1) showed a remarkable paramagnetic effect of the carbonyl group on the chemical shifts of the terminal methylene protons at δ 5.04 (1H, br s) and 5.29 (1H, br s) and for the vinylic proton at 5.28 (1H, br s).

A comparison of the ¹H NMR spectrum with data recorded for other known kaurene type diterpenes [5, 6]and the chemical shifts for **1b** and **1c** suggests that the hydroxy group and the trisubstituted double bond are located at C-12 and C-9, respectively.

A selective catalytic hydrogenation of the terminal methylene of 1c gave 1d, whose ¹H NMR spectrum showed signals for one new methyl group at $\delta 0.9$ (3H, d).

The ¹³C NMR spectrum (Table 2) of the 12β -ethoxyent-kaur-9(11),16-dien-19-oic acid (1) confirms its structural identity. Resonance assignments were made with the aid of off-resonance decoupled (SFORD) spectra, based on comparison of general chemical shift arguments and reported values of related structures as well as those of triand tetracyclic diterpenoids [7–9].

The more polar fractions afforded the 12α -hydroxy-entkaur-16-en-19-oic acid (2), $C_{20}H_{30}O_3$, $[M]^+$ at m/z 318. The IR spectrum exhibited absorptions attributable to hydroxyl (3425 cm⁻¹), carbonyl (1699 cm⁻¹) and terminal methylene (1648 and 850 cm⁻¹) groups. The ¹H NMR spectrum (Table 2) showed signals for the two tertiary methyl groups, a carbinolic proton, a terminal methylene and a trisubstituted double bond.

The position of the hydroxyl group located at C-12 was deduced by comparison of chemical shifts of the ¹H NMR spectra of 2, and with data published in the synthesis of grandiflorine acid [10]. The synthesis of 1b, 1c and compounds from the series 2 had been previously reported. The methyl ester of 1b and the ketone 1c had also been described as isolated from Vellozia caput [11], from Ambrosia hispida [12] and from Viguiera excelsa [5], respectively.

EXPERIMENTAL

Stevia eupatoria Will, was collected in September (1979), 5 km south of Ixlahuaca, State of Mexico. A voucher is on deposit at the Herbarium of the Instituto de Biología (UNAM), México.

Extraction of the whole plant (6 kg) with MeOH yielded material (130 g) which was chromatographed on 600 g of silica gel. The elution with C_6H_6 -EtOAc (9:1) gave 1 which was crystallized from hexane-Me₂CO; (0.01 %) mp 170-172°; [α]_D⁰ + 63.7° (c 0.259; CHCl₃); IR v_{CHCl₃} cm⁻¹: 3200-2700 (OH), 1699 (CO₂H), 1650, 980, 885 (C=CH₂). MS m/z (rel. int.): 344 ([M]⁺, C₂₂H₃₂O₃), 329 [M-Me]⁺ (18.33), 229 [M-C₆H₁₁O₂]⁺ (68.33), 157 [M-C₁₂H₁₁O₂]⁺ (25.8), 141 [M-C₁₄H₁₉O]⁺ (29.16), 107 [M-C₁₄H₂₁O₃]⁺ (25), 93 [M-C₁₅H₂₃O₃]⁺ (27 5). Calc. for C₂₂H₃₂O₃; C, 76.70; H, 9 36; O, 13.93. Found: C, 76.37; H, 9.57; O, 13.95%. Further column chromatography

Table 1. ¹H NMR data of compounds 1-1d and 2 (100 and 60 MHz, CDCl₃, TMS as internal standard)

н	1	1a	1b	1c	1d	2
11	5 34 d (br)	5 32 d (br)	5.25 d (br)	5.82 s (br)	5.85 s	
12			3.82 m			3 84 m
13	2.90 m	2.90 m	2.71 m	3.42 m	2.82 m	2.68 m
17	4.98 s (br)	4.97 s (br)	4.82 s (br)	5.29 s (br)		4.77 s (br)
17'	4.92 s (br)	4.87 s (br)	4 94 s (br)	5.04 s (br)	0.91 d	4 84 s (br)
18	1.26 s	1.17 s	1.16s	1.30 s	1.29 s	1 22 s
20	1.08 s	0.98 s	1.04 s	1.17 s	1.14s	1 12 s
CO₂Me		3655				
21	3.58 m	3.53 m				
22	1.24 t	1.20 t				

J (Hz); 11,12 = 4; 21, 22 = 8; 12, 13 = 2; 13, 14 = 3; 16, 17 = 6.

Table 2. ¹³C NMR chemical shift values and assignments of carbons in compounds 1 and 2 (MHz, CDCl₃, TMS internal standard)

Carbon	1	2	
1	38 25 (t)	40.68 (t)	
2	18.44 (t)	19 05 (t)	
3	29.20 (t)	38.09 (t)	
4	43.39 (s)	43.51 (s)	
5	44.87 (d)	56.86 (d)	
6	20.16 (t)	21.86 (t)	
7	40.77 (t)	33.13 (t)	
8	43 44 (s)	43.66 (s)	
9	159.87 (s)	56.12 (d)	
10	39.03 (s)	38.59 (s)	
11	116.07 (d)	41 23 (t)	
12	80.25 (d)	71.78 (d)	
13	46.42 (d)	51.32 (d)	
4	47.38 (t)	26.17 (t)	
15	44.75 (t)	49.23 (t)	
16	153.29 (s)	152.67 (s)	
17	107.98 (t)	10 4 .76 (t)	
8	23.32 (q)	28.44 (q)	
9	178.46 (s)	180.70 (s)	
20	15 73 (q)	14.5 (s)	
21	64.15 (t)		
22	28.28 (q)		

elution with C₆H₆-EtOAc (6:1) gave the 12*a*-hydroxy-*ent*-kaur-16-en-19-oic acid (2) (0.013%), which was crystallized from hexane-Me₂CO, mp 240-243°; $[\alpha]_{D}^{20}$ -36.5° (*c* 0.200; CHCl₃); IR v_{max}^{CHCl₃} cm⁻¹: 3425 (OH), 1699 (CO₂H), 1648, 850 (C=CH₂). MS *m*/z (rel. int.): 318 (M⁺, C₂₀H₃₀O₃) 107 [M - C₁₂H₁₉O₃]⁺ (100), 303 [M - Me]⁺ (3.9), 300 [M - H₂O]⁺ (25.7), 285 [M - Me - H₂O]⁺ (26.7), 93 [M - C₁₃H₂₁O₂]⁺ (53.4). Calc. for C₂₀H₃₀O₃· C, 75.43; H, 9 50; O, 15 07. Found: C, 75.50; H, 9 32; O, 14.56%.

Methyl ester of 1 The methylation of 200 mg of 1 with CH_2N_2 produced 1a after crystallization from hexane-Me₂CO (190 mg); mp 85-87°; IR $v_{max}^{CHCl_3}$ cm⁻¹. 1725 (methyl ester); 1650, 870 (C=CH₂). MS m/z (rel. int.): 358 ([M]⁺, C₂₂H₃₄O₃) (90.1), 343 [M - Me]⁺ (19.7), 229 [M - C₇H₁₃O₂]⁺ (754), 155 [M - C₁₄H₁₉O]⁺ (213), 107 [M - C₁₅H₂₃O₃]⁺ (34.4), 93 [M - C₁₆H₂₅O₃]⁺ (29.5). Calc. for C₂₃H₃₄O₃; C, 77.05; H, 9.56; O, 13.39 Found: C, 76.59; H, 9 45; O, 13 49%.

Hydrolysis of ether 1 The acid 1 (150 mg), dissolved in Me_2CO , was refluxed with 3 ml conc HCl for 30 min and monitored by TLC. The reaction mixture was diluted with H₂O, neutralized with saturated NaHCO₃ soln and extracted with CHCl₃. After usual work up the residue yielded **1b**, after hexane-Me₂CO crystallization, mp 175-180°; $[\alpha]^{20}$ + 63.81°; IR v ^{KBr}_{max} cm⁻¹: 3440 (OH), 1699 (CO₂H), 1650, 880 (C=CH₂). MS *m*/*z* (rel. int.): 316 ([M]⁺, C₂₀H₂₈O₃) (26.2), 301 [M - Me]⁺ (37.4), 298 [M - H₂O]⁺ (27.8), 283 [M - Me - H₂O]⁺ (29.5), 201 [M - C₆H₁₁O₂]⁺ (27.8), 147 [M - C₁₀H₁₇O₂]⁺ (21.58), 141 [M - C₁₂H₁₅O]⁺ (22.58), 107 [M - C₁₂H₁₇O₃]⁺ (38.7), 93 [M - C₁₃H₁₉O₃]⁺ (45.1), 41 (100). Calc. for C₂₀H₂₈O₃; C, 75.91, H, 8.92; O, 15.17. Found: C, 74.71; H, 8.83; O, 13.70%.

Preparation of 1c. A soln of 1b (100 mg) in Me₂CO (50 ml) was oxidized with 0.4 ml of Jones reagent at -20° , the reaction being monitored by TLC After 30 min the reaction was worked up as usual to give 93 mg of 1c from hexane-Me₂CO crystallization, mp 279-280°, IR $v_{max}^{CHCl_3}$ cm⁻¹. 3200-2800 (OH), 1699 (CO₂H), 1650 (C=O), 1600, 892 (C=CH₂). Calc. for C₂₀H₂₆O₃; C, 76.40; H, 8 34; O, 15.27 Found. C, 75 65; H, 8.33; O, 15.89%

Hydrogenation of 1c A soln of 79.5 mg of 1c in 15 ml of EtOAc was hydrogenated using 79 mg of PtO as catalyst. Filtration and evaporation gave 70 mg of 1d, mp 244–246°; UV λ_{max}^{MeOH} nm: 217 (£3800); IR $\nu_{max}^{CHCl_3}$ cm⁻¹: 3200–2800 (OH), 1690 (CO₂H), 1650 (C=O). Calc. for C₂₀H₂₈O₃; C, 75.91; H, 8.92; O, 15.17. Found: C, 75.23; H, 9.15; O, 15.82%.

Acknowledgement—We thank M. Rosales Hoz for many valuable discussions.

REFERENCES

- 1. Mosetting, E, Beglinger, U., Dolder, F., Lichiti, H., Quitt, P. and Waters, J. W. (1963) J. Am. Chem. Soc. 85, 2305.
- 2 Kohda, H., Kasai, K., Yamasaki, K., Murakami, K. and Tanaka, O. (1976) Phytochemistry 15, 981.
- Kohda, H., Tanaka, O. and Nishi, K. (1976) Chem. Pharm. Bull. 24, 1040.
- Yamasaki, K., Kohda, K., Kobayashi, T., Kaneda, N., Kasai, R., Tanaka, O. and Nishi, K (1977) Chem. Pharm. Bull. 25, 2895
- 5. Bohlman, F. and Levan, N. (1978) Phytochemistry 17, 1957.
- 6. Hanson, J. R. (1967) Tetrahedron 23, 801.
- 7. Herz, W. and Sharma, P. R (1976) J. Org. Chem. 41, 1021.
- 8. Wahlberg, I, Almquist, S., Nishida, T. and Enzell, R. C. (1975) Acta Chem. Scand. B 29, 1047.
- 9 Chalmers, A. A, Gorst-Allman, C P. and Piacenza, L. P. L. (1977) Tetrahedron Letters 1665
- 10. Lewis, J L and McMillan, J. (1980) J Chem. Soc. Perkin Trans 1, 1270
- 11. Pinto, A. C, Prado, S. K. and Pinchin, R. (1981) Phytochemistry 20, 520
- 12 Herz, W, Gage, D and Kumar, K. (1981) Phytochemistry 20, 581