

Clavulanic Acid Biosynthesis: the Stereochemical Course of β -Lactam Formation from Chiral Glycerol

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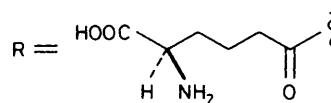
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The overall stereochemical course of β -lactam formation in clavulanic acid was determined to be retention from (1*R*,2*R*)- and (1*S*,2*R*)-[1-³H],[1,3-¹⁴C]glycerol.

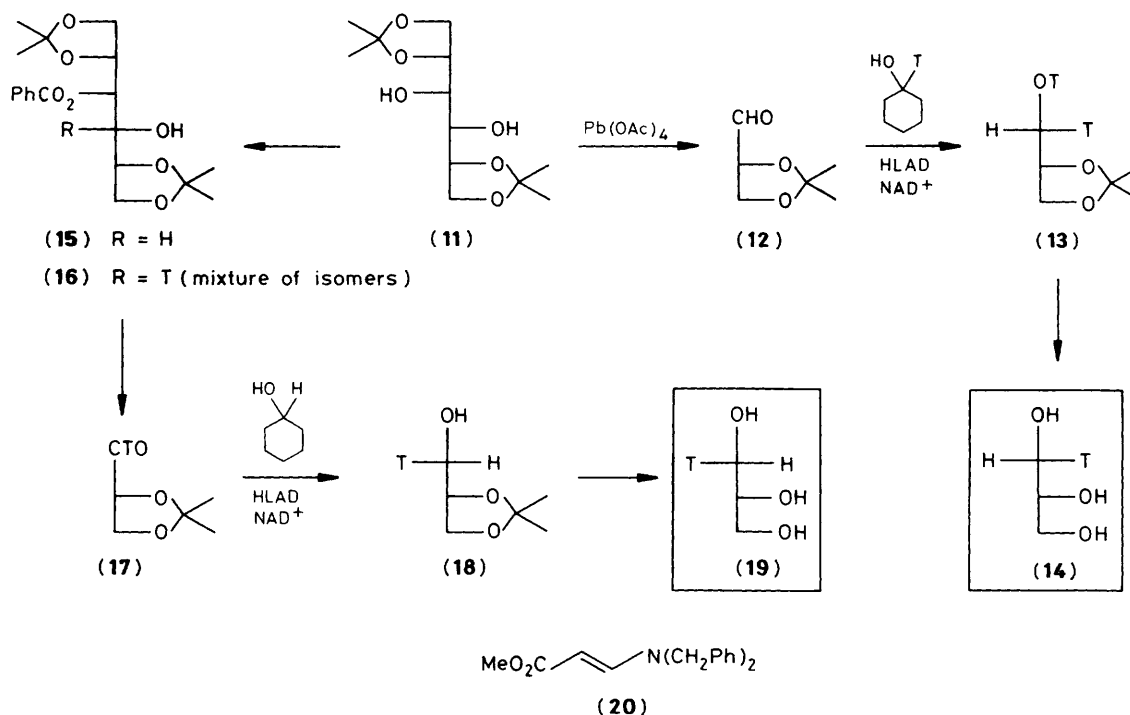
The monocyclic β -lactam ring of nocardicin A (**1**) is assembled from L-serine^{1,2} in a reaction path occurring with inversion of configuration³ and no change in oxidation state at the seryl β -carbon.³ In contrast, the corresponding four-membered rings of penicillin N (**2**) and cephalosporin C (**3**) are derived from L-cysteine in a sequence involving oxidative cyclization of a tripeptide precursor⁴ and overall retention of configuration⁵ at the cysteinyl β -carbon. We record in this communica-

tion a notable parallel to the latter process where the *pro*-(*R*)-hydroxymethylene of glycerol (**4**, -CH_AH_BOH), diastereotopically labelled with tritium, is shown to give rise to clavulanic acid (**10**, R=H) with overall retention of configuration.

The incorporation of [1,3-³H],[1,3-¹⁴C]glycerol (**4**) into the β -lactam carbons (C₃-unit \equiv C-5—7) of clavulanic acid (**10**, R = H) is known to take place with retention of one quarter of



D-Mannitol was converted into its bis-acetonide (**11**) (Scheme 2).⁷ Oxidative cleavage⁷ to the acetonide of D-glyceraldehyde (**12**) and reduction with horse liver alcohol dehydrogenase (HLAD)⁸ in the presence⁹ of [1-³H]cyclohexanol



Scheme 2

Table 1. Incorporation of diastereotopically-labelled glycerols (4) into *p*-bromobenzyl clavulanate (10, R = PBB).

Substrate	$^3\text{H}/^{14}\text{C}$ (% ^3H retained)		
	(14):(19) ^a	(10, R = PBB)	(20)
(1 <i>R</i> ,2 <i>R</i>)-[1- ^3H],[1,3- ^{14}C]Glycerol (14)	12.28	6.71 (55%)	10.73 (87%)
(1 <i>S</i> ,2 <i>R</i>)-[1- ^3H],[1,3- ^{14}C]Glycerol (19)	10.85	0.78 (7%)	0.98 (9%)

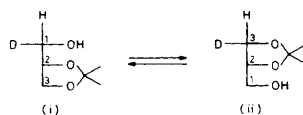
^a Doubly-labelled glycerols were accurately counted as their tris(*p*-nitrobenzoyl) esters.

and NAD^+ gave the (1*R*,2*R*)-alcohol (13).[‡] Mild acid hydrolysis afforded (1*R*,2*R*)-[1- ^3H]glycerol (14). The diastereoisomeric (1*S*,2*R*)-[1- ^3H]glycerol (19) was more difficult to obtain.¹⁰ Attempted preparation of tritiated aldehyde (17) by reduction of (12) with sodium borotritide and reoxidation under a variety of conditions was unsatisfactory. Finally, the

D-mannitol derivative (11) was converted into its monobenzoate (14).¹¹ Oxidation¹¹ of the remaining secondary hydroxy group followed by sodium borotritide reduction gave (16) (3:1 mixture of diastereoisomers favouring the isomer shown). After saponification of the benzoate (68% overall yield from introduction of the radioisotope), (16) was cleaved as above to give the desired tritiated aldehyde (17). Enzymic reduction and acetal hydrolysis (Scheme 2) afforded (1*S*,2*R*)-[1- ^3H]glycerol (19).

The diastereomerically labelled glycerols (14) and (19) were separately combined with [1,3- ^{14}C]glycerol and administered to cultures of *Streptomyces clavuligerus* (ATCC 27064) grown in a triglyceride medium.^{12,13} The results obtained are shown in Table 1. Glycerol labels not only the C₃-unit of clavulanic acid, but also the C₅-unit by circulation of radioisotope around the TCA cycle and incorporation as a C₅-amino acid derived from α -ketoglutarate (Scheme 1).¹³ Significant losses of tritium that attend these latter metabolic events account for the low $^3\text{H}:^{14}\text{C}$ -ratios observed for (10, R = PBB). However, methanolysis of (10, R = PBB) in the presence of dibenzylamine isolates the β -lactam carbons as the crystalline derivative (20) (Table 1).^{13,14} The data in the last column for this derivative show excellent complementarity, the glycerol *pro*-(1*S*) label (4, H_A) being largely lost while the *pro*-(1*R*) tritium (4, H_B) is largely retained in clavulanate.

[‡] While expected, based on ample precedent, the structure assigned to (13) was supported by independent ^1H n.m.r. experiments with the corresponding deuteriated materials. At 400 MHz the methylene hydrogens of glycerol acetonide give four well separated doublets of doublets (C-1: δ 3.59 and 3.74, J_{gem} 11.8 Hz; C-3: δ 3.79 and 4.04, J_{gem} 8.3 Hz). The deuteriated material (i) corresponding to (13) showed



disappearance of the resonance at δ 3.74 and significant broadening of that at the δ 3.58. Equilibration of the acetonide in acetone (cat. dry $p\text{-MeC}_6\text{H}_4\text{SO}_3\text{H}$, 3 Å molecular sieves, room temp., 5 days) gave a mixture of (i) and (ii) where the site of deuteration in the latter could be securely assigned to the (3*S*)-locus [T. D. Inch and N. Williams, *J. Chem. Soc. (C)*, 1970, 263].

In conclusion, the normal metabolic course of glycerol to the glycolytic intermediates (6) and (7) is borne out in the biosynthesis of clavulanic acid in that the *pro*-(*R*) hydroxymethylene of (4, $-\text{CH}_\text{A}\text{H}_\text{B}\text{OH}$) becomes C-5 in (10). The absolute sense of chirality at this centre is maintained through the biosynthetic pathway to label clavulanate with overall retention of stereochemistry at C-5. The diastereotopic tritium labels in (4, H_A and H_B) do not become homotopic, thereby ruling out the intermediacy of, *e.g.*, free pyruvate. The observation of stereochemical retention in the β -lactam ring closure of clavulanic acid is an important parallel to the oxidative cyclizations that characterize penicillin and cephalosporin biosynthesis.^{4,5} The extent of the similarities between these two pathways will be further defined in due course.

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