## A Total Synthesis of Actinomycin D (C1) via Peptide Cyclization Between Proline and Sarcosine1

Syntheses of actinomycin D  $(C_1)^2$  have been reported by Brockmann and Lackner<sup>3</sup>, during which the key step, ring closure of the cyclic peptide lactones, was carried out by lactonization with the use of acetylimidazole/acetylchloride.

This communication concerns a synthesis of actinomycin D (Figure 1) during which a p-nitrophenyl ester intermediate served to cyclize the peptide lactone rings between proline and sarcosine<sup>4</sup>. The synthetic approach is outlined in Figure 2.

Treatment of the mixed anhydride from benzyloxy-carbonyl-L-N-methylvaline and isobutylchloroformate with tert-butyloxycarbonyl-L-threonine and catalytic hydrogenation of the crude product in the presence of palladium black gave the crystalline  $\beta$ -di-depsipeptide derivative, O-(L-N-methylvalyl)-N-tert-butyloxycarbonyl-L-threonine (I), 49%, m.p. 207°,  $[\alpha]_D^{23} + 57.1^\circ$  (c 1, methanol). Anal. calcd. for  $C_{15}H_{28}N_2O_6$  (332.4): C, 54.2; H, 8.49; N, 8.43; O, 28.9. Found: C, 54.2; H, 8.60; N, 8.41; O, 28.9. Condensation of I with benzyloxycarbonylsarcosine by the above mixed anhydride method afforded crystalline O-(benzyloxycarbonylsarcosyl-L-N-methylvalyl)-N-tert-butyloxycarbonyl-L-threonine (II),

52%, m.p. 163°,  $[\alpha]_{25}^{25}$  – 59.7° (c 1, methanol). Anal. calcd. for  $C_{26}H_{39}N_3O_{11}$  (537.6): C, 58.1; H, 7.31; N, 7.82; O, 26.8. Found: C, 58.2; H, 7.46; N, 7.69; O, 27.0.

Benzyloxycarbonyl-D-valyl-Lproline-tert-butyl ester (III) was obtained as colorless prisms [90%, m.p. 92°,  $[\alpha]_{\mathrm{D}}^{16}-19.6^{\circ}$  (c 1, methanol). Anal. calcd. for  $\mathrm{C_{22}H_{32}N_2O_5}$ (404.5): C, 65.4; H, 7.98; N, 6.93. Found: C, 65.4; H, 8.05; N, 7.06] from a reaction of benzyloxycarbonyl-D-valinep-nitrophenyl ester 11 with tert-butyl-L-prolinate 12,18. Catalytic hydrogenation of III in the presence of palladium black and subsequent reaction with the mixed anhydride from II and isobutylchloroformate afforded the  $\beta$ -pentadepsipeptide derivative, O-(benzyloxycarbonylsarcosyl-L-N-methylvalyl)-N-tert-butyloxycarbonyl-L-threonyl-Dvalyl-L-proline-tert-butyl ester (IV), which was purified by counter current distribution in the system: toluenechloroform-methanol-water (5:5:8:2)14, and then precipitated from ether/hexane at low temperature to give a colorless amorphous powder, 85%, m.p. 75-80°,  $[\alpha]_D^{28}$  $-41^{\circ} \pm 1^{\circ}$  (c 1, methanol). Anal. calcd. for  $C_{40}H_{63}N_5O_{11}$ (789.9): C, 60.8; H, 8.04; N, 8.87. Found: C, 60.9; H, 8.08; N, 9.23. A sample for amino acid analysis 15 was hydrolysed with 6N HCl at 110° for 24 h. The following molar ratios

Fig. 1. Structure of actinomycin D  $(C_1)^2$ .

were obtained, valine being taken as 1.0: sarcosine 1.0, proline 1.0, threonine 0.9, N-methylvaline 16 1.0. Cleavage by 4N HCl in dioxane (30 min at room temperature) of the tert-butyloxycarbonyl and tert-butyl ester protecting groups gave O-(benzyloxycarbonylsarcosyl-L-N-methylvalyl)-L-threonyl-D-valyl-L-proline hydrochloride (V), 95%, m.p. 160–165°,  $[\alpha]_D^{24}$  – 45.3° (c 1, methanol). Anal. calcd. for  $C_{31}H_{48}N_5O_9Cl$  (670.2): C, 55.6; H, 7.22; N, 10.4; Cl, 5.29. Found: C, 55.7; H, 7.25; N, 10.2; Cl, 5.17. V was reacted with 2-nitro-3-benzyloxy-4-methyl-benzoylchloride 17 in the presence of N-methylmorpholine. Purification of the crude product by column chromatography on Sephadex LH 20 in methanol afforded O-(benzyloxycarbonylsarcosyl-L-N-methylvalyl)-N-(2-nitro-3-benzyloxy-4-methyl-benzoyl)-L-threonyl-D-valyl-L-proline (VI), 89%, m.p. 115–120°,  $[\alpha]_D^{20}$  – 4.3° (c 0.5, methanol). Anal. calcd. for  $C_{46}H_{58}N_6O_{13}$  (903.0): C, 61.2; H, 6.47; N, 9.31. Found: C, 61.2; H, 6.65; N, 9.40. VI was converted to its p-nitrophenyl ester derivative (VII) with the use of di-pnitrophenyl sulfite in pyridine 18. VII was purified by column chromatography on Sephadex LH 20 in ethyl acetate followed by precipitation from benzene into hexane, 88%, m.p. 110-115° (dec.),  $[\alpha]_D^{20}$  -21.8° (c 0.5, dimethylformamide). Anal. calcd. for C52H61N7O15 (1024.1): C, 61.0; H, 6.00; N, 9.58. Found: C, 61.3; H, 6.59; N. 9.69. Treatment of VII with 4N hydrogen bromide in dioxane 19,20 removed the benzyloxycarbonyl group. The cyclization was carried out at high dilution (c about 0.05) in pyridine for 6 h at 60°21. A crude product was obtained by evaporating the solvent, dissolving the residue in ethyl acetate, successive washing with 1N HCl, and water, drying over MgSO4, and evaporating the solvent. Fractionation was carried out by column chromatography on Sephadex LH 20 in methanol. Evaporation of the fractions comprising the first major peak afforded cyclo-(2-nitro-3-hydroxy-4-methyl-benzoyl)-L-threonyl- ${\tt D-valyl-L-prolyl-sarcosyl-L-} N-methylvalyl \ lactone \ (VIII),$ which was obtained as an amorphous powder by precipitation from ethyl acetate into hexane, 31%,  $[\alpha]_D^{23}$  -14.5° (c 0.5, methanol). VIII was without further characterization converted into actinomycin D by catalytic hydrogenation in the presence of palladium black and subsequent oxidation using potassium ferricyanide22 in a mixture (1:1) of methanol and M/15 phosphate buffer at pH 7.1. Removal of the methanol under reduced pressure, extraction with ethyl acetate, washing of the organic phase with 1 M NaHCO<sub>3</sub>, 1 N HCl, and water, drying over Na<sub>2</sub>SO<sub>4</sub>, and evaporation gave crude actinomycin (IX), which was recrystallized from ethyl acetate by the addition of hexane. From 100 mg of VIII 76 mg (80%) of IX were obtained, orange red prisms, m.p. 240-242°,  $[\alpha]_D^{20}$  -312°  $\pm 10$ ° (c 0.26, methanol). Anal. calcd. for  $C_{62}H_{86}N_{12}O_{16}$  (1255.4): C, 59.3; H, 6.91; N, 13.4. Found: C, 59.3; H, 6.93; N, 13.4. Authentic natural crystalline actinomycin D²³ possessed m.p. 241–243°,  $[\alpha]_D^{20}$  – 323  $\pm$  10° (c 0.26, methanol)²⁴. Microbiological assays²⁵, using Lactobacillus arabinosus (ATCC 8014) and L. fermenti (ATCC 9388) in pantothenate- and thiamine-dependent systems respectively 26, showed that the synthetic material IX is indistinguishable from natural actinomycin D<sup>27</sup>.

Zusammenjassung. Es wird eine Totalsynthese von Actinomycin D  $\{C_1\}$  beschrieben, in der die Schlüsselreaktion, nämlich die Zyklisierung der Pentapeptidlaktonringe, durch eine Nitrophenylester-Synthese zwischen Prolin und Sarkosin ausgeführt wurde. Die Estergruppe zwischen der Carboxylgruppe des N-Methylvalins und der  $\beta$ -Hydroxylgruppe des Threonins wurde durch Reaktion

von Boc-Thr-OH mit dem gemischten Anhydrid aus Z-MeVal-OH und Chlorameisensäure-isobutylester hergestellt

J. MEIENHOFER

Children's Cancer Research Foundation, The Children's Hospital Medical Center and Harvard Medical School, Boston (Massachusetts 02115, USA), 24 May 1968.

- Abbreviations follow the rules of the IUPAC-IUB Commission on Biochemical Nomenclature, in Biochemistry 5, 1445, 2485 (1966); 6, 362 (1967); J. biol. Chem. 241, 2491 (1966).
- <sup>2</sup> Designation D is according to L. C. Vining and S. A. Waksman, Science 120, 389 (1954); designation C<sub>1</sub> is according to H. Brockmann and H. Gröne, Naturwissenschaften 41, 65 (1954).
- <sup>3</sup> H. BROCKMANN and H. LACKNER, Chem. Ber. 101, 1312 (1968); Naturwissenschaften 51, 384 (1964); 51, 435 (1964).
- <sup>4</sup> Ring closure via peptide bond formation between sarcosine and methylvaline during a synthesis of actinomycine C<sub>3</sub> was found by H. BROCKMANN and H. LACKNER, [Chem. Ber. 100, 353 (1967); Naturwissenschaften 47, 230 (1960); 48, 555 (1961)] to proceed in low yield.
- <sup>5</sup> J. R. VAUHGAN JR., J. Am. chem. Soc. 73, 3547 (1951).
- <sup>6</sup> P. A. Plattner, K. Vogler, R. O. Studer, P. Quitt and W. Keller-Schierlein, Helv. chim. Acta 46, 927 (1963).
- E. WÜNSCH and G. WENDLBERGER, Chem. Ber. 97, 2504 (1964).
  K. HOFMANN, R. SCHMIECHEN, R. D. WELLS, Y. WOLMAN and
- <sup>8</sup> K. Hofmann, R. Schmiechen, R. D. Wells, Y. Wolman and N. Yanaihara, J. Am. chem. Soc. 87, 611 (1965).
- <sup>9</sup> E. Schnabel, Justus Liebigs Annln Chem. 702, 188 (1967).
- <sup>10</sup> D. Ben-Ishai and E. Katchalski, J. Am. chem. Soc. 74, 3688 (1952).
- <sup>11</sup> J. MEIENHOFER, J. org. Chem. 32, 1143 (1967).
- <sup>12</sup> G. W. Anderson and F. M. Callahan, J. Am. chem. Soc. 82, 3359 (1960).
- <sup>13</sup> R. Roeske, J. org. Chem. 28, 1251 (1963).
- <sup>14</sup> C. H. Li, J. Meienhofer, E. Schnabel, D. Chung, T.-B. Lo and J. Ramachandran, J. Am. chem. Soc. 82, 5760 (1960); 83, 4449 (1961).
- <sup>15</sup> D. H. SPACKMAN, W. H. STEIN and S. MOORE, Analyt. Chem. 30, 1190 (1958).
- <sup>16</sup> N-Methylvaline produced under standard conditions a very weak ninhydrin reaction [compare: M. Ebata, Y. Takahashi and H. Otsuka, J. Chromat. 25, 1 (1966)] and overlapped with sarcosine. It was, therefore, determined by the method of J. Heilmann, J. Barollier and E. Watzke, Hoppe-Seyler's Z. physiol. Chem. 309, 219 (1957).
- <sup>17</sup> H. Brockmann and H. Muxfeldt, Chem. Ber. 91, 1242 (1958).
- <sup>18</sup> B. ISELIN, W. RITTEL, P. SIEBER and R. SCHWYZER, Helv. chim. Acta 40, 373 (1957).
- <sup>19</sup> D. Ben-Ishai and A. Berger, J. org. Chem. 17, 1564 (1962).
- <sup>20</sup> K. Okawa, Bull. chem. Soc. Japan 30, 976 (1957).
- 21 R. Schwyzer and P. Sieber, Helv. chim. Acta 40, 624 (1957).
- <sup>22</sup> W. G. HANGER, W. C. HOWELL and A. W. JOHNSON, J. chem. Soc. 496 (1958).
- <sup>23</sup> Actinomycin D, lot No. 3008 A-30B, from Lederle Laboratories Division, American Cyanamid Co., Pearl River, N.Y.
- <sup>24</sup> H. Brockmann, Angew. Chem. 66, 1 (1954) reported for natural actinomycin  $C_1$ : m.p. 241-243°,  $[\alpha]_D^{90} 349^\circ \pm 10^\circ$  (c 0.26, methanol).
- <sup>25</sup> G. E. FOLEY, R. E. McCarthy and V. M. Binns, Ann. N.Y. Acad. Sci. 76, 413 (1958).
- <sup>26</sup> G. E. Foley, Antibiotics A. 432 (1955–1956).
- 27 The author wishes to thank Dr. S. Farber for his support of this work, Dr. C. H. Li and Dr. V. Du Vigneaud for helpful discussions, Dr. G. E. Foley for the microbiological assays, Dr. Y. Sano for the preparation of dipeptide (III), and Mrs. Andrea Seki-Viano, Mr. R. Cotton, Mr. A. Trzeciak, and Mrs. Edith Judkins for technical help. This work was supported by Public Health Service Research Grants (No. C-6516 from the National Cancer Institute, No. FR-05526 from the Division of Research Facilities and Resources), National Institutes of Health, by A. and M. Lasker Foundation, New York, and A. T. and V. D. Fuller Cancer Research Unit Grant, American Cancer Society (Massachusetts Division) Inc.