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J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.7b01752 • Publication Date (Web): 15 Aug 2017

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Total Synthesis of β-D-*Ido*-heptopyranosides Related to Capsular Polysaccharides of *Campylobacter jejuni* HS:4

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Table of Contents Graphic:



Abstract:

The 6-deoxy- β -D-*ido*-heptopyranoside related to capsular polysaccharides of *C. jejuni* HS:4 is very remarkable, owing to the unique, multifaceted structural features that have been combined into one molecule which include: (1) the rare *ido*-configuration, (2) the unusual 7-carbon backbone, and (3) the challenging β -(1 \rightarrow 2)-*cis* anomeric configuration. Two distinct strategies toward the total synthesis of this interesting target are reported. The first involved establishment of the β -D-idopyranosyl configuration from β -D-galactopyranosides, prior to a C-6-homologation

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extending the D-hexose to the desired 6-deoxy-D-heptose. However, this approach encountered difficulties due to significantly reduced reactivity of the 6-position of the β -D-idopyranosides, so instead a second strategy was employed which involved first carrying out a 6-homologation on the less flexible D-galactopyranose, followed by a very successful conversion to the desired β -D-*ido*-configuration found in the target heptopyranoside (**2**). This report is the first successful synthesis of the 6-deoxy- β -D-*ido*-heptopyranoside which could possess interesting immunological properties.

Keywords:

Synthesis; idopyranoside; 6-homologation; 6-deoxy-β-D-*ido*-heptopyranoside; 6-deoxy-β-D*galacto*-heptopyranoside.

1. Introduction

Campylobacter jejuni, a gram-negative, spiral-shaped bacterial pathogen, is a leading cause of bacterial infection worldwide.^{1,2} The incidence of *C. jejuni* infection is extremely high, ranging from hyperendemic levels (40/100 children under 5 years old) in developing countries of the world, to endemic levels (0.2–1/100 population) in more developed regions.⁴ The infection is often caused by the consumption of contaminated poultry, water, or raw milk,^{5,6} and symptoms can vary from more severe (e.g. diarrhea, fever, abdominal pain), to less severe (e.g. nausea, headache, muscle pain). Although infection is not usually associated with mortality in developed countries, *C. jejuni* infection can result in a high level of morbidity. In addition, a number of post-infectious sequela can occur and can be very serious; these include the autoimmune Guillain-Barré syndrome (GBS)⁷ or Miller-Fisher syndrome (MFS),⁸ as well as reactive arthritis⁹ among others.¹⁰ Therefore, the development of a vaccine against *C. jejuni* infection has significant importance because of the social and economic impact of the pathogen.¹¹

C. jejuni is known to produce lipooligosaccharides (LOS) which can be highly variable in structure, but often partially resemble human gangliosides.^{12,13} This molecular mimicry is associated with the autoimmune disorders that are caused by *C. jejuni* infection. The bacterium also produces another class of polysaccharides, called capsular polysaccharides (CPS), which possess unique structural features. For example, **Figure 1** shows the repeating structure of *C. jejuni* serotype HS:4 (strain CG8486), the main cause of campylobacteriosis.¹⁴ The repeating structure is a disaccharide that consists of a β -(1 \rightarrow 3)-linked *N*-acetyl-D-glucosamine (GlcNAc) at the reducing end and a very unusual 6-deoxy-D-*ido*-heptopyranose (6-deoxy-*ido*-Hep) with β -anomeric configuration at the non-reducing end. Interestingly, partial modification with *O*-methyl phosphoramidate at the O-2 and O-7 positions of the 6-deoxy-*ido*-heptopyranose residues

was observed, resulting in heterogeneity in the isolated CPS structures. The prevalence of *C*. *jejuni* infection caused by HS:4 strains, as well as the unique structure of 6-deoxy-*ido*-Hep make it an ideal antigen for the development of vaccines against campylobacter infections.¹⁵



Figure 1. Structure of *C. jejuni* CPS containing an unusual 6-deoxy- β -D-*ido*-heptopyranosyl residue. Compounds **1-5** are synthetic analogs containing a β -D-idopyranosyl configuration.

Thus, 6-deoxy-*ido*-Hep derivatives such as compounds 1-2 (Figure 1) represent very attractive synthetic targets, but their total syntheses present a significant challenge due to their unusual 7-carbon backbone which includes a 6-deoxygenation. In addition, installation of the β - $(1\rightarrow 2)$ -*cis*-anomeric configuration constitutes a well-recognized difficulty in carbohydrate chemistry. Furthermore, the *ido*-configuration which places the C-2, C-3, and C-4-hydroxyl groups in an axial position of the pyranoside ${}^{4}C_{1}$ chair also makes them interesting molecules to study for their unique conformational flexibility. Previously, we published a short, scalable

Page 5 of 45

The Journal of Organic Chemistry

synthesis of compound **3** and derivatives starting from β -D-galactopyranosides, by carrying out a double inversion at both C-2 and C-3.¹⁶⁻¹⁸ 2,3-Di-O-sulfonyl-β-D-galactopyranosides were prepared as the key substrates, and were converted to the corresponding 2,3-anhydro-B-Dtalopyranoside intermediates; subsequent nucleophilic attack at C-3 by an alkoxide afforded orthogonally protected β -D-idopyranosides such as compound **3** in a single step from the 2.3-di-O-sulforyl substrates. More recently, we succeeded in 6-homologation of the β -idopyranoside via Swern oxidation and Wittig olefination.¹⁹ However, it was found that the Wittig olefination worked well only with less sterically hindered ylides such as methylenetriphenylphosphorane, to afford fully protected compound 4. By ¹H NMR spectroscopy, it was observed that compound 4 preferentially adopts a ${}^{1}C_{4}$ chair conformation in solution, which orients the vinyl group in a more sterically hindered axial position, creating a substantial difficulty for subsequent hydroboration and preventing the desired primary alcohol from being obtained using either borane-tetrahydrofuran (THF) or 9-BBN as the reagent. We thus altered our strategy to instead focus on synthesis of the corresponding 6-nitrile derivatives, such as compound 5, since reduction of the nitrile functionality could open an avenue to obtain the intermediate aldehyde²⁰ which could be further reduced to the desired 6-deoxy-*ido*-Hep (1).

2. Results and Discussion

2.1 6-Homologation via nitrile substitution

Starting from the previously published 3-*O*-allylated β -D-idopyranoside (6) containing a 4,6diol functionality, a selective O-6-tosylation was carried out in a mixture of anhydrous pyridine and dichloromethane using *p*-toluenesulfonyl chloride; this afforded regioselectively the desired 6-tosylate 7 in ~60% yield. Unfortunately, subsequent attempts at performing an S_N2 substitution with cyanide as a nucleophile in DMF at 80 °C proceeded poorly, and the best isolated yield for desired compound **9** was only 13%. Attempts to improve the yield using either 18-crown-6 or tetra-*n*-butylammonium iodide were both unsuccessful. In fact, when 18-crown-6 was used, a small amount of hydrolyzed 6-alcohol (**6**, up to ~20%) was instead formed, likely the result of a trace amount of water present in the reagents. This was surprising to observe, given the extreme difficulty previously encountered in modifying the C-6 position directly, thus may instead result from attack of the nucleophilic H₂O (or cyanide, or both) at the S atom of the tosylate sulfonate group (as observed for the double inversion of 2,3-di-*O*-sulfonyl- β -D-galactopyranosides). Although conventionally less electrophilic, since the S center is two bonds away from the highly sterically hindered C-6 position, it should be more sterically accessible by a nucleophile.

This difficulty in performing nitrile substitution at C-6 again confirmed the unusual lack of reactivity of the C-6 center in β -D-idopyranosides. We therefore decided to pursue a related substrate that instead utilized 6-iodide as the leaving group, which is known to have improved leaving group ability. Thus, starting from the same diol (6), a direct iodination was carried out in anhydrous dichloromethane using I₂/PPh₃ as the reagent and imidazole as base. ^{21, 22} Unfortunately, the iodination of compound **6** was found to be low yielding (26%), resulting from both unreacted starting material (27%) and the formation of a number of unidentified by-products as indicated by thin-layer chromatography (TLC).



Scheme 1. 6-Homologation via cyanide substitution from methyl β-D-idopyranosides **6** and **10** containing a 4,6-diol functionality.

Alternatively, we attempted to convert an analogous derivative, the 3-*O*-benzylated 4,6-diol (10), to the corresponding 6-iodide (11) as above. The reaction proceeded much better to provide compound 11 in 63% yield. Interestingly, a 1,6-anhydro by-product (12) was also formed from the reaction, which was isolated in 26% yield. Formation of the 1,6-anhydro by-product could be explained by an intramolecular S_N2 attack via an iodide-mediated *O*-demethylation at the anomeric position (Scheme 2). The β -D-idopyranosyl configuration of the substrate may promote this undesired intramolecular attack by facilitating a ring-flip to the 1C_4 chair conformer. Identity of the 1,6-anhydro by-product 12 was confirmed by 1 H NMR spectroscopy of the compound, which revealed the loss of the anomeric methyl group, as well as the presence of a hydroxyl group that correlates with H-4 in the 1 H- 1 H COSY spectrum; additionally, large coupling constants were observed between H-2/H-3 and H-3/H-4 ($J_{2,3} = 8.0$ Hz, $J_{3,4} = 8.1$ Hz).

Furthermore, electrospray ionization high-resolution mass spectrometry (HRMS) also supported the proposed structure (calculated m/z for $[M + NH_4]^+$: 360.1805; found: 360.1802).



Scheme 2. Proposed mechanism for the formation of by-product 12.

The iodinated intermediate **8** was then subjected to treatment with KCN in DMF at 80 °C (**Scheme 1**), but unfortunately although the substitution reaction produced higher yields than with the 6-tosylate (7), the isolated yield for the desired 6-nitrile **9** was still low (~40%). The transformation was observed to be quite sluggish, as large amounts of unreacted starting material were often recovered. Attempts at pushing the reaction further using excess reagent, elevated temperatures, the addition of crown ether (18-crown-6), or phase transfer catalysts (Bu₄NI) to aid in solubility were all unsuccessful. Similarly, we also attempted to carry out the displacement on compound **11** using KCN in DMF at 80 °C to obtain compound **13**. However, by TLC it was found that the expected compound **13** was formed in only low yield, and was therefore not isolated. In an effort to increase reactivity of the substrate, the 6-O-triflate was also prepared and subjected to KCN in anhydrous DMF, but unfortunately only hydrolyzed 6-alcohol (**6**) was recovered from the reaction mixture, similar to what had been observed previously during reaction of the 6-O-tosylate.

The identity of the desired product (9) could be confirmed by the significant shift of the ¹³C and ¹H signals associated with C-6 (20.0 ppm) and H-6 (2.86 and 2.64 ppm) in the NMR spectra,

The Journal of Organic Chemistry

as well as confirmation via HRMS (calculated m/z for $[M + NH_4]^+$: 351.1914; found: 351.1917). Despite the poor yields of nitrile derivatives **9** and **13**, they were subsequently subjected to a reduction with DIBAL-H, as it has been well established in literature that nitrile functionalities can be reduced selectively to an intermediate aldehyde via the imine intermediate²⁰ and could then be further reduced to the primary alcohol (**14** or **15**). Unfortunately, all attempted reduction conditions resulted in a complex product mixture: upon purification, some unreacted starting material was isolated together with other by-products that could not be identified by ¹H NMR spectroscopy. Although it has been reported that in some instances DIBAL-H can cleave benzyl ether groups, this did not appear to be a major pathway of the by-product formation.^{23,24}

The substantial difficulty encountered during 6-iodination and the subsequent substitution with cyanide, as well as the later reduction of the 6-nitrile functionality both appear to be consistent with the reduced reactivity of the 6-position of β -D-idopyranosides in general. Recently, we also reported an unsuccessful 6-homologation via Wittig olefination using a more sterically hindered ylide.¹⁹ We attribute these unsuccessful chemical transformations to the unusual conformational flexibility of β -D-idopyranosides. For example, in the 4C_1 chair conformer, the axial substituent at C-4 sterically shields the C-6 position, while in the 1C_4 chair conformer, the C-6 group occupies an axial position which is inherently more sterically hindered. Thus, in either case, the functional group at the C-6 position of β -D-idopyranosides experiences reduced reactivity because of steric clashes. Previously, Streicher *et al*²⁵ tried to displace the 6tosylate or 6-mesylate of 1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose with a cyanide, and they also encountered significant difficulty because of the steric shielding of the axial 4substituent as is the case in our compounds (7 and 11); the best yield of the 6-nitrile product that they could isolate was only 4.2%, using the 6-mesylate with LiCN in DMF at 110 °C for 17 hours. We thus concluded that the successful synthesis of 6-deoxy-*ido*-Hep would require revision of our initial strategy.

2.2 Revised strategy to synthesize 6-deoxy-ido-heptopyranosides.

Our previous strategy provided excellent results with regards to establishing the β -(1 \rightarrow 2)-*cis*anomeric configuration and also the D-ido-configuration, but failed at 6-homologation. Thus, a logical revision of the synthetic strategy would be to reverse the steps by first completing 6homologation on a β -D-galactopyranosyl moiety, to obtain a 6-deoxy- β -D-galactoheptopyranosyl intermediate; subsequent conversion of the 6-deoxy-β-D-galactoheptopyranoside to the 6-deoxy-β-D-ido-heptopyranoside could afford the desired target. We thought it would be advantageous to carry out the 6-homologation step on the pyranose ring of D-galactose, because of its reduced flexibility. Scheme 3 shows the retrosynthetic route of the new strategy by using readily available 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose (16) as the starting material. We planned to attempt the 6-homologation of D-galactose by Wittig olefination to obtain structure **B**, carry out a subsequent glycosylation (structures $C \rightarrow D$), and then finally convert the heptose from D-galacto- to D-ido-configuration via the 2,3-anhydro-talointermediate (structure E).



Scheme 3. A revised strategy to synthesize 6-deoxy-β-D-*ido*-heptopyranosides from D-galactopyranose by first performing the 6-homologation.

To test the feasibility of the new strategy, we first attempted to synthesize the simple 1,2:3,4di-*O*-isopropylidene- α -D-*galacto*-heptopyranose **19** via Wittig olefination (**Scheme 4**). Thus, compound **16** was first oxidized to the corresponding aldehyde **17** by pyridinium dichromate (PDC)²⁶ along with sodium acetate in dichloromethane, to afford the desired aldehyde in improved yield (82%). Compound **17** was then reacted with methylidenetriphenylphosphorane (**18**, Ph₃P=CH₂, generated in situ by the treatment of methyltriphenylphosphonium bromide with *n*-butyllithium, 1 equiv.) as described previously,²⁷ and the desired alkene **19** was successfully formed and isolated in much improved yield (74%) by column chromatography on silica gel. The structure of compound **19** was confirmed by the three vinylic protons observed at 5.94, 5.37, and 5.28 ppm. However, when alkene **19** was subjected to a hydroboration under standard reaction conditions,^{28,29} no desired primary alcohol **20** was obtained. After work up, several new compounds were indeed formed as evidenced by a number of spots observed via TLC, but upon purification none of them had the characteristic ¹H NMR signals corresponding to the diastereotopic protons H-6a and H-6b in the expected heptose **20**. The unsuccessful hydroboration was consistent with what has been reported previously.¹⁹



Scheme 4. Synthesis of 6-deoxy-1,2:3,4-di-*O*-isopropylidene-α-D-heptopyranoses 20 and 24.

An alternative Wittig olefination with $Ph_3P=CHOBn$ (22) was then carried out using again aldehyde 17. Ylide 22 was obtained via deprotonation of the benzyloxytriphenylphosphonium salt 21 with *n*-butyllithium in THF. After stirring overnight, the desired benzylic enol ether 23 was formed and isolated in ~60% yield as a mixture of ~1:1 *E/Z* isomers (Scheme 4). Previously, Streicher *et al*²⁵ reported an analogous reaction using $Ph_3P=CHOMe$ as an ylide to afford the corresponding alkenes in a similar yield. The two geometric isomers (23E and 23Z) had very similar polarity. Alternatively, the E/Z mixture of benzylic enol ether 23 could be directly subjected to catalytic hydrogenation in methanol using Pearlman's catalyst; after 24

The Journal of Organic Chemistry

hours, the desired 6-deoxy-1,2:3,4-di-*O*-isopropylidene- α -D-*galacto*-heptopyranose **20**²⁵ was obtained and isolated in 75% yield. The ¹H NMR spectrum of compound **20** showed two sets of multiplets at 2.00 and 1.80 ppm which are characteristic for the shielded H-6a and H-6b protons, as well as a second set of diastereotopic protons in a narrow region (3.74-3.88 ppm) which correspond to H-7a and H-7b. In the ¹H-¹³C HSQC correlation spectrum, compound **20** revealed a carbon signal (C-6) at 32.8 ppm which correlated to both H-6a and H-6b, confirming the 6-deoxy functionality. In addition, a more deshielded carbon signal (C-7) at 59.9 ppm correlated to both H-7a and H-7b, confirming the presence of a hydroxyl group at C-7.

During the catalytic hydrogenation step, since two different functionalities (alkene and Obenzyl) were reduced concomitantly, we decided to monitor the reaction progress in order to determine whether a regioselective manipulation of the functional groups would be possible. Indeed, after subjecting the mixture to the same reaction conditions as above for only one hour, we observed the formation of a major product with an $R_{\rm f}$ of 0.64 (20% EtOAc-hexanes). The new compound was only slightly less polar than the starting materials ($R_{\rm f} = 0.62, 20\%$ EtOAc-hexanes), and after stopping the reaction by a simple filtration, the newly formed product was isolated in 96% yield. To our delight, the ¹H NMR spectrum of the new product proved to be compound 24 (Scheme 4), which still had the characteristic benzylic protons at 4.55 and 4.49 ppm as two doublets (J = 11.8 Hz). In addition, two sets of proton signals were observed in a highly shielded region, 1.97 ppm (dddd, 1H, J = 13.9, 9.5, 5.3, 4.9 Hz, H-6a) and 1.90 ppm (dddd, 1H, J = 14.2, 9.8, 5.3, 4.9 Hz, H-6b), which corresponded to the diastereotopic protons H-6a and H-6b of compound 24. As would be expected, the E/Z double bond of the benzylic enol ethers 23 preferentially reacted under hydrogenolysis conditions before cleavage of the O-benzyl group could occur. However, when this reaction was performed on a larger scale (>

2 g), about 20% of product **20** was still formed but could be easily *O*-benzylated again using standard alkylation conditions.

These encouraging results obtained during the 6-homologation of D-galactose prompted us to design a synthetic route to the desired 6-deoxy- β -D-*galacto*-heptopyranoside **29** from compound **24** (Scheme 5).



Scheme 5. Synthesis of 6-deoxy-D-*galacto*-heptopyranosyl imidate donor 27 and subsequent conversion to the corresponding heptopyranosides 29 and 30.

The synthesis began with removal of the two isopropylidene groups of compound 24 in 50% acetic acid - water at 80 °C for 8 hours; the solution was then evaporated and treated with acetic anhydride-pyridine to afford fully acetylated intermediate 25, isolated as an α/β -mixture in 93% yield over two steps. In order to activate the anomeric position for glycosylation, the anomeric *O*-acetate of compound 25 was regioselectively removed using hydrazinium acetate³⁰ in DMF (85% yield), and the obtained hemiacetal 26 was then converted to the desired *O*-trichloroacetimidate 27 by reaction with trichloroacetonitrile in dichloromethane using

Page 15 of 45

The Journal of Organic Chemistry

anhydrous potassium carbonate as base. Since the imidate functionality is often sensitive to the mild acidity of silica gel, the crude reaction mixture was instead subjected to a quick work-up with extraction using ethyl acetate and brine, and then co-evaporated with toluene. This afforded the crude imidate 27 which was used directly for subsequent glycosylation with the previously reported 6-azidohexanol (28).³¹ The alcohol 28 was used in excess (5 equiv.) and the glycosylation carried out at -40 °C in anhydrous dichloromethane using trimethylsilyl trifluoromethanesulfonate (TMSOTf) as a promoter in the presence of 4 Å molecular sieves;^{32,33} after 4 hours, the reaction solution was warmed to ambient temperature. Surprisingly, the expected 6-azidohexyl 2,3,4-tri-O-acetyl-7-O-benzyl-6-deoxy-β-D-galacto-heptopyranoside 29 was not isolated, but instead an analog (30) which had lost the 2-O-acetyl group was obtained. A ¹H-¹H COSY NMR experiment (Figure 2) confirmed the structure of the isolated product: for example, the anomeric proton was observed as a doublet at 4.29 ppm with a large coupling constant ($J_{1,2} = 7.7$ Hz), confirming the β -anomeric configuration. In addition, the H-2 proton was observed at 3.78 ppm as a doublet of doublets, with the lower chemical shift supporting the absence of the expected acetyl group at O-2. On the other hand, the H-3 and H-4 protons were observed at 4.96 and 5.30 ppm, respectively; both protons are significantly more deshielded than H-2, confirming the presence of an acetyl group at both O-3 and O-4. Furthermore, a very weak correlation peak was observed between H-2 and the broad peak at 2.30 ppm, which was assigned to be the hydroxyl attached to C-2. This unexpected result was indeed very beneficial to our synthesis of the final 6-deoxy-β-D-ido-heptopyranoside target, as the free alcohol at C-2 could be selectively activated to generate a good leaving group, which could facilitate formation of the key 2,3-anhydro intermediate (see later).



Figure 2. ¹H-¹H gCOSY spectrum of 6-azidohexyl 3,4-di-*O*-acetyl-7-*O*-benzyl-6-deoxy-β-D*galacto*-heptopyranoside **30** (CDCl₃, 400 MHz, 298 K).

The reason for the regioselective loss of the 2-*O*-acetyl group could be explained by **Scheme 6**. As illustrated, after activation by TMSOTf, imidate **27** should be converted to the reactive oxocarbenium ion, which could be stabilized by participation of the O-2-acetyl group to form a bicyclic oxonium intermediate. Alcohol **28** could attack either the anomeric carbon from the top face (**Path A**) to form expected compound **29**, or the electrophilic C atom of the O-2-acetate (**Path B**) which would lead to the formation of an orthoester. After a proton transfer to the anomeric oxygen, a second molecule of alcohol **28** could attack from the top face, resulting in

The Journal of Organic Chemistry

the formation of the β -glycoside with an orthoacetic acid functionality which would subsequently be hydrolyzed to release the free OH-2 group and afford observed compound **30**. Previously, Nitz *et al*³⁴ also reported a similar 2-*O*-deacetylation when an orthoester was used as a glycosyl donor in the presence of excess allyl alcohol acceptor.



Scheme 6. A proposed pathway for the formation of 6-deoxy-β-D-galacto-heptopyranosides 29

and 30.

It is worth noting that the outcome of the glycosylation reaction varied when different amounts of alcohol (28) were used. For example, when 1.5 equivalents of alcohol (28) were used

during glycosylation with imidate **27**, both the expected 2,3,4-tri-*O*-acetylated product **29** and the 2-*O*-deacetylated product **30** were obtained in a 1.7:1 ratio.

With the unexpected availability of glycoside 30, we proceeded to activate the free OH-2 via mesylation using mesyl chloride in anhydrous pyridine, and the desired 2-O-mesylate 31 was isolated in 91% yield (Scheme 7). Compound 31 could then be directly converted into the 6deoxy- β -D-*ido*-heptopyranoside **34** in a single step, by dissolving the mesylate **31** into benzyl alcohol and treating with potassium *tert*-butoxide: this resulted in a proton exchange between benzyl alcohol and *tert*-butoxide, affording the reactive benzyloxide which catalyzed an Otransesterification to remove the O-3- and O-4-acetates (\rightarrow 32). The free C-3-oxide subsequently carried out an intramolecular attack of the O-2-mesylate, to form the desired 2,3-anhydro-6deoxy- β -D-talopyranoside intermediate (33). Finally, the formed 2,3-anhydro functionality was subjected to nucleophilic attack by the benzyloxide, to form the desired 3,7-di-O-benzylated 6deoxy-β-D-*ido*-heptopyranoside **34**. Using 5 equivalents of potassium *tert*-butoxide and stirring at 60 °C for 10 hours successfully afforded the desired compound 34 in ~60% isolated yield. If the reaction was carried out at a lower temperature and for a shorter time, the key intermediates could instead be isolated. For example, when the reaction was carried out at ambient temperature for 1 hour, the diol 32 was isolated in 75% yield; if the reaction was quenched after 12 hours at ambient temperature, the 2,3-anhydro intermediate **33** was isolated in 86% yield. Apparently, nucleophilic opening of the anhydro functionality of compound 33 by benzyloxide proceeded very slowly at ambient temperature but could be significantly accelerated at higher temperatures (60 °C), and indeed the reaction was very regioselective to afford desired compound 34 as the only regioisomer isolated.³⁵

The Journal of Organic Chemistry

In a similar manner, we also carried out the conversion of O-2-mesylate **31** to the related 3-O-allylated analog **35** in 73% yield using allyl alcohol as the solvent and nucleophile under similar conditions (**Scheme 7**).



Scheme 7. Formation of 6-deoxy- β -D-*ido*-heptopyranoside 2.

Success in constructing the 6-deoxy- β -D-*ido*-heptopyranosyl configurations of compounds **34** and **35** was confirmed by NMR experiments based on their correlation to the unique ³*J* coupling constants associated with the stereochemistry of β -D-*ido*-heptopyranosides. For example, for compound **34**, the anomeric proton was observed at 4.68 ppm as a doublet with a very small coupling constant ($J_{1,2} = 0.7$ Hz); H-3 was observed at 3.91 ppm as a doublet of doublets again with small coupling constants ($J_{2,3} \approx J_{3,4} \approx 2.6$ Hz), and H-5 was observed at 4.03 ppm as a doublet of doublets of doublets ($J_{4,5} < 1$ Hz, $J_{5,6a} = 5.1$ Hz, $J_{5,6b} = 8.2$ Hz). The observation of

small coupling constants between all adjacent protons ($J_{1,2}$, $J_{2,3}$, $J_{3,4}$, $J_{4,5}$) on the pyranose ring is consistent with the small dihedral angles (~60°) between adjacent protons, as expected in the ${}^{4}C_{1}$ *ido*-configuration of the pyranoside. On the other hand, the H-2 and H-4 protons were observed at 3.81 ppm and 3.53 ppm, respectively; analysis of their coupling patterns again afforded small coupling constants (<4.2 Hz) to neighbouring ring protons, as well as coupling to an attached hydroxyl group ($J_{2, OH-2} = 2.0$ Hz; $J_{4, OH-4}$ could not be accurately measured due to overlapping with OCHa of the aglycone chain).

The 3,7-di-O-benzylated 6-deoxy- β -D-ido-heptopyranoside 34 was finally subjected to hydrogenolysis to remove the benzyl protecting groups and concomitantly reduce the azide functionality on the aglycone. The reaction was initially conducted in methanol using Pearlman's catalyst. However, partial N-methylations were observed to afford the desired compound 2 along with some N-methylated by-products. The N-methyl groups could be observed in the NMR spectrum of the crude products: for example, in the ¹H NMR spectrum, a singlet at 2.70 ppm and another singlet at 2.88 ppm were observed, which would correspond to N-methyl groups in the by-products. Electrospray-quadrupole time of flight high resolution mass spectrometry (ESI/Q-TOF HRMS) further confirmed the presence of N-methylated products: we observed peaks at m/z 294.1908, 308.2065, and 322.2218, which correspond respectively to the calculated m/z of desired product 2 ($C_{13}H_{28}NO_6$ [M + H]⁺: 294.1911), and the mono- and di-N-methylated products $(C_{14}H_{30}NO_6 [M + H]^+: 308.1995; C_{15}H_{32}NO_6 [M + H]^+: 322.2155)$. These partial Nmethylations have also been reported recently during the catalytic hydrogenation of another family of oligosaccharide-azides.³⁶ Unfortunately, it was difficult to separate the partially Nmethylated by-products from desired compound 2 which prevented them from being characterized fully.

As an alternative, a Birch reduction was adopted to concomitantly remove the benzyl groups and reduce the azide^{37,38} (Scheme 7). The Birch reduction was carried out at -78 °C in liquid ammonia-THF using sodium metal as the reducing reagent. After one hour, the desired compound 2 was isolated as an acetic acid salt using gel filtration followed by chromatography on reverse-phase C18 silica gel using a gradient of $0 \rightarrow 20\%$ of methanol in water as the eluent (containing 0.2% acetic acid). Figure 3 shows the ¹H-¹³C HSOC NMR spectrum of the isolated compound 2 in D_2O . The anomeric proton (H-1) was observed at 4.72 ppm and is correlated to an anomeric carbon at 99.0 ppm. The other protons of the pyranoside ring, H-2, H-3, H-4 and H-5 protons are observed at 3.61, 3.96, 3.43 and 3.95 ppm, respectively, and they correlate with their attached carbons at 69.4 (C-2), 69.8 (C-3), 69.8 (C-4) and 71.1 (C-5) ppm, respectively. The observed vicinal coupling constants were consistently small ($J_{1,2} = -1$ Hz, $J_{2,3} = 3.5$ Hz, $J_{3,4}$ = 3.5 Hz, $J_{4.5}$ = 1.9 Hz), confirming the β -(1 \rightarrow 2)-*cis*-anomeric configuration as well as the *ido*configuration. The two diastereotopic protons H-6a and H-6b were found in a highly shielded region (1.91 and 1.70 ppm, respectively), confirming the 6-deoxy functionality. Finally, HRMS (ESI/Q-TOF) also revealed a peak at m/z 294.1908 which corresponds to the calculated m/z294.1911 for the expected formula of compound 2 $C_{13}H_{28}NO_6 [M + H]^+$.



Figure 3. ¹H-¹³C gHSQC spectrum of 6-deoxy-β-D-*ido*-heptopyranoside 2 (D₂O, 400 MHz, 298

K).

3. Conclusions

In conclusion, two distinct strategies have been studied to progress toward total synthesis of the challenging 6-deoxy- β -D-*ido*-heptopyranoside related to *C. jejuni* HS:4 CPS. The first strategy involved establishment of the β -D-idopyranosyl configuration from β -Dgalactopyranosides prior to extending the D-hexose to the desired 6-deoxy-D-heptose via C-6homologation. However, this approach was found to be impractical, as the C-6-position of the β -D-idopyranosides was observed to exhibit significantly reduced reactivity, probably caused by an

The Journal of Organic Chemistry

increased steric hindrance of the 6-position which is exacerbated by enhanced conformational flexibility of the β -D-idopyranosyl ring. The second strategy involved carrying out a 6homologation on the less flexible D-galactopyranose to first obtain a 6-deoxy- β -D-galactoheptopyranoside, followed by establishment of the β -D-*ido*-configuration in target heptopyranoside 2. This strategy was found to be very productive, allowing for the successful synthesis of a 6-deoxy- β -D-*ido*-heptopyranoside (2) for the first time. In both strategies, the conversion of β -D-galacto-configuration to β -D-ido-configuration was achieved by a double inversion at the C-2 and C-3 positions via a 2,3-anhydro- β -D-talo-pyranosyl intermediate; opening of the 2,3-anhydro-functionality by an alkoxide was found to be very regio- and stereoselective, with nucleophilic attack occurring exclusively at the C-3 position. The overall transformation of stereochemistry was proven to be very efficient, and required minimal purification. Additionally, our strategy to synthesize β -D-idopyranosides from β -Dgalactopyranosides is very versatile, as it can be successfully carried out on both hexoses and heptoses. This success in synthesis of the 6-deoxy- β -D-*ido*-heptopyranoside (2) should permit us to study its immunological properties in the future.

4. Experimental section

General Methods. All commercial reagents were used as supplied unless otherwise stated. Thin layer chromatography was performed on Silica Gel 60-F254 (E. Merck, Darmstadt) with detection by fluorescence, charring with 5% H_2SO_4 (aq), or a ceric ammonium molybdate solution. Column chromatography was performed on Silica Gel 60 (Silicycle, Ontario) and solvent gradients given refer to stepped gradients and concentrations are reported as % v/v.

Organic solutions were concentrated and/or evaporated to dry under vacuum in a water bath (<60 °C). Molecular sieves were stored in an oven at 100° C and flame-dried under vacuum before use. Amberlite IR-120 (H⁺) ion exchange resin was washed multiple times with MeOH prior to use. All pH values were determined using universal pH paper. Optical rotations were determined in a 5 cm cell at 20 ± 2 °C; $[\alpha]_D^{20}$ values are given in units of 10⁻¹ deg•cm²/g. NMR spectra were recorded on Bruker spectrometers at 400 MHz, and the first-order chemical shifts of ¹H and ¹³C (DEPT-Q) are reported in δ (ppm) and referenced to residual CHCl₃ (δ_H 7.24, δ_C 77.23, CDCl₃), or residual HDO (δ_H 4.79, external acetone δ_C 29.9, D₂O). ¹H and ¹³C NMR spectra were assigned with the assistance of 2D gCOSY and 2D gHSQC experiments. High-resolution ESI-QTOF mass spectra were recorded on an Agilent 6520 Accurate Mass Quadrupole Time-of-Flight LC/MS spectrometer. Elemental analyses were obtained with a Perkin Elmer Series II 2400 CHNS/O Analyzer. All the data were obtained with the assistance of the analytical services of the Department of Chemistry, University of Calgary.

Methyl 3-O-allyl-2-O-benzyl-6-O-tosyl-β-D-idopyranoside (7): Compound **6** (481 mg, 1.48 mmol) and *p*-TsCl (318 mg, 1.67 mmol) were dissolved into dry pyridine (5.0 mL) and left mixing at rt. After 6 h additional *p*-TsCl (43 mg, 0.23 mmol) was added. After 20 h, the reaction was quenched with MeOH (5 mL), then evaporated to dry via co-evaporation with toluene (3 × 5 mL). The crude product was re-dissolved into EtOAc (100 mL), washed with saturated NaCl (aq) solution (3 × 100 mL), dried with Na₂SO₄, filtered, and evaporated to dry. The crude mixture was purified by column chromatography on silica gel using 25% acetone – hexanes to afford the desired product 7 as a colourless syrup (423 mg, 0.884 mmol, 60% yield), as well as a small amount of di-tosylated by-product. R_f 0.66 (EtOAc : hexanes 2:3). [α]_D²⁰: -72.4° (*c* 1.02, CHCl₃).

¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.79 – 7.74 (m, 2H, Ar), 7.31 – 7.24 (m, 7H, Ar), 5.72 (dddd, 1H, J = 17.2, 10.4, 5.6, 5.6 Hz, OCH₂C<u>H</u>=CH₂), 5.13 (dddd, 1H, J = 17.2, 1.6, 1.6, 1.6 Hz, OCH₂CH=C<u>H</u>_aH_b), 5.11 (dddd, 1H, J = 10.4, 1.5, 1.3, 1.3 Hz, OCH₂CH=CH_a<u>H</u>_b), 4.83 (d, 1H, J = 12.1 Hz, PhC<u>H</u>_aH_b), 4.58 (d, 1H, J = 12.1 Hz, PhCH_a<u>H</u>_b), 4.57 (d, 1H, J = 1.0 Hz, H-1), 4.24 (dd, 1H, J = 10.3, 5.1 Hz, H-6a), 4.20 (dd, 1H, J = 10.3, 6.9 Hz, H-6b), 4.02 (ddd, 1H, J = 6.7, 5.2, <1 Hz, H-5), 3.90 (dddd, 1H, J = 12.9, 5.6, 1.4, 1.4 Hz, OC<u>H</u>_aH_bCH=CH₂), 3.86 (dddd, 1H, J = 12.9, 5.7, 1.4, 1.4 Hz, OCH_a<u>H</u>_bCH=CH₂), 3.64 – 3.62 (m, 1H, H-3), 3.55 (dd, 1H, J = 3.4, 0.8 Hz, H-2), 3.50 – 3.48 (m, 4H, OMe and H-4), 3.42 (broad, 1H, 4-OH), 2.40 (s, 3H, Ts). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 144.9 (Ar), 137.4 (Ar), 133.8 (OCH₂<u>C</u>H=CH₂), 132.9 (Ar), 129.9 (Ar), 128.5 (Ar), 128.2 (Ar), 128.14 (Ar), 128.06 (Ar), 117.9 (OCH₂CH=<u>C</u>H₂), 100.8 (C-1), 75.1 (C-3), 74.4 (Ph<u>C</u>H₂), 74.3 (C-2), 73.0 (C-5), 71.2 (O<u>C</u>H₂CH=CH₂), 69.7 (C-6), 66.6 (C-4), 57.2 (OMe), 21.7 (Ts). HRMS (ESI-QTOF) *m*/*z*: [M + Na]⁺ calcd for C₂₄H₃₀O₈SNa 501.1554, found 501.1551.

Methyl 3-O-allyl-2-O-benzyl-6-deoxy-6-iodo-\beta-D-idopyranoside (8): A solution of compound **6** (2.078 g, 6.405 mmol), PPh₃ (2.196 g, 8.374 mmol), and imidazole (0.800 g, 11.8 mmol) in dry CH₂Cl₂ (20 mL) was flushed with Ar, and then cooled to 0 °C in an ice-H₂O bath. I₂ (2.649 g, 10.44 mmol) was added in one portion, and the reaction mixture was slowly warmed back to ambient temperature. After 24 h, a fresh solution of PPh₃ (1.306 g, 4.978 mmol) and I₂ (1.350 g, 5.321 mmol) in dry CH₂Cl₂ (10 mL) was added to the reaction mixture. After an additional 24 h, the solution was evaporated to dry and then redissolved into EtOAc (120 mL). The organic phase was washed with saturated Na₂S₂O₃ (aq) solution (4 × 120 mL), dried with Na₂SO₄, filtered, and evaporated to dry. The crude material was then purified via column chromatography on silica

using $8 \rightarrow 15 \rightarrow 20\%$ acetone – hexanes to afford the pure product **8** as a yellow syrup (721 mg, 1.66 mmol, 26% yield), unreacted starting material (555 mg, 1.71 mmol, 27% recovered). $R_{\rm f}$ 0.85 (EtOAc : toluene 6:4). $[\alpha]_{\rm D}^{20}$: -44° (*c* 0.86, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.34 – 7.28 (m, 5H, Ar), 5.76 (dddd, 1H, J = 17.2, 10.4, 5.7, 5.7 Hz, OCH₂C<u>H</u>=CH₂), 5.16 (dddd, 1H, J = 17.2, 1.6, 1.6, 1.6 Hz, OCH₂CH=C<u>H</u>_aH_b), 5.13 (dddd, 1H, J = 10.4, 1.4, 1.4, 1.4 Hz, OCH₂CH=CH_aH_b), 4.88 (d, 1H, J = 12.2 Hz, PhC<u>H</u>_aH_b), 4.62 (d, 1H, J = 12.2 Hz, PhCH_aH_b), 4.61 (d, 1H, J = 1.0 Hz, H-1), 3.94 – 3.90 (m, 3H, OC<u>H</u>₂CH=CH₂ and H-5), 3.70 – 3.66 (m, 2H, H-3 and H-4), 3.60 (s, 3H, OMe), 3.53 (ddd, 1H, J = 3.1, 1.2, 1.2 Hz, H-2), 3.48 (dd, 1H, J = 12.6, 1.0 Hz, 4-OH), 3.40 (dd, 1H, J = 10.2, 7.7 Hz, H-6a), 3.37 (dd, 1H, J = 10.2, 6.2 Hz, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 137.6 (Ar), 134.0 (OCH₂CH=CH₂), 128.7 (Ar), 128.34 (Ar), 128.26 (Ar), 118.1 (OCH₂CH=<u>C</u>H₂), 101.5 (C-1), 76.0 (C-5), 75.9 (C-3), 74.6 (Ph<u>C</u>H₂), 74.3 (C-2), 71.4 (O<u>C</u>H₂CH=CH₂), 67.8 (C-4), 57.4 (OMe), 4.0 (C-6). HRMS (ESI-QTOF) *m/z*: [M + Na]⁺ calcd for C₁₇H₂₃IO₅Na 457.0482, found 457.0479.

Methyl 3-O-allyl-2-O-benzyl-6-cyano-6-deoxy-\beta-D-idopyranoside (9): Method A: A solution of the 6-*O*-tosyl starting material 7 (121 mg, 0.252 mmol), KCN (76 mg, 1.2 mmol), and 18-crown-6 (299 mg, 1.13 mmol) in dry DMF (1.3 mL) were left mixing at rt. After 48 h, the mixture was temperature was increased to 60 °C. After 24 h of heating, the reaction mixture was evaporated to dry, redissolved into EtOAc (50 mL), washed with saturated NaCl (aq) solution (3 × 50 mL), dried with Na₂SO₄, filtered, and evaporated to dry. The crude material was purified via column chromatography on silica using 15 \rightarrow 25 \rightarrow 40% EtOAc – hexanes to afford the desired product **9** as a colourless syrup (11 mg, 0.032 mmol, 13% yield), the hydrolyzed by-product **6**

The Journal of Organic Chemistry

(16 mg, 0.048 mmol, 19% yield). Attempts at using Bu_4NI instead of 18-crown-6 were also unsuccessful, and resulted in significant formation of the hydrolyzed by-product **6**.

Method B: Compound 8 (113 mg, 0.260 mmol) and KCN (59 mg, 0.91 mmol) were dissolved into dry DMF (1.3 mL) and left mixing at 80 °C. After 48 h, the reaction mixture was diluted with EtOAc (60 mL), washed with saturated NaCl (aq) solution (3×60 mL), dried with Na₂SO₄, filtered, and evaporated to dry. The crude material was purified via column chromatography on silica using $10 \rightarrow 15 \rightarrow 20\%$ EtOAc – hexanes to afford the pure product as a pale yellow syrup (34 mg, 0.10 mmol, 39% yield). Attempts to leave the reaction for longer or at increased temperatures resulted in the formation of a number of uncharacterized by-products. $R_{\rm f}$ 0.56 (EtOAc : hexanes 2:3). $[\alpha]_D^{20}$: -50° (c 0.94, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): δ_H 7.35 – 7.28 (m, 5H, Ar), 5.75 (dddd, 1H, J = 17.0, 10.5, 5.7, 5.7 Hz, OCH₂CH=CH₂), 5.15 (dddd, 1H, J= 17.1, 1.5, 1.5, 1.5, Hz, OCH₂CH=CH₃H_b), 5.14 (dddd, 1H, J = 10.5, 1.3, 1.3, 1.3, Hz, $OCH_2CH=CH_aH_b$, 4.88 (d, 1H, J = 12.1 Hz, Ph CH_aH_b), 4.65 (d, 1H, J = 0.9 Hz, H-1), 4.62 (d, 1H, J = 12.1 Hz, PhCH_aH_b), 4.08 (ddd, 1H, J = 8.6, 5.5, 1.3 Hz, H-5), 3.92 (dddd, 2H, J = 5.7, 1.4, 1.4, <1 Hz, OCH₂CH=CH₂), 3.71 (dd, 1H, *J* = 3.3, 3.3 Hz, H-3), 3.59 (ddd, 1H, *J* = 3.4, 1.0, 1.0 Hz, H-2), 3.58 (s, 3H, OMe), 3.58 (d, 1H, J = 11.7 Hz, 4-OH), 3.46 (dddd, 1H, J = 11.6, 3.1, 1.2, 1.2 Hz, H-4), 2.86 (dd, 1H, J = 16.8, 8.6 Hz, H-6a), 2.64 (dd, 1H, J = 16.8, 5.5 Hz, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 137.4 (Ar), 133.8 (OCH₂CH=CH₂), 128.7 (Ar), 128.4 (Ar), 118.3 (OCH₂CH=CH₂), 117.8 (CN), 101.2 (C-1), 75.3 (C-3), 74.7 (PhCH₂), 74.0 (C-2), 71.6 (OCH₂CH=CH₂), 71.3 (C-5), 67.6 (C-4), 57.4 (OMe), 20.0 (C-6). HRMS (ESI-QTOF) *m/z*: [M + NH_4 ⁺ calcd for C₁₈H₂₇N₂O₅ 351.1914, found 351.1917.

Methyl 2,3-di-O-benzyl-6-deoxy-6-iodo-B-D-idopyranoside (11) and 1,6-anhydro-2,3-di-Obenzyl-β-D-idopyranose (12): A solution of the compound 10 (152 mg, 0.407 mmol), PPh₃ (129 mg, 0.491 mmol), and imidazole (62 mg, 0.91 mmol) in dry CH₂Cl₂ (1.5 mL) was flushed with Ar, and then cooled to 0 °C. I₂ (170 mg, 0.671 mmol) was added in one portion, and the reaction mixture slowly warmed back to ambient temperature. After 24 h, a solution of PPh₃ (218 mg, 0.832 mmol) and I₂ (214 mg, 0.844 mmol) in CH₂Cl₂ (0.5 mL) was added at 0 °C and then the solution warmed back to ambient temperature. After another 24 h, the solution was evaporated to dry and then redissolved into EtOAc (60 mL). The organic phase was washed with saturated $Na_2S_2O_3$ (aq) solution (2 × 60 mL), saturated NaCl (aq) solution (2 × 60 mL), dried with Na_2SO_4 , filtered, and evaporated to dry. The crude material was then purified via column chromatography on silica using $10 \rightarrow 15 \rightarrow 20\%$ acetone – hexanes to afford the pure product 11 as a yellow syrup (124 mg, 0.257 mmol, 63% yield), the 1,6-anhydro by-product 12 (36 mg, 0.11 mmol, 26% vield). Data for **11**: $R_{\rm f}$ 0.70 (EtOAc : toluene 3:2). $[\alpha]_{\rm D}^{-20}$: -64° (*c* 0.96, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.36 – 7.27 (m, 8H, Ar), 7.23 – 7.19 (m, 2H, Ar), 4.85 (d, 1H, J = 12.2 Hz, PhCH_aH_b), 4.67 (d, 1H, J = 0.9 Hz, H-1), 4.61 (d, 1H, J = 12.2 Hz, PhCH_aH_b), 4.49 (d, 1H, J =11.9 Hz, PhCH_aH_b), 4.45 (d, 1H, J = 11.9 Hz, PhCH_aH_b), 3.98 (ddd, 1H, J = 7.6, 6.3, 0.9 Hz, H-5), 3.78 (dd, 1H, J = 3.2, 3.2 Hz, H-3), 3.74 (dddd, 1H, J = 11.5, 3.1, 1.1, 1.1 Hz, H-4), 3.61 (s, 3H, OMe), 3.57 (ddd, 1H, J = 3.2, 1.1, 1.0 Hz, H-2), 3.50 (d, 1H, J = 11.6 Hz, 4-OH), 3.42 (dd, JH, J = 11.6 Hz), 1H, J = 10.1, 7.7 Hz, H-6a), 3.39 (dd, 1H, J = 10.2, 6.2 Hz, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 137.5 (Ar), 137.4 (Ar), 128.7 (Ar), 128.6 (Ar), 128.22 (Ar), 128.16 (Ar), 127.8 (Ar), 101.4 (C-1), 75.92 (C-3), 75.88 (C-5), 74.4 (PhCH₂), 74.1 (C-2), 72.4 (PhCH₂), 67.6 (C-4), 57.2 (OMe), 3.9 (C-6). HRMS (ESI-QTOF) m/z: $[M + Na]^+$ calcd for C₂₁H₂₅IO₅Na 507.0639, found 507.0642. Data for 12: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.37 – 7.26 (m, 10H, Ar), 5.32 (d, 1H, J

= 1.5 Hz, H-1), 4.91 (d, 1H, J = 11.6 Hz, PhC<u>H</u>_aH_b), 4.69 (d, 1H, J = 11.6 Hz, PhCH_a<u>H</u>_b), 4.68 (s, 2H, PhC<u>H</u>₂), 4.36 (ddd, 1H, J = 4.6, 4.6, <1 Hz, H-5), 4.04 (dd, 1H, J = 7.7, <1 Hz, H-6a), 3.82 – 3.78 (m, 1H, H-4), 3.68 (dd, 1H, J = 7.7, 5.0 Hz, H-6b), 3.60 (dd, 1H, J = 8.1, 8.1 Hz, H-3), 3.49 (dd, 1H, J = 8.0, 1.6 Hz, H-2), 2.08 (d, 1H, J = 2.6 Hz, 4-OH). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 138.8 (Ar), 138.1 (Ar), 128.8 (Ar), 128.7 (Ar), 128.21 (Ar), 128.16 (Ar), 128.15 (Ar), 128.1 (Ar), 99.8 (C-1), 82.8 (C-2), 82.5 (C-3), 75.24 (PhCH₂), 75.20 (C-5), 73.0 (PhCH₂), 71.5 (C-4), 65.5 (C-6). HRMS (ESI-QTOF) *m/z*: [M + NH₄]⁺ calcd for C₂₀H₂₆NO₅ 360.1805, found 360.1802.

Methyl 2,3-di-O-benzyl-6-cyano-6-deoxy-\beta-D-idopyranoside (13): The starting material (11, 630 mg, 1.30 mmol) and KCN (674 mg, 10.4 mmol) in dry DMF (10 mL) were heated at 80 °C. After 24 h the reaction was concentrated to half volume via co-evaporation with toluene (2 × 10 mL), then diluted with EtOAc (100 mL). The organic phase was washed with saturated NaCl (aq) solution (3 × 100 mL), H₂O (100 mL), dried with Na₂SO₄, filtered, and evaporated to dry. The crude material was used directly for the following step.

Methyl 2,3-di-O-benzyl-\beta-D-ido-heptopyranoside (15): The crude starting material **13** (1.30 mmol from previous step) was dissolved into dry toluene (20 mL), and cooled to -72 °C. DIBAL-H solution (1 M in toluene, 19 mL, 19 mmol) was added dropwise over 15 min, and after 2 h the reaction mixture slowly warmed up to ambient temperature. After 24 h, the reaction mixture was diluted with CH₂Cl₂ (20 mL) and then quenched with MeOH (30 mL). The crude mixture was evaporated to dry, then redissolved into EtOAc (100 mL), washed with 1.2 M HCl

(aq) (2 \times 100 mL), H₂O (100 mL), dried with Na₂SO₄, filtered, and evaporated to dry. TLC indicated a complex mixture of products had formed, but none were successfully identified.

1,2;3,4-Di-O-isopropylidene-a-D-galactohexodialdo-1,5-pyranose (17): To a suspension of PDC (4.51 g, 0.012 mol) and NaOAc (1.97 g, 0.024 mol) in anhydrous CH₂Cl₂ (50 mL), was added crushed molecular sieves (4 Å, 4.11 g) at ambient temperature. The suspension was stirred for 1 h. A solution of the compound **16** (1.05 g, 0.04 mol) in anhydrous CH₂Cl₂ (2 mL) was added dropwise, and the reaction was continued for another 6 h at ambient temperature. Hexanes (100 mL) were added, and the mixture was filtered through a thin bed of silica gel using 30% EtOAc–hexanes as the eluent to obtain the desired product **17** as a colorless liquid (854 mg, 0.033 mol, 82% yield). $R_{\rm f}$ 0.42 (EtOAc : hexanes 2:3). The ¹H and ¹³C NMR data are consistent with the published data.²⁷

6,7-Dideoxy-1,2:3,4-di-O-isopropylidene- α -D-galacto-hepto-6-enopyranose (19): To a solution of Ph₃PCH₃Br (2.72 g, 7.63 mmol) in anhydrous THF (15 mL), was added NaH (60% oil dispersion, 254 mg, 7.36 mmol) at ambient temperature. The solution was stirred at 50 °C overnight. A solution of the aldehyde 17 (820 mg, 3.18 mmol) in anhydrous THF (2 mL) was added dropwise to the reaction mixture. After stirring for 2 h, methanol (5 mL) was added to quench the reaction. The reaction mixture was concentrated under reduced pressure, and the obtained crude product was redissolved in EtOAc (80 mL), washed with saturated brine (60 mL × 2), dried over anhydrous Na₂SO₄, filtered, and evaporated. The crude mixture was purified by column chromatography on silica gel using a gradient of 5 \rightarrow 15% EtOAc–hexanes as the eluent

to afford the desired alkene **19** as a syrup (615 mg, 2.41 mmol, 74% yield). $R_f 0.62$ (EtOAc : hexanes 1:4). The ¹H and ¹³C NMR data are consistent with the published data.²⁷

Attempted synthesis of 6-deoxy-1,2:3,4-di-O-isopropylidene- α -D-galacto-heptopyranose

(20): To a solution of the alkene 19 (51 mg, 0.21 mmol) in anhydrous THF (2 mL) was added 1M borane THF complex solution (1.0 mL, 1.0 mmol) dropwise. After the reaction mixture was stirred at ambient temperature for 5 h, a 3M solution of NaOH (1.2 mL, 1.1 mmol) and 50% w/w H_2O_2 solution (0.30 mL, 4.2 mmol) were added. The reaction was continued for another 10 h. TLC (EtOAc : hexanes 1:4) showed the disappearance of the starting material, but several spots were formed. Attempted purification of the mixture by column chromatography did not yield the desired product.

(Z)- and (E)-7-O-benzyl-6-deoxy-1,2:3,4-di-O-isopropylidene- α -D-galactohepto-6enopyranose (23): A solution of Ph₃PCH₂OBnCl (21, prepared by heating a solution of benzyl chloromethyl ether (1.0 equiv.) and PPh₃ (1.1 equiv.) in dry toluene at 100 °C for 20 h; 4.68 g, 11.2 mmol) in anhydrous THF (30 mL) was cooled to -78 °C, and a solution of 2.5 M *n*butyllithium in hexanes (3.7 mL, 9.3 mmol) was added. After stirring for 30 min. a solution of the aldehyde **17** (1.21 g, 4.69 mmol) in anhydrous THF (5 mL) was added dropwise, and the reaction mixture was allowed to warm to ambient temperature overnight. Methanol (5 mL) was added to quench the reaction, and the reaction solution was concentrated under reduced pressure. The crude mixture was redissolved in EtOAc (250 mL), and the solution was washed with saturated brine (80 mL × 2), dried over anhydrous Na₂SO₄, filtered, and evaporated. The crude mixture was purified by column chromatography on silica gel using a gradient of 5 \rightarrow 15%

EtOAc-hexanes to afford the desired product 23 as syrup (1.01 g, 2.79 mmol, 60% yield, Z : E =1:1). $R_{\rm f}$ 0.62 (EtOAc : hexanes 1 : 4). Data for Z-isomer of 23: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.39-7.29 (m, 5H, Ar), 6.72 (d, 1H, J = 12.8 Hz, H-7), 5.56 (d, 1H, J = 5.0 Hz, H-1), 5.14 (dd, 1H, J = 12.8, 8.8 Hz, H-6), 4.81 (d, 1H, J = 11.6 Hz, PhCH₂), 4.77 (d, 1H, J = 11.6 Hz, PhCH₂), 4.62 (dd, 1H, J = 7.8, 2.4 Hz, H-3), 4.30 (dd, 1H, J = 5.0, 2.4 Hz, H-2), 4.23 (dd, 1H, J = 8.7, 1.9 Hz, H-5), 4.14 (dd, 1H, J = 7.9, 1.9 Hz, H-4), 1.54 (s, 3H, Me), 1.50 (s, 3H, Me), 1.36 (s, 3H, Me), 1.35 (s, 3H, Me). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 150.6 (C-7), 128.5 (Ar), 128.0 (Ar), 127.8 (Ar), 109.1 (C(Me)₂), 108.4 (C(Me)₂), 99.7 (C-6), 96.6 (C-1), 74.1 (C-4), 71.0 (C-3), 70.9 (PhCH₂), 70.3 (C-2), 66.9 (C-5), 26.2 (Me), 26.1 (Me), 24.9 (Me), 24.4 (Me). Data for *E*-isomer of 23: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.39-7.28 (m, 5H, Ar), 6.22 (dd, 1H, J = 6.2, 1.0 Hz, H-7), 5.54 (d, 1H, J = 4.9 Hz, H-1), 4.94 (ddd, 1H, J = 8.5, 1.0, 0.9 Hz, H-5), 4.85 (d, 1H, J = 12.6Hz, PhCH₂), 4.81 (d, 1H, J = 12.6 Hz, PhCH₂), 4.72 (dd, 1H, J = 8.6, 6.2 Hz, H-6), 4.60 (dd, 1H, J = 7.9, 2.4 Hz, H-3, 4.29 (dd, 1H, J = 5.0, 2.4 Hz, H-2), 4.19 (dd, 1H, J = 7.9, 1.9 Hz, H-4), 1.52 (s, 3H, Me), 1.48 (s, 3H, Me), 1.34 (s, 3H, Me), 1.33 (s, 3H, Me). ¹³C NMR (CDCl₃, 100 MHz): δ_{C} 147.1 (C-7), 128.5 (Ar), 127.9 (Ar), 127.4 (Ar), 109.1 (C(Me)₂), 108.6 (C(Me)₂), 102.8 (C-6), 96.6 (C-1), 74.2 (PhCH₂), 73.4 (C-4), 70.9 (C-2), 70.5 (C-3), 62.2 (C-5), 26.1 (Me), 26.0 (Me), 25.2 (Me), 24.4 (Me). HRMS (ESI-QTOF) m/z: $[M + H]^+$ calcd for C₂₀H₂₇O₆ 363.1808, found 363.1806.

6-Deoxy-1,2:3,4-di-O-isopropylidene- α -D-galacto-heptopyranose (20): To a solution of compound 23 (23.3 mg, 0.064 mmol) in methanol (2 mL) was added Pd(OH)₂ (20% on charcoal, 10 mg). The solution was stirred under a positive atmosphere of hydrogen gas. After 1 day, the mixture was filtered to remove the catalyst and the solution was evaporated to afford the desired

product **20**²⁵ as a colorless syrup (13.1 mg, 0.048 mmol, 75% yield). $R_{\rm f}$ 0.35 (EtOAc : hexanes 2:3). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 5.55 (d, 1H, J = 5.0 Hz, H-1), 4.63 (dd, 1H, J = 7.9, 2.4 Hz, H-3), 4.34 (dd, 1H, J = 5.1, 2.4 Hz, H-2), 4.17 (dd, 1H, J = 7.9, 1.9 Hz, H-4), 4.03 (ddd, 1H, J = 9.0, 3.8, 3.8 Hz, H-5), 3.87-3.75 (m, 2H, H-7a, H-7b), 2.05 (d, 1H, J = 12.0 Hz, OH-7), 2.00 (dddd, 1H, J = 14.8, 10.0, 6.1, 4.4 Hz, H-6a), 1.87 (dddd, 1H, J = 14.5, 9.3, 5.0, 3.8 Hz, H-6b), 1.56 (s, 3H, Me), 1.49 (s, 3H, Me), 1.37 (s, 3H, Me), 1.36 (s, 3H, Me). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 109.2 (<u>C</u>(Me)₂), 108.6 (<u>C</u>(Me)₂), 96.5 (C-1), 73.1 (C-4), 70.9 (C-2), 70.5 (C-3), 65.8 (C-5), 59.9 (C-7), 32.8 (C-6), 26.1 (Me), 26.0 (Me), 24.9 (Me), 24.4 (Me). HRMS (ESI-QTOF) m/z: [M + H]⁺ calcd for C₁₃H₂₃O₆ 275.1489, found 275.1492.

7-O-Benzyl-6-deoxy-1,2:3,4-di-O-isopropylidene-α-D-galacto-heptopyranose (24): To a solution of compound **23** (248 mg, 0.685 mmol) in methanol (3 mL), was added Pd(OH)₂ (20% on charcoal, 22 mg). The solution was stirred under a positive atmosphere of hydrogen for 1 h. The catalyst was filtered off and the organic solution was evaporated to afford the desired product **24** as a syrup (238 mg, 0.657 mmol, 96% yield). $R_{\rm f}$ 0.64 (EtOAc : hexanes 1:4). $[\alpha]_{\rm D}^{20}$: +45.8° (c 1.0, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.37-7.23 (m, 5H, Ar), 5.53 (d, 1H, J = 5.0 Hz, H-1), 4.58 (dd, 1H, J = 7.9, 2.4 Hz, H-3), 4.55 (d, 1H, J = 11.8 Hz, PhCH₂), 4.49 (d, 1H, J = 11.8 Hz, PhCH₂), 4.29 (dd, 1H, J = 5.0, 2.4 Hz, H-2), 4.12 (dd, 1H, J = 7.9, 1.8 Hz, H-4), 4.40 (ddd, 1H, J = 9.0, 3.3, 1.8 Hz, H-5), 3.65 (ddd, 1H, J = 9.3, 9.3, 3.7 Hz, H-7a), 3.60 (ddd, 1H, J = 9.6, 9.4, 5.4 Hz, H-7b), 1.97 (dddd, 1H, J = 13.9, 9.5, 5.3, 4.9 Hz, H-6a), 1.90 (dddd, 1H, J = 14.2, 9.8, 5.3, 4.9 Hz, H-6b), 1.52 (s, 3H, Me), 1.48 (s, 3H, Me), 1.36 (s, 3H, Me), 1.35 (s, 3H, Me). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 128.4 (Ar), 127.6 (Ar), 127.4 (Ar), 109.0 (<u>C</u>(Me)₂), 108.5 (C(Me)₂), 9.6.6 (C-1), 73.1 (C-4), 73.0 (PhCH₂), 70.9 (C-3), 70.7 (C-2), 66.6 (C-7), 64.1

(C-5), 30.8 (C-6), 26.1 (Me), 26.0 (Me), 25.1 (Me), 24.4 (Me). HRMS (ESI-QTOF) m/z: [M + H]⁺ calcd for C₂₀H₂₉O₆ 365.1959, found 365.1966.

7-O-Benzyl-6-deoxy-1,2,3,4-tetra-O-acetyl-D-galacto-heptopyranose (25): A solution of compound 24 (1.02 g, 2.82 mmol) in 50% AcOH – H_2O (8 mL) was heated to 80 °C for 8 h. The reaction mixture was evaporated under reduced pressure, and the obtained residue was acetylated using a mixture of Ac₂O (4 mL) and pyridine (4 mL). After stirring for 10 h at ambient temperature, the mixture was evaporated and co-evaporated with toluene three times. The crude mixture was purified by column chromatography on silica gel using 30% EtOAc-hexanes as the eluent to afford the desired product 25 (α/β : 1:1.8) as a syrup (1.185 g, 2.62 mmol, 93% yield). $R_{\rm f}$ 0.39 (EtOAc : hexanes 2:3). Data for α anomer: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.42-7.33 (m, 5H, Ar), 6.35 (d, 1H, J = 3.3 Hz, H-1), 5.42 (dd, 1H, J = 3.0, 1.3 Hz, H-4), 5.38-5.33 (m, 2H, H-2 and H-3), 4.37 (dd, 1H, J = 8.9, 4.6 Hz, H-5), 4.55-4.48 (m, 2H, PhCH₂), 3.59-3.48 (m, 2H, H-7a and H-7b), 2.17 (s, 3H, Ac), 2.09 (s, 3H, Ac), 2.04 (s, 3H, Ac), 2.02 (s, 3H, Ac), 1.98-1.86 (m, 1H, H-6a), 1.85-1.69 (m, 1H, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 128.4 (Ar), 127.7 (Ar), 127.6 (Ar), 89.8 (C-1), 73.1 (PhCH₂), 69.9 (C-3), 67.9 (C-4), 67.8 (C-5), 66.7 (C-2), 65.4 (C-7), 30.7 (C-6), 20.8 (Ac), 20.7 (Ac), 20.6 (Ac), 20.5 (Ac). Data for β anomer: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.42-7.27 (m, 5H, Ar), 5.70 (d, 1H, J = 7.9 Hz, H-1), 5.38-5.32 (m, 2H, H-2 and H-4), 5.11 (dd, 1H, J = 10.4, 3.4 Hz, H-3), 4.48-4.43 (m, 2H, PhCH₂), 4.04 (ddd, 1H, J = 8.5, 4.8, 1.1 Hz, H-5), 3.59-3.48 (m, 2H, H-7a and H-7b), 2.18 (s, 3H, Ac), 2.13 (s, 3H, Ac), 2.06 (s, 3H, Ac), 2.01 (s, 3H, Ac), 1.98-1.86 (m, 1H, H-6a), 1.85-1.69 (m, 1H, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 128.4 (Ar), 127.7 (Ar), 127.5 (Ar), 92.4 (C-1), 73.2 (PhCH₂), 71.3 (C-5),

The Journal of Organic Chemistry

71.2 (C-3)	, 69.3 (C-2),	68.2 (C-4),	66.5 (C-7	'), 30.8 (C	2-6), 20.8	(Ac), 20.7	(Ac), 20.	7 (Ac),	20.5
(Ac). HRM	IS (ESI-QTO	DF) <i>m/z</i> : [M	$+ \mathrm{NH}_4]^+ \mathrm{c}$	alcd for C	$C_{22}H_{32}NO_{1}$	₀ 470.2021	, found 4	70.2016	

7-O-Benzyl-6-deoxy-2,3,4-tri-O-acetyl-D-galacto-heptopyranose (26): To a solution of compound 25 (560 mg, 1.24 mmol) in anhydrous DMF (4 mL), was added hydrazinium acetate (126 mg, 1.36 mmol), and the mixture was stirred at ambient temperature for 4 h. The mixture was diluted with EtOAc (80 mL), extracted with brine (50 mL \times 2), dried over anhydrous Na_2SO_4 , and evaporated. The crude product was purified by column chromatography on silica gel using 40% EtOAc-hexanes as the eluent to afford the pure product 26 (α/β : 1:2.3) as a syrup (434 mg, 1.06 mmol, 85% vield). $R_{\rm f}$ 0.34 (EtOAc : hexanes 2:3). Data for α anomer: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.38-7.25 (m, 5H, Ar), 5.28 (dd, 1H, J = 2.7, 1.2 Hz, H-4), 5.08-5.01 (m, 2H, H-2 and H-3), 4.60 (broad, 1H, H-1), 4.53-4.46 (m, 2H, PhCH₂), 3.91 (ddd, 1H, J = 8.7, 4.7, 4.7, 4.51.1 Hz, H-5), 3.60-3.44 (m, 2H, H-7a, H-7b), 2.14 (s, 3H, Ac), 2.09 (s, 3H, Ac), 1.99 (s, 3H, Ac), 1.93-1.81 (m, 1H, H-6a), 1.80-1.66 (m, 1H, H-6b). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 127.8 (Ar), 127.7 (Ar), 127.6 (Ar), 95.8 (C-1), 73.0 (PhCH₂), 71.4 (C-3), 70.8 (C-4), 70.2 (C-5), 69.6 (C-2), 65.4 (C-7), 30.8 (C-6), 20.7 (Ac), 20.6 (Ac), 20.5 (Ac). Data for β anomer: ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.38-7.27 (m, 5H, Ar), 5.43-5.38 (m, 2H, H-1 and H-2), 5.34 (dd, 1H, J = 3.4, 1.3Hz, H-4), 5.13 (dd, 1H, J = 10.8, 3.6 Hz, H-3), 4.45-4.36 (m, 2H, PhCH₂), 4.41 (ddd, 1H, J =8.3, 5.0, 1.4 Hz, H-5), 3.58-3.47 (m, 2H, H-7a and H-7b), 2.13 (s, 3H, Ac), 2.08 (s, 3H, Ac), 1.98 (s, 3H, Ac), 1.93-1.81 (m, 1H, H-6a), 1.80-1.66 (m, 1H, H-6b). ¹³C NMR (CDCl₃, 100 MHz): δ_C 128.4 (Ar), 127.8 (Ar), 127.6 (Ar), 90.5 (C-1), 72.8 (PhCH₂), 70.5 (C-4), 68.5 (C-3), 67.7 (C-2), 65.5 (C-7), 65.2 (C-5), 30.6 (C-6), 20.8 (Ac), 20.7 (Ac), 20.6 (Ac). HRMS (ESI-OTOF) m/z: [M $+ NH_4$ ⁺ calcd for C₂₀H₃₀NO₉ 428.1915, found 428.1924.

7-O-Benzyl-6-deoxy-2,3,4-tri-O-acetyl-D-galacto-heptopyranosyl trichloroacetimidate (27): To a solution of compound 26 (362 mg, 0.88 mmol) in anhydrous CH₂Cl₂ (5 mL) was added K₂CO₃ (244 mg, 1.76 mmol) and trichloroacetonitrile (0.5 mL, 4.5 mmol), and the reaction mixture was stirred at ambient temperature for 12 h. The solution was evaporated under reduced pressure. The crude product was dissolved in EtOAc (50 mL), extracted with brine (30 mL × 2) and the organic layer was evaporated and co-evaporated with toluene to afford the crude product 27 (α/β : 1:1) as a colorless syrup. R_f 0.42 (EtOAc : hexanes 2:3). The crude product was used directly for next step without further purification.

6-Azidohexyl 3,4-di-O-acetyl-7-O-benzyl-6-deoxy-β-D-galacto-heptopyranoside (**30**) and 6-Azidohexyl 2,3,4-tri-O-acetyl-7-O-benzyl-6-deoxy-β-D-galacto-heptopyranoside (**29**): A mixture of imidate **27** (136 mg, 0.247 mmol), alcohol **28** (177 mg, 1.24 mmol) and molecular sieves (4 Å, crushed, 500 mg) in anhydrous CH₂Cl₂ (5 mL) was stirred for 1 h at ambient temperature under an atmosphere of argon. The mixture was then cooled down to -40 °C and TMSOTf (50 µL) was added dropwise. After stirring for 4 h, the temperature was gradually warmed up to ambient temperature. A few drops of triethylamine were added to quench the reaction, and the mixture was filtered off through a thin bed of celite, and washed with EtOAc (~40 mL). The organic solution was washed with brine (20 mL), dried over Na₂SO₄, and evaporated. The crude mixture was purified by column chromatography on silica gel using 20% EtOAc–hexane as the eluent to afford the pure product **30** as a syrup (90.2 mg, 0.182 mmol, 77% yield). When a similar reaction of imidate **27** (245 mg, 0.442 mmol) and alcohol **28** (95.1 mg, 0.665 mmol) was carried, compound **29** (93.2 mg, 0.189 mmol, 43% yield) and the 2-O-deacetylated product **30**

(59.8 mg, 0.112 mmol, 25% yield) were obtained. Data for compound 30: $R_{\rm f}$ 0.37 (EtOAc :
hexanes 3:7). $[\alpha]_D^{20}$: +57.1° (<i>c</i> 1.0, CHCl ₃). ¹ H NMR (CDCl ₃ , 400 MHz): δ_H 7.41-7.29 (m, 5H,
Ar), 5.30 (dd, 1H, <i>J</i> = 3.5, 1.1 Hz, H-4), 4.96 (dd, 1H, <i>J</i> = 10.2, 3.5 Hz, H-3), 4.56 (d, 1H, <i>J</i> =
11.9 Hz, $PhCH_2$, 4.46 (d, 1H, $J = 11.9$ Hz, $PhCH_2$), 4.29 (d, 1H, $J = 7.7$ Hz, H-1), 3.89 (dd, 1H,
J = 8.7, 4.1 Hz, H-5), 3.85 (ddd, 1H, $J = 13.3, 6.6, 6.6$ Hz, OC <u>H</u> _a H _b), 3.78 (dd, 1H, $J = 10.2, 7.7$
Hz, H-2), 3.64-3.56 (m, 1H, H-7a), 3.55-3.49 (m, 1H, H-7b), 3.49 (ddd, 1H, J = 13.8, 6.8, 6.8
Hz, OCH _a H _b), 3.29 (t, 2H, $J = 6.8$ Hz, -CH ₂ N ₃), 2.31 (broad, 1H, 2-OH), 2.14 (s, 3H, Ac), 2.07,
(s, 3H, Ac), 1.91 (dddd, 1H, <i>J</i> = 13.9, 9.5, 5.3, 4.9 Hz, H-6a), 1.78 (dddd, 1H, <i>J</i> = 14.0, 9.8, 4.9,
4.9 Hz, H-6b), 1.70-1.56 (m, 4H, $2 \times CH_2$), 1.48-1.35 (m, 4H, $2 \times CH_2$). ¹³ C NMR (CDCl ₃ , 100
MHz): δ _C 170.4 (Ac), 170.3 (Ac), 128.4 (Ar), 127.8 (Ar), 127.7 (Ar), 103.1 (C-1), 73.1 (Ph <u>C</u> H ₂),
73.0 (C-3), 70.0 (CH ₂), 69.9 (C-5), 69.7 (C-4), 69.4 (C-2), 65.6 (C-7), 51.3 (CH ₂), 30.9 (C-6),
29.4 (CH ₂), 28.7 (CH ₂), 26.4 (CH ₂), 25.4 (CH ₂), 20.8 (Ac), 20.7 (Ac). HRMS (ESI-QTOF) <i>m/z</i> :
$[M + NH_4]^+$ calcd for $C_{24}H_{39}N_4O_8$ 511.2762, found 511.2762. Data for compound 29 : R_f 0.55
(EtOAc : hexanes 3:7). $[\alpha]_D^{20}$: +32.3° (c 0.99, CHCl ₃). ¹ H NMR (CDCl ₃ , 400 MHz): δ_H 7.38-
7.28 (m, 5H, Ar), 5.28 (dd, 1H, J = 3.5, 1.1 Hz, H-4), 5.16 (dd, 1H, J = 10.4, 7.9 Hz, H-2), 5.02
(dd, 1H, $J = 10.4$, 3.4 Hz, H-3), 4.53 (d, 1H, $J = 12.0$ Hz, PhCH ₂), 4.44 (d, 1H, $J = 12.0$ Hz,
PhC <u>H</u> ₂), 4.38 (d, 1H, <i>J</i> = 7.9 Hz, H-1), 3.87 (ddd, 1H, <i>J</i> = 8.7, 4.6, 1.3 Hz, H-5), 3.79 (ddd, 1H, <i>J</i>
= 9.6, 6.2, 6.2 Hz, OCH_aH_b), 3.61-3.55 (m, 1H, H-7a), 3.50 (ddd, 1H, J = 9.4, 5.2, 5.2 Hz, H-7b),
3.38 (ddd, 1H, $J = 9.6$, 7.2, 6.3 Hz, OCH _a H _b), 3.25 (t, 2H, $J = 6.9$ Hz, -CH ₂ N ₃), 2.14 (s, 3H, Ac),
2.07, (s, 3H, Ac), 1.98, (s, 3H, Ac), 1.89-1.80 (m, 1H, H-6a), 1.78-1.69 (m, 1H, H-6b), 1.63-1.51
(m, 4H, 2 × CH ₂), 1.42-1.32 (m, 4H, 2 × CH ₂). ¹³ C NMR (CDCl ₃ , 100 MHz): δ_{C} 170.5 (Ac),
170.1 (Ac), 169.4 (Ac), 129.5 (Ar), 128.4 (Ar), 127.7 (Ar), 101.2 (C-1), 73.1 (Ph <u>C</u> H ₂), 71.3 (C-
3), 69.8 (C-4), 69.7 (CH ₂), 69.6 (C-2), 69.2 (C-5), 65.5 (C-7), 51.3 (CH ₂), 30.8 (C-6), 29.3

(CH₂), 28.7 (CH₂), 26.4 (CH₂), 25.4 (CH₂), 20.8 (Ac), 20.7 (Ac), 20.6 (Ac). HRMS (ESI-QTOF) m/z: [M + NH₄]⁺ calcd for C₂₆H₄₁N₄O₉ 553.2868, found 553.2882.

3,4-di-O-acetyl-7-O-benzyl-6-deoxy-2-O-mesyl-β-D-galacto-heptopyranoside 6-Azidohexvl (31): Compound 30 (57.2 mg, 0.116 mmol) was dissolved in anhydrous pyridine (1.0 mL) and methanesulfonyl chloride (20 µL, 0.232 mmol) was added. The mixture was stirred at ambient temperature for 6 h. Methanol (0.5 mL) was added to quench the reaction, and the crude mixture was evaporated and co-evaporated with toluene to dry. The residue was redissolved in EtOAc (~30 mL), and the organic solution was successively washed with aqueous HCl (1 M, 10 mL), aqueous NaOH (1 M, 10 mL) and saturated brine (20 mL), dried over Na₂SO₄, and evaporated. The crude material was purified by column chromatography on silica gel using a gradient of $25 \rightarrow 30\%$ EtOAc- hexanes as the eluent to afford the pure product **31** as a syrup (60.4 mg, 0.106 mmol, 91% yield). $R_{\rm f}$ 0.44 (EtOAc : hexanes 3:7). $[\alpha]_{\rm D}^{20}$: +37.9° (c 1.0, CHCl₃). ¹H NMR $(CDCl_3, 400 \text{ MHz})$: $\delta_H 7.39-7.26 \text{ (m, 5H, Ar)}$, 5.31 (dd, 1H, J = 3.5, 0.8 Hz, H-4), 5.08 (dd, 1H, J= 10.2, 3.5 Hz, H-3, 4.66 (dd, 1H, J = 10.2, 7.9 Hz, H-2), 4.53 (d, 1H, $J = 11.9 \text{ Hz}, \text{PhCH}_2$), 4.44 (d, 1H, J = 7.9 Hz, H-1), 4.47 (d, 1H, J = 11.9 Hz, PhCH₂), 3.88 (dd, 1H, J = 8.7, 4.2 Hz, H-5), 3.80 (ddd, 1H, J = 13.5, 6.8, 6.8 Hz, OCH_aH_b), 3.56 (ddd, 1H, J = 9.0, 8.7, 4.4 Hz, H-7a), 3.49 $(ddd, 1H, J = 9.9, 5.0, 5.0 Hz, H-7b), 3.46 (ddd, 1H, J = 13.4, 6.7, 6.7 Hz, OCH_aH_b), 3.26 (t, 2H, 2H, 2H, 2H)$ J = 6.8 Hz, -CH₂N₃), 3.05 (s, 3H, Ms), 2.14 (s, 3H, Ac), 2.05 (s, 3H, Ac), 1.87 (dddd, 1H, J =13.9, 9.5, 5.3, 4.5 Hz, H-6a), 1.76 (ddd, 1H, J = 14.0, 9.8, 5.2, 5.2 Hz, H-6b), 1.66-1.55 (m, 4H, $2 \times CH_2$, 1.44-1.32 (m, 4H, $2 \times CH_2$). ¹³C NMR (CDCl₃, 100 MHz): δ_C 170.3 (Ac), 169.8 (Ac), 128.4 (Ar), 127.8 (Ar), 127.7 (Ar), 100.4 (C-1), 77.5 (C-2), 73.1 (PhCH₂), 70.7 (C-5), 70.0 (C-4), 69.9 (CH₂), 69.8 (C-3), 66.4 (C-7), 51.3 (CH₂), 39.0 (Ms), 30.7 (C-6), 29.4 (CH₂), 28.7

(CH₂), 26.4 (CH₂), 25.4 (CH₂), 20.6 (Ac), 20.5 (Ac). HRMS (ESI-QTOF) m/z: [M + NH₄]⁺ calcd for C₂₅H₄₁N₄O₁₀S 589.2538, found 589.2532.

-Azidohexyl 7-O-benzyl-6-deoxy-2-O-mesyl- β -D-galacto-heptopyranoside (32): To a solution of compound **31** (33.5 mg, 0.058 mmol) in benzyl alcohol (1.0 mL) was added KOBu-t (20 mg) at ambient temperature, and the reaction mixture was stirred for 1 h until TLC showed the disappearance of the starting material. The reaction mixture was diluted with EtOAc (30 mL) and the organic solution was washed with brine (20 mL), dried over anhydrous Na_2SO_4 and evaporated. The crude mixture was then purified by column chromatography on silica gel using 50% EtOAc-hexanes to afford the pure compound 32 as a syrup (21.1 mg, 0.044 mmol, 75% yield). $R_{\rm f}$ 0.26 (EtOAc : hexanes 1:1). $[\alpha]_{\rm D}^{20}$: +47.3° (c 0.98, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.39-7.26 (m, 5H, Ar), 4.54 (d, 1H, J = 11.9 Hz, PhCH₂), 4.47 (d, 1H, J = 11.9 Hz, PhCH₂), 4.45 (dd, 1H, J = 9.4, 7.9 Hz, H-2), 4.34 (d, 1H, J = 7.9 Hz, H-1), 3.83 (dd, 1H, J = 5.2, 3.4 Hz, H-4), 3.81-3.76 (m, 1H, OCH_aH_b), 3.73 (dd, 1H, J = 9.4, 3.5 Hz, H-3), 3.66 (dd, 1H, J =7.2, 5.9 Hz, H-5), 3.63-3.55 (m, 2H, H-7a, H-7b), 3.45 (ddd, 1H, J = 9.5, 6.7, 6.7 Hz, OCH_aH_b), 3.25 (t, 2H, J = 6.8 Hz, -CH₂N₃), 3.12 (s, 3H, Ms), 2.06 (dddd, 1H, J = 13.2, 7.7, 5.3, 3.9 Hz, H-6a), 1.96 (dddd, 1H, J = 14.7, 8.0, 5.2, 5.2 Hz, H-6b), 1.66-1.52 (m, 4H, 2 × CH₂), 1.43-1.32 (m, 4H, $2 \times CH_2$). ¹³C NMR (CDCl₃, 100 MHz): δ_C 128.4 (Ar), 127.8 (Ar), 127.7 (Ar), 100.0 (C-1), 81.8 (C-2), 73.2 (PhCH₂), 72.1 (C-3), 71.4 (C-5), 70.7 (C-4), 69.6 (CH₂), 65.8 (C-7), 51.3 (CH₂), 38.5 (Ms), 30.7 (C-6), 29.4 (CH₂), 28.7 (CH₂), 26.4 (CH₂), 25.4 (CH₂). HRMS (ESI-QTOF) m/z: $[M + NH_4]^+$ calcd for C₂₁H₃₇N₄O₈S 505.2327, found 505.2326.

6-Azidohexvl 2,3-anhvdro-7-O-benzvl-6-deoxv-B-D-talo-heptopyranoside (33): To a solution of the compound 32 (72 mg, 0.143 mmol) in benzyl alcohol (1.0 mL) was added KOBu-t (52 mg, 0.377 mmol), and the reaction was continued at ambient temperature for 12 h. The reaction mixture was diluted with EtOAc (30 mL) and washed with brine (20 mL \times 2). The organic layer was dried over anhydrous Na_2SO_4 , and evaporated. The crude mixture was then purified by column chromatography on silica gel using a gradient of $7 \rightarrow 15\%$ EtOAc-CH₂Cl₂ as the eluent to afford the pure product 33 as a syrup (49.7 mg, 0.123 mmol, 86% yield). $R_{\rm f}$ 0.42 (EtOAc : hexanes 1:1). $\left[\alpha\right]_{D}^{20}$: +30.5° (c 1.0, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.38-7.26 (m, 5H, Ar), 4.70 (d, 1H, J = 0.2 Hz, H-1), 4.54 (d, 1H, J = 12.1 Hz, PhCH₂), 4.45 (d, 1H, J = 12.1 Hz, PhCH₂), 3.87 (ddd, 1H, J = 13.0, 6.5, 6.5 Hz, OCH_aH_b), 3.72 (broad, 1H, H-4), 3.66-3.55 (m, 2H, H-7a and H-7b), 3.57 (t, 1H, J = 0.9 Hz, H-3), 3.53 (dd, 1H, J = 5.4, 2.7 Hz, H-5), 3.47 (ddd, 1H, J = 13.6, 6.8, 6.8 Hz, OCH_aH_b), 3.30 (d, 1H, J = 3.8 Hz, H-2), 3.29 (t, 2H, J = 6.8 Hz, - CH_2N_3), 2.43 (d, 1H, J = 11.7 Hz, OH-4), 1.97 (dddd, 1H, J = 18.0, 13.9, 9.5, 5.5 Hz, H-6a), 1.91 (dddd, 1H, J = 18.5, 12.9, 9.3, 5.5 Hz, H-6b), 1.73-1.55 (m, 4H, 2 × CH₂), 1.49-1.36 (m, 4H, 2 × CH₂). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 128.4 (Ar), 127.7 (Ar), 127.6 (Ar), 97.4 (C-1), 74.2 (C-5), 73.0 (PhCH₂), 69.6 (CH₂), 66.0 (C-7), 62.9 (C-4), 71.2 (C-5), 54.2 (C-3), 52.9 (C-2), 51.4 (CH₂), 31.0 (C-6), 29.7 (CH₂), 29.5 (CH₂), 26.5 (CH₂), 25.6 (CH₂). HRMS (ESI-QTOF) m/z: $[M + NH_4]^+$ calcd for C₂₀H₃₃N₄O₅ 409.2445, found 409.2433.

6-Azidohexyl 3,7-di-O-benzyl-6-deoxy-β-D-ido-heptopyranoside (34): To the solution of the compound 33 (35 mg, 0.089 mmol) in benzyl alcohol (1 mL) was added KOBu-t (30 mg, 0.22 mmol). The reaction mixture was stirred for 12 h at 60 °C. The reaction mixture was diluted with EtOAc (30 mL) and washed with saturated brine (20 mL). The organic layer was dried over

anhydrous Na₂SO₄, filtered, and evaporated. The crude material was then purified by column chromatography on silica gel using a gradient of $7 \rightarrow 10\%$ EtOAc-CH₂Cl₂ as the eluent to afford the pure product 34 as a syrup (29.6 mg, 0.059 mmol, 67% yield). Compound 34 could also be synthesized from compound **31** in 63% yield by reaction with KOBu-t in benzyl alcohol by gradually raising the temperature to 60 °C. $R_{\rm f}$ 0.45 (EtOAc : hexanes1:1). $\left[\alpha\right]_{\rm D}^{20}$: -52.5° (c 1.0, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.38-7.25 (m, 10H, Ar), 4.68 (d, 1H, J = 0.7 Hz, H-1), 4.63 (d, 1H, J = 11.8 Hz, PhCH₂), 4.58 (d, 1H, J = 11.8 Hz, PhCH₂), 4.55 (d, 1H, J = 11.9 Hz, PhCH₂), 4.49 (d, 1H, J = 11.9 Hz, PhCH₂), 4.03 (dd, 1H, J = 8.5, 5.2 Hz, H-5), 3.91 (t, 1H, J =2.6 Hz, H-3), 3.86 (ddd, 1H, J = 9.6, 6.6, 6.6 Hz, OCH_aH_b), 3.78 (broad, 1H, H-2), 3.69-3.63 (m, 2H, H-7a and H-7b), 3.58-3.46 (m, 3H, H-4, OCH_aH_b, OH-4), 3.26 (t, 2H, J = 6.8 Hz, $-CH_2N_3$), 2.74 (broad, 1H, OH-2), 2.12 (dddd, 1H, J = 18.0, 13.9, 9.5, 5.5 Hz, H-6a), 1.98 (dddd, 1H, J = 18.5, 12.9, 9.3, 5.5 Hz, H-6b), 1.66-1.56 (m, 4H, $2 \times CH_2$), 1.44-1.32 (m, 4H, $2 \times CH_2$). ¹³C NMR (CDCl₃, 100 MHz): δ_C 128.5 (Ar), 128.4 (Ar), 127.9 (Ar), 127.7 (Ar), 127.6 (Ar), 127.5 (Ar), 98.4 (C-1), 76.4 (C-3), 73.0 (PhCH₂), 72.3 (PhCH₂), 71.2 (C-5), 69.1 (CH₂), 68.9 (C-2), 68.0 (C-4), 66.3 (C-7), 51.4 (CH₂), 31.0 (C-6), 29.7 (CH₂), 29.4 (CH₂), 26.5 (CH₂), 25.6 (CH₂). HRMS (ESI-QTOF) m/z: $[M + Na]^+$ calcd for C₂₇H₃₇N₃O₆Na 522.2575, found 522.2567.

6-Azidohexyl 3-O-allyl-7-O-benzyl-6-deoxy-β-D-ido-heptopyranoside (35): To the solution of the compound **33** (156 mg, 0.397 mmol) in allyl alcohol (1.5 mL) was added KOBu-*t* (101 mg, 0.901 mmol). The reaction mixture was stirred for 12 h at 60 °C until TLC showed the disappearance of the starting material. The reaction mixture was diluted with EtOAc (30 mL) and washed with brine (20 mL × 2). The organic layer was dried over anhydrous Na₂SO₄, filtered, and evaporated. The crude material was then purified by column chromatography on

silica gel using a gradient of $5 \rightarrow 10\%$ EtOAc-CH₂Cl₂ as the eluent to afford the pure product 35 as a syrup (129.7 mg, 0.289 mmol, 73% yield). $R_{\rm f}$ 0.46 (EtOAc : hexanes 1:1). $[\alpha]_{\rm D}^{20}$: -62.1° (c 1.01, CHCl₃). ¹H NMR (CDCl₃, 400 MHz): $\delta_{\rm H}$ 7.39-7.25 (m, 5H, Ar), 5.87 (dddd, 1H, J = 21.7, 15.9, 10.5, 5.5 Hz, $-OCH_2-CH=CH_2$), 5.28 (ddd, 1H, J = 17.1, 3.2, 1.6 Hz, $-OCH_2-CH=CH_2$), 5.20 (ddd, 1H, J = 10.3, 2.7, 1.5 Hz, -OCH₂-CH=CH₂), 4.64 (d, 1H, J = 1.1 Hz, H-1), 4.55 (d, 1H, J = 12.1 Hz, PhCH₂), 4.49 (d, 1H, J = 12.1 Hz, PhCH₂), 4.08 (dddd, 1H, J = 15.5, 5.6, 1.5, 1.5 Hz -OCH₂-CH=CH₂), 4.04 (dddd, 1H, J = 15.7, 5.5, 1.4, 1.4 Hz -OCH₂-CH=CH₂), 3.98 (dd, 1H, J = 8.5, 5.1 Hz, H-5), 3.85 (dd, 1H, J = 12.9, 6.6 Hz, OCH_aH_b), 3.82 (dd, 1H, J = 2.1, 1.7 Hz, H-3), 3.77 (broad, 1H, H-2), 3.67-3.60 (m, 2H, H-7a and H-7b), 3.51-3.43 (m, 3H, H-4, $OCH_{a}H_{b}$, OH-4), 3.26 (t, 2H, J = 6.9 Hz, $-CH_{2}N_{3}$), 2.77 (d, 1H, J = 2.0 Hz, OH-2), 2.09 (dddd, 1H, J = 18.0, 13.9, 9.5, 5.5 Hz, H-6a), 1.94 (dddd, 1H, J = 18.5, 12.9, 9.3, 5.5 Hz, H-6b), 1.66-1.56 (m, 4H, 2 × CH₂), 1.38-1.32 (m, 4H, 2 × CH₂). ¹³C NMR (CDCl₃, 100 MHz): $\delta_{\rm C}$ 134.2 (-OCH₂-CH=CH₂), 128.4 (Ar), 127.6 (Ar), 127.5 (Ar), 117.4 (-OCH₂-CH=CH₂), 98.4 (C-1), 76.1 (C-3), 73.0 (PhCH₂), 71.2 (-OCH₂-CH=CH₂), 71.1 (C-5), 69.1 (CH₂), 68.9 (C-2), 68.0 (C-4), 66.3 (C-7), 51.3 (CH₂), 31.0 (C-6), 29.4 (CH₂), 28.7 (CH₂), 26.4 (CH₂), 25.6 (CH₂). HRMS (ESI-QTOF) m/z: $[M + Na]^+$ calcd for C₂₃H₃₅N₃O₆Na 472.2418, found 472.2400.

6-Aminohexyl 6-deoxy-β-D-ido-heptopyranoside acetic acid salt (2): To the solution of the compound **34** (11 mg, 22 μmol) in methanol/CH₂Cl₂ (2 mL/0.5 mL) was added palladium hydroxide (20% Pd(OH)₂ on charcoal; 15 mg), acetic acid (2 drops) and water (5 drops). The solution was stirred under positive pressure hydrogen gas for 2 days. The mixture was filtered and evaporated to afford the product **2** as a colorless syrup (6.2 mg, 21 μmol, 96% yield). $R_{\rm f}$ 0.21 (MeOH : CH₂Cl₂ 3:7). HRMS (ESI-QTOF, **2**) m/z: [M + H]⁺ calcd for C₁₃H₂₈NO₆

294.1911, found 294.1908. The ¹H NMR experiment showed the presence of partially Nmethylated side products. HRMS (ESI-QTOF, mono-N-methylated) m/z: $[M + H]^+$ calcd for $C_{14}H_{30}NO_6$ 308.2073, found 308.1995. HRMS (ESI-QTOF, N,N-dimethylated) m/z: $[M + H]^+$ calcd for C₁₅H₃₂NO₆ 322.2230, found 322.2155. Birch reduction: Liquid NH₃ (~20 mL) was collected in a flask with the help of a dry-ice condenser to -78 °C. Sodium metal (~44 mg) was added, and the solution turn dark blue. A solution of the compound 34 (16 mg, 32 µmol) in THF (1 mL) was added dropwise, and the solution was stirred at -78 °C for 1 h. MeOH (0.5 mL) was added to quench the reaction, and the solution was allowed to warm up to ambient temperature. After the evaporation of ammonia, the mixture was redissolved in MeOH (~5 mL) and neutralized with AcOH. The solution was evaporated to dryness and the residue was subjected to a gel filtration on Sephadex LH-20 using MeOH as the eluent to afford the pure product 2 which was further purified by reverse-phase chromatography on Sep-Pak (C18) using $0 \rightarrow 20\%$ MeOH- H_2O (containing 0.2% AcOH) as the eluent. Compound 2 was obtained as syrup (8.9 mg, 31 μ mol, 95% yield). $[\alpha]_D^{20}$: -9.7° (c 0.37, MeOH). ¹H NMR (D₂O, 400 MHz): δ_H 4.72 (s, 1H, H-1), 3.97 (t, 1H, J = 3.6 Hz, H-3), 3.94 (dd, 1H, J = 3.9, 1.8 Hz, H-4), 3.79 (ddd, 1H, J = 9.9, 6.8, 6.8 Hz, OCH_aH_b), 3.69-3.64 (m, 2H, H-7a and H-7b), 3.61 (d, J = 2.6 Hz, H-2), 3.60-3.54 (m, 1H, $OCH_{a}H_{b}$, 3.43 (dd, 1H, J = 1.7, 1.7 Hz, H-5), 2.88 (t, 2H, J = 7.4 Hz, $-CH_{2}N_{3}$), 1.91 (ddd, 1H, J = 14.9, 10.3, 5.3, 5.3 Hz, H-6a), 1.70 (dddd, 1H, J = 14.8, 7.5, 7.5, 3.9 Hz, H-6b), 1.60-1.47 (m, 4H, 2 × CH₂), 1.36-1.25 (m, 4H, 2 × CH₂). ¹³C NMR (CDCl₃, 100 MHz): δ_{C} 98.9 (C-1), 71.1 (C-5), 69.8 (C-3, C-4), 69.7 (OCH_aH_b), 69.4 (C-2), 58.1 (C-7), 39.4 (CH₂NH₂), 32.4 (C-6), 28.5 (CH₂), 26.7 (CH₂), 25.3 (CH₂), 24.6 (CH₂). HRMS (ESI-QTOF) m/z: $[M + H]^+$ calcd for C₁₃H₂₈NO₆ 294.1911, found 294.1908.

Supporting Information

¹H, ¹³C, ¹H-¹H gCOSY and ¹H-¹³C gHSQC NMR spectra are provided for all synthesized compounds.

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