

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 15 (2005) 149–151

5'-Homoaristeromycin. Synthesis and antiviral activity against orthopox viruses

Minmin Yang and Stewart W. Schneller*

Department of Chemistry and Biochemistry, Auburn University, Auburn, AL 36849, USA

Received 9 September 2004; revised 6 October 2004; accepted 6 October 2004 Available online 28 October 2004

Abstract—An efficient synthesis of 5'-homoaristeromycin has been developed. This permitted an extensive antiviral analysis, which found potent activity toward vaccinia, cowpox, and monkeypox viruses. For comparative purposes, 5'-homoadenosine was made available by a newly designed route and found to be inactive. © 2004 Elsevier Ltd. All rights reserved.

Inhibitors of *S*-adenosyl-L-homocysteine (AdoHcy) hydrolase have shown promise as antiviral agents^{1,2} by disrupting essential viral macromolecular methylation processes.² Carbocyclic nucleosides ³ represent a prominent class of compounds whose antiviral potential has been traced to such an effect.¹ Within that group, carbocyclic adenosine (aristeromycin, 1) is at the center of these investigations⁴ but its promise is limited by a toxicity arising from 5'-phosphate formation.⁵

Structural modifications of **1** with the aim of reducing phosphate-based toxicity have yielded meaningful drug candidates.⁶ An approach not explored, however, is extension of the C-5' hydroxymethyl side chain by a methylene group to provide the C-5' homolog of aristeromycin (**2**). This analog can be expected⁷ to have displaced the phosphate-susceptible hydroxyl from the phosphate-transfer zone in the kinases responsible for metabolism to **1** to its nucleotides. In support of this, **2** has been reported⁸ to be inactive against HSV-1 and HSV-2, possibly, due to its failure to be phosphorylated (Fig. 1).

To investigate 2 more thoroughly as a possible antiviral agent a more practical synthesis of it was necessary. For comparative antiviral purposes, 5'-homoadenosine (3) was also sought by a much more efficient way than exists

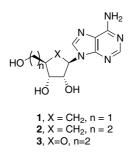


Figure 1.

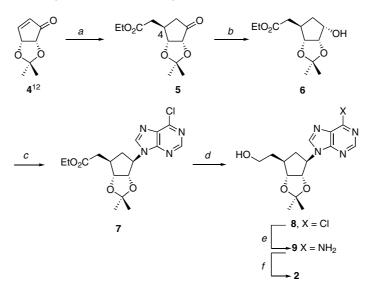
in the literature.⁹ The results of this effort are communicated here.

Existing methods^{8,10,11} for preparing 5'-homoaristeromycin (2) suffer from too many steps, limited scale-up, low yields and, in one case, resulting in a racemic product. Therefore, an efficient and stereoselective synthesis of 2 was needed. Starting from enone 4^{12} (Scheme 1), 1,4-addition of ethyl trimethylsilylacetate followed by in situ cleavage of the trimethylsilyl group furnished, stereoselectively, the ketone ester 5 as the only product. The stereochemistry at C-4 of 5 was derived from the fact (i) that a 1,4-addition to the concave structure of **5** can be expected¹³ to give a β (up) product and (ii) that 5 was converted into the known 2. Reduction of 5 with sodium borohydride provided the coupling precursor 6, which has been reported via a more tedious way.14 Mitsunubo coupling reaction of 6 with 6-chloropurine furnished 7. Selective reduction of 7 with diisobutylaluminum hydride (DIBAL) yielded the desired alcohol 8.

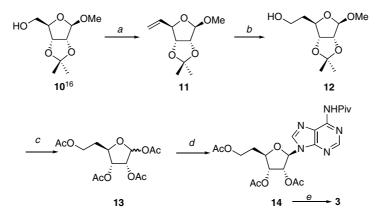
Keywords: Carbocyclic nucleosides; Nucleoside homologation; Monkeypox.

^{*}Corresponding author. Tel.: +1 334 844 5737; fax: +1 334 844 5748; e-mail: schnest@auburn.edu

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter @ 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2004.10.019



Scheme 1. Reagents and conditions: (a) (i) ethyl (trimethylsilyl)acetate, *n*-BuLi, DIPA, HMPA/THF, -78 °C, (ii) KF, EtOH/H₂O, rt, 84% for two steps from 4; (b) NaBH₄, MeOH, ice temp, 100%; (c) 6-chloropurine, Ph₃P, DIAD, THF, 0–50 °C, 52%; (d) DIBAL, CH₂Cl₂, -50 °C, 80%; (e) NH₃/MeOH, 94%; (f) HCl/MeOH, 95%.



Scheme 2. Reagents and conditions: (a) (i) DMSO, DIPEA, SO₃-Py, CH₂Cl₂, 90%, (ii) Ph₃PCH₃Br, *t*-BuOK, Et₂O, 75%; (b) 9-BBN, THF, then NaOH, H₂O₂, 97%; (c) (i) 70% AcOH, 85°C; (ii) Ac₂O, DMAP, pyridine, 85% for two steps from 12; (d) Adpiv, HMDS, TMSCl, TMSOTf, 60%; (e) NH₃/MeOH, 91%.

Ammonolysis of 8 (to 9) followed by hydrolytic deprotection smoothly afforded 2 in good overall yield.¹⁵

The known syntheses of 6'-homoadenosine⁹ (3) either involved many steps^{9a} or suffer a low yield of the final product.^{9b,c} Our plan (Scheme 2) envisioned beginning with homologation of 10.¹⁶ Side chain oxidation of 10 followed by Wittig olefination afforded 11. Submitting 11 to regioselective hydroboration with 9-BBN followed by oxidative hydrolysis, smoothly provided 12 in high yield. Hydrolysis of 12 with acetic acid produced a tetrol, which was fully protected with acetic anhydride to provide the anomeric acetate 13. The coupling reaction of 13 with *N*-pivaloyl protected adenine under Vorbrüggen glycosylation conditions yielded the desired N-9 product 14 as the only isolated product. Deprotection of 14 with ammonia furnished 3 in good overall yield¹⁷ from D-ribose. Compounds 2 and 3 were evaluated against a wide variety of both DNA viruses and RNA viruses.¹⁸ From this, very significant effects were seen for 2 toward vaccinia (IC₅₀ 1.2µg/mL), cowpox (IC₅₀ 0.12µg/mL), and monkeypox (IC₅₀ 0.12µg/mL) viruses, all in Vero 76 cells with $CC_{50} > 100\mu$ g/mL. This observation is particularly noteworthy since it is well known that vaccinia is susceptible to AdoHcy hydrolase inhibitors but cowpox was thought not to be.^{18e} In any case, details of this investigation and the, possibly, less notable activity of 2 toward other viruses¹⁸ will be forthcoming, including its potency toward variola.¹⁹ Analog 3 was inactive in all of the assays employed.¹⁸

Acknowledgements

This research was supported by funds from the NIH (AI 56540). We are also indebted to the following individu-

als for providing the antiviral data: Dr. Erik De Clercq, the Rega Institute, Leuven Belgium; Dr. Earl Kern, University of Alabama at Birmingham, Birmingham, AL; Dr. Brent Korba, Georgetown University, Washington, DC; Dr. Robert Sidwell, Utah State University, Logan, UT; and, Dr. John Huggins, U.S. Army Medical Research Institute of Infectious Diseases Frederick, MD.

References and notes

- 1. De Clercq, E. Nat. Rev. Drug Discovery 2002, 1, 13.
- 2. Liu, S.; Wolfe, M. S.; Borchardt, R. T. Antiviral Res. 1992, 19, 247.
- Rodriguez, J. B.; Comin, M. J. Mini-Rev. Med. Chem. 2003, 3, 95.
- 4. Yuan, C.-S.; Liu, S.; Wnuk, S. F.; Robins, M. J.; Borchardt, R. T. Adv. Antiviral Drug Des. **1996**, 2, 41.
- (a) Bennett, L. L., Jr.; Allan, P. W.; Rose, L. M.; Comber, R. N.; Secrist, J. A., III. *Mol. Pharmacol.* **1986**, *29*, 383;
 (b) Bennett, L. L.; Bowdon, B. J.; Allan, P. W.; Rose, L. M. *Biochem. Pharmacol.* **1986**, *35*, 4106.
- (a) Hasobe, M.; Liang, H.; Ault-Riche, D. B.; Borcherding, D. R.; Wolfe, M. S.; Borchardt, R. T. Antiviral Chem. Chemother. 1993, 4, 245; (b) Rajappan, V. P.; Schneller, S. W.; Williams, S. L.; Kern, E. R. Bioorg. Med. Chem. 2002, 10, 883.
- Shuto, S.; Obara, T.; Saito, Y.; Andrei, G.; Snoeck, R.; De Clercq, E.; Matsuda, A. J. Med. Chem. 1996, 39, 2392.
- 8. Jones, M. F.; Roberts, S. M. J. Chem. Soc., Perkin Trans. 1 1988, 11, 2927.
- (a) Robins, M. J.; Guo, Z. Q.; Wnuk, S. F. J. Am. Chem. Soc. 1997, 119, 3637; (b) Kappler, F.; Hampton, A. In Nucleic Acid Chemistry: Improved and New Synthetic Procedures Methods and Techniques; Townsend, L. B., Tipson, R. S., Eds.; Wiley: New York, 1991; p 240; (c) Mikhailov, S. N.; Padyukova, N. S.; Karpeiskii, M. Y.; Kolobushkina, L. I.; Beigelman, L. N. Collect. Czech. Chem. Commun. 1989, 54, 1055; (d) Hollmann, J.; Schlimme, E. Liebigs Ann. Chem. 1984, 1, 98.

- Siddiqi, S. M.; Chen, X.; Schneller, S. W. Nucleosides Nucleotides 1993, 12, 267.
- 11. Kapeller, H.; Baumgartner, H.; Griengl, H. Monatsh. Chem. 1997, 128, 191.
- Yang, M.; Ye, W.; Schneller, S. W. J. Org. Chem. 2004, 69, 3993.
- 13. Wolfe, M. S.; Anderson, B. L.; Borcherding, D. R.; Borchardt, R. T. J. Org. Chem. **1990**, 55, 4712.
- Matsugi, M.; Gotanda, K.; Ohira, C.; Suemura, M.; Sano, A.; Kita, Y. J. Org. Chem. 1999, 64, 6928.
- 15. Selected data for **2**: mp 178–179 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 8.20 (s, 1H), 8.11 (s, 1H), 7.17 (s, 2H), 4.90 (d, J = 6.27 Hz, 1H), 4.68 (d, J = 4.63 Hz, 1H), 4.57 (m, 1H), 4.43 (t, J = 5.12 Hz, 1H), 4.35 (m, 1H), 3.71 (m, 1H), 3.46 (m, 2H), 2.24 (m, 1H), 1.97 (m, 1H), 1.74 (m, 2H), 1.58 (m, 1H);⁸ ¹³C NMR (100 MHz, CDCl₃) δ 156.0, 152.1, 149.6, 140.2, 119.3, 74.9, 74.3, 59.9, 59.4, 40.0, 37.4, 32.5. Anal. Calcd for C₁₂H₁₇N₅O₃: C, 51.60; H, 6.14; N, 25.08. Found: C, 51.39; H, 6.18; N, 24.81.
- Ugarkar, B. G.; DaRe, J. M.; Kopcho, J. J.; Browne, C. E., III; Schanzer, J. M.; Wiesner, J. B.; Erion, M. D. J. Med. Chem. 2000, 43, 2883.
- 17. Selected data for **3**: mp 223–224 °C (lit. ⁹c 231.5–232.5 °C); ¹H NMR (250 MHz, DMSO- d_6) δ 8.27 (s, 1H), 8.10 (s, 1H), 7.24 (s, 2H), 5.79 (d, J = 5.3 Hz, 1H), 5.36 (d, J = 5.7 Hz, 1H), 5.11 (d, J = 5.1 Hz, 1H), 4.61 (m, 1H), 4.44 (t, J = 5.1 Hz, 1H), 4.03 (m, 1H), 3.93 (m, 1H), 3.42 (m, 2H), 1.76 (m, 2H); ¹³C NMR (62.9 MHz, DMSO- d_6) δ 156.1, 152.6, 149.4, 139.9, 119.2, 87.4, 81.0, 73.4, 72.9, 57.5, 36.6. Anal. Calcd for C₁₁H₁₅N₅O₄: C, 46.97; H, 5.38; N, 24.90. Found: C, 46.85; H, 5.48; N, 24.63.
- For leading references on the procedures used for the assays see (a) Ref. 5; (b) Siddiqi, S. M.; Chen, X.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1994, 37, 551; (c) Seley, K. L.; Schneller, S. W.; Korba, B. Nucleosides Nucleotides 1997, 16, 2095; (d) http://www.usu.edu/iar/ Brochure/brochure.html (September 6, 2004); and; (e) Baker, R. O.; Bray, M.; Huggins, J. W. Antiviral Res. 2003, 57, 13.
- 19. Bray, M.; Roy, C. J. J. Antimicrob. Chemother. 2004, 54, 1.