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Morphology Control of Fluorescent Nanoaggregates by Co-Self-Assembly of Wedge- and Dumbbell-Shaped Amphiphilic Perylene Bisimides

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Perylene bisimides (PBIs) have been extensively studied as building blocks for functional supramolecular architectures in nonpolar solvents through hydrogen bonding, metal ion coordination, and $\pi - \pi$ stacking interaction. In contrast, little attention was paid to the self-assembly behavior of PBIs in aqueous solution.² Amphiphilic molecules, consisting of hydrophobic and hydrophilic moieties, self-assemble into highly organized aggregates with various morphologies such as spherical micelles, wormlike micelles, spherical and hollow vesicles, planar bilayers, nanotubes, and others.3 The formation of these morphologies depends on solvent environments, molecular structures and shapes, as well as the relative fraction of hydrophilic and hydrophobic parts.⁴ On the other hand, the stabilization of self-assembled superstructures through covalent linkage, for example, by polymerization, is of great interest for the practical application of supramolecular chemistry.⁵ By covalent stabilization, the aggregate dimension and shape can be captured, and aggregate strength can be increased.⁶ Here, we report for the first time that the nanoaggregates of perylene bisimides with particular morphology can be obtained by self-assembly of differently shaped amphiphilic PBIs in aqueous solution.

Chart 1

In this work, the wedge- and dumbbell-shaped amphiphilic perylene bisimides PBI **1–4** (Chart 1) were synthesized and fully characterized (see details in Supporting Information). The absorption and fluorescence spectra of molecularly dissolved PBI **1–4** display a mirror-image relationship with a small Stokes' shift (5–6 nm) in "good" solvents such as CH₂Cl₂ and THF (Figure S-15 in Supporting Information). Upon addition of the "bad" solvent water into the THF solution, a gradual decrease in fluorescence intensity at 500–600 nm was observed as exemplified for PBI **1** (Figure 1), and a new broad and structureless fluorescence band appears at 600–800 nm owing to excimer formation. These observations imply that monomeric PBI **1** self-assembles into fluorescent multimolecular aggregates in aqueous solution.

For the wedge-shaped PBI 1, Israelachvili's critical packing parameter $(P_c)^8$ was calculated to be 0.252, implying that spherical micelles should be favorably formed in polar solvents $(P_c < ^1/_3)$ for micelle formation). The theoretical prediction was confirmed by transmission electron microscopic (TEM) studies. A large number of spherical micelles with the diameter of 4–6 nm were observed for PBI 1 in THF-containing water (2%, v/v) as shown in Figure 2a. The micelles were formed with a narrow polydispersity and a

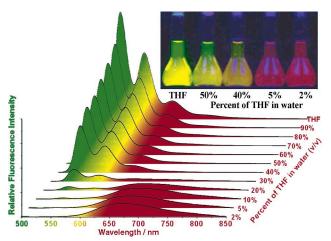


Figure 1. Fluorescence spectra of PBI 1 in THF and THF-containing water, $\lambda_{\rm ex}=490$ nm, [PBI 1] = 2×10^{-6} M. The inset shows a photograph of PBI 1 in THF and THF-containing water under UV-light irradiation.

high degree of curvature due to the conical structure. The same aggregation behavior was also observed for PBI **2**. In contrast, dumbbell-shaped PBI **4** self-assembled into rod aggregates with a diameter of 4 nm (Figure 2b). The formation of well-defined aggregates of these PBIs may be attributed to the $\pi-\pi$ stacking interaction between the perylene cores along one-dimensional long axis without curvature, and stabilized by surrounding hydrophilic chains as shown in Scheme 1(bottom).

More interestingly, hollow vesicles were observed for the co-selfassembled system of PBI 1 and PBI 3 ([PBI 1]/[PBI 3] = 8/1 in molar ratio) in THF-containing water (2%, v/v) as shown in Figure 2c. Spherical vesicles were formed with the average diameter of 94 nm and the wall thickness of 7-8 nm, which is approximately twice the length of a single optimized molecule PBI 1 (3.3 nm) or PBI 3 (4.0 nm) in water. The aggregation number for the inner and outer layers were calculated to be approximate 2.9×10^5 and 3.2×10^5 , respectively, from the bilayer volume divided by the volume of the single molecule. For the co-self-assembled system with higher PBI 3 content ([PBI 1]/[PBI 3]= 4/1), bilayer vesicles were observed with a larger average diameter of 133 nm in THF-containing water (2%, v/v) as shown in Figure 2e. The size distribution of these coaggregates also became broader at higher PBI 3 content as measured from TEM (Figure 2g,h) and dynamic light scattering (DLS) (Figure S-24), and these large vesicles emit red fluorescence (Figure 2f).

The observed variation of the aggregates' shape and size can be rationalized by considering spontaneous curvature. When dumbbell-shaped PBI 3 is co-self-assembled with wedge-shaped PBI 1, the average hydrophobic part increases, and the interface between the hydrophilic and hydrophobic parts changes from a curved interface to a more flat one. The resulting decrease in spontaneous curvature releases the strain in the initially formed micelles with a high curvature, leading to micelle growth and a transition to vesicles as

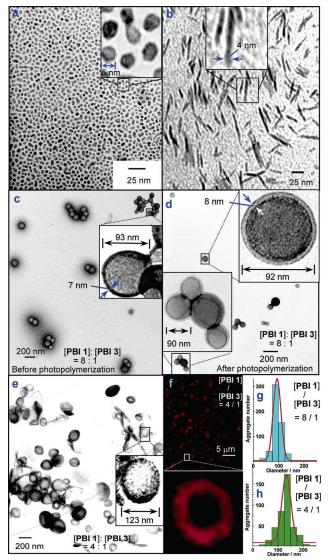
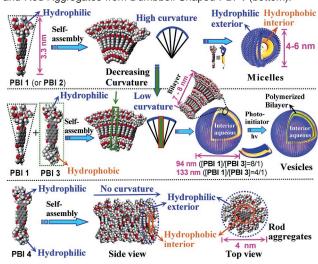


Figure 2. TEM images of self-assembled PBI 1 (a), PBI 4 (b), and before (c) and after (d) photopolymerization of co-aggregates of PBI 1 and PBI 3 in a 8:1 molar ratio; TEM (e) and confocal fluorescence images (f) of co-aggregates of PBI 1 and PBI 3 in a 4:1 molar ratio in THF-containing water (2%, v/v); and size distribution obtained from 548 co-aggregates ([PBI 1]/[PBI 3] = 8/1) (g), and from 402 co-aggregates ([PBI 1]/[PBI 3] = 4/1) (h). [PBI 1] = 0.5 mg/mL (4.32 × 10⁻⁴ M); [PBI 4] = 0.5 mg/mL.

illustrated in Scheme 1 (top and middle). It is interesting to note that such mechanism for the control of self-assembly by spontaneous curvature is present in biology. A typical example is the deformation of flat lipid membranes by protein coating into transport spherical vesicles.⁹

To stabilize these nanoaggregates, the in situ photopolymerization was performed in an organized state using 2,2-dimethoxy-2-phenylacetophenone as photoinitiator under UV irradiation at 350 nm. This wavelength was chosen because the photoinitiator has a maximum absorption at 350 nm, where perylene chromophores have a very weak absorption. After photopolymerization, the vesicles from co-self-assembly retained their original shapes as shown in Figure 2d. The photopolymerization was confirmed by infrared (IR) studies. The vibration band at 968.1 cm⁻¹ (characteristic vibration of $\delta(=CH_2)$ wagging mode) disappears after photopolymerization (Figure S-23). These vesicles do not change their morphology after the addition of PBI 3, confirming that the vesicle structures are locked by photopolymerization.

Scheme 1. Schematic Illustration for the Formation of Micelles from Wedge-Shaped PBI 1 (top), Bilayer Vesicles from the Co-self-assembly of PBI 1 and Dumbbell-Shaped PBI 3 (middle), and Rod Aggregates from Dumbbell-Shaped PBI 4 (bottom).



In summary, the aggregate morphologies of amphiphilic PBIs are dependent on their molecular shapes. The co-self-assembly of wedge-shaped PBI 1 and dumbbell-shaped PBI 3 generated hollow vesicles owing to the changes in spontaneous curvature. The bilayer structures of the vesicles could be stabilized by in situ photopolymerization. The morphology changes by co-self-assembly of different molecular architectures provide a guideline for the rational design of particular morphology such as vesicles.

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Supporting Information Available: Synthetic details, spectroscopic characterization, UV—vis absorption, fluorescence and 2D NMR spectra of amphiphilic perylene bisimides, TEM images, and DLS data of aggregates. This material is available free of charge via the Internet at http://pubs.acs.org.

References

- (1) Würthner, F. Chem. Commun. 2004, 1564-1579.
- (2) (a) Abdalla, M. A.; Bayer, J.; R\u00e4dler, J. O.; M\u00fcllen, K. Angew. Chem., Int. Ed. 2004, 43, 3967-3970. (b) Wang, W.; Wan, W.; Zhou, H. H.; Niu, S. Q.; Li, A. D. Q. J. Am. Chem. Soc. 2003, 125, 5248-5249.
- (3) (a) Percec, V.; Dulcey, A. E.; Balagurusamy, V. S. K.; Miura, Y.; Smidrkal, J.; Peterca, M.; Nummelin, S.; Edlund, U.; Hudson, S. D.; Heiney, P. A.; Hu, D. A.; Magonov, S. N.; Vinogradov, S. A. Nature 2004, 430, 764-768. (b) Antonietti, M.; Förster, S. Adv. Mater. 2003, 15, 1323-1333. (c) Hill, J. P.; Jin, W. S.; Kosaka, A.; Fukushima, T.; Ichihara, H.; Shimomura, T.; Ito, K.; Hashizume, T.; Ishii, N.; Aida, T. Science 2004, 304, 1481-1483. (d) Hartgerink, J. D.; Beniash, E.; Stupp, S. I. Science 2001, 294, 1684-1688.
- (4) (a) Uzun, O.; Sanyal, A.; Nakade, H.; Thibault, R. J.; Rotello, V. M. J. Am. Chem. Soc. 2004, 126, 14773–14777. (b) Arnt, L.; Tew, G. N. J. Am. Chem. Soc. 2002, 124, 7664–7665. (c) Percec, V.; Ahn, C. H.; Ungar, G.; Yeardley, D. J. P.; Möller, M.; Sheiko, S. S. Nature 1998, 391, 161–164.
- (5) Müller, A.; O'Brien, D. F. Chem. Rev. 2002, 102, 727-757.
- (6) (a) Yamamoto, T.; Fukushima, T.; Yamamoto, Y.; Kosaka, A.; Jin, W.; Ishii, N.; Aida, T. J. Am. Chem. Soc. 2006, 128, 14337–14340. (b) Zhang, X.; Li, Z. C.; Li, K. B.; Lin, S.; Du, F. S.; Li, F. M. Prog. Polym. Sci. 2006, 31, 893–948.
- (7) Würthner, F.; Chen, Z.; Dehm, V.; Stepanenko, V. Chem. Commun. 2006, 1188–1190.
- (8) (a) Israelachvili, J. *Intermolecular and Surface Forces*, 2nd ed.; Academic Press: San Diego, CA, 1991. (b) $P_{\rm c}=v/al$, where v is the effective molecular volume, a is the occupied area by hydrophilic chains, and l is optimal molecular length.
- (9) (a) Bigay, J.; Gounon, P.; Robineau, S.; Antonny, B. Nature 2003, 426, 563–566. (b) Lippincott-Schwartz, J.; Liu, W. Nature 2003, 426, 507–508.

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