the extract washed with H<sub>2</sub>O and dried (Na<sub>2</sub>SO<sub>4</sub>). Concentration of the Et<sub>2</sub>O extract left a colorless solid which was recrystallized from Me<sub>2</sub>CO to give 5 (400 mg, 72%) as a colorless solid: mp 124°; ir and nmr as expected. Anal. (C<sub>29</sub>H<sub>49</sub>IO) C, H. **19-Iodositost-5-en-3** $\beta$ -01-<sup>125</sup>I. A solution of 5 (50 mg) and Na<sup>125</sup>I

19-Iodositost-5-en- $3\beta \cdot 6\dot{1}^{.125}I$ . A solution of 5 (50 mg) and Na<sup>125</sup>I (3.8 mCi) in Me<sub>2</sub>CO (4 ml) was heated to gentle reflux for 4 hr. The solution was allowed to cool and cold H<sub>2</sub>O was added slowly when a solid separated. The solid was collected by filtration and recrystallized from Me<sub>2</sub>CO to give 32 mg of 19-iodo- $\beta$ -sitosterol-<sup>126</sup>I with a specific activity of 66.0  $\mu$ Ci/mg (55% exchange). The using CHCl<sub>3</sub>-EtOH (1:1) ( $R_{\rm f}$  0.64) or C<sub>6</sub>H<sub>6</sub>-EtOAc (1:1) ( $R_{\rm f}$  0.43) showed a single spot co-incident with a single radioactive peak appearing on the radiochromatogram scan.

Tissue Distribution Studies. Radioiodinated steroids were given by intraperitoneal injection to immature male Sprague-Dawley albino rats weighing 175-200 g. The dose administered was approximately 50  $\mu$ Ci per rat and the vehicle used was 90% EtOH (0.2-0.3 ml). Groups of two animals were killed by exsanguination through the ventricle 24, 48, 96, and 144 hr after the injection. The major organs such as liver and kidney were excised, weighed, and homogenized. These organs were washed thoroughly with isotonic saline to remove blood, dried, and minced with scissors. Minced tissue was then placed in a homogenizer tube containing 20 ml of H<sub>2</sub>O in the case of liver and 3 ml of H<sub>2</sub>O in the case of kidney. Homogenates were not prepared for small organs such as adrenal and thyroid. Liver and kidney homogenates and entire adrenal, thyroid, and several heparinized blood samples were placed in scintillation counting vials. To each vial 0.5 ml of 10% NaOH solution was added and left overnight and then heated for at least 10 min at 60° in a water bath to complete digestion. The vials were allowed to cool and 5 drops each of glacial HOAc and 30% H<sub>2</sub>O<sub>2</sub> solution were added. Ten milliliters of thixotropic liquid-counting system<sup>10</sup> was then added to each vial and the contents were shaken using a vortex mixer. The vials were kept in a cool dark place for at least 4 hr before counting. Radioactivity was assayed in a Beckman LS-200 liquid scintillation spectrometer. Sufficient counts were accumulated to reduce the probable error of counting to less than 5%. All counts were corrected for quench by using <sup>125</sup>I-quench standards curves.

## References

- G. N. Holcomb, C. M. Boyd, R. E. Counsell, W. H. Beierwaltes, R. A. Szczesniak, D. R. Murty, and G. A. Bruno, J. Pharm. Sci., 60, 390 (1971).
- (2) (a) R. E. Counsell, V. V. Ranade, R. J. Blair, W. H. Beierwaltes, and P. A. Weinhold, *Steroids*, 16, 318 (1970); (b) R. J. Blair, W. H. Beierwaltes, L. M. Lieberman, C. N. Boyd, R. E. Counsell, P. A. Weinhold, and V. M. Varma, *J. Nucl. Med.*, 12, 176 (1971).
- (3) L. M. Lieberman, W. H. Beierwaltes, J. W. Conn, A. N. Ansari, and H. Nishiyama, N. Engl. J. Med., 285, 1387 (1971).
- (4) J. W. Conn, R. Morita, E. L. Cohen, W. H. Beierwaltes, W. J. McDonald, and K. Herwig, Arch. Int. Med., 129, 417 (1972).
- (5) W. H. Beierwaltes, L. M. Lieberman, A. N. Ansari, and H. Nishiyama, J. Amer. Med. Ass., 216, 275 (1971).
- (6) R. Morita, L. M. Lieberman, W. H. Beierwaltes, J. W. Conn, A. N. Ansari, and H. Nishiyama, J. Clin. Endocrinol. Metab., 34, 36 (1972).
- (7) T. Nagai, B. A. Solio, and C. S. Koh, J. Nucl. Med., 9, 576 (1968).
- (8) E. Denot, C. Casas-Campillo, and P. Crabbe, Eur. J. Steroids, 2, 495 (1967).
- (9) A. Bowers, R. Villotti, J. A. Edwards, E. Denot, and D. Halpern, J. Amer. Chem. Soc., 84, 3204 (1962).
  (10) E. P. Frenkel, B. E. Whalley, C. T. Knorp, and D. R. Korst,
- (10) E. P. Frenkel, B. E. Whalley, C. T. Knorp, and D. R. Korst, J. Lab. Clin. Med., 59, 174 (1962).

## Substituted Tetralines. 5. Analgesic Properties of Some Diastereoisomeric N,N-Dimethyl-4-phenyl-1,2,3,4-tetrahydro-2-naphthylamines<sup>1</sup>

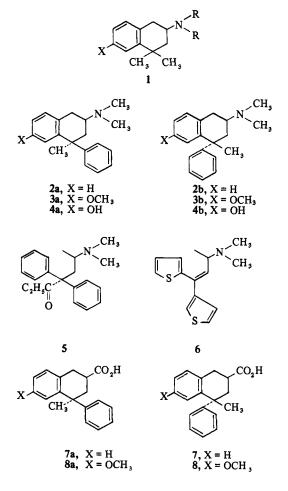
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In a series of recent publications the synthesis and analgesic potency of some derivatives of general structure 1were reported.<sup>2-4</sup> As part of a continuing study of structure-activity relationships directed toward the investigation of the effect of 4 substitutions in this system, the diastereoisomeric N,N-dimethyl-4-phenyl-1,2,3,4-tetrahydro-2naphthylamines 2a and 2b and their 6-methoxyl and 6hydroxyl derivatives 3a,b and 4a,b, respectively, were prepared and tested for analgesic activity.

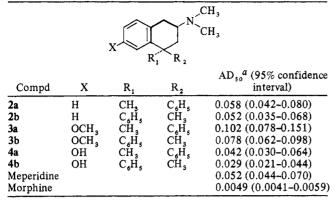
Substitution of a phenyl group for methyl in 1 provides analogs of the diaryl analgesics, *e.g.*, methadone (5) and thiambutene (6), wherein the conformation of one of the phenyl rings is partially restricted. If it is assumed that the tetralin ring system (present in virtually all potent analgesics having rigid structures<sup>5</sup>) serves in the role as a determinant of analgesic potency proposed by Beckett and Casy,<sup>6</sup> then 4-phenyl-substituted tetralins might be useful for investigating the importance of the second phenyl group of diaryl analgesics.

**Chemistry**. The amines **2a**, **2b**, **3a**, and **3b** were prepared from the corresponding 4-methyl-4-phenyl-1,2,3,4-tetrahydro-2-naphthoic acids **7a**, **7b**, **8a**, and **8b** by means of a modified Curtius procedure,<sup>7</sup> followed by Eshweiler-Clarke methylation<sup>8</sup> of the primary amines. The synthesis, separation, and proof of stereostructure of the diastereoisomeric acids, **7** and **8**, have been reported previously.<sup>9</sup> Treatment of the *N*,*N*-dimethyl-6-methoxy-4-methyl-4phenyl-1,2,3,4-tetrahydro-2-naphthylamines **3a** and **3b** with 48% HBr gave **4a** and **4b**, respectively.



**Biological Results.** The analgesic potencies of the amines were determined in white albino mice (Carnworth Farm CF No. 1 strain) by the hot-plate procedure of Eddy and Leimbach.<sup>10</sup> In the complete assay procedure for each compound, five groups of six mice each were screened at dosage levels of 0.025, 0.050, 0.075, and 0.100 mmol/kg and a normal

Table I. Analgesic Potencies of Tetrahydro-2 naphthylamines



<sup>a</sup>Expressed as mmol/kg.

saline control was administered intraperitoneally. Relative potencies, expressed as the  $AD_{50}$  (the dose of compound that doubled the normal reaction time in 50% of the mice tested), were calculated by the method of Litchfield and Wilcoxon.<sup>11</sup> From the data (Table I) it is seen that compounds 2a and 2b are comparable in potency to meperidine in mice. Substitution of a 6-hydroxy group (4a and 4b), corresponding to the 3-hydroxy in morphine, increases potency, whereas 6-methoxyl substitution (3a and 3b) decreases potency.

The lack of significant differences in activity between any of the diastereoisomeric (cis and trans) pairs of amines (2, 3, or 4) indicated that analgesic activity is apparently not dependent on a single specific conformation/configuration of the 4-phenyl group of these compounds. This result is paralleled in previous studies of conformationally restricted analgesics of the meperidine,<sup>12</sup> prodine,<sup>13</sup> and benzomorphan<sup>14</sup> types. The observed minimal conformational/configurational requirements for these compounds may be rationalized in terms of different modes of interaction of analgesic agents with receptors<sup>15</sup> and/or multiple receptors with different structural specifications.<sup>16</sup>

## Experimental Section<sup>7</sup>

cis-N.N-Dimethyl-4-methyl-4-phenyl-1.2.3.4-tetrahydro-2naphthylamine Hydrochloride (2a). A mixture of 8 g (0.03 mol) of  $7a^9$  and freshly distilled SOCl<sub>2</sub> was refluxed 4 hr. The excess SOCl<sub>2</sub> was removed in vacuo (aspirator) leaving the acid chloride  $[\nu_{max} (CHCl_3) 1802 \text{ cm}^{-1}]$ . The acid chloride was dissolved in 50 ml of dry reagent grade acetone. The solution was chilled in an ice bath and, with vigorous stirring, a chilled solution of 2 g (0.03)mol) of NaN<sub>3</sub> in 6 ml of  $H_2O$  was added over a period of 1 min. The mixture was stirred for 45 min, then 100 ml of cold H<sub>2</sub>O was added, and the crude azide was extracted with dry xylene. The extract was washed once with saturated NaCl solution and then filtered through anhydrous Mg<sub>2</sub>SO<sub>4</sub>. Rearrangement of the azide to the isocyanate was accomplished by applying vacuum (aspirator) to the xylene solution contained in a 500-ml flask and stirring vigorously with a magnetic stirrer The evolution of N<sub>2</sub> was in evidence for a period of about 3 hr. The vacuum was then removed, the flask fitted with a reflux condensor, and the solution heated to the boiling point. Xylene was then distilled off in vacuo yielding the isocyanate  $[\nu_{max} (CHCl_3) 2272 \text{ cm}^{-1}]$ . The nmr spectrum exhibited a single CH<sub>3</sub> absorption of  $\delta$  1.72 ppm, indicating that no epimerization had taken place. Concentrated HCl (50 ml) was cautiously added to the isocyanate and, after the exothermic reaction subsided, the brownish solution was refluxed overnight. Distilled H<sub>2</sub>O was then added and the solution extracted with ether. The acidic aqueous solution was then evaporated and the residue dissolved in 10% KOH solution and extracted twice with Et<sub>2</sub>O. The ether extracts

†Melting points (uncorrected) were determined on a Fisher-Johns melting block. Microanalyses were performed by the Galbraith Laboratories, Knoxville, Tenn., and by the Weiler and Strauss Microanalytical Laboratory, Oxford, England. Ir spectra were obtained on Beckman IR-5 and IR-33 spectrophotometers. Nmr spectra were obtained with a Varian T-60 spectrometer. were combined and dried over anhydrous  $K_2CO_3$  and the Et<sub>2</sub>O was distilled to yield the primary amine. The crude primary amine was then dissolved in 10 ml (0.2 mol) of 90% HCOOH, 10 ml of 37% HCHO was then added, and the mixture refluxed for 12 hr. After the mixture cooled, 5 ml of concentrated HCl was added and the mixture evaporated *in vacuo* (aspirator) to dryness. To the residue was added 10% KOH and the mixture was extracted twice with Et<sub>2</sub>O. The Et<sub>2</sub>O solution of the amine causing 4.0 g (45%) of the HCl salt **2a** to precipitate as yellowish white needles, mp 243-245° [from CHCl<sub>3</sub>-(CH<sub>3</sub>CH<sub>2</sub>)<sub>2</sub>O]. Anal. (C<sub>19</sub>H<sub>24</sub>CIN) C, H, N.

trans-N,N-Dimethyl-4-methyl-4-phenyl-1,2,3,4-tetrahydro-2naphthylamine Hydrochloride (2b). Following the same procedure employed for the preparation of 2a, 8 g of 7b<sup>9</sup> gave 3.8 g (45%) of 2b as yellowish white needles, mp 246-248° [from CHCl<sub>3</sub>-(CH<sub>3</sub>CH<sub>2</sub>)<sub>2</sub>O]. Anal. (C<sub>19</sub>H<sub>24</sub>ClN) C, H, N.

cis-N,N-Dimethyl-6-methoxy-4-methyl-4-phenyl-1,2,3,4-tetrahydro-2-naphthylamine Hydrochloride (3a). Similar procedures employed for the synthesis of 2a and 2b were used to prepare 3a and 3b. Thus, 8.5 g (0.02 mol) of 8a<sup>9</sup> was converted to the acid chloride [ $\nu_{max}$  (CHCl<sub>3</sub>) 1792 cm<sup>-1</sup>] and thence to the azide, which was then rearranged to the isocyanate [ $\nu_{max}$  (CHCl<sub>3</sub>) 2260 cm<sup>-1</sup>]. The isocyanate was hydrolyzed in 90% HCOOH (concentrated HCl appeared to cause some cleavage of the methoxyl group in a previous reaction) to a primary amine which was then N,N-dimethylated and converted to the HCl salt, giving 4.0 g (40%) of 3a, mp 188-189° (sealed tube) following recrystallization [CHCl<sub>3</sub>-(CH<sub>3</sub>CH<sub>2</sub>)<sub>2</sub>O] Anal. (C<sub>20</sub>H<sub>26</sub>ClNO) C, H, N. trans-N,N-Dimethyl-6-methoxy-4-methyl-4-phenyl-1,2,3,4-tetra-

trans. N.N. Dimethyl-6-methoxy-4-methyl-4-phenyl-1,2,3,4-tetrahydro-2-naphthylamine Hydrochloride (3b). Following procedures for the synthesis of 3a, 8.5 g (0.025 mol) of 8b<sup>9</sup> gave 4.0 g (40%) of 3b, mp 223° (sealed tube) following recrystallization [CHCl<sub>3</sub> – (CH<sub>3</sub>CH<sub>2</sub>)<sub>2</sub>O]. Anal. (C<sub>20</sub>H<sub>26</sub>CINO) C, H, N.

cis-N,N-Dimethyl-6-hydroxy-4-methyl-4-phenyl-1,2,3,4-tetrahydro-2-naphthylamine Hydrobromide (4a). A solution of 1.65 g (0.005 mol) of 3a in 15 ml of 48% HBr was refluxed under a stream of N<sub>2</sub> until evolution of CH<sub>3</sub>Br (as evidenced by a cloudy appearance to the N<sub>2</sub> stream) ceased. After about 1 hr of reflux, H<sub>2</sub>O and excess HBr were removed *in vacuo* (aspirator) and the residue was recrystallized (EtOH-EtOAc) giving a nearly quantitative yield of 4a as colorless needles, mp 293°. Anal. (C<sub>19</sub>H<sub>24</sub>BrNO) C, H, N.

trans-N,N-Dimethyl-6-hydroxy-4-methyl-4 phenyl-1,2,3,4-tetrahydro-2-naphthylamine Hydrobromide (4b). Following the procedure for the synthesis of 4a, 1.65 g (0.005 mol) of 3b gave a quantitative yield of 4b as pale yellow needles, mp 318°. Anal. ( $C_{19}H_{24}BrNO$ ) C, H, N; C: calcd, 62.98; found, 63.22.

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## References

- Presented in part at the 163rd National Meeting of the American Chemical Society, Boston, Mass., April 1972.
- (2) A. R. Martin, A. P. Parulkar, D. J. Gusseck, L. J. Anderson, G. L. Grunewald, and A. I. White, *J. Pharm. Sci.*, 58, 340 (1969).
- (3) D. E. Green, A. R. Martin, and A. I. White, *ibid.*, **59**, 526 (1970).
- (4) V. S. Pai, A. P. Parulkar, A. R. Martin, and A. I. White, *ibid.*, 60, 201 (1971).
- (5) A. H. Beckett and A. F. Casy, Progr. Med. Chem., 4, 171 (1965).
- (6) A. H. Beckett and A. F. Casy, J. Pharm. Pharmacol., 6, 986 (1954).
- (7) P. A. Smith, "Organic Reactions," Collect. Vol. III, Wiley, New York, N. Y., 1946, p 337.
- (8) A. Burger and W. Yost, *J. Amer. Chem. Soc.*, 70, 2198 (1948).
  (9) E. M. Kandeel, L. J. Anderson, J. H. Block, A. I. White, and
- A. R. Martin, J. Pharm. Sci., 61, 1231 (1972).
- (10) N. B. Eddy and D. Leimbach, J. Pharmacol. Exp. Ther., 107, 385 (1953).
- (11) J. T. Litchfield, Jr., and F. Wilcoxon, ibid., 96, 99 (1949).
- (12) P. S. Portoghese, A. A. Mikhail, and H. J. Kupferberg, J. Med. Chem., 11, 219 (1968).
- (13) E. E. Smissman and M. J. Steinman, ibid., 9, 455 (1966).
- (14) A. E. Jacobson and E. L. May, ibid., 7, 409 (1964).
- (15) P. S. Portoghese, ibid., 8, 147 (1965).
- (16) S. Archer and L. S. Harris, Progr. Drug Res., 8, 262 (1965).