## Application of FITC-labeled Ternatin on Its Cellular Localization in 3T3-L1 Murine Preadipocytes

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The cellular localization of a potent fat-accumulation inhibitor (–)-ternatin was investigated. FITC-labeled ternatin, a chemical probe for current study, was synthesized from a highly bioactive ternatin analogue using Click Chemistry, which was found to be bioactive. The treatment of 3T3-L1 murine preadipocytes with synthetic probe revealed that (–)-ternatin is localized in a specific organelle of 3T3-L1 cells.

(–)-Ternatin (1, Figure 1) is a highly N-methylated cyclic heptapeptide that was isolated from the mushroom *Coriolus versicolor* in our continuing search for potential antiobesity drugs from natural sources.<sup>1</sup> The in vitro biological evaluation revealed that 1 potently inhibited fat-accumulation against 3T3-L1 murine adipocytes. In addition, we also demonstrated the inhibitory effect of fat accumulation in vivo, which led to suppression of body-weight gain in high-fat-diet induced obese mice by treatment with 1 (5 mg/kg/day) for 5 weeks.<sup>2</sup> Therefore, 1 could be considered as a plausible lead compound for antiobesity drugs. However, both the biological mechanism of 1 and its cellular target still remained unknown.

As an initial effort toward bioorganic studies on 1, we recently reported its structure–activity relationships, which revealed key amino acid residues (IIe<sup>1</sup> and Leu<sup>4</sup>) responsible for potent bioactivity.<sup>3</sup> These SAR profiles enabled further chemical modification, i.e., installation of functional groups, in appropriate positions in the chemical structure of 1. In a previous paper, we reported [*N*Me-D-ProGly<sup>6</sup>]-ternatin (2, Figure 1) as a highly bioactive analogue of 1, which was applied to the synthesis of biotin-labeled ternatin as a chemical probe for identification of bio-molecules that bind to 1.<sup>4</sup> As a part of our on-going bio-organic studies on 1, we describe here the synthesis and biolog-



(-)-Ternatin (1): R = Me (NMe-D-Ala<sup>6</sup>)

[NMe-D-ProGly<sup>6</sup>]-Ternatin (2): R =

Figure 1. Structures of 1 and 2.



Scheme 1. a) 50% TFA/CH<sub>2</sub>Cl<sub>2</sub>; b) FITC, NEt<sub>3</sub>, DMF, 59% in 2 steps; c) 2 (1.0 equiv), CuSO<sub>4</sub>, sodium ascorbate, *t*-BuOH, H<sub>2</sub>O, 58%; d) 50% TFA/CH<sub>2</sub>Cl<sub>2</sub>; e) FITC, NEt<sub>3</sub>, DMF, quant. in 2 steps.

ical evaluation of FITC-labeled ternatin **3**, and its cellular localization in 3T3-L1 murine preadipocytes.

On the basis of the synthetic route to biotin-labeled compounds<sup>4</sup> which feature the Cu-catalysed Huisgen reaction (Click Chemistry),<sup>5</sup> we thought that **3** could be synthesized from three components, 2, FITC (fluorescent isothiocyanate), and linker 4, which is shown in Scheme 1. First, compound 4, prepared from triethylene glycol in five steps, was subjected to Boc deprotection under acidic conditions. The resulting amine (as a TFA salt) was coupled with FITC in the presence of NEt<sub>3</sub> to provide the FITC-linker conjugate 5. Finally, the Cu-catalyzed Huisgen reaction of 5 with alkyne 2 using CuSO<sub>4</sub> and sodium ascorbate in t-BuOH/H2O generated 3. After HPLC purification of the reaction mixture, the desired 3 was obtained in 58% yield. On the other hand, another FITC-linker conjugate 7 which bears a methyl ester group instead of the peptide core in 3 was synthesized as a negative compound in our study. The Boc deprotection of 6 followed by coupling with FITC afforded 7.

Table	1.	Fat-a	ccumula	tion	inhibito	ory e	ffect	s of	comp	ounds	1-3
and 7,	an	d cell	viability	of 3	3T3-L1	adi	pocy	tes <sup>a</sup>	_		

Compound	Fat-acumulation inhibitory effect: IC <sub>50</sub> /µM	Cell viability: IC <sub>50</sub> /µM
(–)-Ternatin (1)	$0.027\pm0.003$	$0.28\pm0.03$
2	$0.019\pm0.001$	>5.2 <sup>b</sup>
3	$23 \pm 1.5$	>75 <sup>b</sup>
7	>164 <sup>b</sup>	>164 <sup>b</sup>

<sup>a</sup>Values are means of quadruplicate determinations. <sup>b</sup>Not tested at higher concentrations.

The in vitro fat-accumulation inhibitory effects for synthetic compounds **3** and **7** were then assessed (Table 1). The bioassay consisted of the treatment of confluent 3T3-L1 preadipocytes with each sample and insulin (an inducer of adipogenesis), and further incubation for 7 days. After this period, control cells were differentiated into mature adipocytes. Both the rates of fat accumulation and cell viability were calculated to identify undesired fat-accumulation inhibition caused by cytotoxicity.

On the basis of the results, **3** showed moderate inhibitory effect on fat accumulation with an IC<sub>50</sub> value 23  $\mu$ M and no significant cytotoxicity at tested concentrations (IC<sub>50</sub> > 75  $\mu$ M), which suggested that **3** was potent enough to be applied for further bio-organic investigation. Meanwhile, **7** did not show any inhibitory effect or cytotoxicity (IC<sub>50</sub> > 164  $\mu$ M).

Visualization of the intracellular localization of synthesized compounds was conducted using fluorescence microscopy (Eclipse TE200; Nikon, Tokyo, Japan), which is shown in Figure 2.<sup>6</sup> For this examination, we employed 3T3-L1 murine preadipocytes. Our recent research demonstrated that **1** shows its inhibitory effects against fat-accumulation by suppressing the differentiation of 3T3-L1 cells in the early stage. Therefore, **1** was thought to show its biological effects in 3T3-L1 murine preadipocytes.

By the fluorescence microscopic analysis, a certain rate of 3T3-L1 cells treated with **3** showed strong fluorescence under UV (Figure 2b). The pattern of the fluorescence was partial and spot-like inside the cells. On the other hand, no fluorescence was observed when cells were treated with **7** (Figure 2d). These results suggested that **3** was incorporated into cells and localized in a specific organelle of 3T3-L1 cells. It was confirmed that the organelle where **3** accumulated was not identical to nucleus by **3** and DAPI co-staining analysis (data not shown). The part of localization is now under investigation. Thus, FITC-labeled (–)-ternatin was demonstrated to localize in a specific organelle of 3T3-L1 cells, and then, (–)-ternatin is likely to function there to show its biological activity.

In summary, we synthesized FITC-conjugated ternatin **3** using Click Chemistry, and examined its inhibitory effect against the fat accumulation in 3T3-L1 cells. Next, we surveyed its intracellular localization in 3T3-L1 cells using fluorescence microscopic analysis. **3** was observed to localize in a specific organelle of 3T3-L1 cells, and thus, **3** will be a useful tool for the analysis of the mechanism of action and the identification of the target molecules of **1**.



**Figure 2.** Cellular localization of FITC-labeled ternatin **3** in 3T3-L1 murine preadipocytes analyzed by fluorescence. (a) Cells treated with **3** for 18 h; (b) fluorescence image of a; (c) cells treated with **7** for 18 h; (d) fluorescence image of c; (e) control cells (no additive); (f) fluorescence image of e.

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- 6 3T3-L1 preadipocytes were inoculated at  $5 \times 10^4$  cells/well in 24-well plates, and incubated with 25  $\mu$ M of 3 or 7 for 18 h. After washing, cells were analyzed by phase contrast (left panels) and fluorescence (right panels) microscopy. The cells were photographed at a magnification of  $200 \times$ .