

Article

Subscriber access provided by UB + Fachbibliothek Chemie | (FU-Bibliothekssystem)

Unexpected Observation of the Dimroth Rearrangement in the Ribosylation of 4-Aminopyrimidines

Ramil Y. Baiazitov, Nadiya Sydorenko, Hongyu Ren, and Young-Choon Moon

J. Org. Chem., Just Accepted Manuscript • Publication Date (Web): 11 May 2017

Downloaded from http://pubs.acs.org on May 12, 2017

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



The Journal of Organic Chemistry is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Unexpected Observation of the Dimroth Rearrangement in the Ribosylation of 4-Aminopyrimidines

Ramil Y. Baiazitov,* Nadiya Sydorenko, Hongyu Ren, and Young-Choon Moon.

PTC Therapeutics, Inc., 100 Corporate Ct., South Plainfield, NJ 07080.

rbaiazitov@ptcbio.com

Abstract: A method for the preparation of 1-(*N*-ribofuranosyl)-6-imino-1,6-dihydropyrimidin-4amines **3** or 4-(*N*-ribofuranosyl)-6-aminopyrimidines **4** via glycosylation of 4-aminopyrimidines **2** or **5** is described. Silylated 4-aminopyrimidines **2** or **5** upon ribosylation with **1** provide products **3**. When intermediates **3** contain a strongly electron-withdrawing group, such as C(4)-Cl or C(5)-NO₂, they rearrange to products **4** in the presence of aqueous ammonia. A mechanism is proposed that involves a ring-opening/ring-closing (Dimroth) rearrangement.

Graphical abstract:



Introduction

Clitocine (Figure 1) was first isolated from the mushroom *Clitocybe inversa* in 1986.¹ The structure of this adenosine analog has been confirmed by the synthesis of the molecule and X-ray crystal structure spectroscopy.² Clitocine attracted significant attention from the scientific community because in addition to strong insecticidal activity¹, this compound also exhibits antitumor³ and read-through⁴ activities. These pharmacological properties prompted several research groups to optimize the synthesis of clitocine and its analogs³⁻⁵.



Figure 1. Structure of clitocine.

Clitocine and its analogs have been prepared following two major routes (Scheme 1). In the first route, the ribofuranosylamine *i* can react as the nucleophile with a 4-chloropyrimidine *ii* as the electrophile by an SN_{Ar} mechanism with formation of the nucleoside core.⁵ Unfortunately, nucleophile *i* is not thermally stable and is known to decompose via extrusion of ammonia and formation of the bisglycosylamine.⁶ This instability severely limits the reaction scope such that only highly activated electrophiles such as 5-nitro-, 5-sulfonyl, or 5-alkoxycarbonyl-4-chloropyrimidines, are suitable substrates. An additional drawback of this approach is that *i* consists of a mixture of anomers at C(1) resulting in the formation of mixtures of α - and β -anomeric products *iii*.

In the second route, condensation of a ribofuranosyl electrophile (such as iv) with a 4aminopyrimidine nucleophile, v, can produce the desired analog by a Vorbrüggen reaction⁷ in the presence of a Lewis acid (Scheme 1). In this route, aminopyrimidine v is usually silylated prior

The Journal of Organic Chemistry

to reaction to increase both solubility and reactivity. Surprisingly, this method has been used to prepare only a limited number of 4-(*N*-ribofuranosyl)-6-amino- (or 6-hydroxy-) pyrimidines.⁸ Moreover, it has been demonstrated that the 4-(*N*-ribofuranosyl)-6-aminopyrimidine *vii* is not the kinetic product of this reaction but rather the dearomatized 1-(*N*-ribofuranosyl)-6-imino-1,6-dihydropyrimidin-4-amine (*vi*, Y = NH, or the tautomer), which isomerizes to product *vii* upon exposure to silica gel or in acetic acid.^{8b} No mechanism for this rearrangement has been proposed.

Scheme 1



Isomerization such as from *vi* to *vii* is not common. There are numerous examples of the Vorbrüggen reaction with 4-aminopyrimidines where the endocyclic nitrogen atom, usually opposite to the exocyclic amino group, formed the glycosylic bond with the sugar residue and no isomerization followed.⁹ Apparently, the endocyclic nitrogen atom is the most nucleophilic one. Products in which the exocyclic amino group forms the glycosylic bond are rare and only a few such products could be found in literature. Moreover, it is not clear if these products were formed directly or as a result of a similar isomerization.^{8a,d} The phenomenon of a secondary isomerization requires an explanation.

In this report, we demonstrate that silylated 4,6-diaminopyrimidines during Vorbrüggen reaction attack the ribofuranoside donor by the endocyclic nitrogen atom opposite to the exocyclic amino group to form 1-(*N*-ribofuranosyl)-6-imino-1,6-dihydropyrimidin-4-amines *vi*. Depending on the nucleophile substitution, a rearrangement of *vi* can take place via the pyrimidine ring opening/closure (i. e., Dimroth rearrangement¹⁰) catalized by a nucleophile, such as ammonia. This rearrangement is facile only when the intermediate is activated by an electron-withdrawing substituent on the pyrimidine, such as a nitro-group at C(5). This limitation helps to explain the scarcity of the literature examples of such reactions. However, we found that even the intermediates without C(5) activation can isomerize when one of the NH₂-groups in the nucleophile is replaced by an electron-withdrawing Cl-group, which provides the required activation. This electron-withdrawing Cl-group overcomes this limitation and enables preparation of 4-(N-ribofuranosyl)-6-aminopyrimidines via Vorbrüggen reaction using 4-chloro-6-aminopyrimidines as the nucleophiles, followed by reaction with ammonia and in situ Dimroth-like rearrangement to provide products *vii*.

Results and Discussion

1. Reactions between Silylated 4,6-Diaminopyrimidines 2 and Ribofuranoside 1. Only a very limited number of clitocine analogs were prepared by Vorbrüggen reaction. The analogs of clitocine in which the C(5)-nitro group is replaced by hydrogen or chlorine atoms (4a, X = H; 4b, X = Cl; Table 1) were targeted first. It was expected that using conditions similar to the previously published clitocine preparation procedure (Table 1, entry 4), 4,6-diaminopyrimidine 2a and 2b will be converted to 4a (X = H) and 4b (X = Cl). The procedure^{8c} involved silylation

of the 4,6-diaminopyrimidine 2c (Table 1, X = NO₂) with HMDS, followed by Vorbrüggen reaction and optional isomerization on silica gel.^{8b}



Table 1. Reactions between 4,6-Diaminopyrimidines 2 and Ribofuranoside 1

Entry	Х	Conditions	Temp, time ^{<i>a</i>}	Product	Yield, % ^b
1	Н	BSA/TMSOTf	rt, 8 h	3a	39
2	Н	BSA/TMSOTf	60 °C, 50 min	3 a	85
3	Cl	TEA/TMSOTf	rt, 1.5 h	3b	86
4 ^{<i>c</i>}	NO ₂	HMDS/TMSOTf	rt, 18 h	4c	72
5	NO ₂	TEA/TMSOTf	rt, 16 h	4c	92

^aTemperature and time in the presence of TMSOTf. ^bIsolated yield. ^cTaken from Ref 8c.

However, in our hands, the control reaction between **1** and **2c** was irreproducible and the product of ribosylation often could not be isolated in pure form after chromatography on silica gel as judged by HPLC and NMR analyses. Moreover, the reaction was difficult to monitor by TLC or HPLC, perhaps, because of the relatively fast conversion of **3c** into **4c** during the analysis. Serendipitously, it was found that when the sample for the HPLC-MS analysis was dissolved in acetonitrile containing ammonium hydroxide, the HPLC-peak corresponding to the product **4c** could be clearly seen and no peak corresponding to **3c** could be observed, suggesting very fast isomerization from **3c** to **4c**. This fast isomerization was further confirmed when ammonium hydroxide was used for the reaction quench and upon isolation of **4c**.

In the optimized procedure, 4,6-diaminopyrimidine 2c (Table 1, X = NO₂) was combined with ribofuranoside 1 in the presence of TMSOTf (2 equiv.) and triethylamine (1 equiv) to form 4c in 92% yield upon quench with ice-cold acetonitrile-ammonium hydroxide mixture and column chromatography (Table 1, entry 5). This improved procedure for the synthesis of 4c was very reproducible in our hands.

Next, similar reactions with other 4,6-diaminopyrimidines were attempted. Unexpectedly, silylated 4,6-diaminopyrimidine **2a** (Table 1, X = H) combined with ribofuranoside **1** in the presence of TMSOTf to form **3a** as a single β -isomer (Table 1, entries 1 and 2) instead of the anticipated **4a**. Heating to 60 °C accelerated the reaction and improved the isolated yield of **3a** to 85%.

The structure of **3a** was established using 1D and 2D-NMR spectroscopy (Figure 2). For example, the ¹H-¹³C-HMBC spectrum (see Supporting Information, page S11) clearly shows coupling (the cross-peaks) that is comparable in magnitude between H(1') of the furanose ($\delta = 6.43$, d, J = 5.0 Hz, 1H) and both C(2) and C(4) of pyrimidine ($\delta = 154.0$ (C(4)), 148.7 (C(2))). Additionally, 2D-NOESY experiment (see Supporting Information, page S13) demonstrated through-space correlation between H(2) of the pyrimidine ring with both H(1') and H(2') of the furanose. The signal of H(1') in **3a** is a doublet, as opposed to the broad singlet (doublet of doublets in several other analogs) observed in **4a** (c.f. Figure 3).



Figure 2. Structural assignment of 3a

The Journal of Organic Chemistry

Reaction of the 5-chloropyrimidine **2b** proceeded in an analogous fashion to provide **3b** in 86% yield (Table 1, entry 3).¹¹ In neither case could the product of isomerization **4** be observed by HPLC or NMR analysis. Longer treatment of **3a** with ammonium hydroxide led to removal of the benzoate group at C(2') and C(3') without causing isomerization of the pyrimidine ring.¹²

2. Dimroth Rearrangement from 3 to 4. The described above failure to prepare 4a and 4b was unexpected. It was demonstrated previously ^{8b} that a similar analog 3c could be isolated and isomerized to 4c in the presence of silica gel or acetic acid, Scheme 2. The reaction mechanism was not proposed, although the acidic conditions suggest that it may involve breaking of the $C(1')-N^b$ bond in the intermediate 3c and making of the $C(1')-N^a$ bond instead in N^a-4c. This may proceed, for example, via ionization at C(1'). The lability of $C(1')-N^b$ in an aminal, such as 3c, especially under acidic conditions (such as in the presence of acetic acid or silica gel as in Ref 8b) supports this mechanism.

Scheme 2



On the other hand, it is possible, that isomerization of 3c to 4c (Scheme 3) is initiated by a nucleophilic attack onto the pyrimidine ring (NuH may be HOH or H₂NH, etc.) with a concominant ring opening to form a short-lived intermediate *xii*. Rotation of the bond in *xii* indicated on the scheme and extrusion of the nucleophilic catalyst regenerates the pyrimidine ring in N^b-4c. The difference between N^a-4c and N^b-4c is in the position of the nitrogen atoms: atoms N^a and N^b exchange positions within the pyrimidine ring.

Scheme 3



To establish which mechanism is operative, ribosylation with ¹⁵N-labelled pyrimidine **8** was conducted (Scheme 4). This intermediate was prepared by treatment of 4,6-dichloro-5-nitropyrimidine **7** with ¹⁵N-labelled ammonium hydroxide. The pyrimidine **8** was silylated and glycosylated with **1** in the presence of tin (IV) chloride. After quenching with NH₄OH, product **10** was isolated in 47 % yield.

Scheme 4



The structure of **10** was established by inspection of the magnitude of the ¹H-¹⁵N coupling constants from ¹H-NMR spectroscopy.¹³ Thus, the absolute values for the ¹J (¹H-¹⁵N) couplings in arylamines (Aryl-¹⁵NH) are expected to be around 90 Hz. Typical values for the endocyclic ²J (¹H-¹⁵N) couplings are between 0 and 20 Hz; and in pyridines it is close to 17 Hz.¹³ By comparison, for the ³J (¹H-¹⁵N) couplings the absolute values are typically between 0 and 5

The Journal of Organic Chemistry

Hz.^{14,15} ¹H-NMR analysis of **10** revealed that each of the NH₂ protons is a doublet with the characteristically large ¹J (¹H-¹⁵N) coupling constants of 90 and 95 Hz. However, the other NH signal shows a characteristically smaller ³J (¹H-¹⁵N) coupling constant value of 3.9 Hz. This suggests that the N-atom attached to the anomeric center is no longer ¹⁵N-labelled. Instead, the ring N(1)-atom is labeled, which is further supported by the characteristic ²J (¹H(2)-¹⁵N(1)) coupling constant value of 16 Hz. By comparison, in **8** this H(2) is a singlet (⁴J (¹H-¹⁵N) ~ 0 Hz).

The position of the labelled ¹⁵N-atoms in structure of **10** supports the mechanism presented in Scheme 3. This result proves that the isomerization of a kinetic product **9** (or **3c**) into the final product **10** (or **4c**) is possible through the pyrimidine ring opening when the ring is activated enough. This attack by a nucleophilic catalyst onto **3c** is faster than onto **3a** and **3b**, which contain the less electrophilic pyrimidine rings. As a consequence, **3a** and **3b** do not isomerize under these conditions. This result does not preclude the possibility of an alternative mechanism that may be operative when the rearrangement is conducted in the presence of silica gel or acetic acid.

3. Reactions between Silylated 4-Chloro-6-Aminopyrimidines and Ribofuranoside 1. Another serendipitous discovery, which also involves Dimroth rearrangement, enabled the preparation of the desired 4a as well as several other analogs. Silylation (BSA/TMSOTf) of 4chloro-6-aminopyrimidine (5a, X= H, Table 2, entry 1) followed by treatment with 1 afforded exocyclic product 4a (X = H) as a single β -isomer after quenching the mixture with ammonium hydroxide. This ribosylation is much faster than the formation of the endocyclic product 3a during the reaction with 2a. Complete conversion is observed after only 1 h at room temperature in the presence of TMSOTf (2 equiv). Upon quenching into an ice-cold mixture of acetonitrile/ammonium hydroxide (approx. 10/1) **4a** was isolated in 89% yield.





Entry	Х	Conditions	Time, temp	Product	Yield, %
1	Н	BSA/TMSOTf	rt, 1 h	4a	89
2	Cl	BSA/TMSOTf	0 °C, 1 h	$4\mathbf{b}^{a}$	65
3	Me	BSA/TMSOTf	0 °C, 1 h	4d	100
4	OMe	HMDS/SnCl ₄	0 °C, 1h	4e	73
5	F	HMDS/SnCl ₄	0 °C, 2.5 h	4f - β^b	26
6	F	BSA/TMSOTf	rt, 1 h	4f- β / 4f- α ~ 2.7	34 (+ 45 % of 3f)

^{*a*}*Endo*-product **3b** was observed by reversed-phase LC-MS¹⁶ but not isolated (~ 10 % relative to the exoproduct, UV-254 nm). *Exo*-product **4b**: M+H⁺ = 589, 1.41 min; endo-product **3b**: M+H⁺ = 589, 1.17 min. ^{*b*}*Endo*-product **3f** was observed by LC-MS but not isolated (~ 30 % peak area relative to the exo-product, UV-254 nm). *Exo*-product **4f**: M+H⁺ = 573, 1.34 min; endo-product **3f**: M+H⁺ = 573, 1.16 min.

The structure of **4a** was established via 1D and 2D-NMR spectroscopy. For example, the COSY spectrum (see Supporting Information, pages S32, S33) shows a coupling between the N(H) linking the two rings and H(1') of the furanose. A similar cross-peak in the COSY spectrum of **3a** is absent (see Supporting Information, page S10). Unlike the HMBC-spectrum of **3a**, there is no coupling in HMBC spectrum of **4a** between H(1') and any of the aromatic carbon

The Journal of Organic Chemistry

atoms (see Supporting Information, page S34). The N(H) signal is a doublet in DMSO- d_6 and the H(1') signal is a doublet of doublets in some analogs (a broad signal in others).

The reaction scope is quite broad as illustrated by the examples in Table 2. Substrates bearing electron donating methyl- and methoxy-groups at C(5) of the pyrimidine are viable (**2d**, entry 3 and **2e**, entry 4). As entry 4 in Table 2 demonstrates, HMDS can be used for silylation instead of BSA, and SnCl₄ can replace TMSOTf as the Lewis acid.^{17,18}

Substrates bearing electron-withdrawing groups at C(5) are less viable. For example, while the 5-chloropyrimidine (entry 2) provided product **4b** in 65% yield, a trace amount of, presumably, product **3b** could be observed by LC-MS analysis of the crude reaction mixture. Reaction of the 5-fluoropyrimidine was even more complex (Table 2, entries 5 and 6). Thus silylation of pyrimidine **5f** with BSA followed by ribosylation using TMSOTf as the Lewis acid (entry 6) afforded **4f** in 34% yield as a mixture of two epimers at C(1') together with a 45% of **3f**. Use of the HMDS/SnCl₄ combination (Table 2, entry 5) afforded the pure β -isomer of **4f** albeit in only 26% yield. The α -isomer was not observed in this case; **3f** was also formed, but could not be isolated in pure form.

4. Proposed Mechanism of the Reaction between the Silylated Aminopyrimidines with Ribofuranoside 1. Formation of the different products upon a small change (3 from 2 vs. 4 from 5) of the nucleophile structures requires an explanation. It may seem that in silylated 6-chloro-4-aminopyrimidines 5 the most nucleophilic nitrogen atom is the exocyclic one, while in the 4,6-diaminopyrimidine 2 the most nucleophilic nitrogen atom is the endocyclic one. However, another explanation is possible, which involves the initial formation of the same product of the endocyclic attack in both cases, followed by isomerization, which is possible only when the chlorinated nucleophile is used.

For example, the reaction can start with the attack of the N(3) atom of the silvlated

pyrimidine *ix* (Scheme 5), onto the cationic intermediate *viii* from the convex face with formation of intermediate *x*. If the intermediate *x* contains the electron-withdrawing chlorogroup at C(4) (Z = CI), then the pyrimidine ring in *x* may be electronically activated enough to be opened with ammonia during the subsequent reaction quench providing intermediate *xi* by the attack at C(2). Upon Cl-atom displacement in *xi* with ammonia and C=C bond isomerization, intermediate *xii* forms, which cyclizes with extrusion of ammonia and regeneration of the aromatic pyrimidine 4. This process is very similar to the isomerization from 3c to 4c, Scheme 3. In each case the dearomatized intermediate is electronically activated towards attack by ammonia followed by Dimroth rearrangement and aromatization. The difference is in the nature and position of the activating group: C(4)-Cl in *x vs.* C(5)-NO₂ in 3c. Formation of 3f (Table 2, entries 5 and 6) may be explained by the alternative attack with ammonia at C(4) of the chlorinated intermediate *x* (Z = CI). Similar to 3a or 3b (but unlike 3c), pyrimidine 3f cannot be opened with ammonia at the conditions studied because it is not activated enough.

Scheme 5



The Journal of Organic Chemistry

If the mechanism described in Scheme 5 is operative, then the bold nitrogen atom in 4, was not originaly there but comes from the ammonia used in the quench. To confirm the proposed mechanism, ¹⁵N-labelled ammonia was used for the reaction quench, Scheme 6. As would be expected from the mechanistic proposal, the product $4a^{-15}N$ (which was isolated in 48% yield) incorporated the label at the endocyclic N(2) position.

Scheme 6



5. The Structure Elucidation of 4a-¹⁵N. The position of the ¹⁵N-atom in 4a-¹⁵N was determined by ¹H-NMR analysis (Figure 3). ¹⁵N-NMR spectra of several pyrimidines 4 in CD₃CN also were recorded. The analysis of the ¹⁵N-NMR spectra is complicated because we were unable to reference them according to IUPAC recommendations. The ¹H-NMR spectrum (DMSO- d_6) of 4a-¹⁵N features a doublet for H(C(2)) due to the ²J (¹H-¹⁵N) coupling to the neighboring ¹⁵N-atom. The magnitude of this coupling (15.1 Hz) is within the range for the absolute values typical for the endocyclic ${}^{2}J$ (${}^{1}H{}^{-15}N$) couplings (between 0 and 20 Hz; and in pyridines it is close to 17 Hz).¹³ In contrast, in the ¹⁴N-analog **4a** this signal is a barely resolved doublet (J = 0.6 Hz) at 500 MHz. The magnitude (15.1 Hz) of this ¹H-¹⁵N coupling in 4a-¹⁵N is consistent with the structure $4a^{-15}N$. This value (15.1 Hz) is matched by the ¹⁵N-NMR data¹⁹ of 4a-¹⁵N, which exhibits a doublet at 223 ppm (J = 15.6 Hz). Additionally, the signal for H-N connected to C(4) is a doublet of doublets (J = 2.2, 9.5 Hz, in the unlabeled analog 4a this signal is a doublet, J=9.1 Hz). The magnitude of the smaller coupling constant (2.2 Hz) is consistent with ${}^{3}J({}^{1}H-{}^{15}N)$, further confirming the structure. Unfortunately, the signal for H(C(5)) could not be analyzed due to an overlap; $H_2N(C(6))$ is a broad singlet.



Figure 3. ¹H-NMR spectra of 4a and 4a-¹⁵N confirming the position of the ¹⁵N-atom.

ACS Paragon Plus Environment

Conclusions

A method of preparation of 1-(N-ribofuranosyl)-6-imino-1,6-dihydropyrimidin-4-amines **3** and 4-(N-ribofuranosyl)-6-aminopyrimidines **4** via glycosylation of 4-aminopyrimidines is presented. Upon Vorbrüggen reaction with a ribofuranosyl donor **1** silylated 4-aminopyrimidines attack by the endocyclic nitrogen atom, opposite to the exocyclic amino group. When sufficiently activated and in the presence of ammonium hydroxide, the kinetic product **3** can isomerize to **4**. The activation can be provided by C(4)-Cl or C(5)-NO₂ groups. The formation of **4** from **3** involves ring-opening of the intermediate. Extension to other substrates is possible.²⁰

Experimental Section

¹H and ¹³C NMR spectra were recorded on a Bruker 500 MHz NMR spectrometer (500 MHz ¹H, 126 MHz ¹³C) in methanol- d_4 , acetone- d_6 , or DMSO- d_6 . Data are reported in the following order: chemical shift in ppm (δ); multiplicities are indicated (br (broadened), s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet)); coupling constants, *J*, are reported in hertz (Hz); integration is provided and assignment when possible. ¹H and ¹³C NMR assignments are corroborated by 2D experiments (COSY, HMQC, HMBC, and NOESY) when relevant. Spectra are available in the SI. HRMS analysis (ESI Positive) was performed on an LTQ Orbitrap Discovery mass spectrometer. UPLC-MS analysis was conducted using a column Acquity UPLC HSS C18 1.8 μ M; Solvent A: 0.1 % aqueous formic acid, Solvent B: acetonitrile; gradient from A:B=95:5 to A:B=5:95 over 2 min or similar. Analytical thin-layer chromatography was performed on silica gel plates with F-254 indicator. Visualization was accomplished by UV light. Column chromatography was performed with silica gel. Anhydrous solvents were purchased from Acros. All commercially available reagents were purchased and used without

further purification. All temperatures refer to the external aluminum heat block temperature unless otherwise noted, all reactions were conducted in test tubes equipped with a stir bar, a septum, and a nitrogen inlet through a needle.

(2R,3R,4R,5R)-2-((6-Amino-5-nitropyrimidin-4-yl)amino)-5-

((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (4c). To 5-nitro-2,6diaminopyrimidine (2c, 465 mg, 3 mmol) in a test tube cooled in a room temperature water bath was added acetonitrile (6 mL) and triethylamine (0.42 mL, 3 mmol), followed by dropwise addition of TMSOTf (1.08 mL, 6 mmol). The mixture was stirred at room temperature for 1 h, then cooled in ice bath and 1 (1.51 g, 3 mmol) was added in one portion. The reaction mixture was stirred at room temperature for 16 h, then was quenched with an ice-cold mixture of acetonitrile and ammonium hydroxide (10/1, 20 mL), partitioned between water and ethyl acetate, washed with brine, dried with MgSO₄, concentrated and purified by chromatography on silica gel (gradient hexanes/ethyl acetate from 1/0 to 60/40 to provide the title compound 4c as a white solid material (1.65 g, 92 %). The ¹H-NMR spectrum of this material matched previously reported data.^{8c}

(2R, 3R, 4R, 5R)-2-(4-Amino-6-iminopyrimidin-1(6H)-yl)-5-

((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (**3a**). To 4,6-diaminopyrimidine (**2a**, 110 mg, 1 mmol) was added (3R,4R,5R)-2-acetoxy-5-((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl dibenzoate **1** (504 mg, 1 mmol, 1 equiv), the tube was flushed with nitrogen, and acetonitrile (3 mL) was added, followed by bis(trimethylsilyl)acetamide (BSA, 0.5 mL, 2 mmol). The reaction mixture was heated to 90 °C under nitrogen for 10 min, then cooled in an ice bath. TMSOTf (0.36 mL, 2 mmol) was added and the contents were heated to 60 °C for 1 hour, when

the UPLC/MS analysis showed complete consumption of 1. The reaction mixture was guenched into an ice-cold mixture of acetonitrile (20 mL) and ammonium hydroxide (2 mL), extracted with toluene, washed with water, brine, dried with MgSO₄, concentrated and purified by chromatography on silica gel (gradient dichloromethane/ethyl acetate from 1/0 to 0/1, then ethyl acetate/trimethylamine (50/1, then 20/1), then ethyl acetate/methanol $\sim 10/1$ to provide an oily residue. The material was triturated with ether while cooling in an ice bath, diluted with equal volume of pentane, filtered, and washed with pentane, then dried under vacuum to provide the title compound **3a** as a white solid material (473 mg, 85 %): TLC (streaking in EtOAc/MeOH \sim 10/1) Rf ~ 0.44-0.72; ¹H NMR (500 MHz, methanol-d₄) δ = 8.58 (s, 1H, H(2)), 8.09 (dd, J=1.3, 8.2 Hz, 2H), 8.04 (dd, J=1.1, 8.4 Hz, 2H), 7.97 (dd, J=1.3, 8.5 Hz, 2H), 7.68 - 7.60 (m, 3H), 7.52 $(t, J=7.9 \text{ Hz}, 2\text{H}), 7.48 - 7.40 \text{ (m, 4H)}, 6.36 \text{ (d, } J=5.0 \text{ Hz}, 1\text{H}, \text{H}(1')), 6.05 \text{ (t, } J=6.0 \text{ Hz}, 1\text{H}, 1^{-1})$ H(2')), 5.94 (t, J=5.8 Hz, 1H, H(3')), 5.78 (s, 1H, H(5)), 4.97 (td, J=3.6, 5.4 Hz, 1H, H(4')), 4.86 (t, J=4.4 Hz, 2H, H(5')); ¹H NMR (500 MHz, DMSO-d₆) ²¹ δ = 8.72 (s, 1H, H(2)), 8.22 - 8.04 (m, 6H, includes NH and NH₂), 7.99 (dd, J=7.6, 15.4 Hz, 4H), 7.79 - 7.64 (m, 3H), 7.58 (t, J=7.6 Hz, 2H), 7.52 (g, J=7.8 Hz, 4H), 6.47 (d, J=5.0 Hz, 1H, H(1')), 6.08 (t, J=5.7 Hz, 1H, H(2')), 5.98 (t, J=6.0 Hz, 1H, H(3')), 5.73 (s, 1H, H(5)), 4.97 - 4.76 (m, 3H) ¹³C NMR (126 MHz, DMSO-d₆) $\delta = 165.5, 164.56, 164.54, 162.2$ (C(6)), 154.0 (C(4)), 148.7 (C(2)), 134.1, 134.00, 133.7, 129.49, 129.45, 129.3, 129.1, 128.84, 128.7, 128.5, 128.3, 87.7, 80.9 (C(5)), 80.1 (C(4')), 73.4 (C(2')), 69.9 (C(3')), 63.5 (C(5')). One signal is missing, probably, due to overlap. UPLC: 1.05 min, 555 $[M+H]^+$, also 445, 341, 201; HRMS (ESI) $[M+H]^+$ calcd for $C_{30}H_{27}N_4O_7^+$ 555.1874, found 555.1857.

(2R,3R,4R,5R)-2-(4-(l4-Azanyl)-5-chloro-6-iminopyrimidin-1(6H)-yl)-5-((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (**3b**). 5-chloropyrimidine-4,6-diamine

2b (144 mg, 1.0 mmol) and trimethylamine (0.14 mL, 1.0 mmol) were mixed in dry acetonitrile (2 mL) under N₂. The mixture was cooled to 0 °C in an ice-water bath and trimethylsilyl trifluoromethanesulfonate (0.4 mL, 2.2 mmol) was added slowly. The reaction was warmed up to room temperature and allowed to stir for 1 hour at room temperature. It was then cooled to 0 °C in an ice-water bath and 1 (504 mg, 1.0 mmol) was added after which the reaction mixture was stirred at room temperature for 1.5 hours until UPLC showed complete consumption of 1. It was then guenched with 20 mL 10% NH₄OH/acetonitrile solution at 0 °C, concentrated and purified using silica gel column chromatography (eluent 20% ethyl acetate/hexane to 100% ethyl acetate) to yield the title compound **3b** as a white solid material (505 mg, 86 %). ¹H NMR (500 MHz, methanol-d₄) $\delta = 8.64$ (s, 1H, H(2)), 8.11 - 8.07 (m, 2H), 8.05 - 7.98 (m, 4H), 7.68 - 7.62 (m, 3H), 7.54 - 7.49 (m, 2H), 7.48 - 7.42 (m, 4H), 6.46 (d, J=4.1 Hz, 1H, H(1')), 6.07 (dd, J=3.8, 5.7) Hz, 1H, H(2')), 5.95 (t, J=6.0 Hz, 1H, H(3')), 5.01 (td, J=3.8, 6.0 Hz, 1H, H(4')), 4.87 (dd, J=1.7, 3.8 Hz, 2H, H(5')); ¹H NMR (500 MHz, acetone) $\delta = 8.95$ (s, 1H, H(2)), 8.24 - 8.06 (m, 6H, includes NH₂), 8.06 - 7.99 (m, 2H), 7.87 (br. s, 1H), 7.72 - 7.67 (m, 2H), 7.65 (tt, J=1.3, 7.6 Hz, 1H), 7.60 - 7.37 (m, 6H), 6.81 (d, J=3.8 Hz, 1H, H(1')), 6.27 (dd, J=3.8, 5.4 Hz, 1H, H(2')), 6.13 (dd, J=5.7, 6.6 Hz, 1H, H(3')), 5.24 (ddd, J=2.8, 4.0, 6.7 Hz, 1H, H(4')), 5.03 (dd, J=4.1, 12.9 Hz, 1H, HH(5')), 4.94 (dd, J=2.8, 12.9 Hz, 1H, HH(5')); ¹³C NMR (126 MHz, Acetone) $\delta =$ 166.69, 166.65, 165.8, 160.1 C(6), 152.4 C(4), 146.9 (C(2)), 135.2, 134.8, 134.5, 130.9, 130.7, 130.6, 129.8, 129.72, 129.71, 129.6, 123.4, 120.9, 91.29 (C5), 91.25 (C(1')), 82.5 (C(4')), 75.5 (C(2')), 70.6 (C(3')), 63.9 (C5')); UPLC: 1.39 min, 589 [M+H]⁺, also 297, 282; HRMS (ESI) $[M+H]^+$ calcd for $C_{30}H_{26}CIN_4O_7^+$ 589.1485, found 589.1461.

An alternative procedure for (2R,3R,4R,5R)-2-(4-(14-Azanyl)-5-chloro-6-iminopyrimidin-1(6H)-yl)-5-((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (**3b**). To **1** (504 mg, 1.0

mmol), 5-chloropyrimidine-4,6-diamine **2b** (144 mg, 1.0 mmol, 1 equiv.) and N,Obis(trimethylsilyl)acetamide (0.5 mL, 2.0 mmol, 2 equiv.) in dry acetonitrile (3mL) were heated to 90 °C under N₂ for 10 minutes after which clear solution was formed. The reaction mixture was then cooled in an ice-water bath and trimethylsilyl trifluoromethanesulfonate (0.36 mL, 2.0 mmol) was added slowly. The reaction mixture was stirred for 1.5 hours at room temperature and then heated to 60 °C for 2.5 hours until UPLC analysis showed complete consumption of **1**. The reaction was then quenched with 20 mL 10% NH₄OH/acetonitrile solution at 0 °C, concentrated and purified using silica gel column chromatography (eluent 20% ethyl acetate/hexane to 100% ethyl acetate) to yield the title compound **3b** as an off-white solid material (124 mg, 21%). The ¹H-NMR and ¹³C-NMR spectra of this material match the one obtained via a different procedure (see above).

(2R,3R,4R,5R)-2-((6-Aminopyrimidin-4-yl)amino)-5-((benzoyloxy)methyl)tetrahydrofuran-

3,4-diyl Dibenzoate (4a). To 4-chloro-6-aminopyrimidine (5a, 130 mg, 1 mmol) was added 1 (504 mg, 1 mmol, 1 equiv.), the tube was flushed with nitrogen, and acetonitrile (3 mL) was added, followed by bis(trimethylsilyl)acetamide (BSA, 0.25 mL, 1 mmol, 1 equiv.). The reaction mixture was heated to 90 °C under nitrogen for 10 min, then cooled in an ice bath. TMSOTF (0.36 mL, 2 mmol, 2 equiv.) was added and the contents were stirred at room temperature for 1 hour, when the UPLC/MS analysis showed almost complete consumption of both starting materials. The reaction mixture was quenched into an ice-cold mixture of acetonitrile (20 mL) and ammonium hydroxide (2 mL), extracted with toluene/ethyl acetate ~ 1/1, washed with water, brine, dried with MgSO₄, concentrated and purified by chromatography on silica gel (gradient dichloromethane/ethyl acetate from 1/0 to 0/1, then ethyl acetate/methanol ~ 20/1 to 10/1 to provide the title compound **4a** as a white solid material (495 mg, 89 %): ¹H NMR (500 MHz,

DMSO-d₆) $\delta = 8.05 - 8.02$ (m, 2H), 7.96 (d, *J*=0.6 Hz, 1H, H(2)), 7.91 - 7.88 (m, 2H), 7.87 - 7.84 (m, 2H), 7.83 (d, *J*=9.1 Hz, 1H, NH), 7.70 - 7.60 (m, 3H), 7.54 - 7.50 (m, 2H), 7.49 - 7.41 (m, 4H), 6.33 (s, 2H, NH₂), 6.04 (br. s., 1H, H(1')), 5.80 (dd, *J*=4.4, 5.7 Hz, 1H, H(3')), 5.62 - 5.56 (m, 2H, H(2') and H(5)), 4.59 - 4.49 (m, 3H, H(4' and 5')); ¹³C NMR (126 MHz, DMSO-d₆) $\delta = 165.5$, 164.8, 164.7, 163.9 (C(4) or C(6)), 161.5 (C(4) or C(6)), 157.7 (C(2)), 133.8, 133.7, 133.5, 129.34, 129.28, 129.23, 129.20, 128.78, 128.71, 128.67, 128.65, 128.58, 83.9 (C(5)), 83.4 (C(1')), 77.3 (C(4')), 73.7 (C(2')), 71.1 (C(3')), 64.3 (C(5')); UPLC/MS: 1.16 min, 555 [M+H]⁺; HRMS (ESI) [M+H]⁺ calcd for C₃₀H₂₇N₄O₇⁺ 555.1874, found 555.1849.

(2R, 3R, 4R, 5R)-2-((6-Amino-5-methylpyrimidin-4-yl)amino)-5-

(*(benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate* (**4d**) was prepared following the same procedure as for **4a**. Yield: 574 mg (100 %) of white solid material. TLC R_f ~ 0.39 (EtOAc); ¹H NMR (500 MHz, DMSO-d₆) δ = 8.11 (s, 1H, H(2)), 8.03 - 7.99 (m, 2H), 7.93 - 7.89 (m, 2H), 7.85 - 7.82 (m, 2H), 7.79 (d, *J*=8.8 Hz, 1H, NH), 7.69 - 7.59 (m, 3H), 7.53 - 7.45 (m, 4H), 7.44 - 7.39 (m, 2H), 6.88 (br. s., 2H, NH₂), 6.26 (dd, *J*=5.0, 9.1 Hz, 1H, H(1')), 5.88 (t, *J*=5.7 Hz, 1H, H(3')), 5.84 (dd, *J*=4.7, 6.0 Hz, 1H, H(2')), 4.63 - 4.55 (m, 2H, H(4'+5a')), 4.51 (dd, *J*=4.3, 11.2 Hz, 1H, H(5b')), 1.89 (s, 3H, Me); ¹³C NMR (126 MHz, DMSO-d₆) δ = 165.5, 164.8, 164.7, 158.7, 157.7, 150.9 (C(2)), 133.9, 133.8, 133.5, 129.32, 129.30, 129.28, 129.22, 128.8, 128.71, 128.68, 128.63, 128.59, 91.3 (C(5)), 84.0 (C(1')), 77.3 (C(4')), 73.8 (C(2')), 70.8 (C(3')), 63.9 (C(5')), 8.9 (Me). UPLC: 1.18 min, 569 [M+H]⁺; HRMS (ESI) [M+H]⁺ calcd for C₃₁H₂₉N₄O₇⁺ 569.2031, found 569.2005.

(2R,3R,4R,5R)-2-((6-Amino-5-methoxypyrimidin-4-yl)amino)-5-((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (**4e**).

The Journal of Organic Chemistry

Mixture of 6-chloro-5-methoxypyrimidin-4-amine **5e** (159 mg, 1.0 mmol), $(NH_4)_2SO_4$ (159 mg, 1.2 mmol) and pyridine (0.6 mL) in hexamethyldisilazane (5 mL) under inert atmosphere (N_2) was heated to 140 °C in a sealed tube over a period of 1 h. The reaction mixture was cooled and concentrated under reduced pressure. To the crude silvlated pyrimidine was added 1 (504 mg, 1.0 mmol) and dry acetonitrile (3 mL). The resulting mixture was cooled in an ice-water bath and SnCl₄ (1.0 M solution in CH₂Cl₂, 2 mL, 2 mmol) was added dropwise. After 1 h at 0 °C the UPLC analysis showed complete consumption of 1. The mixture was then guenched with 20 mL of 10% NH₄OH/acetonitrile solution at 0 $^{\circ}$ C. The white precipitate was removed by filtration, the filtrate was concentrated and purified using silica-gel column chromatography (eluent 10% ethyl acetate/hexane to 100% ethyl acetate) to yield the title compound 4e as a white solid material (428 mg, 73%). ¹H NMR (500 MHz, Acetone-d₆) δ = 8.11 (d, J=7.88 Hz, 2 H), 7.99 (dd, J=14.03, 8.04 Hz, 4 H), 7.86 (s, 1 H, H(2)), 7.57 - 7.68 (m, 3 H), 7.50 (t, J=7.57 Hz, 2 H), 7.45 (q, J=7.88 Hz, 4 H), 6.99 (d, J=9.60 Hz, 1 H, NH), 6.43 (dd, J=9.60, 6.00 Hz, 1 H, H(1')), 5.96 (dd, J=5.99, 4.41 Hz, 1 H, H(3')) 5.92 (dd, J=6.30, 6.00 Hz, 1 H, H(2')), 5.75 (br. s., 2 H, NH₂), 4.71 (dd, J=11.03, 3.47 Hz, 1 H, H(5'a)), 4.65 (q, J=4.40 Hz, 1 H, H(4')), 4.62 (dd, J=11.00, 4.40 Hz, 1 H, H(5'b)), 3.59 (s, 3 H, OMe). ¹³C NMR (126 MHz, Acetone) $\delta =$ 166.6, 166.0, 165.9, 157.5 (C(4) or C(6)), 154.9 (C(4) or C(6)), 153.7 (C2)), 134.40, 134.35, 134.1, 130.9, 130.5, 130.42, 130.41, 130.35, 130.29, 129.5, 129.4 (may be two overlapping signals), 123.6 (C(5)), 84.9 (C(1')), 79.1 (C(4')), 75.2 (C(2')), 72.5 (C(3')), 65.2 (C(5')), 58.9 (OMe); UPLC/MS: 1.25 min, 585 $[M+H]^+$; HRMS (ESI) $[M+H]^+$ calcd for $C_{31}H_{29}N_4O_8^+$ 585.1980, found 585.1984.

((benzovloxy)methyl)tetrahydrofuran-3,4-divl Dibenzoate (4b) was prepared following the same

(2R,3R,4R,5R)-2-((6-Amino-5-chloropyrimidin-4-yl)amino)-5-

procedure as for **4a**. Yield: 378 mg (65 %), a white solid material. TLC Rf ~ 0.38 (EtOAc/hexanes ~ 1/1); ¹H NMR (500 MHz, DMSO-d₆) δ = 8.03 (dd, *J*=1.3, 8.2 Hz, 2H, H(2)), 7.94 (s, 1H), 7.90 (dd, *J*=1.3, 8.2 Hz, 2H), 7.85 - 7.81 (m, 3H, contains NH), 7.69 - 7.59 (m, 3H), 7.51 (t, *J*=7.7 Hz, 2H), 7.47 (t, *J*=7.9 Hz, 2H), 7.41 (t, *J*=7.9 Hz, 2H), 6.82 (br. s., 2H, NH₂), 6.25 (dd, *J*=4.7, 9.1 Hz, 1H, H(1')), 5.91 - 5.86 (m, 2H, H(2'+3')), 4.62 - 4.48 (m, 3H, H(4'+5')); ¹³C NMR (126 MHz, DMSO-d₆) δ = 165.5, 164.8, 164.7, 159.4 (C(6) or C(4)), 156.4 (C(6) or C(4)), 154.8 (C(2)), 133.8, 133.7, 133.5, 129.4, 129.30, 129.27, 129.22, 128.8, 128.71, 128.68, 128.66, 128.62, 91.8 (C(5)), 83.7 (C(1')), 77.3 (C(4')), 73.7 (C(2') or C(3')), 70.8 (C(2') or C(3')), 64.0 (C(5')); UPLC/MS: 1.39 min, 589 [M+H]⁺; HRMS (ESI) [M+H]⁺ calcd for C₃₀H₂₆ClN₄O₇⁺ 589.1485, found 589.1457.

(3R, 4R, 5R)-2-((6-Amino-5-fluoropyrimidin-4-yl)amino)-5-

((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (**4f**, mixture α/β) and (2R,3R,4R,5R)-2-(4-Amino-5-fluoro-6-iminopyrimidin-1(6H)-yl)-5-((benzoyloxy)methyl)tetrahydrofuran-3,4-

diyl Dibenzoate (**3f**) were prepared following the same procedure as for **4a**. Purification by chromatography (gradient ethyl acetate/hexanes ~ 1/4 to 1/0, then to ethyl acetate/methanol 5/1) provided the less polar material, which was dissolved in ether and precipitated by adding slowly the same volume of hexanes, then filtered, and washed with hexanes to provide a white solid material containing **4f** (mixture $\alpha/\beta \sim 1/2.7$). Yield: 194 mg (34 %). ¹H NMR (500 MHz, Acetone-d₆, integration of the mixture is normalized, signals of the minor components are set as 1H, see the supporting information for the spectrum) $\delta = 8.13 - 8.06$ (m, 7H), 8.04 - 7.92 (m, 15H), 7.85 (d, *J*=1.3 Hz, 2.7H), 7.81 (d, *J*=1.6 Hz, 1H), 7.68 - 7.56 (m, 11H), 7.54 - 7.39 (m, 23H), 7.24 (d, *J*=9.1 Hz, 2.7H), 6.74 (dd, *J*=5.0, 10.1 Hz, 1H), 6.41 (dd, *J*=6.0, 9.5 Hz, 2.7H), 6.28 (d, *J*=8.8 Hz, 1H), 6.01 (t, J=5.0 Hz, 1H), 5.99 - 5.92 (m, 10H), 5.88 (t, J=5.7 Hz, 2.7H). The

more polar material (**3f**, 370 mg, 64%) was dissolved in 2 mL of ethyl acetate, diluted with 20 mL of ether, cooled in an ice bath, diluted with 20 mL of hexanes, stirred, then filtered and dried under vacuum to provide **3f** as a white solid material. Yield 255 mg (45 %). ¹H NMR (500 MHz, DMSO-d₆) δ = 8.61 (s, 1H), 8.48 - 8.15 (m, 4H, NH₂+NH+extra H), 8.02 (dd, *J*=1.3, 8.5 Hz, 2H), 7.95 (dt, *J*=1.1, 8.1 Hz, 4H), 7.72 - 7.64 (m, 3H), 7.54 (t, *J*=7.7 Hz, 2H), 7.48 (q, *J*=7.6 Hz, 4H), 6.49 (d, *J*=4.7 Hz, 1H, H(1')), 6.05 (dd, *J*=5.0, 6.0 Hz, 1H, H(2')), 5.95 (t, *J*=5.8 Hz, 1H, H(3')), 4.92 - 4.87 (m, 1H, H(4')), 4.85 - 4.75 (m, 2H, CH₂(5')); ¹³C NMR (126 MHz, DMSO-d₆) δ = 165.5, 164.51, 164.49, 150.7 (d, *J*=9.1 Hz), 144.9 (d, *J*=20.0 Hz), 144.2 (d, *J*=6.4 Hz, C(2)), 134.1, 134.0, 133.7, 129.5, 129.42, 129.32, 129.0, 128.8, 128.7 (two overlapping signals), 128.42, 128.37, 126.4 (d, *J*=237.1 Hz, C(5)), 88.1 (C1')), 80.4 (C(4')), 73.9 (C(2')), 69.8 (C(3')), 63.4 (C(5')), (four extra signals observed between 116 and 125 ppm. Cannot explain); UPLC/MS: 1.14 min, 573 [M+H]⁺ also 445, 201; HRMS (ESI) [M+H]⁺ calcd for C₃₀H₂₆FN₄O₇⁺ 573.1780, found 573.1750.

(2R, 3R, 4R, 5R)-2-((6-Amino-5-fluoropyrimidin-4-yl)amino)-5-

(*(benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate* (**4f**, β -isomer). To 2-chloro-3-fluoro-4-amino-pyrimidine (**2f**, 148 mg, 1 mmol) was added ammonium sulfate (7 mg, 5 mol%), HMDS (3 mL), and the mixture was heated under nitrogen at 135 °C (external temperature) overnight (15 hours). A clear solution was not formed. Pyridine (1 mL) was added and a clear solution was formed immediately. After 30 min the mixture was concentrated under vacuum. To the crude silylated pyrimidine was added **1** (504 mg, 1 mmol), acetonitrile (3 mL), the reaction mixture was cooled in an ice bath and treated with SnCl₄ (1M solution in DCM, 2 mL), which led to instantaneous dissolution. After 3 h at this temperature, the reaction mixture was quenched into ice-cold mixture of acetonitrile (20 mL) and ammonium hydroxide (2 mL). The reaction

mixture was diluted with toluene and filtered through Celite, concentrated and purified by chromatography twice. First purification used a gradient ethyl acetate/hexanes $\sim 1/4$ to 1/0, then acetate/methanol ~ 5/1.Second purification used to ethvl а gradient ethvl acetate/dichloromethane ~ 1/4 to 1/0, then to ethyl acetate/methanol ~ 5/1. The product (352 mg, 0.61 mmol) was dissolved in ether and precipitated with pentane, then washed with pentane to provide the title compound as a white solid material (147 mg, 26 %, single beta-isomer). The endo-isomer **3f** was also present in the more polar fractions; however, it could not be isolated clean. Data for 4f: ¹H NMR (500 MHz, DMSO-d₆) $\delta = 8.07$ (d, J=9.1 Hz, 1H, NH), 8.05 - 8.01 (m, 2H), 7.93 - 7.88 (m, 2H), 7.87 - 7.82 (m, 2H), 7.81 (d, J=1.6 Hz, 1H), 7.69 - 7.58 (m, 3H),7.55 - 7.49 (m, 2H), 7.49 - 7.44 (m, 2H), 7.44 - 7.39 (m, 2H), 6.67 (br.s, 2H, NH₂), 6.19 (dd, J=5.4, 9.5 Hz, 1H, H(1')), 5.89 - 5.84 (m, 1H, H(3')), 5.78 (t, J=5.7 Hz, 1H, H(2')), 4.62 - 4.48 (m, 3H, H(4' and 5')); 13 C NMR (126 MHz, DMSO-d₆) δ = 165.5, 164.8, 164.7, 152.2 (d, J=10.0 Hz, C(2)), 151.7 (d. J=8.2 Hz, C(4) or C(6)), 148.3 (d. J=7.3 Hz, C(4) or C(6)), 133.8, 133.7. 133.5, 129.4, 129.3, 129.24, 129.20, 130.1 (d, J=246.1 Hz, C(5)), 128.8, 128.7 (broad, three signals may overlap here), 128.6, 83.1 (C(1')), 77.3 (C(4')), 73.7 (C(2')), 70.9 (C(3')), 64.1 (C(5')); UPLC/MS: 1.36 min, 573 $[M+H]^+;$ HRMS (ESI) $[M+H]^+$ calcd for $C_{30}H_{26}FN_4O_7^+$ 573.1780, found 573.1750.

(2R,3R,4R,5R)-2-((6-Aminopyrimidin-4-yl-3-15N)amino)-5-

((benzoyloxy)methyl)tetrahydrofuran-3,4-diyl Dibenzoate (4a-¹⁵N). To 2a (130 mg, 1 mmol) was added 1 (504 mg, 1 mmol), acetonitrile (3 mL), BSA (0.25 mL), and the mixture was heated to 90 °C for 10 minutes, then cooled in an ice bath. TMSOTf was added (0.36 mL, 2 mmol) and the reaction was continued at room temperature for 25 minutes, then quenched into stirred ice-cold

mixture of acetonitrile (20 mL), water (5 mL), ¹⁵NH₄Cl (98 % ¹⁵N, 550 mg, 10 mmol), and K₂CO₃ (1,380 mg, 10 mmol). The mixture was diluted with toluene, washed with water, brine, $(MgSO_4)$ concentrated and purified by chromatography dried (gradient ethvl acetate/dichloromethane $\sim 0/1$ to 1/0). The isolated material was dissolved in ether, cooled in an ice bath, and precipitated by addition of hexanes. The solid material was washed with pentane and dried under vacuum to provide the title compound as a white solid material (159 mg, 0.29 mmol, 29 %). The mother liquor was partially concentrated in a flow of nitrogen at rt to provide additional amount of the title compound (105 mg, 0.19 mmol). ¹H NMR (500 MHz, DMSO-d₆) δ $= 8.05 - 8.00 \text{ (m, 2H)}, 7.96 \text{ (d, } J=15.1 \text{ Hz}, 1\text{H}, \text{H}(2)), 7.92 - 7.88 \text{ (m, 2H)}, 7.87 - 7.84 \text{ (m, 2H)}, 7.87 - 7.88 \text{ (m, 2H$ 7.83 (dd, J=2.2, 9.5 Hz, 1H, NH), 7.70 - 7.60 (m, 3H), 7.55 - 7.50 (m, 2H), 7.49 - 7.41 (m, 4H), 6.34 (s, 2H, NH₂), 6.04 (br. s., 1H, H(1')), 5.80 (dd, J=4.4, 5.7 Hz, 1H, H(3')), 5.62 - 5.55 (m, 2H, H(2') and H(5)), 4.59 - 4.47 (m, 3H, H(4') and H(5')); ¹³C NMR (126 MHz, DMSO-d₆) $\delta =$ 165.5. 164.8. 164.7. 163.9 (d. J=1.8 Hz, C(4 or 6)), 161.5 (d. J=4.5 Hz, C(4 or 6)), 157.8. 133.83, 133.76, 133.5, 129.4, 129.28, 129.25, 129.23, 128.8, 128.73, 128.71, 128.66, 128.6, 83.9 (C(5)), 83.4 (C(1')), 77.3 (C(4')), 73.7 (C(2')), 71.1 (C(3')), 64.3 (C(5')); UPLC/MS: 1.15 min, $[M+H]^+$; HRMS (ESI) $[M+H]^+$ calcd for $C_{30}H_{27}N_3^{15}NO_7^+$ 556.1845, found 556.1820.

5-Nitropyrimidine-4,6-diamine-¹⁵N₂ (8). To ¹⁵NH₄Cl (460 mg, 8.4 mmol) was added acetonitrile and the mixture was cooled in an ice bath. Triethylamine (2.1 mL, 15 mmol) was added, followed by 4,6-dichloro-5-nitropyrimidine (7, 386 mg, 2.0 mmol) and the mixture was stirred in a sealed tube at room temperature for 6 hours, then diluted with water and filtered, washed the solid material with water and acetonitrile, then dried to provide 8 as a light brown colored material (281 mg, 1.8 mmol, 90 %). ¹H NMR (500 MHz, DMSO-d₆) δ = 8.49 (d, *J*=92.4 Hz, 2H, 2 x NH^aH^b), 8.42 (d, *J*=91.4 Hz, 2H, 2 x NH^aH^b), 7.87 (s, 1H, H(2)). Satisfactory ¹³C-

NMR spectrum could not be obtained due to poor solubility of the material. See HSQC spectrum in the Supporting Information, page S102) for the approximate location of the C(2) signal (158.5 ppm); HRMS (ESI) [M+H]+ calcd for $C_4H_5N_3^{15}N_2O_2^+$ 158.0457, found 158.0448.

 $(2R, 3R, 4R, 5R)-2-((4-(Amino-^{15}N)-5-nitropyrimidin-6-yl-1-^{15}N)amino)-5-$

((benzovloxy)methyl)tetrahydrofuran-3,4-divl Dibenzoate (10). To 8 (159 mg, 1 mmol) was added ammonium sulfate (7 mg, 0.05 equiv), HMDS (3 mL), pyridine (1 mL), and the mixture was heated under nitrogen in a 130 °C heat block. After 5 hours a clear solution was formed and the reaction mixture was concentrated under vacuum using rotovap, then under high vacuum. To this crude silvlated aminopyrimidine was added 1 (454 mg, 0.9 mmol), acetonitrile (3 mL), TMSOTf (0.36 mL, 2 mmol) and the mixture was left at room temperature overnight (18 hours) and then quenched with ammonium hydroxide (0.5 mL), partitioned between EtOAc and brine, the organic layer was dried with $MgSO_4$, concentrated and purified by chromatography (gradient ethyl acetate/dichloromethane $\sim 0/1$ to 1/3). The isolated material was recrystallized from isopropanol to provide the title product 10 (281 mg, 47 %). Additional recrystallization from methanol provided analytically pure 10 (147 mg) as a lightly colored yellow solid material. ¹H NMR (500 MHz, DMSO-d₆) δ = 9.63 (dd, J=3.9, 8.4 Hz, 1H, HN), 8.63 (d, J=91.1 Hz, 1H, NH_aH_b), 8.56 (d, J=93.6 Hz, 1H, NH_aH_b), 8.04 (d, J=15.8 Hz, 1H, H(2)), 8.02 - 8.00 (m, 2H), 7.92 - 7.88 (m, 2H), 7.87 - 7.83 (m, 2H), 7.69 - 7.60 (m, 3H), 7.53 - 7.40 (m, 7H), 6.37 (dd, J=4.9, 8.4 Hz, 1H, H(1')), 6.00 (dd, J=5.0, 6.0 Hz, 1H, H(2')), 5.91 (t, J=5.4 Hz, 1H, H(3')), 4.67 - 4.61 (m, 2H, H(4'+5'a)), 4.60 - 4.54 (m, 1H, H(5'b)), ¹³C NMR (126 MHz, DMSO-d₆) $\delta =$ 165.5, 164.7, 164.6, 159.1 (d, J=2.7 Hz, (C(2))), 158.5 (dd, J=1.8, 21.8 Hz, (C(6))), 156.5 (d, J=3.6 Hz, (C(4))), 133.8, 133.7, 133.5, 129.29, 129.27, 129.22, 128.8, 128.68, 128.67, 128.63, 128.5, 112.5 (C(5)), 83.9 (C(1')), 78.0, 74.0 (C(2')), 71.0 (C(3')), 63.8 (C(5')). One Bz signal is

missing, probably due to overlap. UPLC: 1.43 min, 602 $[M+H]^+$, also 445, 201; HRMS (ESI) $[M+H]^+$ calcd for $C_{30}H_{25}N_3^{15}N_2O_9^+$ 602.1666, found 602.1637.

Acknowledgment. The authors are thankful to Mrs. Jane Yang for analytical support and to Prof. Scott E. Denmark from the University of Illinois, Urbana-Champaign for discussions.

Supporting Information Available. Spectral data for all new compounds. This material is available free of charge via the Internet at <u>http://pubs.acs.org</u> or from <u>rbaiazitov@ptcbio.com</u>. **References**

- (1) Kubo, I.; Kim, M.; Wood, W. F.; Naoki, H. Tetrahedron Lett. 1986, 27, 4277.
- Moss, R. J.; Petrie, C. R.; Meyer, R. B.; Dee Nord, L. Jr.; Willis, R. C.; Smith, R. A.;
 Larson, S. B.; Kini, G. D.; Robins, R. K. J. Med. Chem. 1988, 31, 786.
- (3) (a) Fortin, H.; Tomasi, S.; Delcros, J.; Bansard, J.; Boustie, J. *ChemMedChem* 2006, *1*, 189. (b) Ren, G.; Zhao, Y. P.; Yang, L.; Fu, C. X. *Cancer Lett.* 2008, *262*, 190. (c) Shin, H.; Min, C. *Clitocine and its Analogues*, In *Modified Nucleosides*; Herdewijn, P., Ed.; Wiley-VCH: Weinheim, 2008, Chap. 22, 567. (d) Liu, F. Y.; Ren, G.; Fu, C. X.; Wu, P. CN 101333236, 2008; *Chem. Abstr.* 2008, *150*, 106103. (e) Liu, F. Y.; Guo, T. D.; Wu, S. H.; Wu, P.; Feng, G. P. CN 101347442, 2009; *Chem. Abstr.* 2009, *150*, 206311.
- (4) (a) Wilde, R. G.; Kennedy, P. D.; Almstead, N. G.; Welch, E. M.; Takasugi, J. J.; Friesen, W. J. Nucleoside compounds and their use for treating cancer and diseases associated with somatic mutations. WO 2004009609, 2004; *Chem. Abstr.* 2004, *140*, 128608. (b) Wilde, R. G.; Almstead, N. G.; Welch, E. M.; Beckmann, H. Use of nucleoside compounds for nonsense suppression and the treatment of genetic diseases. WO2004009610, 2004; *Chem. Abstr.* 2004, *140*, 122839.

- (5) (a) Lee, C.-H.; Daanen, J. F.; Jiang, M.; Yu, H.; Kohlhaas, K. L.; Alexander, K.; Jarvis, M. F.; Kowaluk, E. L.; Bhagwaty, S. S. *Bioorg. Med. Chem. Lett.* 2001, *11*, 2419–2422;
 (b) Kamikawa, T.; Fujie, S.; Yamagiwa, Y.; Kim, M.; Kawaguchi H. *Chem. Commun.* 1988, 195-196.
- (6) (a) Linek, K.; Alföldi, J. Carb. Res., 1987, 164, 195-205; (b) Linek, K.; Alföldi, J.;
 Defaye, J. Carb. Res., 1993, 247, 329-335.
- (7) Vorbbrüggen, H.; Ruh-Pohlenz, C. Synthesis of Nucleosides in Organic Reactions 2000, 55, DOI: 10.1002/0471264180.or055.01.
- (8) (a) Kusano, S.; Hattori, K.; Imoto, S.; Nagatsugi, F. *Nucleic Acids Symposium Series*,
 2009, 53, 169-170; (b) Choi, H.; Choi, B. S.; Chang, J. H.; Lee, K. W.; Nam, D. H.; Kim,
 Y. K.; Lee, J. H.; Heo, T.; Shin, H.; Kim, N.-S. *Synlett*, 2005, 1942-1944; (c) Moss, R. J.;
 Petrie, C. R.; Meyer Jr., R. B.; Nord, L. D.; Willis, R. C.; Smith, R. A. Larson, S. B.;
 Kini, G. D.; Robins, R. K. *J. Med. Chem.* 1988, *31*, 786-790; (d) Rokos, H.; Pfleiderer,
 W. *Chem. Ber.* 1971, *104*, 748-769.
- (9) For example: (a) Rajeev, K. G.; Broom, A. D. Org. Lett., 2000, 2, 3595-3598; (b) Gosselin, G.; Bergogne, M.-C.; Rudder, J. D.; Clercqm E. D.; Imbach, J.-L. J. Med. Chem., 1987, 30, 982-991; (c) Ciceri, S.; Ciuffreda, P.; Grisenti, P.; Ferraboschi, P. Tetrahedron Lett., 2015, 56, 5909-5913; (d) Chien, T.-C.; Chen, C.-S.; Yu, F.-H.; Chern, J.-W. Chem. Pharm. Bull, 2004, 52, 1422—1426; (e) Kovaliov, M.; Segal, M.; Fischer, B. Tetrahedron, 2013, 69, 3698-3705.
- (10) For example, reviews: (a) El Ashry, E. S. H.; El Kilany, Y.; Rashed, N.; Assafir, H. Adv. *Het. Chem.* 1999, 75, 79-165; (b) Fujii, T.; Itaya, T. *Heterocycles* 1998, 48, 359-390.
 Relevant applications in chemistry: (c) Macon, J. B.; Wolfenden, R. *Biochem.* 1968, 7,

The Journal of Organic Chemistry

3453-3458; (d) Brown, D. J; Harper, J. S. J. Chem. Soc. 1965, 5542-5551; (e) Brown, D.

J; Harper, J. S. J. Chem. Soc. 1963, 1276-1284.

- (11) BSA/TMSOTf combination also worked; however, the yield was lower (21 %)
- (12) The product could not be isolated sufficiently pure.
- (13) Witanowski, M.; Stefaniak, L.; Webb, G. A. Nitrogen NMR spectroscopy in Annual reports on NMR spectroscopy, G. A. Webb, Ed., Volume 11b, **1981**
- (14) Although there may be a trend that the absolute value of ${}^{3}J$ (${}^{1}H{-}{}^{15}N$) is lower in heterocycles than that of ${}^{2}J$ (${}^{1}H{-}{}^{15}N$), this is not always true and cautious analysis is required.
- (15) Unfortunately, there is no clear trend for the ¹³C-¹⁵N couplings. For example, the magnitude of ³J (¹³C-¹⁵N) in pyridine is larger (-3.85 Hz) than ²J (¹³C-¹⁵N) coupling (+2.53 Hz) or ¹J (¹³C-¹⁵N) coupling constant (+0.62 Hz). As a consequence, the ¹³C-NMR coupling constants could not be used for the structure confirmation.
- (16) See the *Experimental Section* for details.
- (17) Various combinations of silylating agents (HMDS, BSA, BSTFA, TMSOTf/triethylamine) and Lewis acids (TMSOTf or SnCl₄) in dichloromethane, acetonitrile or 1,2-dichloroethane can often be used. Moreover, 2,3,5-triacetate analog of 1 can be used (86 % yield, not shown).
- (18) Attempted debenzoylation of several analogs 4 with ammonia or NaOMe in methanol led to formation of a material that was a complex mixture of products according to NMR analysis. The mass spectrometry analysis suggested that a compound with the expected mass is present. TLC analysis shows a single spot with the retention different from either animopyrimidine or ribose. We believe that upon deprotection the product undergoes

isomerization via mutarotation and, like many other reducing furanosides, may exist as a mixture of several interconverting forms: α/β -furanose, α/β -pyranose, as well as in the open form (imine).

- (19) The sample was referenced to the ¹⁵N signal at natural abundance of CD₃CN, which was set as 244 ppm. See Supporting Information, page S113
- (20) Preliminary data (not shown) suggest that 2-chloro-3-nitro-4-aminopyridine reacts similarly to provide the deazo-analog of 3. Use of 6-chloro-N-methylpyrimidin-4-amine and ammonium hydroxide quench may provide the N(6)-Me analog of 4a. When 5-H was used for the reaction and NH2Me used for the quench, the Me could be introduced to the N(4) of 3a.
- ¹H-NMR spectra of this compound in DMSO or acetone show an extra proton between
 7.9 amd 8.2 ppm. This proton is absent in the spectrum taken in methanol.