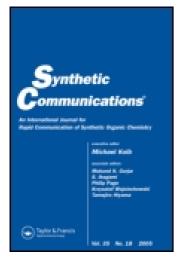
This article was downloaded by: [University of Bristol] On: 25 February 2015, At: 15:27 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry

Publication details, including instructions for authors and subscription information:

http://www.tandfonline.com/loi/lsyc20

Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides

Ahmed F. Khattab^a

^a Chemistry Department, Faculty of Science, Monoufiya University, Shebien EI-Koam, Egypt Published online: 16 Aug 2006.

To cite this article: Ahmed F. Khattab (2006) Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides, Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry, 36:8, 1097-1107, DOI: <u>10.1080/00397910500498994</u>

To link to this article: http://dx.doi.org/10.1080/00397910500498994

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at http://www.tandfonline.com/page/terms-and-conditions

Synthetic Communications[®], 36: 1097–1107, 2006 Copyright © Taylor & Francis Group, LLC ISSN 0039-7911 print/1532-2432 online DOI: 10.1080/00397910500498994



Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides

Ahmed F. Khattab

Chemistry Department, Faculty of Science, Monoufiya University, Shebien El-Koam, Egypt

Abstract: Acyclonucleosides of pyrimidine were prepared by condensing appropriately silylated 5-substituted pyrimidines with an acyclic side chain in the form of an acetylated haloalkoxyalcohol and subsequent removal of the protecting acetyl group in base. Also, acyclonucleosides with a 1-ethoxymethyl side chain have been investigated. The activity against hepatitis B virus (HBV) has been tested.

Keywords: Pyrimidine acyclonucleosides, substituted pyrimidines

The synthesis and biological evaluation of modified acyclonucleosides have been very active research areas for a number of years. Several pyrimidine acyclonucleosides are presently known as potent antiviral agents.^[1–5] Among them, acyclic nucleoside of the 1-[(2-hydroxyethoxy)methyl]-6-(phenylthio) thymine (HEPT) type has shown high selectivity toward HIV-1,^[6,7] and it was found that replacement of the sulfur atom with a methylene group resulted in a new series of potent HEPT analogues, of which MKC-442 (6-benzyl-1-ethoxymethyl-5-isopropyluracil) was found to be extremely potent.^[8,9] Therefore, a series of new pyrimidine acyclonucleosides are still of great interest as potential new active compounds with less prominent side effects.

We have previously reported the condensation between methyl 2-deoxy-3,5-di-*O*-toluoyl-*D*-pentofuranoside and silylated pyrimidines gave anomeric mixtures of 2'-deoxynucleosides, which can be envisaged as formal analogues of nucleic acids components.^[10] As a continuation of our interest in this area,

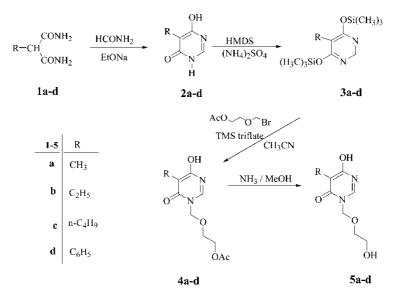
Received in the U.K. June 27, 2005

Address correspondence to Ahmed F. Khattab, Chemistry Department, Faculty of Science, Monoufiya University, Shebien El-Koam, Egypt. E-mail: khattab2000@ yahoo.com

we now report the synthesis and anti-HBV activity of a new series of pyrimidine acyclonucleosides in which the carbohydrate moiety has been replaced by an acyclic side chain^[11]

RESULTS AND DISCUSSION

The starting 4-hydroxy-6(1H)-pyrimidinones 2a-d have been prepared on condensation of malondiamide derivatives 1a-d with formamide in the presence of sodium ethoxide by the method of Hull.^[12] Silvlation of 2a-d with 1,1,1,3,3,3-hexamethyldisilazane (HMDS) in the presence of a catalytic amount of ammonium sulfate was performed according to Wittenburg^[13] to give the derivatives 3a-d. The latter were condensed with 2-acetoxyethoxy methyl bromide^[14] using the trimethylsilyl trifluoromethanesulfonate (TMS triflate) using method of Vorbrüggen et al.^[15] in anhydrous acetonitrile to give acyclonucleosides (4a-d) in 65-76% yield. After their chromatographic purification, treatment of these nucleosides with methanolic ammonia at room temperature for 24 h, followed by chromatographic purification, resulted in complete deprotection of the hydroxyl group, and the corresponding 1-(2-hydroxyethoxymethyl)pyrimidine derivatives (5a-d) were obtained in 82-90% yield (Scheme 1). The structures of the new compounds were determined on the basis of their elemental microanalyses and spectral data, which are in agreement with their proposed structures

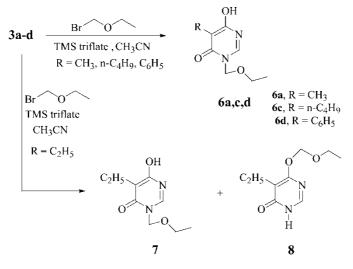


Scheme 1.

Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides

Similarly, condensation of bromomethyl ethyl ether with the silvlated pyrimidines 3a-b, using TMS triflate as the Lewis acid catalyst, gives the alkylated products 6-8. As reported in Scheme 2, N-alkylated products 6a,c,d were obtained, on condensation of bromomethyl ethyl ether with 3a,c,d, as the sole products in fairly good yields. However, in the case of alkylation of 5-ethylpyrimidine derivative 3b, column chromatographic purification of the reaction mixture gave two products. The less polar product appeared to be the required N-alkylated product (30% yield). The ¹H NMR spectrum of the more polar product (25% yield) showed the presence of ethoxymethyl moiety, which shifted downfield compared with that of N-alkylated product 7. The methylene singlet signal appeared at δ 5.49 ppm for the product 8, compared to δ 5.24 ppm for N-alkylated product 7. In addition, the ¹³C NMR spectrum showed the chemical shift at C-1 of acyclic part of 7 at δ 74.38 ppm and that of compound 8 at δ 90.28 ppm. This indicated that the alkylation of 3b occurred on the nitrogen atom to give N-alkylated product 7 and on the oxygen atom to give O-alkylated product 8, respectively. The FAB-mass spectra of 7 and 8 showed the same protonated positive molecular ion peak $(M + H)^+$ at m/z 199. An attempt to synthesize the acyclonucleosides 4–7 by simple alkylation of the sodium salt of the pyrimidines 2a-d with the appropriate halomethyl ethers according to the method of Sasaki et al.^[16] was unsuccesful.

Preliminary viral screening against HBV indicated that compounds **4a** and **4c** showed moderate viral replication inhibition and low cytotoxicity. Compounds **5a**, **5d**, and **6a** showed high inhibition with moderate cytotixicity,



Scheme 2.

Downloaded by [University of Bristol] at 15:27 25 February 2015

Compound (10 µM)	Inhibition (%)			
	1 week	2 weeks	3 weeks	Cytotoxicity (%)
4a	24.6	18.3	11.1	4.8
4c	26.7	14.6	9.3	4.6
5a	80.5	78.7	72.3	15.2
5d	78.7	75.2	70.4	13.0
6a	80.6	77.5	70.6	14.1
6c	79.5	75.4	73.5	4.2
6d	80.4	78.5	71.1	4.8
7	85.8	83.4	80.5	2.5
8	75.2	80.5	70.2	3.4

Table 1. Inhibition of HBV replication by selected compounds

and compounds **6c**, **6d**, **7**, and **8** showed high inhibition with low cytotoxicity (Table 1).

EXPERIMENTAL

Melting points were determined on a Boetius melting-point apparatus and are uncorrected. Elemental analysis were carried out in the Microanalytical Laboratories of the Faculty of Science, Cairo University. ¹H and ¹³C NMR spectra were recorded on a Bruker AC 250 FT-NMR spectrometer at 250 MHz for ¹H NMR and 62.9 MHz for ¹³C NMR. Mass spectra were recorded using fast atom bombardment (FAB) on a Kratos Ms 50-spectrometer. Analytical TLC plates 60 F_{254} and silica gel (0.040–0.063 mm) were purchased from Merck. Anhydrous CH₃CN was distilled from P₂O₅, followed by distillation from CaH. All other solvents were used after distillation and drying. Viral screening against HBV has been done at National Liver Institute, Monoufiya University, Shebien El-Koam, Egypt.

General Procedure for the Preparation of Compounds (2a-c)

A mixture of methyl malondiamide **1a** or ethyl malondiamide **1b** and/or n-butyl malondiamide **1c** (10 mmol), formamide (15 mmol), and sodium ethoxide [prepared by dissolving 0.92 g (40 mmol) of sodium metal in 30 ml of ethanol] was refluxed for 12 h. The mixture was evaporated to dryness under reduced pressure, and the obtained residue was dissolved in water and acidified with 4M HCl. The solid product was filtered off, dried, and recrystallized from the proper solvent to afford **2a**, **2b**, and **2c**, respectively, as white prisms in 71-83% yields. Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides

Data

4-Hydroxy-5-methylpyrimidin-6(1*H***)-one (2a)**. White prisms from methanol, (yield 83%): mp >320°C (reported^[17] yield 52%, mp >330°C). ¹H NMR (DMSO-d₆), (δ ppm): 1.77 (s, 3H, CH₃), 7.93 (s, 1H, C2-H), 11.63 (sb, 1H, OH), 12.01 (sb, 1H, NH). ¹³C NMR (DMSO-d₆), (δ ppm): 7.68 (CH₃), 97.98 (C5), 146.30 (C2), 163.07, 164.38 (C4 and C6). Anal. cacld. for C₅H₆N₂O₂: C, 47.62; H, 4.80; N, 22.21. Found: C, 47.85; H, 5.12; N, 21.90.

5-Ethyl-4-hydroxypyrimidin-6(1*H***)-one (2b).** White prisms from methanol, (yield 71%): mp >320°C. ¹H NMR (DMSO-d₆), (δ ppm): 0.97 (t, 3H, J = Hz, CH₃CH₂), 2.31 (q, 2H, J = 7.3 Hz, CH₂CH₃), 7.92 (s, 1H, C2-H), 11.61 (sb, 1H, OH), 11.90 (sb, 1H, NH). ¹³C NMR (DMSO-d₆), (δ ppm) 12.45 (CH₃), 15.49 (CH₂), 104.26 (C5), 146.58 (C2), 163.01, 164.06 (C4 and C6). Anal. calcd. for C₆H₈N₂O₂: C, 51.42; H, 5.75; N, 19.99. Found: C, 51.12; H, 5.80; N, 20.25.

5-Butyl-4-hydroxypyrimidin-6(1*H***)-one (2c).** White prisms from ethanol, (yield 77%): mp 288–290°C (reported^[18] yield 39%, mp 282°C). ¹H NMR (DMSO-d₆), (δ ppm): 0.87 (t, 3H, J = 7.0 Hz, CH₃), 1.23–1.44 (m, 4H, CH₃CH₂CH₂CH₂), 2.25–2.31 (t, 2H, J = 7.2 Hz, CH₃CH₂CH₂CH₂CH₂), 7.89 (s, 1H, C2-H), 11.60 (sb, 1H, OH), 12.20 (sb, 1H, NH). ¹³C NMR (DMSO-d₆), (δ ppm): 13.83 (CH₃), 21.81, 22.11, 29.70 (3 × CH₂), 102.88 (C5), 146.51 (C2), 162.34, 164.28 (C4 and C6). Anal. calcd. for C₈H₁₂N₂O₂: C, 57.13; H, 7.19; N, 16.66. Found: C, 57.43; H, 7.54; N, 16.47.

4-Hydroxy-5-phenylpyrimidin-6(1H)-one (2d)

This compound was prepared according to ref. 12.

General Procedure for the Preparation of Compounds (4a–d)

A mixture of pyrimidine derivatives 2a-d (10 mmol), 1,1,1,3,3,3-hexamethyldisilazane (HMDS) (30 ml), and ammonium sulfate (30 mg) was heated under reflux for 6 h. The mixture was concentrated at room temperature under reduced pressure to obtain the silylated derivatives 3a-d as pale yellow oil. Anhydrous MeCN (20 ml) was added, and the solution was stirred at -25° C. TMS triflate (2.35 g, 10.5 mmol) was added to the mixture followed by the dropwise addition of the 2-acetoxyethoxy methyl bromide (20 mmol) in anhydrous MeCN (5 ml). The reaction mixture was stirred at -25° C for 2 h and then at room temperature for 1 h. The reaction mixture was diluted with 200 ml of CH₂Cl₂ and washed with ice-cold saturated aqueous NaHCO₃ (4 × 100 ml). The organic phase was separated, washed with cold water (4 × 100 ml), dried over Na₂SO₄, and evaporated to obtain

the crude products, which were chromatographed on a silica-gel column using methanol/chloroform (5-10%) as eluent.

Data

1-[(2-Acetoxyethoxy)methyl]-4-hydroxy-5-methylpyrimidin-6(1*H***)-one (4a). White solid (yield 68%), mp 104–106°C. ¹H NMR (DMSO-d₆), (δ ppm): 1.79 (s, 3H, CH₃), 1.98 (s, 3H, CH₃CO), 3.63–3.75 (m, 2H, CH₂), 4.07–4.11 (m, 2H, CH₂), 5.28 (s, 2H, NCH₂O), 8.27 (s, 1H, C2-H), 11.35 (sb, 1H, OH). ¹³C NMR (DMSO-d₆), (δ ppm): 8.05 (CH₃), 20.46 (<u>C</u>H₃CO), 62.83 (CH₂), 67.02 (CH₂), 74.71 (NCH₂O), 97.25 (C5), 149.88 (C2), 162.32 (C4), 164.85 (C6), 170.16 (<u>C</u>OCH₃). FAB MS: m/z (%) 243 (M + 1, 100), 154 (36). Anal. cacld. for \overline{C}_{10}H_{14}N_2O_5: C, 49.58; H, 5.82; N, 11.56. Found: C, 49.32; H, 6.10; N, 11.78.**

1-[(2-Acetoxyethoxy)methyl]-4-hydroxy-5-ethylpyrimidin-6(1*H***)-one (4b). White solid (yield 65%), mp 106–108°C,. ¹H NMR (CDCl₃), (δ ppm): 1.12 (t, 3H, J = 7.3 Hz, CH₃CH₂), 2.08 (s, 3H, CH₃CO), 2.51 (q, 2H, J = 7.3 Hz, CH₂CH₃), 3.85–3.89 (m, 2H, CH₂), 4.20–4.24 (m, 2H, CH₂), 5.26 (s, 2H, NCH₂O), 8.07 (s, 1H, C2-H), 11.32 (sb, 1H, OH). ¹³C NMR (CDCl₃), (δ ppm): 11.92 (CH₃), 16.15 (CH₂), 20.56 (CH₃CO), 62.63 (CH₂), 68.22 (CH₂), 75.46 (NCH₂O), 106.31 (C5), 147.42 (C2), 162.29 (C4), 164.08 (C6), 170.60 (CH₃CO). FAB MS: m/z (%) 257 (M + 1, 100), 154 (47). Anal. calcd. for C₁₁H₁₆N₂O₅: C, 51.56; H, 6.29; N, 10.93. Found: C, 51.85; H, 6.50; N, 11.22.**

1-[(2-Acetoxyethoxy)methyl]-4-hydroxy-5-n-butylpyrimidin-6(1*H***)-one (4c). White solid (yield 65%), mp 78–80°C,. ¹H NMR (CDCl₃), (δ ppm): 0.86–0.91 (t, 3H, J = 7.0 Hz, CH₃CH₂CH₂), 1.21–1.41 (m, 4H, CH₃CH₂CH₂CH₂), 1.96 (s, 3H, CH₃CO), 2.23–2.30 (t, 2H, J = 6.8 Hz, CH₃CH₂CH₂CH₂), 3.64–3.73 (m, 2H, CH₂), 4.12–4.23 (m, 2H, CH₂), 5.24 (s, 2H, NCH₂O), 8.26(s, 1H, C2-H), 11.25 (sb, 1H, OH). ¹³C NMR (CDCl₃), (δ ppm): 13.66 (CH₃), 20.43 (CH₃CO), 22.01, 22.15, 28.46 (3 × CH₂), 62.81 (CH2), 68.11 (CH₂), 74.37 (NCH₂O), 103.88 (C5), 146.43 (C2), 161.73, 163.64 (C6), 170.35 (COCH3). FAB MS: m/z (%) 285 (M + 1, 100), 154 (32). Anal. calcd. for C₁₃H₂₀N₂O₅: C, 54.92; H, 7.09; N, 9.85. Found. C, 55.13; H, 7.21, N, 10,12.**

1-[(2-Acetoxyethoxy)methyl]-4-hydroxy-5-phenylpyrimidin-6(1*H***)-one (4d). White solid (yield 76%), mp 136–138°C, ¹H NMR (CDCl₃), (\delta ppm): 2.02 (s, 3H, CH₃), 3.81–3.85 (m, 2H, CH₂), 4.15–4.18 (m, 2H, CH₂), 5.33 (s, 2H, NCH₂O), 7.30–7.63 (m, 5H, Ar-H), 8.04 (s, 1H, C₂-H), 11.46 (sb, 1H, OH). ¹³C NMR (CDCl₃), (\delta ppm): 20.56 (CH₃), 62.78 (CH₂), 68.45 (CH₂), 75.81 (NCH₂O), 104.63 (C5), 127.52, 127.80, 130.09, 130.49 (Ar-C), 148.68 (C2), 161.31 (C4), 163.84 (C6), 170.73 (COCH₃). FAB MS: m/z**

(%) 305 (M + 1, 100), 189 (32), 154 (58). Anal. calcd. for $C_{15}H_{16}N_2O_5$: C, 59.21; H, 5.30; N, 9.21. Found: C, 59.52; H, 5.11; N, 9.50.

General Procedure for the Preparation of (5a–d)

A mixture of 4a, 4b, 4c, and/or 4d (5 mmol), methanol (25 ml), and concentrated aqueous ammonia (25%, 25 ml) was stirred at room temperature for 24 h. The solvent was evaporated under reduced pressure, and the residue was purified on a silica-gel column using 5-10% methanol in diethyl ether to give 5a-d in 82-90% yields.

Data

4-Hydroxy-1-[(2-hydroxyethoxy)methyl]-5-methylpyrimidin-6(1*H***)-one (5a). Colorless oil (yield 87%). ¹H NMR (DMSO-d₆), (\delta ppm): 1.77 (s, 3H, CH₃), 3.54–3.61 (m, 2H, CH₂), 3.75–3.84 (m, 2H, CH₂), 4.53 (sb, 1H, OH), 5.16 (s, 2H, NCH₂O), 8.07 (s, 1H, C2-H), 11.26 (sb, 1H, OH). ¹³C NMR (DMSO-d₆), (\delta ppm): 8.04 (CH₃), 61.78 (CH₂), 68.04 (CH₂), 72.62 (NCH₂O), 98.01 (C5), 147.86 (C2), 161.23 (C4), 163.58 (C6). Anal. calcd. for C₈H₁₂N₂O₄: C, 48.00; H, 6.04; N, 13.99. Found: 48.32; H, 5.95; N, 14.12.**

5-Ethyl-4-hydroxy-1-[(2-hydroxyethoxy)methyl]pyrimidin-6(1*H***)-one (5b). Colorless oil (yield 85%). ¹H NMR (DMSO-d₆), (\delta ppm): 1.11 (t, 3H, J = 7.1, CH₃CH₂), 2.44 (q, 2H, J = 7.3 Hz, CH₂CH₃), 3.75–3.79 (m, 2H, CH₂), 4.11–4.17 (m, 2H, CH₂), 4.61 (sb, 1H, OH), 5.24 (s, 2H, NCH₂O), 7.96 (s, 1H, C2-H), 11.27 (sb, 1H, OH). ¹³C NMR (DMSO-d₆) (\delta ppm): 11.75 (CH₃), 15.45 (CH₂), 61.62 (CH₂), 66.34 (CH₂), 73.34 (NCH₂O), 105.41 (C5), 146.44 (C2), 162.48 (C4), 164.03 (C6). Anal. calcd. for C₉H₁₄N₂O₄: C, 50.46; H, 6.59; N, 13.08. Found: C, 50.13; H, 6.50; N, 13.34.**

5-Butyl-4-hydroxy-1-[(2-hydroxyethoxy)methyl]pyrimidin-6(1*H***)-one (5c).** Colorless oil (yield 82%). ¹H NMR (DMSO-d₆), (δ ppm) 0.85–0.89 (t, 3H, J = 7.2 Hz, CH₃), 1.11–1.13 (m, 4H, 2 × CH₂), 2.11–2.19 (t, 2H, J = 7.3 Hz, CH₂), 3.44–3.51 (m, 2H, CH₂), 4.07–4.11 (m, 2H, CH₂), 4.62 (sb, 1H, OH), 5.21 (s, 2H, NCH₂O), 7.97 (s, 1H, C2-H), 11.23 (sb,1H, OH). ¹³C NMR (DMSO-d₆), (δ ppm): 13.54 (CH₃), 22.21, 22.43, 29.21 (3 × CH₂), 62.75 (CH₂), 68.46 (CH₂), 72.44 (NCH₂O), 102.65 (C5), 144.56 (C2), 161.76 (C4), 164.03 (C6). Anal. calcd. for C₁₁H₁₈N₂O₄: C, 54.43; H, 7.49; N, 11.56. Found: C, 54.57; H, 7.22; N, 11.12.

4-Hydroxy-1-[(2-hydroxyethoxy)methyl]-5-phenylpyrimidin-6(1*H***)-one (5d). Colorless oil (yield 90%). ¹H NMR (DMSO-d₆), (\delta ppm): 3.41–3.47 (m, 2H,**

CH₂), 3.58–3.62 (m, 2H, CH₂), 4.63 (sb, 1H, OH), 5.11 (s, 2H, NCH₂O), 7.21–7.32 (m, 5H, Ar-H), 7.91 (s, 1H, C2-H), 11.23 (sb, 1H, OH). ¹³C NMR (DMSO-d₆) (δ ppm) 60.06 (CH₂), 69.19 (CH₂), 71.58 (NCH₂O), 104.22 (C5), 126.52, 127.60, 130.20, 130.41 (Ar-C), 147.62 (C2), 161.41 (C4), 163.72 (C6). Anal. calcd. for C₁₃H₁₄N₂O₄: C, 59.54; H, 5.38; N, 10.68. Found: C, 59.33; H, 5.51, N, 10.44.

General Procedure for the Preparation of Compounds (6-8)

A solution of chloromethyl ethyl ether (1.86 g, 20 mmol) in anhydrous MeCN (10 ml) was added to a stirred solution of the silylated pyrimidines 3a-d in anhydrous MeCN at -25° C. The stirring was continued for 2 h at -25° C and for 1 h at room temperature, then the reaction mixture worked up as previously described. The compounds were purified by silica-gel column chromatography with methanol in chloroform (0–5%). *N*-Alkylated products **6a**-**c** were obtained as the sole products in 70–75% yield. *N*-Alkylated product **7** and its *O*-alkylated counterpart **8** mixture were obtained and separated. Fractions with *N*-alkylated product **7** were eluated faster than their *O*-alkylated counterpart **8**.

Data

1-(Ethoxymethyl)-4-hydroxy-5-methylpyrimidin-6(1*H***)-one (6a**). White solid (yield 73%), mp 156–158°C. ¹H NMR (DMSO-d₆), (δ ppm): 1.11 (t, 3H, *J* = 7.0 Hz, CH₃), 1.79 (s, 3H, CH₃), 3.53 (q, 2H, *J* = 7.0 Hz, CH₂), 5.24 (s, 2H, NCH₂O), 8.26 (s, 1H, C2-H), 11.42 (sb, 1H, OH). ¹³C NMR (DMSO-d₆), (δ ppm): 8.07 (CH₃), 14.76 (CH₃), 64.11 (CH₂), 74.40 (NCH₂O), 97.24 (C5), 148.83 (C2), 162.28 (C4), 164.79 (C6). FAB MS: m/z (%). 185 (M+1, 100). 154 (16). Anal. calcd. for C₈H₁₂N₂O₃: C, 52.17; H, 6.57; N, 15.21. Found: C, 52.43; H, 6.25; N, 15.11.

5-Butyl-1-(ethoxymethyl)-4-hydroxypyrimidin-6(1*H***)-one (6c). White solid (yield 70%), mp 98–100°C ¹H NMR (DMSO-d₆) (\delta pmm): 0.85–0.90 (t, 3H,** *J* **= 7.0 Hz, CH₃CH₂CH₂CH₂CH₂), 1.08–1.13 (t, 3H,** *J* **= 7.0 Hz, CH₃CH₂O), 1.25–1.38 (m, 4 H, 2 × CH₂), 2.28–2.34 (t, 2H,** *J* **= 7.1 Hz, CH₂), 3.49–3.58 (q, 2H,** *J* **= 6.8 Hz, OCH₂CH₃), 5.24 (s, 2H, NCH₂O), 8.25 (s,1H, C2-H), 11.27 (sb, 1H, OH). ¹³C NMR (DMSO-d₆) (\delta ppm): 13.76 (CH₃), 14.78 (CH₃), 22.04, 22.16, 29.58 (3 × CH₂), 64.16 (OCH₂CH₃), 74.36 (NCH₂O), 102.03 (C5), 149.05 (C2), 162.03 (C4), 164.78 (C6). FAB MS: m/z (%) 227 (M + 1, 100). Anal. calcd. for C₁₁H₁₈N₂O₃: C, 58.39; H, 8.02; N, 12.38. Found: C, 58.15; H, 8.35; N, 12.11.**

1-(Ethoxymethyl)-4-hydroxy-5-phenylpyrimidin-6(1*H***)-one (6d). White solid (yield 75%), mp 218–220°C. ¹H NMR (DMSO-d₆), (\delta ppm): 1.12 (t, 3H,** *J* **= 3.6 Hz, CH₃), 3.58 (q,** *J* **= 7.0 Hz, CH₂), 5.28 (s, 2H, NCH₂O), 7.21–7.49 (m, 5H, Ar-H), 8.44 (s, 1H, C2-H), 11.33 (s, 1H, OH). ¹³C NMR (DMSO-d₆) \delta 14.80 (CH₃), 64.35 (CH₂), 74.73 (NCH₂O), 101.91 (C5), 126.22, 127.16, 130.33, 132.59 (Ar-C), 150.33 (C2), 160.66 (C4), 164.21 (C6). FAB MS: m/z (%) 247 (M + 1, 100), 189 (10), 176 (20). Anal. calcd. for C₁₃H₁₄N₂O₃: C, 63.40; H, 5.73; N, 11.38. Found: C, 63.70; H, 5.54; N, 11.53.**

1-(Ethoxymethyl)-5-ethyl-4-hydroxypyrimidin-6(1*H***)-one (7). White solid (yield 30%), mp 160–162°C. ¹H NMR (DMSO-d₆), (δ ppm): 0.97 (t, 3H, J = 7.3 Hz, CH₃), 1.11 (t, 3H, J = 7.0 Hz, CH₃), 2.32 (q, 2H, J = 7.4 Hz, CH₂), 3.54 (q, 2H, J = 7.0 Hz, CH₂), 5.24 (s, 1H, NCH₂O), 8.25 (s, 1H, C2-H), 11.33 (sb, 1H, OH). ¹³C NMR (DMSO-d₆), (δ ppm): 12.31 (CH₃), 14.78 (CH₃), 15.87 (CH₂), 64.17 (CH₂), 74.38 (NCH₂O), 103.36 (C5), 149.09 (C2), 161.86 (C4), 164.56 (C6). FAB MS: m/z (%) 199 (M + 1, 100), 169 (16), 154 (20). Anal. calcd. for C₉H₁₄N₂O₃: C, 54.53; H, 7.12; N, 14.13. Found: C, 54.60; H, 7.42; N, 14.45.**

4-(Ethoxymethoxy)-5-ethylpyrimidin-6(1*H***)-one (8). White solid (yield 30%), mp 118–120°C. ¹H NMR (DMSO-d₆), (\delta ppm): 0.99 (t, 3H, J = 7.3 Hz, CH₃), 1.13 (t, 3H, J = 7.0 Hz, CH₃), 2.35 (q, 2H, J = 7.2 Hz, CH₂), 3.67 (q, 2H, J = 7.0 Hz, CH₂), 5.49 (s, 1H, OCH₂O), 8.01 (s, 1H, C2-H). ¹³C NMR (DMSO-d₆), (\delta ppm): 12.38 (CH₃), 14.90 (CH₃), 15.51 (CH₂), 64.41 (CH₂), 90.28 (OCH₂O), 108.05 (C5), 146.84 (C2), 162.82 (C4), 163.63 (C6). FAB MS: m/z (%) 199 (M + 1, 100). Anal. calcd. for C₉H₁₄N₂O₃: C, 54.53; H, 7.12; N, 14.13. Found: C, 54.85; H, 7.25; N, 14.50.**

BIOLOGICAL ACTIVITY STUDIES

The required cell line was made by transfection of Hep G2-cells with a plasmid containing multiple tandem copies of HBV genome (subtype ayw).^[19] The 2.2.15 cell line was maintained in RPMI-1640 (Glutamax) culture media containing 100 IU/ml nystatin $+380 \,\mu\text{g/ml}$ G418 (geneticin). The transfected Hep G2-2.2.15 cell line was kept in tissue culture flask at $37^{\circ}\text{C} + 5\%$ CO₂. Maintenance media were added to the cell culture together with the tested compounds (final concentration = $10 \,\mu\text{M}$). The supernatant liquid was collected after 1, 2, and/or 3 weeks by asbiration of the media from culture flask and washing the cells twice by PBS. Ten percent versene/trypsin was added, and the cells were incubated for 1 min at 37°C .^[20] The DNA replication was estimated by the PCR (polymerase chain reaction) technique. The PCR reaction mixture contained 14 μ L of extracted supernatant, 4 mmol/L of MgCl₂, 10 μ mol/L of DIG-11-dUTP,

190 μ mol/L of dTTP, 200 μ mol/L of dATP, dGTP, 1,5 U Taq polymerase, 20 mmol/L of HCl (pH 8.4), 50 mmol/L of KCl, 1 μ mol/L of HCID-1 primer (5'GGAAAGAAGTCAGAAGGCA3'), and 1 μ mol/L of HCID-2 (5'TTGGGGGAGGAGATTAGGTT3'), in a total volume of 50 μ L. PCR reaction conditions were 32 cycles of 1 min at 94°C, 30 s at 58°C, and 30 s at 72°C + 3 s for each cycle in a thermal circler as described in literature.^[21] The percentage inhibition could be calculated by the relation between the plank experiment (containing maintenance media without the tested compounds) and the results obtained after the periods. The percentage cytotoxicity could be estimated by the relation between the number of the living and dead cells after 3 weeks as counted by the Haemocytometer.

ACKNOWLEDGEMENTS

DANIDA and Danish Ministry of Foreign Affairs are gratefully acknowledged for financial support through the project "Development of New Drugs against Hepatitis" at Monoufiya University, and E. B. Pedersen is thanked for making spectral measurements at Chemistry Institute, Syddansk University, Odense M, Denmark.

REFERENCE

- Schaeffer, J. H.; Beauchamp, L.; De Miranda, P.; Elion, G.; Bauer, G. D.; Collins, P. *Nature*. **1978**, 272, 583.
- Pan, B.-C.; Chen, Z.-H.; Piras, G.; Dutschman, G. E.; Rowe, E. C.; Cheng, Y.-C.; Chu, S.-H. J. Heterocycl. Chem. 1994, 31, 177.
- Zeid, I. F.; Abdel-Rahman, A.-H. A.; Abdel-Megied, E.-S. A.; El-Etrawy, S. H.A. Nucleosides Nucleotides 1999, 18 (1), 95.
- Krivonogov, V. P.; Myshkin, V. A.; Sivkova, G. A.; Grebenkova, N. A.; Srubillin, D. V.; Kozlova, G. G.; Abdrakhmanov, I. B.; Mannapova, R. T.; Spirikhin, L. V.; Tolstikov, G. A. *Pharm. Chem. J.* **2001**, *35* (8), 411.
- El-Essawy, A. F.; El-Brollosy, R. N.; Pedersen, B. E.; Nielsen, C. J. *Heterocycl. Chem.* 2003, 40, 213.
- Miyasaka, T.; Tanaka, H.; Baba, M.; Hayakawa, H.; Walker, R. T.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1989, 32, 2507.
- Tanaka, H.; Baba, M.; Hayakawa, H.; Sakamaki, T.; Miyasaka, T.; Ubasawa, M.; Takashima, H.; Sekiya, K.; Nitta, I.; Shigeta, S.; Walker, R. T.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1991, 34, 349.
- Baba, M.; Shigeta, S.; Yuasa, S.; Takashima, H.; Sekiya, K.; Ubasawa, M.; Tanaka, H.; Miyasaka, T.; Walker, R. T.; De Clercq, E. Antimicrob. Agents Chemother. 1994, 38, 688.
- Baba, M.; Tanaka, H.; Miyasaka, T.; Yuasa, S.; Ubasawa, M.; Walker, R. T.; De Clereq, E. Nucleosides Nucleotides 1995, 14, 575.
- Khattab, A. F.; Abdel Megied, A. E.-S.; Pedersen, E. B. Nucleosides Nucleotides Nucleic Acids. 2003, 22 (1), 99.

Synthesis of New 5-Substituted Pyrimidine Acyclonucleosides

- 11. El-Essawy, F. A.; Khattab, A. F. J. Heterocycl. Chem. 2004, 41, 311.
- 12. Hull, R. J. Chem. Soc. 1951, 487, 2214.
- 13. Wittenburg, E. Z. Chem. 1964, 4 (3), 303.
- 14. Robins, J. M.; Peter, W. P. Can. J. Chem. 1982, 60, 547.
- 15. Vorbrüggen, H.; Krolikiewicz, K.; Bennua, B. Chem. Ber. 1981, 114, 1234.
- 16. Sasaki, T.; Minamoto, K.; Suzuki, T.; Yamashita, S. Tetrahedron 1980, 36, 865.
- 17. Prystaš, M. Coll. Czech. Chem. Commun. 1967, 32, 4241.
- Buděšínský, Z.; Roubínek, F.; Svátek, E. Coll. Czech. Chem. Commun. 1965, 30, 3730.
- 19. Sells, M. A.; Chen, M. L.; Acs, G. Proc. Natl. Acad. Sci. USA 1987, 84, 1005.
- Doong, S. L.; Tasi, C. H.; Schinazai, R. F.; Liotta, D. C.; Cheng, Y. C. Proc. Natl. Acad. Sci. USA 1991, 88, 8495.
- 21. Kobra, B. E.; Gerin, J. L. Antiviral Res. 1992, 19, 55.