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Chirality Transfer from Chiral Monoamines to an *m*-Phthalic Diamide-Linked Zinc Bisporphyrinate with a Benzylamide Substituent

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S Supporting Information

ABSTRACT: An *m*-phthalic diamide-linked bisporphyrin with a benzylamide substituent has been designed and synthesized. It has two types of carbonyl groups. In the solution of this zinc bisporphyrinate, these carbonyl groups are involved in the formation of two different Zn– O coordination interactions: one is formed between neighboring zinc bisporphyrinates; another is formed within zinc bisporphyrinate. The chirality sensing abilities of this zinc porphyrinate to a number of chiral monoamines have been examined. When zinc bisporphyrinate was mixed with a series of chiral monoamines, the signs of the circular dichroism spectra for the chiral monoamines of the same handedness with an aryl group as the substituent are just opposite to those with an alkyl group as the substituent. NMR studies reveal that stepwise coordinations lead to 1:1 and 1:2 host–guest complexes. The structure of the 1:1 host–guest complex was confirmed by crystallography, it is the first time that a 1:1



host-guest complex formed between zinc bisporphyrinate and a chiral monoamine has been crystallographically characterized. The structure reveals that there is an intramolecular hydrogen bond between the amide oxygen and the coordinated NH₂. We further investigated the chirality transfer mechanism by density functional theory calculations. Our studies suggest that the interactions between the linker and guests in this bisporphyrin system are crucial in the chirality transfer process, and the nature of the bulkiest substituent of chiral monoamines makes a difference. For R-type guests, with an alkyl group, the steric repulsion makes the conformer A more energetically favorable, which leads to the anticlockwise twist and negative Cotton effect. However, with an aryl group, the π - π interaction makes the conformer B more energetically favorable, which leads to the clockwise twist and positive Cotton effect.

INTRODUCTION

Porphyrins have been widely used in chirality recognition, induction, and transfer in recent years.¹⁻⁵ Because porphyrins have distinct spectroscopic properties, such as a red-shifted and intense UV-vis absorption, circular dichroism (CD) spectroscopy provides a convenient method for studying porphyrin systems. Bisporphyrins have an advantage in these studies especially for chiral guests without chromophores, such as diamines, diols, and amino alcohols, because they can provide two strong chromophores. However, for monodentate guests, such as monoamines, studies are usually more difficult because there is only one coordination site. Only a limited number of porphyrin systems have been reported for chiral monoamines.⁶⁻¹² Some studies reported that these chiral guests need to be converted to their chemical derivatives before measurement.⁷⁻⁹ However, a few systems can have chiralitysensing ability for monoamines without chemical derivations. ${}^{6,10-12}_{6,10-12}$

In order to have effective chirality sensing, host-guest interactions generally require at least two-point fixation. Diamines or diols could provide two coordination sites to satisfy this requirement in bisporphyrin systems. However, for monoamines, it is usually more difficult because the amine group can only provide one binding site. So, (an)other interaction(s) is (are) required. For example, in the ethanebridged bisporphyrin system developed by Borovkov and coworkers, besides the coordination interaction, the short linkage causes steric interactions between the ethyl groups of the porphyrin and the substituents of the ligand, which result in the induced supramolecular chirality.^{6,10,11} For the biphenolbridged metal-free bisporphyrin system developed by Borhan and co-workers, there are both hydrogen-bonding and steric interactions between the monoamines and host molecule, which leads to stereodifferentiation.¹²

It is still a great challenge to design bisporphyrin systems suitable for chiral monoamines that can lead to two-point or multipoint interactions between the hosts and guests. If the linker in the bisporphyrin is close to the coordination site, there will also be interactions between the linker and guest when the

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Scheme 1. Synthetic Route for Compound III and Its Zinc Complex



guest is coordinated to the metalloporphyrin. That could provide two-point or multipoint interactions. Then the chirality could be transferred from the guest to the corresponding host– guest complexes.

We have been working on this in recent years and have recently developed several amide-linked zinc bisporphyrins and trisporphyrinate.¹³⁻¹⁷ These have all shown chirality transfer ability from chiral monoamines to their host-guest complexes. In the case of zinc trisporphyrinate, we obtained the crystal structure of its 1:3 host-guest complex, and the crystal structure reveals that the porphyrin subunits adopt a trans configuration. However, it is difficult to explain the corresponding chirality transfer mechanism for such a multichromophore system in low symmetry. On the other hand, for the zinc bisporphyrinate system having only two porphyrin subunits, the corresponding chirality transfer mechanism could be relatively easy to investigate. However, our previous studies did not get any crystal structures of their host-guest complexes for the bisporphyrin system, so we did not obtain an accurate host-guest binding mode. Herein, in order to obtain more insight on the chirality transfer mechanism for these systems, we have designed and synthesized the bisporphyrin III, as shown in Scheme 1. The bisporphyrin III is derived from *m*phthalic diamide-linked bisporphyrin, and it has both features of previously studied amide-linked bisporphyrin and trisporphyrin: two porphyrin subunits and three amide groups. These amide groups can be classified as types A and B, which could have different roles in the chirality transfer process. We have studied their chiral-sensing abilities to five chiral monoamines (3-methyl-2-butylamine, the guest 1; 3,3-dimethyl-2-butylamine, the guest 2; 1-cyclohexylethylamine, the guest 3; 1phenylethylamine, the guest 4; 1-(2-naphthyl)ethylamine, the guest 5). We obtained the crystal structure of the host-guest crystal structure of $[Zn_2-III] \cdot 1R$. Further investigation on the chirality transfer mechanism was made by UV-vis and NMR spectroscopy and density functional theory (DFT) calculations.

EXPERIMENTAL SECTION

Material and Physical Methods. All reagents were obtained from commercial sources without further purification unless otherwise noted. Triethylamine (Et₃N) was distilled over potassium hydroxide, and methylene chloride was treated with CaH₂ before use. Zinc 5-(2aminophenyl)-10,15,20-triphenylporphyrinate was synthesized according to reported methods.¹⁴ Elemental analyses (carbon, hydrogen, and nitrogen) were performed with an Elementar Vario EL III analytical instrument. ¹H NMR spectra were recorded at room temperature, using an Agilent 400 MHz spectrometer in CDCl₃ with tetramethylsilane (TMS) as the internal standard; chemical shifts are expressed in ppm relative to TMS (0 ppm). Two-dimensional NMR spectra were recorded on a Bruker AVANCE 600 MHz NMR instrument. UV-vis spectra were recorded with a Shimadzu UV-3150 spectrometer. CD spectra were recorded on a PMT model J-815 spectropolarimeter at 298 K. Scanning conditions were as follows: date pitch = 0.2 nm; bandwidth = 2 nm; response time = 1 s; scanning speed = 50 nm/min.

CD and UV–Vis Measurements. UV–vis titration experiments were carried out as follows. Measurements were performed by adding different aliquots of the optically active monoamine solution to the host solution in methylene chloride at room temperature. Then UV– vis spectra were recorded after each addition.

For CD measurements, the background spectrum was taken from 380 to 460 nm with a scan rate of 100 nm/min at 25 $^{\circ}$ C. A zinc bisporphyrinate solution was injected into a 1.0 cm quartz cuvette. Then guests were added to the above solution to form the corresponding complexes. CD spectra were measured after several minutes. The resultant CD spectra recorded in millidegrees were normalized on the basis of the concentrations of zinc bisporphyrinates.

A solid CD spectrum was measured with the following method: KBr pellets were prepared by grinding a single crystal sample with solid KBr and applying great pressure to the dry mixture. The thickness of the KBr pellet was about 0.3 mm. The CD spectrum was then recorded.

¹H NMR Titrations. For ¹H NMR titration experiments, portions of a solution of chiral monoamine in $CDCl_3$ were added to the solution of $[Zn_2$ -III] in $CDCl_3$ in a 5-mm-o.d. NMR tube, and NMR spectra were recorded after each addition.

Preparation of the Free Base Bisporphyrin I. A solution of 5nitroisophthalic acid (0.069 g, 0.36 mmol) in thionyl chloride (8 mL)

was refluxed under nitrogen for 4 h, followed by the removal of excess reagent under reduced pressure. The residual white solid was redissolved in CH₂Cl₂ (30 mL), and Et₃N (150 µL, 1.1 mmol) was added to the solution described above and stirred for 10 min in an ice bath. Zinc 5-(2-aminophenyl)-10,15,20-triphenylporphyrinate (0.50 g, 0.78 mmol) was added to the solution under a nitrogen atmosphere overnight in an ice bath. Then it was washed with water, and the organic layer was collected and evaporated to dryness under vacuum. A purple solid was obtained and purified by silica gel chromatography (pure CH₂Cl₂; 0.27 g, 52% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.82 (d, J = 8.9 Hz, 8H), 8.71 (d, J = 4.4 Hz, 4H), 8.53 (d, J = 4.5 Hz, 4H), 8.18 (d, J = 7.4 Hz, 6H), 8.10 (dd, J = 16.4, 7.0 Hz, 8H), 7.96 (d, J = 7.4 Hz, 2H), 7.83–7.76 (m, 6H), 7.71 (dd, J = 12.7, 6.9 Hz, 10H), 7.61 (t, J = 6.9 Hz, 4H), 7.49 (t, J = 7.5 Hz, 2H), 7.30 (s, 2H), 6.62 (s, 2H), 6.58 (s, 1H), -2.91 (s, 4H). ¹³C NMR (151 MHz, DMSO): δ 163.71, 145.79, 141.55, 138.32, 136.42, 136.32, 136.28, 134.66, 134.50, 134.37, 132.18, 131.64, 129.37, 128.46, 128.33, 127.39, 127.28, 127.21, 126.43, 125.43, 123.28, 120.64, 120.13, 116.10, 40.39, 40.25, 40.11, 39.97, 39.84, 39.70, 39.56, 31.36, 22.47, 14.37. UV-vis [CH₂Cl₂; λ_{max}] nm $(\log \epsilon, \text{ cm}^{-1} \text{ M}^{-1})$]: 418 (5.86), 517 (4.59), 553 (4.07), 594 (4.16). Anal. Calcd for C₉₆H₆₃N₁₀O₄: C, 81.17; H, 4.47; N, 9.86. Found: C, 81.20; H, 4.50; N, 9.88.

Preparation of the Free Base Bisporphyrin II. The free base bisporphyrin I (0.50 g, 0.35 mmol) was dissolved in 20 mL of concentrated HCl, and 2.3 g of SnCl₂·2H₂O was added. The brightgreen solution was stirred at room temperature for 45 min and then heated at 65 °C for 30 min. After the solution was cooled in an ice bath, concentrated ammonia was added to bring the suspension to pH 10. The brown-violet mixture was stirred for 1 h with 50 mL of CHC1₃. The organic layer was separated, washed twice with water, and dried over Na₂SO₄. After filtration, the solvent was removed on a rotary evaporator. The purple solid was obtained and purified by column chromatography (silica, pure CH₂Cl₂; 0.39 g, 80% yield). ¹H NMR (400 MHz, DMSO): δ 8.92 (s, 2H), 8.74 (t, J = 22.5 Hz, 8H), 8.62 (s, 8H), 8.19 (s, 2H), 8.13 (s, 6H), 8.04 (t, J = 9.9 Hz, 4H), 7.98 $(d, I = 6.9 \text{ Hz}, 2\text{H}), 7.82 (d, I = 6.2 \text{ Hz}, 6\text{H}), 7.77 (s, 8\text{H}), 7.66 (s, 8\text$ 6H), 7.61–7.55 (m, 2H), 7.52 (d, J = 7.7 Hz, 2H), 5.66 (s, 1H), 5.54 (s, 2H), 3.90 (s, 2H), -3.02 (s, 4H). ¹³C NMR (151 MHz, DMSO): δ 166.04, 147.23, 141.63, 141.58, 138.77, 136.27, 135.77, 135.36, 134.56, 131.56, 129.18, 128.50, 128.37, 127.43, 127.33, 126.00, 124.63, 120.62, 120.10, 116.27, 114.39, 113.06, 40.37, 40.23, 40.09, 39.96, 39.82, 39.68, 39.54. UV-vis [CH₂Cl₂; λ_{max} nm (log ε , cm⁻¹ M⁻¹)]: 417 (5.77), 517 (4.05), 551 (3.82), 529 (3.96). Anal. Calcd for C₉₆H₆₅N₁₀O₂: C, 82.92; H, 4.71; N, 10.07. Found: C, 82.95; H, 4.74; N, 10.09.

Preparation of the Free Base Bisporphyrin III. The reaction was performed under anaerobic conditions. The free base bisporphyrin II (0.50 g, 0.36 mmol) was dissolved in anhydrous methylene chloride (50 mL). Et₃N (150 μ L, 1.07 mmol) was added to the solution described above, the mixture was stirred for 15 min in an ice bath, and then benzoyl chloride (46 μ L, 0.40 mmol) was added. The mixture was slowly warmed to room temperature, and the reaction was monitored by thin-layer chromatography (TLC). After 8 h, the reaction was complete. The solution was rotor-evaporated to dryness under vacuum. The purple solid was obtained and purified by column chromatography (silica, pure CH₂Cl₂; 0.26 g, 48% yield). ⁱH NMR (400 MHz, CDCl₃): δ 8.88–8.79 (m, 4H), 8.76 (d, J = 4.0 Hz, 4H), 8.69 (d, J = 4.0 Hz, 4H), 8.55 (d, J = 3.7 Hz, 4H), 8.25-8.15 (m, 6H), 8.11 (s, 4H), 7.99 (d, J = 6.7 Hz, 4H), 7.88 (d, J = 7.4 Hz, 2H), 7.83-7.73 (m, 6H), 7.69 (s, 10H), 7.54 (s, 4H), 7.44 (t, J = 7.4 Hz, 2H), 7.39 (s, 2H), 7.06 (t, J = 7.3 Hz, 1H), 6.54 (t, J = 7.5 Hz, 2H), 6.24 (s, 1H), 6.16 (s, 2H), 5.78 (s, 1H), 5.67 (d, J = 7.6 Hz, 2H), -2.86 (s, 4H). ¹³C NMR (151 MHz, DMSO): δ 163.54, 163.34, 142.13, 141.75, 138.03, 137.43, 135.09, 135.03, 134.72, 134.59, 134.51, 132.63, 131.95, 130.97, 129.38, 127.89, 127.72, 127.70, 126.83, 126.76, 126.60, 125.71, 123.07, 120.97, 120.93, 120.69, 120.57, 119.45, 112.64, 77.28, 77.07, 76.86. UV–vis $[CH_2Cl_2; \lambda_{max}, nm (\log \epsilon, cm^{-1} M^{-1})]: 417 (5.53), 515$ (4.10), 552 (3.16), 590 (2.54). Anal. Calcd for C103H69N10O3: C, 82.77; H, 4.65; N, 9.37. Found: C, 82.81; H, 4.68; N, 9.32.

Preparation of [Zn₂-III]. The free base bisporphyrin III (0.50 g, 0.33 mmol) was dissolved in a mixture of CHCl₃ (150 mL) and CH₂OH (50 mL). Zn(CH₃COO)₂ (0.24 g 1.32 mmol) was added to the solution described above and refluxed for 2 h. Then it was extracted with water, and the organic layer was collected and evaporated to dryness under vacuum. The purple solid was obtained and purified by silica gel chromatography (99:1 CH₂Cl₂/methanol; 0.51 g, 94% yield). ¹H NMR (400 MHz, CDCl₃): δ 8.85 (s, 8H), 8.61 (d, J = 4.3 Hz, 4H), 8.37 (d, J = 4.3 Hz, 4H), 8.19 (s, 6H), 8.04 (d, J = 6.9 Hz, 2H), 7.91 (d, J = 7.3 Hz, 2H), 7.85–7.72 (m, 8H), 7.66 (d, J = 16.4 Hz, 10H), 7.49 (t, J = 7.5 Hz, 8H), 7.09 (s, 3H), 6.82 (s, 2H), 6.56 (t, J = 7.4 Hz, 2H), 5.79 (s, 1H), 5.06 (s, 2H), 4.18 (d, J = 7.5 Hz, 2H), 3.78 (s, 1H). ¹³C NMR (151 MHz, DMSO): δ 161.38, 150.18, 149.91, 149.13, 146.06, 146.04, 145.42, 142.92, 137.62, 135.43, 134.42, 134.01, 133.46, 132.58, 132.01, 131.89, 130.83, 130.48, 130.04, 128.94, 127.45, 127.17, 126.34, 126.22, 123.91, 123.49, 121.48, 120.90, 120.24, 119.26, 118.11, 112.14, 96.90, 93.15, 77.19, 76.98, 76.77, 31.57, 29.67, 14.10, 1.01. UV-vis [CH₂Cl₂; λ_{max} nm (log ε , cm⁻¹ M⁻¹)]: 422 (5.60), 556 (4.21), 597 (3.04). Anal. Calcd for C₁₀₃H₆₅N₁₀O₃Zn₂: C, 76.30; H, 4.04; N, 8.64. Found: C, 76.35; H, 3.99; N, 8.69.

Preparation of Crystals of [Zn₂-III]-**1R.** [Zn₂-**III**] (50 mg, 0.030 mmol) was dissolved in CH₂Cl₂ (1 mL), 200 μ L of a 0.2 M solution of (*R*)-2-amino-3-methylbutane in anhydrous CH₂Cl₂ was added to the solution described above, and the mixture was stirred for ~3 min. Then it was transferred to 8 mm × 250 mm glass tubes. *n*-Hexane was added as a nonsolvent at room temperature. After 3 months, purple crystals were obtained, which were then isolated by filtration, washed with *n*-hexane, and dried under vacuum. (11 mg, 21% yield). Anal. Calcd for C₁₀₈H₇₈N₁₁O₃Zn₂: C, 75.92; H, 4.60; N, 9.02; Found: C, 75.82; H, 4.65; N, 9.05.

X-ray Structure Determination. The measurements of single crystals were performed on a Bruker APEX-II CCD X-ray diffractometer by using graphite-monochromated Mo K α (λ = 0.71073 nm). The structure was determined by direct methods and refined on F^2 using the full-matrix least-squares method with *SHELXTL*, version 2014.¹⁸ All non-hydrogen atoms were refined anisotropically; all hydrogen atoms were theoretically added and rode on their parent atoms. In the structure, the asymmetric unit contains badly disordered solvent molecules. *SQUEEZE*¹⁹ was used to model all disordered solvates. The electron count within the interporphyrin voids was 825e for [Zn₂-III]-1R (corresponding roughly to five molecules of methylene chloride per [Zn₂-III]). Details of the crystal parameters, data collection, and refinements are summarized in Table 1. Complete crystallographic details, atomic coordinates, anisotropic thermal parameters, and fixed hydrogen-atom coordinates are given in the CIF file (CCDC 1531702).

Computational Methods. All calculations are on the basis of the crystal structural data. To simplify the calculations, we only consider a single bisporphyrin molecule, and we did not calculate the aggregated species. We first performed DFT calculations on the free host [Zn₂-III]. Then we did calculations on the corresponding 1:1 ([Zn₂-III]·IR and [Zn₂-III]·4R) and 1:2 ([Zn₂-III]·(1R)₂ and [Zn₂-III]·(4R)₂) complexes. Full optimizations on these structures were performed by DFT at the level of B3LYP/6-31G* using the *Gaussian09* suite of programs.²⁰ We employed DFT with no symmetry constraints to investigate the optimized geometries.

RESULTS AND DISCUSSION

CD Spectral Studies. In this study, five types of chiral monoamines were used as guests, and they have different substituents X. Each host and each enantiopure amine were mixed in methylene chloride, and the corresponding CD spectra were recorded. Because the formation constants for binding monoamines to zinc porphyrinates are in general around 10^3 L/mol²¹ and the concentrations of porphyrin used in our CD measurements are around 10^{-6} mol/L, a large excess of guests are required to completely convert the host to the corresponding host–guest complex. In our experiments, more

Table 1. Crystal Data and Structural Refinement Data of $[Zn_2-III]$ ·1R

crystal	$[Zn_2-III]\cdot 1R$		
chemical formula	C ₁₀₈ H ₇₈ N ₁₂ O ₃ Zn ₂		
fw	1722.56		
wavelength (Å)	0.71073		
temperature (K)	223(2)		
cryst syst	monoclinic		
space group	C2		
a (Å)	36.298(3)		
b (Å)	15.1343(11)		
c (Å)	26.262(2)		
α (deg)	90		
β (deg)	133.414(4)		
γ (deg)	90		
V (Å ³)	10479.7(15)		
Ζ	4		
density (Mg/m ³)	1.092		
abs coeff (mm ⁻¹)	0.509		
<i>F</i> (000)	3576.0		
data collection θ range (deg)	2.41-27.51		
index ranges	$-47 \le h \le 47, -19 \le k \le 19, -34 \le l \le 34$		
reflns collected	301609		
R _{int}	0.0622		
indep reflns	24236		
data/restraints/param	24236/92/1181		
GOF on F ²	1.024		
$\mathrm{R1}^{a} \left[I > 2\sigma(I) \right]$	0.0346		
wR2	0.0871		
peak/hole (e/ų)	0.491/-0.462		
Flack parameter	0.032(6)		
^a R1 = $(F_o - F_c)/F_{o}$; wR2 = $w(F_o^2 - F_c^2)^2/w(F_0^2)^2$] ^{1/2} .			

than 1000 equiv of guests was generally used. Their titration CD spectra and the corresponding plots of the CD intensity as a function of the equivalents of guests are provided in the Supporting Information. These figures show that the CD intensities reached a maximum at a large excess of guests, which suggested that all zinc bisporphyrinates were converted to the corresponding host-guest complexes. The spectra in Figure 1 show clear signals with typical bisignate Cotton effects in the Soret band region. The CD spectral data are listed in Table 2. These results reveal the following features: (1) For a pair of enantiomers of chiral amines, the CD spectra showed similar shapes and intensities but opposite signs. (2) More importantly, when the substituent X is an aryl group or an alkyl group, the signs of the resulting CD are different. For the R-type chiral monoamines, when X is an alkyl group, such as 1R, 2R, and 3R, the longer-wavelength peak of the Soret band is negative and the shorter-wavelength peak is positive. However, when X is an aryl group, such as 4R and 5R, the signs of CD are just opposite to those with X as an alkyl group. As follows, we have investigated the chirality transfer mechanism through UV-vis spectra, NMR studies, the crystal structure, and DFT calculations.

UV-vis Spectral Studies and Self-Aggregation Process for the Host. UV-vis spectra of $[Zn_2-III]$ were measured at different concentrations, as shown in Figure 2. The spectra of $[Zn_2-III]$ show one strong Soret band at 421 nm and one shoulder at 431 nm, which are similar to those reported for *m*phthalic diamide-linked zinc bisporphyrinates¹⁴ but different from those for the oxalic amide-linked species.¹³ For comparison, the free base bisporphyrin III was also measured by UV–vis spectroscopy. The spectrum is provided in Figure S11, which does not show such a shoulder. This suggests that zinc is involved in the formation of such a shoulder. Generally, the coordination of zinc porphyrin to oxygen or nitrogen leads to bathochromic shifts compared to four-coordinate zinc porphyrins.²² In our case, such a shoulder at longer wavelength could be caused by the Zn–O coordination interactions.

As shown in Figure 2, when the concentration of $[Zn_2-III]$ increased, the intensity of the shoulder increased, and the intensity of the peak at 421 nm deceased. Such a concentration-dependent spectral change suggests that the Zn–O bond is formed between neighboring zinc bisporphyrinates, and aggregation through the coordination interactions occurs in solution. Such a coordination mode was actually found in our previous studies.^{14,16} So, the following equilibrium (1) exists in solution. $[Zn_2-III]_n$ is the aggregated species, and $[Zn_2-III]$ is the single zinc bisporphyrinate molecule.

$$[\operatorname{Zn}_2\operatorname{-III}]_n \rightleftharpoons n[\operatorname{Zn}_2\operatorname{-III}]$$
(1)

UV–vis spectroscopic titrations also have been performed between $[Zn_2$ -III] and five chiral monoamines. A representative example of the spectral change of $[Zn_2$ -III] induced by the addition of 1R is shown in Figure 3. (Others are provided in Figures S12–S15.) Before the addition, the spectrum of $[Zn_2$ -III] shows one strong Soret band at 421 nm and one shoulder at 431 nm. During the titration, the electronic spectra showed that the intensity of the band at 421 nm decreased and the intensity at 431 nm increased with increasing the ligand concentrations.

When guests are mixed with zinc bisporphyrinate, the following two equilibria could be in the solution because there are two zinc binding sites:

$$[\operatorname{Zn}_2\operatorname{-III}] + L \stackrel{K_1}{\rightleftharpoons} [\operatorname{Zn}_2\operatorname{-III}] \cdot L$$
(2)

$$[Zn_2-III] \cdot L + L \stackrel{K_2}{\rightleftharpoons} [Zn_2-III] \cdot (L)_2$$
(3)

where L represents a guest.

However, because of the self-aggregation process [equilibrium (1)] in solution, the equilibria in solution became more complicated. The binding constants for the above two equilibria are difficult to determine. In fact, in the titration spectra, we did not observe clear isosbestic points. This is possibly due to the self-aggregation process. Self-aggregation through Zn–O bonds is further confirmed by NMR studies.

NMR Studies. For the free base bisporphyrin III and its complex [Zn₂-III], the ¹H NMR spectra are presented in Figure 4. In this bisporphyrin, all protons are aromatic protons except the NH protons. The resonances for the aromatic protons are generally located in the 7-9 ppm region.¹⁶ However, for the bisporphyrin III and its complex $[Zn_2-III]$, some resonances are located in the 3-7 ppm region. Because the linker is between two porphyrin subunits, the ring current effect of porphyrin could cause such upfield shifts. So, these resonances are assigned to the protons of the linker. Detailed assignments are based on ¹H and ¹H-¹H COSY NMR, which are presented in the Supporting Information. Compared with the free base bisporphyrin III, most signals for the zinc bisporphyrinate [Zn2-III] remain unchanged, while the resonances of H12, H(14,16), and H(22,26) shift remarkably upfield to 3.80, 5.08, and 4.28 ppm, respectively. Such upfield shifts suggest that these protons are even much closer to the



Figure 1. CD spectra of a solution of $[Zn_2-III]$ (1.75 × 10⁻⁶ mol/L) and a large excess of guests in methylene chloride at 298 K. (A) 2500 equiv of 1S, (dashed line) or 1R (solid line); (B) 1600 equiv of 2S (dashed line) or 2R (solid line); (C) 1535 equiv of 3S (dashed line) or 3R (solid line); (D) 1340 equiv of 4S (dashed line) or 4R (solid line); (E) 1550 equiv of 5S (dashed line) or 5R (solid line).

porphyrin plane in zinc bisporphyrinate than in free base bisporphyrin, which could be due to coordination of the carbonyl oxygen atom to the neighboring zinc porphyrinate. In addition, ¹H NMR spectra at different concentrations have also been measured. In these spectra, most resonances do not change, but the resonance of H12 shows slight shifts.

On the basis of these NMR spectral changes, we proposed one possible mode for the self-aggregation of $[Zn_2-III]$, as shown in Figure 5. In the structural formula, there are two types of carbonyl groups. In solution, both carbonyl oxygen atoms are coordinated to zinc: the A-type carbonyl group involves "intermolecular" interactions; the B-type carbonyl group involves "intramolecular" Zn–O coordination interactions. Such coordination interactions cause H12, H(14,16), and H(22,26) to locate above the adjacent porphyrin plane, and the ring current effect causes their resonances to shift upfield compared with the free base bisporphyrin.

As is also shown in Figure 5, when the B-type carbonyl oxygen atom is coordinated to zinc, coordination interactions lead to the adjacent phenyl ring above the plane of the subunit P1. H22 and H26 are closer to the shielding cone of the porphyrin ring than H23 and H25, which is similar to that in the crystal structure of $[Zn_2-III] \cdot 1R$ (vide infra). As a result, the resonances of H22 and H26 shift more upfield than those of H23 and H25.

On the other hand, "intermolecular" interactions are concentration-dependent, while "intramolecular" interactions are not. When we change the concentrations, the resonances of these protons behave differently. The proton H12 is close to the A-type carbonyl group, and "intermolecular" interactions lead to its shift upon changes in the concentration. On the

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Table 2. Observed CD Spectral Data of the Complexes of $[Zn_2-III]$ (1.75 × 10⁻⁶ M) with Chiral Monoamines^{*a*} in Methylene Chloride at 298 K

	$\lambda_{ m max}$ nm ($\Delta \varepsilon$, cm ⁻¹ M ⁻¹)		
	first cotton	second cotton	$A_{obs}^{\ b}$
1R	432 (-142)	423 (+70)	-212
15	431 (+143)	421 (-68)	+211
2R	431 (-234)	424 (+160)	-394
28	431 (+248)	423 (-150)	+398
3R	433 (-50)	425 (+68)	-118
38	433 (+53)	424 (-70)	+123
4R	431 (+197)	419 (-160)	+357
4 S	431 (-193)	419 (+163)	-356
5R	430 (+498)	422 (-452)	+950
55	429 (-483)	420 (+443)	-926

^{*a*}The corresponding host–guest ratios are the same as those in Figure 1. ^{*b*}A_{obs} = $\Delta \varepsilon_1 - \Delta \varepsilon_2$. This value represents the total amplitude of the experimentally observed CD couplets.



Figure 2. UV–vis spectra of $[Zn_2$ -**III**] in 0.1 mm, 1 mm, and 1.0 cm quartz cells at different concentrations in methylene chloride. The intensities are normalized on the basis of the concentrations of $[Zn_2$ -**III**]: (A) 1.5×10^{-7} mol/L; (B) 6.0×10^{-7} mol/L; (C) 1.5×10^{-6} mol/L; (D) 1.5×10^{-5} mol/L; (E) 1.5×10^{-4} mol/L.



Figure 3. UV-vis spectral change of $[Zn_2-III]$ (1.50×10^{-6} M) upon the addition of 1R as the host-guest molar ratios are from 1:0 to 1:850.

other hand, the protons close to the B-type carbonyl group, such as H24, H(22,26), and H(23,25), do not show clear shifts because the B-type carbonyl group involves "intramolecular" Zn–O coordination interactions. "Intermolecular" interactions can result in self-aggregation, as shown in equilibrium (1), which may lead to the formation of a dimer, trimer, or even polymer. It is likely that there are several aggregated species

with different "n" values in solution, and there could also be equilibria between them, which makes the equilibria more complicated in solution.

¹H NMR titration experiments were performed between [Zn₂-III] and 1R. When the guest was added to the [Zn₂-III] solution, there were some resonances located in the -4 to 0 ppm region, as shown in Figure S20, which is similar to our studies on previously reported amide-linked bisporphyrin systems. Obviously, the large upfield shifts of these protons are due to coordination of the guests to zinc porphyrinate. The detailed titration spectra shown in Figure 6 gave us more information on the coordination interactions. During titration, the resonances of the protons of the linker in the host have shown remarkable shifts. When the host-guest ratio is below 1:1, the resonance of H12 has the largest shift; when the ratio is over 1:1, the resonance of H(22,26) has the largest shift. When the guest is in a large excess, the resonance pattern is very similar to that of the free base porphyrin III. This suggests that the NMR spectral changes during titration are caused by replacing the Zn–O bonds with the Zn–N (from amine) bonds. While the structural formula suggests that H12 is close to the A-type carbonyl, H(22,26) is close to the B-type carbonyl. The above studies suggest that, during titration, the A-type carbonyl oxygen atom was first replaced by one monoamine to form the 1:1 complex, and then the B-type carbonyl oxygen atom was replaced by another monoamine to form the 1:2 host-guest complex.

The ¹H NMR titration experiments were also performed between $[Zn_2-III]$ and 4R. Their spectra (Figure S22) show changes similar to those for 1R. This suggests that 4R is also coordinated to $[Zn_2-III]$ in a stepwise fashion, leading to the formation of 1:1 and 1:2 host-guest complexes during titration.

On the basis of the ¹H NMR titration results, Job plots for the titration of **1R** or **4R** into a [Zn₂-**III**] solution are shown in Figures S23 and S24. The plots showed a broad peak around 0.3–0.5 mole fraction but not at 0.33, as was expected for a 1:2 host–guest complex. This is possibly due to the limitation of Job plots. Actually, there were some reports on the limited applicability of this method.^{23–25} Because there are several equilibria and too many species (including the aggregated species) in solution, the Job plot method is not suitable for this system.

X-ray Crystal Structure of $[Zn_2-III]\cdot 1R$. In order to understand the chirality transfer mechanism, we need to obtain the binding mode of the host–guest complexes. One of the best ways to get the binding mode is through the crystal structure. We tried to obtain crystal structures for the host– guest complexes. For one of the alkyl-substituted guests, 1R, we obtained single crystals of $[Zn_2-III]\cdot 1R$.

The structure of $[Zn_2-III] \cdot IR$ was solved in a chiral space group, C2. One asymmetric unit contains one zinc bisporphyrinate molecule. The bisporphyrin host molecule has two zinc porphyrinate subunits that are linked by an *m*phthalic diamide group. Interestingly, both zinc atoms are fivecoordinate, but their axial ligations are different. As shown in Figure 7, for the subunit P1, the axial ligand is the N1 atom of the chiral guest. The ligand adopts an "inside" binding mode (the ligand is facing the linker). For the subunit P2, the axial ligation is from the B-type carbonyl oxygen O013 of the amide group of the linker. Because of coordination, the phenyl rings composed of C21, C22, C23, C24, C25, and C26 are over the plane of the porphyrin subunit P2, and C22/C26 and their



Figure 4. ¹H NMR spectra for compound III (A; 3.3×10^{-3} M) and [Zn₂-III] at different concentrations: (B) 4.6×10^{-3} M; (C) 1.2×10^{-3} M; (D) 0.92×10^{-3} M; (E) 0.45×10^{-3} M. The inset shows the proton numbering scheme of the linker in the host. The arrows show the shifts of the relevant protons.



Figure 5. Proposed binding mode for [Zn₂-**III**]. The A-type carbonyl group involves "intermolecular" interactions, and the B-type carbonyl group involves "intramolecular" Zn–O coordination interactions.

protons H22/H26 are closer to the porphyrin center than C23/C25 and H23/H25 (vide supra). So, this coordination mode is consistent with our NMR studies. Besides the coordination interaction, the guest **1R** also involves hydrogen-bonding interactions. There is an intramolecular hydrogen bond between N1 and the carbonyl oxygen O012. The corresponding distance N1…O012 is 2.919(4) Å, and the bond angle N1–H1B…O012 is 159°. These interactions could be the major factors stabilizing the "inside" binding mode for the 1:1 complex.

The C5–C15–C5'–C15' torsion angle can be used to present the dihedral angle between the two transition moments of the chromophores for the effective exciton coupling interactions in the bisporphyrin system.^{26,27} In this structure, two porphyrin subunits form an anticlockwise twist with the C5–C15–C5'–C15' torsion angle (here it is C1M3–C1M1–C2M1–C2M3) of –147°. On the basis of the exciton chirality

method,²⁸ the anticlockwise twist will lead to a negative Cotton effect. This is consistent with our experimental results.

Solid-State CD Spectrum. For $[Zn_2-III] \cdot 1R$, we also did CD measurements on several single-crystal samples. For these measurements, the signals were all weak. However, they did show CD signals in the Soret band region. One of them is given in Figure 8. The CD spectrum shows the same sign and similar shape as the solution CD spectra. This confirms the chirality of the single crystal.

Computational Studies on the Chirality Transfer Mechanism. In order to gain more insight into the chirality transfer mechanism for this system, we did further investigations by DFT calculations. On the basis of the crystal structure, we did calculations on the host, 1:1 and 1:2 host– guest complexes for the guests **IR** and **4R**. All of the optimized structures are displayed in Figures 9 and 10. We found that, in all of these optimized structures, except the zinc atoms coordinated by amines, other zinc atoms are all coordinated by the amide oxygen atom. Such a coordination mode was actually found for zinc trisporphyrinate as well.²⁹

As shown in Figure 9A, for the free host [Zn₂-III], the optimized structure is in low symmetry, which leads to a conformer with a noncentrosymmetric (chiral) configuration. Such conformers exist as a pair of enantiomers, namely, the conformers A and B. For the conformer A, the two porphyrin subunits adopt an anticlockwise twist; for the conformer B, the two porphyrin subunits adopt a clockwise twist. However, the solution consists of a racemic mixture, which is optically inactive.

When the chiral guest $\mathbf{1R}$ is mixed with the host, both conformers A and B could interact with it and form the corresponding 1:1 complexes, the conformer A·**1R** and the conformer B·**1R**. Their optimized structures are shown in Figure 9B. In both cases, coordination and hydrogen-bonding interactions are maintained like those in the crystal structure. The calculations suggest that the conformer A·**1R** is more energetically favorable, which is 1.96 kJ/mol lower than the conformer B·**1R**.



Figure 6. ¹H NMR spectral changes of $[Zn_2-III]$ (4.4 × 10⁻³ M) upon the addition of 1R in CDCl₃ at 298 K as the host–guest molar ratios are from 1:0 to 1:8.2.



Figure 7. ORTEP view for $[Zn_2-III] \cdot IR$ with 50% probability thermal ellipsoids. A hydrogen bond is shown as a dashed line. The arrows show the direction of the effective transition moments along the C5… C15 axis (here labeled as C1M1…C1M3 and C2M1…C2M3). Some phenyl rings and some hydrogen atoms have been omitted for clarity.

What structural differences cause the difference in energy for these two conformers? For the chiral monoamines used in this work, the chirogenic carbon center is bonded to four different groups: NH₂, CH₃, H, and X. For the case of **1R**, X is an isopropyl group $[CH(CH_3)_2]$, an alkyl substituent. Because of the coordination interaction, the position of NH₂ is fixed. Then the major differences are caused by the orientations of the other three groups. The bulkiness order is $H < CH_3 < CH(CH_3)_2$. When the guest **1R** is coordinated to zinc porphyrinate, two parts of the steric repulsions have to be considered: (1) One is



Figure 8. CD spectrum of the single crystal of $[Zn_2-III] \cdot 1R$ in the KBr pellet.

between the guest and the porphyrin plane. (2) Another is between the guest and the linker. In order to minimize the steric repulsions between the guest and the porphyrin plane, in both conformers $A \cdot IR$ and $B \cdot IR$, the least bulky group (hydrogen) is facing the porphyrin plane and the bulkiest group (isopropyl) is away from the porphyrin plane. However, as shown in Figure 9B, the orientations of the isopropyl groups in two conformers are different. In the conformer $A \cdot IR$, it is away from the linking phenyl. However, in the conformer $B \cdot IR$, it tilts toward the linking phenyl. The former orientation may cause less steric repulsion between the guest and the linker and lower its energy. Therefore, the conformer $A \cdot IR$ should be the major contributor to the CD spectra.

In the conformer A·**1R**, the two porphyrin subunits adopt an anticlockwise twist with a C5-C15-C5'-C15' torsion angle of -144° . On the basis of the exciton chirality method,²⁸ this anticlockwise twist will lead to a negative Cotton effect. This is also consistent with our experimental results.

When the 1:1 complex further reacts with a guest, both conformers $A \cdot (1R)_2$ and $B \cdot (1R)_2$ of the 1:2 complexes will form. As shown in Figure 9C, in the optimized structures, the two guest molecules are not equivalent. One is similar to that in the 1:1 complex; besides the coordination interaction, it also forms a hydrogen bond with the A-type carbonyl oxygen atom. The second replaces the coordination site of the B-type



Figure 9. Chirality transfer mechanism based on DFT calculations for **1R**. Hydrogen atoms of the chiral carbon atoms are marked by red circles; arrows show the orientations of the isopropyl groups. (A) Host [Zn₂-**III**], which exists as a pair of enantiomers in solution. The conformer A forms an anticlockwise twist, and the conformer B forms a clockwise twist. (B) Optimized 1:1 host–guest complexes for [Zn₂-**III**]·**1R**. The conformer A·**1R** is more energetically favorable, which leads to the anticlockwise twist. (C) Optimized 1:2 host–guest complex for [Zn₂-**III**]·(**1R**)₂. The conformer A·(**1R**)₂ is more energetically favorable, which leads to the anticlockwise twist.

carbonyl oxygen atom but does not form any hydrogen bond. The calculations suggest that the conformer $A \cdot (\mathbf{1R})_2$ is more energetically favorable, which is 3.96 kJ/mol lower than the conformer $B \cdot (\mathbf{1R})_2$. Similar to the case of their 1:1 complex, the difference could be also caused by the orientations of the isopropyl groups. Besides the different orientations of the isopropyl group of the first guest coordinated in the subunit P1, the isopropyl group of the second guest coordinated in the subunit P2 also adopts different orientations in both conformers. As shown in Figure 9C, in the conformer $A \cdot (\mathbf{1R})_2$, the isopropyl group is away from the benzylamide substituent. However, in the conformer $B \cdot (\mathbf{1R})_2$, it tilts toward this substituent. The former orientation may cause less steric repulsions and lower its energy. Therefore, the conformer A.

Figure 10. Chirality transfer mechanism based on DFT calculations for **4R**. Hydrogen atoms of the chiral carbon atoms are marked by red circles. (A) Host $[Zn_2-III]$ existing as a pair of enantiomers, the conformers A and B. (B) Optimized 1:1 host–guest complexes for $[Zn_2-III]\cdot 4\mathbf{R}$. The conformer B·4R is more energetically favorable, which leads to the clockwise twist. (C) Optimized 1:2 host–guest complex for $[Zn_2-III]\cdot(4\mathbf{R})_2$. The conformer B· $(4\mathbf{R})_2$ is more energetically favorable, which leads to the clockwise twist.

 $(1R)_2$ should be the major contributor to the CD spectra. In the conformer A· $(1R)_2$, the two porphyrin subunits adopt an anticlockwise twist with a C5–C15–C5′–C15′ torsion angle of –157°. On the basis of the exciton chirality method,²⁸ this anticlockwise twist will lead to a negative Cotton effect. This is also consistent with our experimental results.

Why are the signs of the CD signals different when the substituent is an aryl group compared to an alkyl group? In the calculations, (R)-1-phenylethylamine $(4\mathbf{R})$ was used as the guest with an aryl substituent. When the chiral guest $4\mathbf{R}$ is mixed with the host, this leads to 1:1 and 1:2 complexes. For the 1:1 complexes, the optimized structures are the conformer A·4R and the conformer B·4R, as shown in Figure 10B. In both cases, hydrogen bonds are maintained. However, different from the case of 1R, there are also $\pi - \pi$ interactions between the phenyl ring of the guest and the phenyl ring of the linker. Such types of $\pi - \pi$ interactions were also found in the reported

amide-linked zinc trisporphyrinate complex.¹⁷ The calculations suggest that the conformer $B \cdot 4\mathbf{R}$ is more energetically favorable, which is 6.48 kJ/mol lower than the conformer A·4**R**.

For the chiral guest 4R, X is a phenyl group. Because of the coordination and $\pi - \pi$ interactions, the positions of NH₂ and the phenyl group are fixed, and then the major differences between the conformer A·4R and the conformer B·4R are caused by the orientations of the methyl group and the hydrogen atom. The bulkiness order is H < CH₃. In their optimized structures, as shown in Figure 10B, the methyl group is facing the porphyrin plane in the conformer $A \cdot 4R$, while it is away from the porphyrin plane in the conformer B·4R. Such an orientation may cause less steric repulsion for the conformer B-4R and, hence, lower its energy. Therefore, the conformer $B \cdot 4R$ could be the major contributor for the CD signal. In the conformer $B \cdot 4R$, the two porphyrin subunits adopt a clockwise twist with a C5-C15-C5'-C15' torsion angle of 154° . On the basis of the exciton chirality method,²⁸ the above conformer will lead to a positive Cotton effect.

When both conformers of the 1:1 complex further react with a guest, both conformers $A \cdot (4R)_2$ and $B \cdot (4R)_2$ of the 1:2 complexes will form. As shown in Figure 10C, the two guest molecules in the optimized structures are not equivalent. One is similar to that in the 1:1 complex; besides coordination interaction, it also involves hydrogen-bonding and $\pi - \pi$ interactions. The other one only involves coordination and $\pi - \pi$ interactions but does not form any hydrogen bonds. The calculations suggest that the conformer $B \cdot (4R)_2$ is more energetically favorable, which is about 0.92 kJ/mol lower than the conformer $A \cdot (4\mathbf{R})_2$. Similar to the case of their 1:1 complex, the difference could also be caused by the orientations of the methyl group and hydrogen, as shown in Figure 10C. The methyl group is facing the porphyrin plane in the conformer A. $(4\mathbf{R})_2$, while it is away from the porphyrin plane in the conformer $B(4R)_2$. Such an orientation may cause less steric repulsion for the conformer $B \cdot (4R)_2$ and, hence, lower its energy. In the conformer $B(4R)_2$, the two porphyrin subunits adopt an anticlockwise twist with a C5-C15-C5'-C15' torsion angle of 168°. On the basis of the exciton chirality method,²⁸ this clockwise twist will lead to a positive Cotton effect. This is also consistent with our experimental results.

CONCLUSION

In conclusion, we have designed and synthesized a new mphthalic diamide-linked zinc bisporphyrinate. It can function as a chirality probe for a series of chiral monoamines. For the chiral monoamines with the same handedness, the signs of their CD signals for those with an aryl group as the substituent (X) are opposite to those with an alkyl group as the substituent. NMR studies suggest that stepwise coordination leads to the 1:1 and 1:2 complexes. The crystal structure of a 1:1 hostguest complex reveals that this bisporphyrin adopts a trans configuration and the NH₂ of the amine is involved in both coordination and hydrogen-bonding interactions. DFT calculations suggest that, in the host-guest complexes, the nature of the bulkiest group X makes a difference. When X is an alkyl group, the steric repulsion causes the conformer B-1R to be more energetically favorable, which leads to a clockwise twist; when X is an aryl group, it can form $\pi - \pi$ interactions with the phenyl ring of the linker, which causes the conformer A·4R to be more energetically favorable and leads to an anticlockwise twist. Our studies suggest that, in this bisporphyrin system, the interactions between the linker and guests are crucial, and we

could control chirality transfer by adjusting the linker. Further studies on the functionalization of the linker are under investigation.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.inorg-chem.7b00815.

CD, UV-vis, and NMR spectra, calculation data, relative energies, and Cartesian coordinates (PDF)

Accession Codes

CCDC 1531702 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

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