Preliminary communication

Conjugation of L-ascorbic acid and D-glucose

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The biochemical activity of L-ascorbic acid (vitamin C) (1) has led to the synthesis of numerous analogs and derivatives of this compound. From a different viewpoint, 1 has been considered a useful synthetic precursor to many molecules of potential biological utility on account of its inherent, varied chemical functionality. Thus, L-threonolactone has been prepared¹ from 1. Also, Jones *et al.*² have succeeded in transforming 1 into 3-hexuloses, and Brimacombe *et al.*³ have attempted to synthesize spirolactones from 1. As part of a program directed toward the transformation of 1 into complex carbohydrates, we report the synthesis and the proof of structure of the coupling product of 1 with 2,3,4,6tetra-O-acetyl- α -D-glucopyranosyl bromide (3). Another stimulus for the present study was the potential biological activity of the coupling product; indeed, 1 exists in many natural materials in the form of a more complicated compound, namely ascorbigen, which exhibits antiscorbutic activity and which releases 1 on hydrolysis. Ascorbigen was first isolated⁴ pure from cabbage and savoy and synthesized⁵ from 1 and 3-(hydroxymethyl)indole.

In the present work, the coupling was accomplished by treatment of 3 with L-ascorbate anion (2) in N,N-dimethylformamide at room temperature for 8 h. 3-O-(2,3,4,6-Tetra-O-acetyl- β -D-glucopyranosyl)-L-ascorbic acid (4) was obtained in 22% yield* by extraction and subsequent chromatography on silica gel. The major product was 2,3,4,6tetra-O-acetyl-D-glucose⁶ (39%), and the starting material 3 was also recovered in 10% yield. When coupling was performed by using conditions of phase-transfer catalysis (compound 3, sodium ascorbate, tetra-*n*-butylammonium bromide, and benzene), 4 was obtained in a lower yield (16%). The ¹H-n.m.r. spectrum (60 MHz) of 4 in chloroform-*d* showed a oneproton doublet at τ 4.30 ($J_{1'2'}$ 7.0 Hz) attributable to the anomeric proton (H-1'). The large coupling constant is consistent with the β -glycosidic linkage. The i.r. spectrum (Nujol) indicated that 4 has an α,β -unsaturated lactone ring [1770, 1740 (overlap with acetate carbonyl groups), 1660 cm⁻¹]. The u.v. spectrum provided further evidence for the presence of a conjugated system in 4 (λ_{max}^{MeOH} 227 nm). A positive (green) ferric chloride test in methanol indicated the presence of the enolic hydroxyl group. On the basis of the foregoing data, the coupling product was assigned the structure shown by formula 4. However, the compound could not be obtained completely pure, because decomposition occurred in

^{*}Partial decomposition occurred during chromatography.





носн

I HOCH₂

7b









DMF = N, N-dimethylformamide

3 + 7

solution. The decomposition is presumably attributable to the autohydrolysis of the glycosidic linkage caused by the acidic proton of the hydroxyl group at C-2. This hydroxyl group could be protected by treatment of 4 with diazomethane to yield 2-O-methyl-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-ascorbic acid (5) almost quantitatively; m.p. 143.5–144°, $[\alpha]_D^{23}$ –10.5° (c 0.20, methanol). Anal. Calc. for C₂₁H₂₈O₁₅: C, 48.47; H, 5.42. Found: C, 48.21; H, 5.69. The spectral data indicated that 5 has a β -glycosidic linkage and an α_{β} -unsaturated lactone ring; ¹H-n.m.r. data: (CDCl₃, 100 MHz): τ 4.27 (1H, d, $J_{1',2'}$ 7.0 Hz, H-1'), 5.22 (1H, s, H-4), 6.06 (3H, s, OCH₃), 7.91, 7.93, and 7.94 (12H, OCOCH₃); ¹³C-n.m.r. data: (p.p.m. downfield from Me₄Si, CDCl₃, 15.09 MHz): δ 170.6, 169.9 (OCOCH₃), 169.4 (C-1), 152.9 (C-3), 123.9 (C-2), 98.7 (C-1'), 75.9 (C-4), and 20.6 (OCOCH₃); ν_{max}^{KBr} 3410, 1770, 1740, and 1660 cm⁻¹; λ_{max}^{MeOH} 227 nm. Compound 5 showed a negative ferric chloride test and released formaldehyde⁷ on periodate oxidation; these results indicated that the two enolic hydroxyl groups were substituted and that vicinal hydroxyl groups were present at C-5 and C-6.

Although the coupling under the present conditions was expected to occur at O-3 of 1 on the basis of the different acidities of the hydroxyl groups on C-2 and C-3, in order to prove the coupling position unambiguously, 3-O-methyl-2-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-ascorbic acid (8) was synthesized. Selective methylation of 1 with diazomethane below -20° provided 3-O-methyl-L-ascorbic acid⁸ (6). Treatment of 3 with 3-O-methyl-L-ascorbate anion (7) in *N*,*N*-dimethylformamide at room temperature for 10 h gave 8 as an amorphous solid in 17% yield; $[\alpha]_D^{23} - 7.5^{\circ}$ (c 0.20, methanol): ¹H-n.m.r. data: (CDCl₃, 100 MHz): τ 4.67 (1H, d, $J_{1',2'}$ 6.9 Hz, H-1'), 5.26 (1H, d, $J_{4,5}$ 1.6 Hz, H-4), 5.81 (3H, s, OCH₃), 7.87, 7.92, 7.96, and 7.99 (12H, OCOCH₃); ¹³C-n.m.r. data: (downfield from Me₄Si, CDCl₃, 15.09 MHz): δ 170.5, 170.0, 169.5 (OCOCH₃), 168.6 (C-1), 160.5 (C-3), 117.8 (C-2), 98.6 (C-1'), 76.0 (C-4), and 20.6 (OCOCH₃); ν_{max}^{KBr} 3430, 1775, 1750, and 1680 cm⁻¹; $\lambda_{max}^{\text{MeOH}}$ 229 nm. *Anal.* Calc. for C₂₁H₂₈O₁₅: C, 48.47; H, 5.42. Found: C. 48.68; H, 5.29. Compound 8 showed also a negative ferric chloride test and released form-aldehyde on periodate oxidation. The characterization of compound 8 thus corroborated the assignment of the structure of the regioisomer 5.

As 2 and 7 have ambident nucleophilic characters, and 2a and 2b (ref. 9) and 7a and 7b (ref. 10) were proposed as resonance structures, respectively, *C*-glycosidation might also have been expected. However, *C*-glycosidation products were not detected under the reaction conditions employed in the present study, a result which presumably can be explained by effects of the medium. In general, *C*-alkylation is favored in media of high hydrogen-bonding power¹¹ and *O*-alkylation predominates in such polar aprotic solvents as *N*,*N*-dimethylformamide and dimethyl sulfoxide¹². This effect of the medium has been observed previously¹³ in benzylation of 1.

To afford a deacetylated conjugate, compound 5 was treated with sodium methoxide at room temperature. The reaction was complete within 20 min to provide syrupy 3-O- $(\beta$ -D-glucopyranosyl)-2-O-methyl-L-ascorbic acid (9). Decomposition occurred when the reaction time was prolonged. It is known that enol glycosides are usually sensitive to alkali¹⁴. The ¹H-n.m.r., i.r., and u.v. spectra of 9 were consistent with the assigned structure: ¹H-n.m.r. data: (methanol- d_4 , 60 MHz): τ 4.53 (1H, d, $J_{1',2'}$ 6.2 Hz, H-1'), 5.02 (1H, d, $J_{4,5}$ 1.2 Hz, H-4), and 5.74 (3 H, s, OCH₃); ν_{max}^{film} 3380, 1775, 1745, and 1680 cm⁻¹; λ_{max}^{MeOH} 230 nm. Reacetylation of 9 with acetic anhydride in pyridine gave 5,6-di-*O*-acetyl-2-*O*methyl-3-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)-L-ascorbic acid (10) in 84% yield as an amorphous solii; $[\alpha]_D^{23}$ -45.8° (c 0.24, chloroform); ¹H-n.m.r. data: (CDCl₃, 60 MHz): τ 4.37 (1H, d, $J_{1',2'}$ 7.0 Hz, H-1'), 5.12 (1H, d, $J_{4,5}$ 1.8 Hz, H-4), 6.06 (3H, s, OCH₃), 7.76, 7.90, and 7.95 (18H, OCOCH₃); ν_{max}^{film} 1775, 1760, and 1690 cm⁻¹; λ_{max}^{MeOH} 229 nm. *Anal.* Calc. for C₂₅H₃₂O₁₇: C, 49.67; H, 5.34. Found: C, 49.64; H, 5.44. Similarly, deacetylation of 8 yielded 2-*O*-(β -D-glucopyranosyl-3-*O*-methyl-L-ascorbic acid (11); ¹H-n.m.r. data: (methanol- d_4 , 60 MHz): τ 4.95 (1H, $J_{1',2'}$ 6.0 Hz, H-1') and 5.69 (3H, s, OCH₃); ν_{max}^{film} 3340, 1765, 1740, and 1655 cm⁻¹; λ_{max}^{MeOH} 232 nm. Reacetylation of 11 gave 5,6-di-*O*acety¹-3-*O*-methyl-2-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-glucopyranosyl)-L-ascorbic acid (12) in 86% yield; m.p. 117–118°, $[\alpha]_D^{23}$ -66.7° (c 0.18, chloroform); ¹H-n.m.r. data: (CDCl₃, 60 MHz): τ 4.72 (1H, d, $J_{1',2'}$ 6.9 Hz, H-1'), 5.20 (1H, d, $J_{4,5}$ 2.5 Hz, H-4), 6.22 (3H, s, OCH₃), 7.87, 7.91, 7.93, 7.95, and 7.98 (OCOCH₃); ν_{max}^{film} 1770, 1750, and 1680 cm⁻¹; λ_{max}^{MeOH} 232 nm. *Anal.* Calc. for C₂₅H₃₂O₁₇: C, 49.67; H, 5.34. Found: C, 49.38; H, 5.31. Acetylation of 5 and 8 also provided 10 and 12, respectively.

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