TERPENOIDS AND RELATED COMPOUNDS—XI¹ CHEMICAL INVESTIGATION OF *ALEURITES MONTANA* AND THE STRUCTURE OF ALEURITOLIC ACID—A NEW TRITERPENE ACID*

D. R. MISRA and H. N. KHASTGIR

Chemistry Department, North Bengal University, Darjeeling, West Bengal, India

(Received in UK 17 January 1970; accepted for publication 12 March 1970)

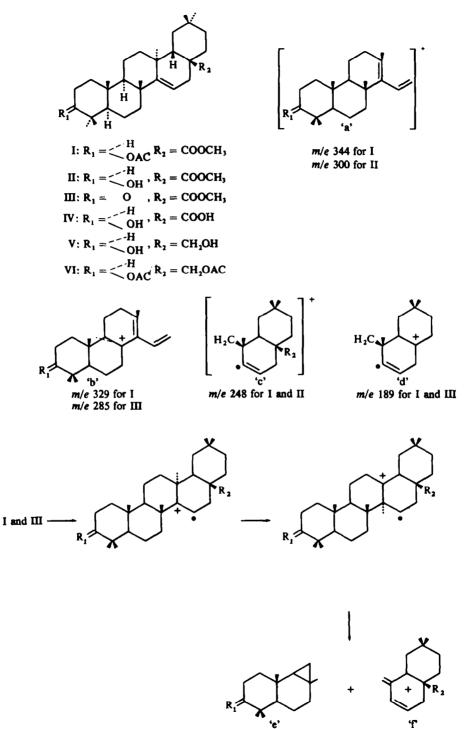
Abstract—The isolation of friedelin, β -sitosterol, betulinic acid and a new triterpene acid, aleuritolic acid, $C_{30}H_{48}O_3$ from the bark of *Aleurites montana* has been described. The new acid is present as its acetate. On the basis of physical and chemical evidences, structure IV is suggested for aleuritolic acid.

Aleurites montana² (Euphorbiaceae) is a tree of moderate height called Tung in Bengali.

The benzene extract of the bark of A. montana was separated into the acid and the neutral fractions. The neutral fraction on chromatography first yielded friedelin, m.p. $259-61^{\circ}$, $(\alpha)_{\rm D}-36^{\circ}$, identical with an authentic sample³ (m.m.p. and IR). The more polar component, $C_{29}H_{50}O$, m.p.135-7°, $(\alpha)_{\rm D}-40^{\circ}$ was identified as β -sitosterol.⁴ The acid fraction on esterification with diazomethane and subsequent chromatography first yielded the new compound, 3β -acetoxy methyl aleuritolate (I) m.p. $241-43^{\circ}$, $(\alpha)_{\rm D}+23.08^{\circ}$. The corresponding acid, named aleuritolic acid (IV) has been shown to be olean-14(15)-en-3 β -ol-28-oic acid on the basis of chemical and physical evidence. The more polar component m.p. $220 - 22^{\circ}$, $(\alpha)_{\rm D}+1\cdot4^{\circ}$, isolated from the chromatogram has been identified as methyl betulinate.⁵

Acetoxy methyl aleuritolate (I) $C_{33}H_{52}O_4$, (M⁺, 512), (α)_D + 23.08°, no UV absorption above 220 mµ, v_{max} 1735 cm⁻¹ (broad peak, -O -COCH₃ and -COOCH₃), 1245 cm⁻¹ (-O-COCH₃), 820 cm⁻¹ (trisubstituted double bond). Its NMR spectrum showed the presence of seven tertiary Me groups between 0.8 to 1.05 ppm, one acetoxy group at 2.04 ppm and one carbomethoxy group at 3.58 ppm. It showed a multiplet centered at 5.50 ppm indicating the presence of a trisubstituted double bond. Hydrolysis of acetoxy methyl alcuritolate (I) with 5% methanolic KOH furnished methyl aleuritolate II, m.p. 208–10°, $(\alpha)_{\rm p}$ + 11.11°, $\nu_{\rm max}^{\rm KBr}$ 3480 cm⁻¹ (---OH), 1735 cm⁻¹ (-COOCH₃), 820 cm⁻¹ (trisubstituted double bond), NMR signals at 5.50 ppm (multiplet, 1H, trisubstituted double bond), 3.54 ppm (-COOCH₃) and signals for seven tertiary Me groups. Methyl aleuritolate on prolonged treatment with 10% and 15% methanolic KOH under refluxing conditions gave back the starting material indicating the hindered nature of the carboxyl group. Methyl aleuritolate (II) on CrO_3 -pyridine oxidation⁶ gave the ketoester (III) $C_{31}H_{48}O_3$, (M⁺, 468), $(\alpha)_{\rm D}$ + 11.76°, v KBr 1705 cm⁻¹ (6-membered ring ketone), 1735 cm⁻¹ (trisubstituted double bond), NMR signals at 5.58 ppm (multiplet, 1H, trisubstituted double bond),

^{*} A part of the work has been communicated to J. Ind. Chem. Soc. as short communication.



m/e 262 for I m/e 220 for III

m/e 233

3.58 ppm (--COOCH₋₃) and signals for seven tertiary Me protons. Hydrolysis of methyl ester II with potassium tertiary butoxide in DMSO⁷ furnished the new acid IV, m.p. 300-302° (dec.), $v \frac{CHCl_1}{max}$ 3420 cm⁻¹ (--OH), 1700 cm⁻¹ (--COOH), 820 cm⁻¹ (trisubstituted double bond) for which the name aleuritolic acid is proposed.

Acetoxy methyl aleuritolate (I) showed yellow coloration with TNM and consumed one mole equivalent of perbenzoic acid indicating the presence of one double bond. Acid isomerization⁸ of I gave acetyl methyl oleanolate m.p. 219–20°, $(\alpha)_D$ 58.82°⁹ (m.m.p. and IR). This fact coupled with the NMR data establishes that aleuritolic acid (IV) contains a modified oleanane skeleton with a double bond, a β —OH at C— 3 and a —COOH group at C—17.

The position of the double bond at 14–15 position in aleuritolic acid (IV) was established from the mass spectral fragmentation pattern of acetoxy methyl aleuritolate (I) and methyl aleuritolonate (III). Compounds I and III exhibited mass peaks 'a' at m/e 344 and m/e 300 respectively. These ion peaks are accompanied by peaks 15 mass units lower 'b', which are formed by the loss of allylically activated Me group at C-8. The spectrum of I exhibited in addition, a peak at m/e 284 ('a'—CH₃COOH) and at m/e 269 ('b'—CH₃COOH). In addition to species 'a' and its further decomposition products, the spectra of I and III showed a very abundant peak 'c' at m/e 284 derived from rings D and E. Furthermore, fragment 'c' loses the substituent at C-17 giving rise to a fragment 'd' at m/e 189, ('c'—COOCH₃). The appearance of small but prominent peaks at m/e 262 and 220, 'e' derived from compounds I and III respectively and peaks at m/e 233, 'f' accompanied by a peak at m/e 174 ('f'— COOCH₃) can be explained by reactions shown in the accompanying chart. This type of fragmentation is consistent with the mass spectral data of Δ^{14} -taraxerene derivatives reported by Djerassi *et al.*¹⁰

The above data are compatible with structure IV for alcuritolic acid and co-relation with a suitable member of oleanane series was considered. LAH reduction of acetoxy methyl alcuritolate(I) furnished the corresponding diol (V) m.p. 265–67° which was identified as myricadiol¹¹⁻¹⁴ (m.m.p. and IR). Acetylation of the diol gave myricadiol diacetate VI, m.p. 251–53°, $(\alpha)_D$ –3° identical with an authentic sample of myricadiol diacetate (m.m.p. and IR).

EXPERIMENTAL

M.p's are uncorrected. Petroleum used throughout the investigation had b.p. 60-80°. Optical rotations refer to solns. in CHCl₃. IR spectra were recorded in Perkin-Elmer model 337 spectrophotometer. NMR spectra were determined in CDCl₃ on a Varian A-60 spectrometer using TMS as internal standard.

Extraction of the bark of A. montana. The dried ground bark of A. montana (2 Kg) was extracted with benzene in a Soxhlet apparatus for 18 hr. The gummy residue obtained after the removal of benzene was separated into the acid and neutral fractions. The neutral gummy fraction (4.6 gm) obtained after the evaporation of ether was chromatographed over alumina (200 gm, deactivated with 8 ml of 10% aqueous AcOH). Elution with petroleum yielded a fraction m.p. 244–48° (0.85 gm), which on crystallization from CHCl₃-MeOH mixture afforded pure friedelin, m.p. 259–61°, (α)_D -36° identical with an authentic specimen (m.m.p. and IR). (Found: C, 84·18; H, 11·76. Calc. for C₃₀H₃₀O: C, 84.44; H, 11·81%). Further elution of the column with petroleum: benzene (3:2) gave a solid (1·21 gm) m.p. 130–32° which on crystallization from CHCl₃-MeOH mixture gave pure β -sitosterol, m.p. 135–37°, (α)_D -40°, acetate, m.p. 128–29°, (α)_D -38°, identical with an authentic specimen (m.m.p. and IR). (Found: C, 81·47; H, 11·84. Calc for C₃₁H₃₂O₂: C, 81·52; H, 11·48%).

Hydrolysis of acetoxy methyl aleuritolate (I) and preparation of methyl aleuritolate (II) To a soln of I (0.20 gm) in benzene (10 ml), 10% methanolic KOH (30 ml) was added and the reaction mixture was refluxed for 4 hr. After usual work up and crystallization from CHCl₃-MeOH it gave pure II, m.p. 208-10°, $(\alpha)_D + 11 \cdot 11°$, $\nu \underset{\text{Mex}}{\text{Mex}} \text{ cm}^{-1}$ (--OH), 1735 cm⁻¹ (--COOCH₃), 820 cm⁻¹ (trisubstituted double bond). (Found: C, 78.92; H, 10.56. C₃₁H₅₀O₃ reqires: C, 79.10; H, 10.71%).

 CrO_3 -Pyridine oxidation of methyl aleuritolate (II) and preparation of methyl aleuritolonate (III). A soln of II (0.20 gm) in pyridine (6 ml) cooled to 15° was added to a CrO_3 -Pyridine complex,⁶ prepared from CrO_3 (0.20 gm) and pyridine (2 ml) at 15° and the reaction mixture was allowed to stand at room temp for 18 hr. Excess of CrO_3 was decomposed by addition of MeOH (5 ml). The mixture was then digested with EtOAc and filtered. The filtrate after working up as usual, yielded a solid (0.19 gm), which was chromatographed over activated alumina (20 gm). Elution with petroleum afforded a solid (0.12 gm), m.p. 171-74°, which after crystallization from $CHCl_3$ -MeOH mixture furnished III (0.08 gm), m.p. 174-76°, (α)_D + 11.76°, ν max 1705 cm⁻¹ (six membered ring ketone), 1735 cm⁻¹ (--COOCH₃), 820 cm⁻¹ (trisubstituted double bond). (Found: C, 79.41; H, 10.32. $C_{31}H_{45}O_3$ requires: C, 79.48; H, 10.45%).

Hydrolysis of methyl aleuritolate and preparation of aleuritolic acid (IV). To a normal soln of t-BuOK in t-BuOH (prepared from 0.4 gm of K in 10 ml dry t-BuOH) a soln of methyl aleuritolate (0.15 gm) in DMSO (10 ml) was added and the reaction mixture was heated on an oil bath at 105° for 4 hr.⁷ The reaction mixture was then cooled, diluted with water and acidified with dil HCl. The solid that separated out was taken up in CHCl₃, which after being washed with water was dried (Na₂SO₄). Removal of the solvent gave an amorphous solid which after crystallization from CHCl₃-MeOH mixture gave a solid, m.p. 300-302° (dec), $v_{\text{CHC}^3}^{\text{CHC}_3}$ 3400 cm⁻¹ (--OH), 1700 cm⁻¹ (COOH), 820 cm⁻¹ (trisubstituted double bond). (Found: C, 78.84; H, 10.38. C₃₀H₄₄O₃ requires: C, 78.94; H, 10.52%).

Acetyl aleuritolic acid. Aleuritolic acid IV on acetylation by Ac_2O -Pyridine method gave the acetyl derivative which on crystallization from CHCl₃-MeOH mixture gave crystalline acetyl aleuritolic acid m.p. 278-81°. (Found: C, 79.32; H, 10.11. $C_{32}H_{50}O_4$ requires: C, 79.66; H, 10.37%).

Preparation of aleuritolonic acid from III. Methyl aleuritolonate (0.15 gm) was hydrolysed by the same method described above.⁷ The product after crystallization from CHCl₃-MeOH mixture gave crystalline aleuritolonic acid, m.p. 280-82°. (Found: C, 79.20; H, 10.30. $C_{30}H_{46}O_3$ requires: C, 79.29; H, 10.18%).

Isomerization of acetyl methyl aleuritolate and preparation of acetyl methyl oleanolate. Compound I (0.20 gm) was isomerized by heating for 15 min with cone HCl and AcOH. The crystalline solid obtained after usual work up on crystallization from CHCl₃-MeOH mixture afforded pure acetyl methyl oleanolate (0.17 gm), m.p. 219-20°, $(\alpha)_{\rm D}$ + 58.82° identical with an authentic sample (m.m.p. and IR). (Found: C, 76.86; H, 9-83. Calc for C₃₃H₃₂O₄; C, 77.34; H, 10.15%).

Perbenzoic acid titration of acetyl methyl aleuritolate I. Compound I (0.0548 gm) was titrated with I.22 N perbenzoic acid in CHCl₃. It took up one mole equivalent of perbenzoic acid within 24 hr with no further uptake indicating the presence of one double bond.

LAH reduction of methyl aleuritolate (II). To a soln of II (0.15 gm) in dry dioxan (15 ml) was added LAH (0.075 gm) and the mixture was heated on a water bath for 4 hr. After the reaction, excess LAH was destroyed by careful addition of moist ether and then with a satd Na₂SO₄aq. The ethereal soln was washed with water and dried (Na₂SO₄). On removal of the solvent, a solid was obtained which after crystallization from CHCl₃-MeOH gave pure crystals of V, m.p. 265-7°, identical with an authentic sample (m.m.p. and IR). (Found: 81.23; H, 10.51. Calc for C₃₀H₃₀O₂; C, 81.39; H, 11.38%). Myricadiol diacetate VI. Acetylation of V with Ac₂O-pyridine in the usual manner afforded pure crystalline VI, m.p. 251-52°, $(\alpha)_D - 3^\circ$, identical with an authentic sample. (Found: C, 78-04; H, 9-89. Calc for C₁₄H₁₄O₄: C, 77-56; H. 10-26%).

Acknowledgements—The authors' thanks are due to Prof. J. N. Chatterjee of Patna Science College, Patna for IR spectra and to Dr. T. R. Govindachari of Ciba Research Centre, Goregaon, Bombay for NMR spectra recorded in the paper. Thanks are also due to Prof. P. Sengupta of Kalyani University, Kalyani, West Bengal and to Prof. P. K. Bose of Bose Institute, Calcutta for authentic samples of acetyl methyl oleanolate and myricadiol and myricadiol diacetate respectively. One of us (DRM) is indebted to East India Pharmaceutical Works, Calcutta for the award of a fellowship.

REFERENCES

- ¹ Terpenoids and related compound—Part X, D. R. Misra and H. N. Khastgir, J. Ind. Chem. Soc. 46, 843 (1969)
- ² a J. D. Hooker, Flora of British India Vol. V, p. 239, reprint (1956);
 ^b Kirtikar and Basu, Indian Medicinal Plants (2nd Edition) Vol. III; p. 2247. Published by L. M.
 - Basu (1963); ^c D. Prain, Bengal Plants Vol. II, p. 705. Botanical Survey of India (1963)
- ³ ^a Drake and Jacobsen, J. Am. Chem. Soc. 57, 1570, 1834 (1935);
- ^b P. R. Jefferies, J. Chem. Soc. 473 (1954)
- ⁴ Sir Ian Heilbron, A. H. Cook, H. M. Bunbury and D. H. Hey, *Dictionary of Organic Compounds* Vol. V, p. 2902, Eyre and Spottiswoode, Revised edition (1965)
- ⁵ ^a G. S. Davy, T. G. Halsall and E. R. H. Jones, J. Chem. Soc. 2696 (1951);
 ^b C. Djerassi, L. H. Liu, E. Farkas, A. E. Lippman, A. J. Lemin, L. E. Geller, R. N. McDonald and B. J. Taylor, J. Am. Chem. Soc. 77, 1200 (1955)
- ⁶ G. I. Poos, G. E. Arth, R. E. Beyler and L. H. Sarett, Ibid., 75, 427 (1953)
- ⁷ F. C. Chang and N. F. Wood, Tetrahedron Letters No. 40, 2969 (1964)
- ⁸ J. M. Beaton and F. S. Spring, J. Chem. Soc. 2131 (1955)
- ⁹ J. Simonsen and W. C. J. Ross, *The Terpenes* Vol. V, p. 201. Cambridge University Press, Cambridge (1957)
- ¹⁰ H. Budzikiewiez, J. M. Wilson and C. Djerassi, J. Am. Chem. Soc. 85, 3688 (1963)
- ¹¹ A. A. Ryabinin and L. G. Matyukhina, Dokl. Akad. Nauk SSSR 129, 125 (1958)
- ¹² A. A. Ryabinin and L. G. Matyukhina, Ibid. 131, 316 (1959)
- ¹³ Buddha Dev Paul and P. K. Bose, J. Ind. Chem. Soc. 44, 659 (1967)
- ¹⁴ K. P. Agarwal, A. C. Roy and M. L. Dhar, Ind. J. Chem. 1, 28 (1963)