Synthesis of $(2\underline{S}, 3\underline{S}, 5\underline{S})$ -3-Hydroxy-5-methyl-2-pyrrolidinecarboxylic Acid, a Component of Actinomycin Z₁

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 $(2\underline{S},3\underline{S},5\underline{S})$ -3-Hydroxy-5-methyl-2-pyrrolidinecarboxylic acid and the enantiomer, the relative configurations of which are the same as that of the natural compound found in actinomycin Z_1 were synthesized and the optical properties were determened.

3-Hydroxy-5-methyl-2-pyrrolidinecarboxylic acid (3-hydroxy-5-methylproline) is an unusual imino acid identified as a component of a peptide antibiotic actinomycin Z_1 .¹⁾ The four diastereomers of the imino acid were synthesized by Mauger et al. and their relative stereochemistries were determined by NMR data and epimerization studies.²⁾ It has been demonstrated that 3-hydroxy-5-methylproline obtained from the hydrolyzate of actinomycin Z_1 was identical with one of the synthetic isomers which has 2,3-trans and 2,5-cis relative stereochemistry, however, the absolute configuration (2<u>S</u>,3<u>S</u>,5<u>S</u> or 2<u>R</u>,3<u>R</u>,5<u>R</u>) remains unknown.

As a part of our efforts toward the synthesis of biologically active β -hydroxy- α -amino acid derivatives,³) we planned to synthesize $(2\underline{S},3\underline{S},5\underline{S})$ -3-hydroxy-5-methylproline (<u>1</u>) in order to reveal the optical properties based on the retrograde pathway $A \rightarrow B \rightarrow C \rightarrow D + E$.



The main features are (i) [3+2] dipolar cycloaddition⁴⁾ of <u>D</u> and <u>E</u>, (ii) reductive cleavage of an isoxazole ring to a β -hydroxyketone^{5a)} (<u>C</u> \rightarrow <u>B</u>), and (iii) reductive cyclization (<u>B</u> \rightarrow <u>A</u>). The present communication describes an efficient synthesis of (2<u>S</u>,3<u>S</u>,5<u>S</u>)-3-hydroxy-5-methylproline (<u>1</u>) and the stereoisomers (<u>2</u>-<u>6</u>) along these lines.⁶)

Reaction of L-vinylglycine $\underline{7}^{7}$ (Z: benzyloxycarbonyl) and nitrile oxide $\underline{8}$ generated in situ (nitroethane, p-chlorophenylisocyanate,⁸⁾ Et₃N, benzene, 25 °C, 19 h, then reflux, 6.8 h) afforded a 2.3:1 mixture of <u>threo</u>- and <u>erythro</u>-isoxazolines,⁹⁾ 9, $[\alpha]_{D}$ -0.69° (c 0.57, CHCl₃) and <u>10</u>, $[\alpha]_{D}$ +71° (c 0.51, CHCl₃), respectively in 53% yield. These diastereomers were separated by column



chromatography on silica gel (hexane/ethyl acetate, 2:1) and, the stereochemistry of the isolated compounds was determined by converting them to the corresponding proline derivatives shown below. Hydrogenolytic cleavage of isoxazoline 10 according to the Curran's method^{5b)} (Raney-Ni, B(OH)₃, MeOH/H₂O, H₂) afforded a β -hydroxyketone <u>12</u>, [α]_D -6.5° (c 0.71, MeOH) in 88% yield. After removal of the amino protecting group of $\underline{12}$ (H₂/10% Pd-C, EtOH), the resulting cyclic imine $\underline{13}$ was reduced (NaBH₃CN, EtOH, pH 5)¹⁰⁾ to a diastereomeric mixture of 3-hydroxy-5methylproline esters (14 and 15). The mixture was hydrolyzed (0.15 mol Ba(OH)2, 20 °C, 4 h) without separation and the hydrolyzate was subjected to a Dowex 50W x 4 column chromatography eluting with ammonia-formate buffer (pH 2.70) to afford (2<u>S</u>,3<u>S</u>,5<u>S</u>)-3-hydroxy-5-methylproline <u>1</u>, mp 247-249 °C (dec.), [a]_D -17° (c 0.50, H_2O) and $(2\underline{S}, 3\underline{S}, 5\underline{R})$ -isomer 2, mp 252-255 °C (dec.), $[\alpha]_D$ -10° (c 0.48, H_2O) in 44% and 26% yields (from <u>12</u>), respectively. The ¹H-NMR data of 1 and 2 were in accord with those reported about racemic 1 and 2, respectively.²⁾



The <u>threo</u>-isoxazoline <u>9</u> was independently converted by the same series of reactions to 3-hydroxy-5-methylprolines <u>via</u> β -hydroxyketone <u>11</u>,¹¹⁾ [α]_D +2.8° (c 0.71, MeOH) (<u>9 + 11</u>, 83% yield). A mixture of (2<u>S</u>,3<u>R</u>,4<u>S</u>)- and (2<u>S</u>,3<u>R</u>,4<u>R</u>)-isomers (<u>3/4</u>, 2:1) was obtained in 23% yield (from <u>11</u>) together with (2<u>R</u>,3<u>R</u>,4<u>S</u>) isomer <u>5</u> and (2<u>R</u>,3<u>R</u>,4<u>R</u>)-isomer 6 (7% and 5% yields, respectively).¹²⁾ These results

suggest that epimerization attributable to a tautomerism of the cyclic imine intermediate occurred at the C-2 position. The $(2\underline{R},3\underline{R},5\underline{S})$ -isomer <u>6</u> and $(2\underline{R},3\underline{R},5\underline{S})$ -isomer <u>5</u> exhibited the same spectral properties as those of <u>1</u> and <u>2</u>, respectively except the opposite optical rotations, <u>6</u>, $[\alpha]_D$ +18° (c 0.32, H₂O), <u>5</u>, $[\alpha]_D$ +10° (c 0.20, H₂O).



In conclusion, we demonstrated an efficient synthesis of optically active 3-hydroxy-5-methylprolines (1-6) and clarified their optical properties, which will serve to elucidate the absolute configuration of the natural compound from actinomycin z_1 .¹³⁾

The present methodology seems to be very useful for preparations of β -hydroxy- α -amino and imino acids especially considering the wide variation of molecules containing nitryl oxide groups.

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- 11) In this case, Kozikowski's procedure using $AlCl_3$ in the place of $B(OH)_3$ was used. See Ref. 5a.
- 12) The mixture of <u>3</u> and <u>4</u> could not be separated under the chromatographic condition (Dowex 50w x 4, pH 2.70 buffer) however, the structures and the ratio were confirmed by comparison of ¹H-NMR spectrum of the mixture with those of racemic 3 and 4.
- 13) The natural 3-hydroxy-5-methylproline from actinomycin Z₁ was not obtained in sufficient quantity for optical rotation. Private communication from Dr. A. B. Mauger.
- 14) ¹H-NMR data: <u>9</u> (100 MHz, CDCl₃) δ⁻¹.80 (3H, br s), 2.69-3.06 (2H, m), 3.68 (3H, s), 4.39 $(1H, dd, J=2.0, 10.0 Hz, H_{(\alpha)})$, 4.79-5.59 $(2H, m, H_{(\beta)})$ and NH), 5.03 (2H, s), 7.24 (5H, s). <u>10</u> (100 MHz, CDCl₃) & 1.94 (3H, br s), 3.07 (2H, br d, J=8.0 Hz), 3.69 (3H, s), 4.40 (1H, dd, J=4.0, 8.2 Hz, $H_{(\alpha)}$), 4.78 (1H, dt, J=4.0, 8.0 Hz, H_(B)), 5.09 (2H, s), 5.63 (1H, br d, J=8.2 Hz), 7.30 (5H, <u>11</u> (100 MHz, CDCl₃) δ 2.13 (3H, s), 2.68 (2H, br d, J=6.2 Hz), 3.35 (1H, s). br d, J=3.2 Hz, OH), 3.73 (3H, s), 4.34 (1H, br d, J=9.2 Hz, $H_{(\alpha)}$), 4.47-4.83 (1H, m, H_(B)), 5.11 (2H, s), 5.65 (1H, br d, J=9.2 Hz, NH), 7.32 (5H, s). 12 (100 MHz, CDCl₃) δ 2.14 (3H, s), 2.74 (2H, br d, J=5.6 Hz), 3.55 (1H, br d, J=4.2 Hz, OH), 3.71 (3H, s), 4.08-4.45 (2H, m, H_(α), H_(β)), 5.06 (2H, s), 5.70 (1H, br d, J=8.0 Hz, NH), 7.27 (5H, s). $1 (500 \text{ MHz}, D_2 \text{O}) \delta$ 1.51 (3H, d, J=6.7 Hz, Me), 1.76 (1H, ddd, J=4.3, 11.5, 14.1 Hz, H_(4a)), 2.20 (1H, ddd, J=1.0, 6.0, 14.1 Hz, H_(4b)), 4.07 (1H, br s, H₍₂₎), 4.07 (1H, ddd, J=6.0, 6.7, 11.5 Hz, $H_{(5)}$), 4.68 (1H, br dd, J=1.0, 4.3 Hz, $H_{(3)}$). 2 (500 MHz, D_2O) δ 1.48 (3H, d, J=6.8 Hz, Me), 1.74 (1H, dddd, J=1.0, 4.6, 7.4, 14.0 Hz, H_(4b)), 2.44 (1H, ddd, J=5.9, 8.0, 14.0 Hz, H_(4a)), 3.93 (1H, ddd, J=6.8, 7.4, 8.0 Hz, H₍₅₎), 4.03 (1H, dd, J=1.0, 3.1 Hz, H₍₂₎), 4.62 (1H, ddd, J=3.1, 4.6, 5.9 Hz, H₍₃₎).
- 15) 13 C-NMR data: <u>1</u> (25 MHz, D₂O) δ 17.4 (q), 39.7 (t), 56.0 (d), 69.9 (d), 74.6 (d), 172.0 (s). <u>2</u> (25 MHz, D₂O) δ 17.9 (q), 38.9 (t), 55.0 (d), 68.1 (d), 73.8 (d), 171.9 (s).

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