



## Thiophenyl oxime-derived phosphonates as nano-molar class C beta-lactamase inhibitors reducing MIC of imipenem against *Pseudomonas aeruginosa* and *Acinetobacter baumannii*

Qiang Tan<sup>a,\*</sup>, Aimie M. Ogawa<sup>b</sup>, Susan L. Raghoobar<sup>b</sup>, Douglas Wisniewski<sup>b</sup>, Lawrence Colwell<sup>a</sup>, Young-Whan Park<sup>b</sup>, Katherine Young<sup>b</sup>, Jeffrey D. Hermes<sup>b</sup>, Frank P. DiNinno<sup>a</sup>, Milton L. Hammond<sup>a</sup>

<sup>a</sup> Department of Medicinal Chemistry, Merck & Co., Inc., Rahway, NJ 07065, USA

<sup>b</sup> Department of Infectious Diseases, Merck & Co., Inc., Rahway, NJ 07065, USA

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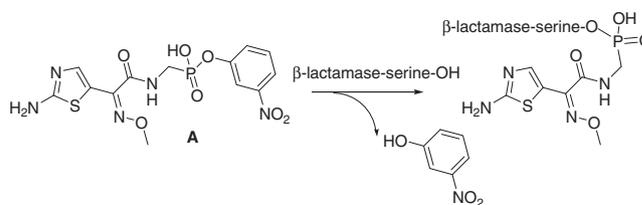
### ABSTRACT

The preparation and characterization of a series of thiophenyl oxime phosphonate beta-lactamase inhibitors is described. A number of these analogs were potent and selective inhibitors of class C beta-lactamases from *Pseudomonas aeruginosa* and *Enterobacter cloacae*. Compounds **3b** and **7** reduced the MIC of imipenem against an AmpC expressing strain of imipenem-resistant *P. aeruginosa*. A number of the title compounds retained micromolar potency against the class D OXA-40 beta-lactamase from *Acinetobacter baumannii* and at high concentrations compound **3b** was shown to reduce the MIC of imipenem against a highly imipenem-resistant strain of *A. baumannii* expressing the OXA-40 beta-lactamase. In mice compound **3b** exhibited pharmacokinetics similar to imipenem.

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Beta-lactamases, enzymes that catalyze the chemical degradation of beta-lactam antibiotics, are a leading cause of the fading efficacy of beta-lactam antibiotics, particularly in problematic Gram-negative organisms.<sup>1</sup> Co-administration of beta-lactam antibiotics with beta-lactamase inhibitors (BLI's) has been a time honored strategy to restore the clinical efficacy. Beta-lactamases have been categorized into four classes:<sup>2</sup> classes A, C and D are serine-based hydrolases; class B comprises zinc-metallo hydrolases. Unfortunately, the currently marketed beta-lactamase inhibitors, such as clavulanate, tazobactam and sulbactam, are only effective on a subset of class A beta-lactamases. Thus there is an important medical need to develop new beta-lactamase inhibitors to combat the growing threat of antibiotic resistance.

Recently phosphonate **A** was reported as a class C beta-lactamase inhibitor. Incubation of compound **A** with a susceptible beta-lactamase results in an irreversible phosphorylation of the active site serine.<sup>3</sup> While compound **A** is a valuable proof-of-principle



compound, it possesses a number of liabilities among which are moderate potency, and a problematic nitro-phenol leaving group. Moreover, it has not been reported whether compound **A** improves the efficacy of beta-lactam antibiotics against resistant bacterial strains.

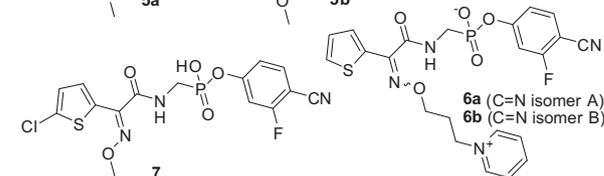
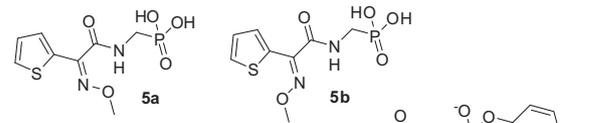
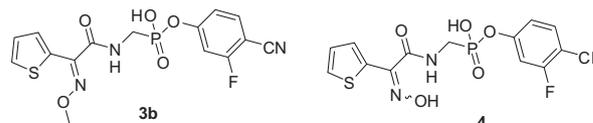
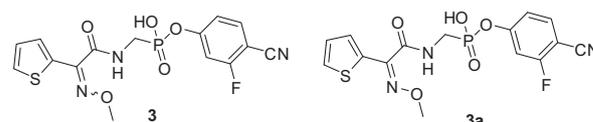
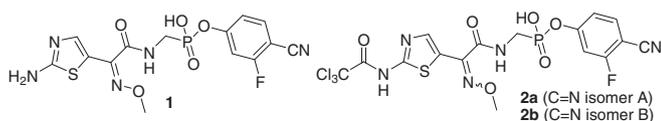
We selected **1**, a close analog of compound **A**, as an initial target for our studies, and tested it against class C as well as class A and D beta-lactamases. Similar to compound **A**, **1** carries a good phenolic leaving group; but unlike compound **A**, **1** does not have a nitrophenol as a leaving group. As Table 1 shows, **1** is a selective class C beta-lactamase inhibitor, with a low  $\mu\text{M}$   $\text{IC}_{50}$  for AmpC and P99. Initial SAR by capping the amino group as trichloroacetamide (**2a** & **2b**) did not improve potency.

\* Corresponding author.

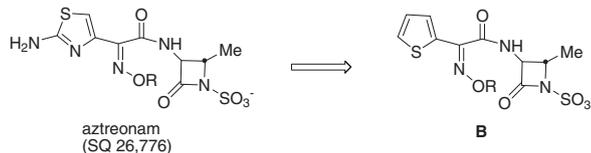
E-mail address: qiang\_tan@merck.com (Q. Tan).

**Table 1**  
IC<sub>50</sub> (μM) against different classes (bold letter in parenthesis) of beta-lactamases

Compound	TEM-1 (A) ( <i>Pseudomonas aeruginosa</i> )	SHV-5 (A) ( <i>Klebsiella pneumoniae</i> )	AmpC (C) ( <i>Pseudomonas aeruginosa</i> )	P99 (C) ( <i>Enterobacte cloacae</i> )	OXA-40 (D) ( <i>Acinetobacter baumannii</i> )
<b>1</b>	>50	>50	1.5	0.80	>50
<b>2a</b>	>50	25	3.0	0.34	>12.5
<b>2b</b>	>50	15	3.0	0.71	>12.5
<b>3</b>	>50	>50	0.0096	0.039	>50
<b>3a</b>	>50	>12.5	0.046	0.10	26.
<b>3b</b>	28	22	0.0075	0.032	17
<b>4</b>	23	27	0.20	0.10	21
<b>5a</b>	>50	>50	6.5	21	>50
<b>5b</b>	>50	>50	3.2	3.0	>50
<b>6a</b>	17	>50	0.021	0.087	27
<b>6b</b>	>50	>50	0.087	0.17	>50
<b>7</b>	16	-	0.0050	0.019	31



A carries an amino-thiazole oxime moiety common to many 2nd and 3rd generation Cephlosporins (Fig. 1), a design<sup>3</sup> intended to reduce recognition by beta-lactamases. This perhaps explains its moderate potency. Thus to improve potency we replaced the amino-thiazole with a thiophene (**3**~**7**), a design inspired by the following:



Aztreonam, a narrow spectrum monobactam antibiotic, carries a similar amino-thiazole oxime moiety. However, it has been reported that certain thiophene analogs such as compound **B** were significantly better than aztreonam in acting as a beta-lactamase inhibitor and synergizing the antibacterial activity of ceftazidime.<sup>4</sup>

Syntheses of thiophene analogs (**3**~**7**, Fig. 2) are straightforward. The published general chemistry<sup>3</sup> gave access to both oxime isomers **14** (Scheme 1). A slight modification as shown in Scheme 2

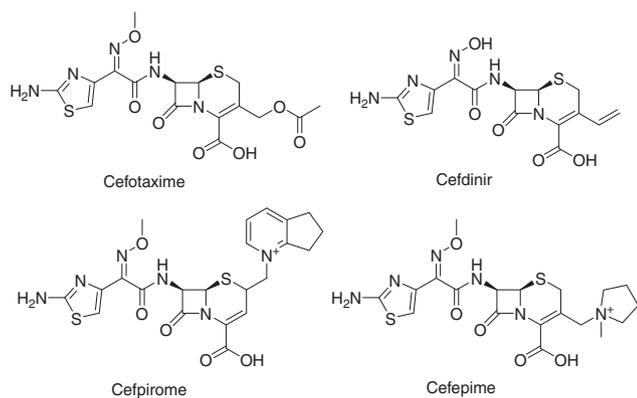
**Figure 2.** Thiophenyl oxime-derived phosphonate as novel BLI's.

provided isomer **3b** stereoselectively, employing oxime C=N bond isomerization to yield a single isomer **17** during TMSBr-induced ester cleavage of isomer mixture **16**.<sup>5</sup> Conveniently, as shown next, **3b** is the more potent isomer.

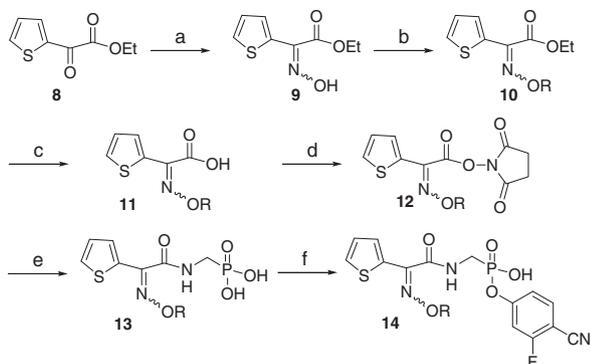
As shown in Table 1 replacement of the amino-thiazole with a thiophene resulted in a significant improvement in potency against the two class C b-lactamases tested. The Z oxime stereoisomer **3b** is more potent than the E isomer, **3a**. The desmethyl analog **4** is slightly less potent, as are **6a** & **6b**, compounds designed to modify the overall molecular charge. Compounds **5a** & **5b** confirm the presence of the phenolic leaving group is necessary for inhibitory activity, consistent with the mechanism of action. The chloro thiophene **7** exhibited potency similar to **3b**.

All of these compounds are much less potent on class A and D enzymes, with double-digit μM or more IC<sub>50</sub>. Noteworthy is that the best class C compounds, **3b** & **7**, also possesses the best class A and class D activities, suggesting that it might be possible to design an optimal compound with broad spectrum inhibitory activity.

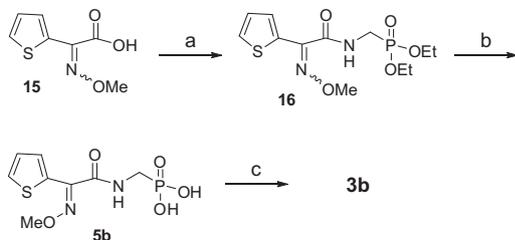
Compounds **3b** and **7** were evaluated for their ability to synergize the antimicrobial activity of imipenem against an AmpC producing strain of imipenem-resistant *Pseudomonas aeruginosa*. As shown in Table 2 co-administration with up-to-100 μM **3b** or **7**



**Figure 1.** Representative cephalosporins carrying amino-thiazole oxime moiety.



**Scheme 1.** Reagents and conditions: (a) hydroxyamine hydrochloride, pyridine, EtOH; gave ca. 1:1 mixture of oxime isomers; (b) cesium carbonate, DMF, RBr or RI; (c) KOH, MeOH, THF, water; (d) EDC, hydroxysuccinimide, dioxane, DMF; (e) (aminomethyl)phosphonic acid, sodium bicarbonate, water, DMF, EtOH, THF; (f) ArOH, trichloroacetonitrile, pyridine, 80 °C.



**Scheme 2.** Reagents and conditions: (a) HATU, Hunig's base, diethyl (aminomethyl)phosphonate hydrochloride, DMF; (b) TMSBr, DCM; (c) ArOH, trichloroacetonitrile, pyridine, 80 °C.

**Table 2**  
MIC of Imipenem against *Pseudomonas aeruginosa* (CL5701) in the presence of BLI

Imipenem MIC (μg/mL)	With <b>3b</b> (μM)	With <b>7</b> (μM)
20	0	0
10	25	12.5
5	50	25
2.5	>100	100

reduced the imipenem MIC against *P. aeruginosa* in a dose-dependent fashion. In terms of both dose-response and maximum MIC reduction, **7** is slightly more potent than **3b** in this assay (Table 1). In a control experiment, neither compound **3b** or **7** alone displayed antibiotic activity at maximum dose of 100 μM.

Highly resistant *Acinetobacter baumannii* strains are known to resist the activity of beta-lactam antibiotics by a variety of mecha-

nisms including the high level expression of both class C and class D beta-lactamases. As a proof of principle experiment we evaluated the ability of compounds **3b** and **7** to synergize the activity of imipenem against the highly imipenem-resistant strain of *A. baumannii* (CL6188).<sup>6</sup> *A. baumannii* (CL6188) expresses multiple beta-lactamases of class A, C and D. For this strain, compound **3b** was able to significantly lower the MIC of imipenem from 128 to 16 μg/mL. Interestingly, compound **7** was ineffective. No antibiotic activity was observed with either inhibitor alone at 100 μM.

Taking all the data together we decided to take a further look at **3b**. This compound in mice at 2.5 mg/kg i.v. dosing gave the following pharmacokinetics:

Half life 0.54 h; AUC(Norm) 0.98 μM/h/kg/mg; Clearance 43 mL/min/kg; volume of distribution 1 L/kg.

Its half life matches or exceeds the imipenem mouse half life;<sup>7</sup> together with the MIC reduction data, **3b** warrants further study as a potential beta-lactamase inhibitor in combination with imipenem.

In summary, we discovered a series of novel thiophenyl oxime-derived phosphonates as potent and selective class C beta-lactamase inhibitors, with weak class A and class D activities. The most potent compounds displayed suitable in vitro and PK profile suggesting further studies of their potential to enhance the efficacy of beta-lactam antibiotics, in particular imipenem, against resistant bacterial strains. These studies and further SAR exploration will be reported in due course.

## References and notes

- Matagne, A.; Dubus, A.; Galleni, M.; Frere, J.-M. *Nat. Prod. Rep.* **1999**, *16*, 1.
- Hall, B. G.; Barlow, M. J. *Antimicrob. Chemother.* **2005**, *55*, 1050.
- Nukaga, M.; Kumar, S.; Nukaga, K.; Pratt, R. F.; Knox, J. R. *J. Biol. Chem.* **2004**, *279*, 9344.
- (a) Maiti, S. N.; Setti, E. L.; Phillips, O. A.; Reddy, A. V. N.; Micetich, R. G.; Singh, R.; Higashitani, F.; Kunugita, C.; Nishida, K.; Uji, T. *PCT Int. Appl.* **1998**, WO 9847895 A1.; (b) Setti, E. L.; Maiti, S. N.; Phillips, O. A.; Reddy, A. V. N.; Micetich, R. G.; Higashitani, F.; Kunugita, C.; Nishida, K.; Uji, T.; Higashitani, F.; Hyodo, A.; Unemi, N.; Maiti, S. N.; Phillips, O. A.; Spevak, P.; Atchison, K. P.; Salama, S. M.; Atwal, H.; Micetich, R. G. *Antimicrob. Agents Chemother.* **1999**, *43*, 1895; (d) Danes, C.; Navia, M. M.; Ruiz, J.; Marco, F.; Jurado, A.; Jimenez de Anta, M. T.; Vila, J. *J. Antimicrob. Chemother.* **2002**, *50*, 261.
- <sup>1</sup>H NMR (CD<sub>3</sub>OD, 500 MHz) δ (ppm): Compound **5b** 7.91 (dd, *J* = 4.0, 1.1 Hz, 1H), 7.68 (dd, *J* = 5.1, 1.1 Hz, 1H), 7.11 (dd, *J* = 5.1, 4.0 Hz, 1H), 4.12 (s, 3H), 3.73 (d, *J* = 12.6 Hz, 2H). NOE: irradiation of -OMe protons at 4.12 ppm gave NOE signal at 7.91 ppm corresponding to proton at thiophene C-3. Compound **3b** 7.95 (d, *J* = 3.4 Hz, 1H), 7.70 (d, *J* = 4.3 Hz, 1H), 7.54 (t, *J* = 8.2 Hz, 1H), 7.14 (t, *J* = 4.5 Hz, 1H), 6.74 (dd, *J* = 8.5, 2.2 Hz, 1H), 6.70 (dd, *J* = 11.3, 2.2 Hz, 1H), 4.15 (s, 3H), 3.77 (d, *J* = 12.6 Hz, 2H).
- (a) Montefour, K.; Frieden, J.; Hurst, S.; Helmich, C.; Headley, D.; Martin, M.; Boyle, D. A. *Crit. Care Nurse* **2008**, *28*, 15; (b) Perez, F.; Endimiani, A.; Bonomo, R. A. *Exp. Rev. Anti-Infect. Ther.* **2008**, *6*, 269; Bou, G.; Martinez-Beltran, J. *Antimicrob. Agents Chemother.* **2000**, *40*, 428; **2006**, *50*, 2280.; (d) Bou, G.; Oliver, A.; Martinez-Beltran, J. *Antimicrob. Agents Chemother.* **2000**, *44*, 1556.
- An independent PK study reported imipenem half life in mice to be approximately 17 min. (Fluckiger, U.; Segessenmann, C.; Gerber, A. U. *Antimicrob. Agents Chemother.* **1991**, *35*, 1905.)