Featured Article

Subscriber access provided by University of Rochester | River Campus & amp; Miner Libraries

Synthesis of illudinine from dimedone and identification of activity as a monoamine oxidase inhibitor (MAOI)

Robert Gaston, Werner J. Geldenhuys, and Gregory B. Dudley

J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.0c01301 • Publication Date (Web): 24 Aug 2020

Downloaded from pubs.acs.org on August 26, 2020

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.

is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

Synthesis of illudinine from dimedone and identification of activity as a monoamine oxidase inhibitor (MAOI)

Robert Gaston, Jr.,[†] Werner J. Geldenhuys,[‡] Gregory B. Dudley^{†,*}

[†]C. Eugene Bennett Department of Chemistry and [‡]Department of Pharmaceutical Sciences, West Virginia University, Morgantown, West Virginia 26505-6045, United States.

ABSTRACT: The fungal metabolite illudinine is prepared in seven steps and ca. 55% overall yield from dimedone using an "open and shut" (ring-opening and ring-closing) strategy. Tandem ring-opening fragmentation and olefination of dimedone establishes alkyne and vinyl-arene functionality linked by a neopentylene tether. Oxidative cycloisomerization then provides the illudinine framework. The key innovation in this second-generation synthesis of illudinine is the use of the nitrile functional group, rather than an ester, as the functional precursor to the carboxylic acid of illudinine. The small, linear nitrile (C=N) is associated with improved selectivity, π -conjugation, and reactivity at multiple points in the synthetic sequence relative to the carboxylic acid ester. Preliminary assays indicate that illudinine and several related synthetic analogues are monoamine oxidase inhibitors (MAOIs), which is the first reported indication of biological activity associated with this natural product. Illudinine was found to inhibit monoamine oxidase B (MAO-B) with an IC₅₀ of 18 ± 7.1 µM in preliminary assays.

INTRODUCTION

Illudinine is the alkaloid congener of illudalic acid, with both being natural sesquiterpene metabolites of the toxic jack o'lantern mushroom (*Omphalotus illudens*; aka *Clitocybe illudens*).¹ After initially being identified as "the inactive acidic compound", illudalic acid later emerged from high-throughput screening as "the first potent, selective" inhibitor of the human leukocyte common antigen-related (LAR) receptor-type protein tyrosine phosphatase (PTP),² which has implications for drug discovery related to insulin resistance, diabetes, obesity, CNS injury, drug addiction, and various cancers.³ No biological activity has been identified previously for illudinine.



Illudalic acid and illudinine (Figure 1) are members of the illudalane sesquiterpene family of natural products. Other illudalanes include the alcyopterosins⁴ and other pterosins,⁵ armiloid A,6 echinolactones,7 fomajorin D,8 granulolactone9 and dihydrogranuloinden,¹⁰ incarnatins and incarnalactones,¹¹ onitin,¹² russujaponols,¹³ sterostreins,¹⁴ and several unnamed illudalanes.¹⁵ The illudalanes are biosynthetically related to the illudins and illudanes, and some illudalanes may be artifacts of the illudin/illudane isolation process.¹⁶ The illudalanes have a of known biological activities, but varietv their pharmacological development has suffered from limited access to synthetic derivatives. For example, medicinal chemistry efforts associated with the cytotoxicity of the alcyopterosins and the phosphatase inhibition activity of illudalic acid have focused on simplified synthetic analogues in which the fused gem-dimethylcyclopentane region has been truncated,¹⁷ removed,¹⁸ and/or replaced¹⁹ with something more synthetically accessible. In all cases, these synthetically expedient structural modifications resulted in loss of potency. Improved synthetic access to the illudalanes themselves and to analogues that incorporate the desirable features of the fused gemdimethylcyclopentane region are needed to advance drug discovery.

The deceptively simple illudalane carbon skeleton presents challenges for modern synthetic methodology and strategies for chemical synthesis. The bicyclo[4.3.0]nonane carbocyclic core includes a quaternary center and is often embodied as a 2,2-dimethyl-indane with full substitution around the aromatic ring (cf. Figure 1). The various illudalane sesquiterpenes have been popular targets for target-oriented synthesis, especially in recent years;²⁰ discussion here focuses on illudinine.

Figure 1. Selected illudalane sesquiterpenes.

Scheme 1. Woodward and Hoye synthesis of illudinine (15 steps, 4.6% overall from indane).



Scheme 2. Teske and Deiters synthesis of illudinine (8 steps, 17% overall from diyne 3; 14 steps, 6.9% from isophorone).



The first synthesis of illudinine (1) (along with illudalic acid) was published in 1977 by Woodward and Hoye in a 15-step route from indane (4.6% overall yield, Scheme 1).²¹ This work highlights the close structural relationship between illudinine and illudalic acid, including the demonstration that illudinine can arise from condensation of the methyl ether of illudalic acid with ammonia (eq 1).



In 2008, Teske and Deiters developed an elegant nickelcatalyzed cyclotrimerization method (cf. $4\rightarrow 5$) that enabled an 8-step synthesis of illudinine from diyne **3** (17% overall yield, Scheme 2),²² which itself is made from isophorone.²³ These previous syntheses of illudinine underscore the challenges associated with crafting the fused *gem*-dimethylcyclopentane ring. The Woodward approach was to anneal a dimethylcyclopentanone through a series of substitution reactions, followed by deoxygenation. Deiters made use of 4,4dimethyl-1,6-heptadiyne (**3**), which is an attractive building block for the synthesis of many illudalanes except that it is typically prepared in 6 steps from isophorone,²³ starting with Eschenmoser–Tanabe fragmentation (\rightarrow **2**).²⁴

We have been developing anionic fragmentation²⁵ processes for making alkynes,²⁶ including in tandem with olefination for preparing tethered enynes.²⁷ This methodology provides access to neopentylene-tethered (NPT) 1,6-enynes in three steps from dimedone (Scheme 3, top). Like 1,6-diyne **3** (Scheme 2), NPT enynes are attractive building blocks for chemical synthesis. We have shown how NPT enynes derived from dimedone can advance synthetic methodology,²⁸ streamline previous syntheses of hirsutene and illudol,²⁷ support concise syntheses of alcyopterosin A^{20a} and illudinine,²⁹ and facilitate production of synthetic illudalic acid and analogues.^{20j} Our approach to illudinine begins with tandem fragmentation and Knoevenagel-type condensation of **6** with **7** to prepare pyridyl-enynes **8**, then cycloisomerization to access isoquinoline systems *en route* to the natural product (Scheme 3, bottom).

Scheme 3. Tandem fragmentation / olefination methodology and application to the synthetic approach to illudinine.



Here we report our synthesis of illudinine from dimedone, building on preliminary communications and with a key tactical innovation compared to our first-generation approach. We recapitulate the first iteration of this approach (Scheme 3, bottom, EWG = CO_2Et), updating it to include a recent focused examination of the microwave-mediated oxidative cycloisomerization.³⁰ For the revised approach, we used a nitrile (C=N) functional group instead of an ethyl ester as the precursor to the carboxylic acid of illudinine (EWG = CN). The nitrile requires harsher conditions for late-stage hydrolysis to the

carboxylic acid, but illudinine itself is robust to hydrolysis.³¹

Scheme 4. First-generation synthesis of illudinine (8 steps, 14-22% yield from dimedone).



The nitrile is also smaller and more electron-withdrawing than the ester, including by virtue of better π -conjugation owing to its limited conformational freedom. These attributes of the nitrile translated into several improvements in reaction outcomes, resulting in an overall synthetic production process that is shorter (7 steps vs. 8 steps) and affords a higher overall yield (58% vs. 14-22%) from dimedone. We also report biological activity of illudinine for the first time; it is a monoamine oxidase inhibitor (MAOI), with an IC₅₀ of ~18 µM for monoamine oxidase B (MAO-B) (*vide infra*).

RESULTS AND DISCUSSION

First-generation synthesis.^{29,30} The synthesis of illudinine as previously reported is outlined in Scheme 4. The hydroxycycloalkenyl triflate (6) needed for fragmentation / olefination can be reached in two steps from dimedone (cf. Scheme 3, top). Tandem fragmentation / olefination is mediated by two equivalents of strong base to deprotonate the two substrates at low temperature. As the reaction mixture warms, the triflate undergoes anionic fragmentation to 3,3-dimethyl-5-hexynal (not shown), which undergoes Knoevenagel-type condensation in situ with pyridine 7a. Vinyl pyridine 8a is obtained as a ca. 2:1 mixture of Z:E isomers. From here, an oxidative cycloisomerization involving an inverse-demand dehydro-Diels-Alder reaction was envisioned based on recent precedent from the Brummond lab in the "normal" electron-demand series,³² but inverse-demand Diels-Alder processes can be complicated by competing reaction pathways.³³ In this case, heating 8a at temperatures up to 240 °C resulted in thermal decomposition with no evidence of the desired product (10, X = H). However, bromo-alkyne 9 proved to be a competent substrate, providing modest to good yields of bromoisoquinoline 11. Bromo-alkyne dienophiles are known,³⁴ and computational data suggests that the bromine substituent weakens the alkyne π -bond,³⁵ which thus facilitates the cycloaddition process.

Nonetheless, this oxidative cycloisomerization has a narrow reactivity window, with product formation and decomposition competing at the reaction temperatures required for reactivity. In our preliminary investigations, we examined microwave (MW) and conventional (CONV) heating methods interchangeably, and we found that MW heating for 12 h at 200 °C provided a viable 40% yield of the isoquinoline. A similar yield could be obtained by CONV heating for 24 h, but the isolated yields declined thereafter linked to product decomposition.

We later reexamined this reaction systematically, focusing on temperature, time, and heating mode (MW v CONV).³⁰ Yields were consistently highest at 200 °C and 24 h compared with temperatures and times above and below these values. It was also found that solutions of the substrate heat more efficiently in the microwave than solutions of the product (faster and to

higher temperature at constant MW power), and that isolated yields of product were consistently higher under MW compared to CONV heating, despite essentially identical temperature profiles and other reaction variables. For example, MW heating at 200 °C for 24 h produced bromo-isoquinoline **11** in 62% yield, compared with 39% under CONV heating. Possible interpretations and strategic applications of this observed phenomena are discussed elsewhere.³⁶

Replacement of the bromide with methoxide to provide illudinine ethyl ester was challenging, as discussed in our preliminary report. Direct substitution was unsuccessful in our hands, but the Maloney–Fier conditions³⁷ introduced a hydroxy substituent (11 \rightarrow X=OH) in 89% yield. Methylation of this hydroxy-isoquinoline (like other hydroxy-isoquinolines³⁸) was highly selective for (undesired) *N*-methylation under most conditions, but a protocol involving TMS-diazomethane^{38a} was carefully optimized to reverse selectivity to 70:30 favoring *O*-methylation and produced illudinine ethyl ester in 63% isolated yield. Saponification as previously reported provided illudinine in essentially quantitative yield.²⁹

Our synthetic approach to illudinine had thus been reduced to practice as an 8-step synthesis in ca. 22% overall yield from dimedone (Scheme 4). These metrics compare favorably to previous syntheses (Scheme 1 and 2), but issues of selectivity and yield remained to be addressed, in particular with the thermal oxidative cycloisomerization and subsequent bromideto-methoxide conversion.



Figure 2. *left*: the *Z*- and *E*-isomers of **8a**; *Z*-**8a** is aligned for intramolecular Diels–Alder cycloaddition. *right*: perspective depiction of a conformation of bromo-isoquinoline **11** in which the ester is aligned perpendicular to the isoquinoline plane.

Considerations for process improvement. One concern we aimed to address was the 2:1 mixture of *Z*:*E* enyne isomers **8a** generated in the tandem fragmentation / olefination, which preceded the oxidative cycloisomerization. The *Z*-isomer of **8a** is appropriately disposed for intramolecular cycloaddition, whereas the *E*-isomer is not (Figure 2, left). Recovered **8a** from incomplete cycloisomerization reactions retained the ca. 2:1 ratio, suggesting that this (thermodynamic) ratio is maintained during the high temperature reaction by *in situ Z*/*E* isomerization, in which case the starting *Z*:*E* ratio may be moot. However, a careful chromatographic separation of **8a** provided fractions that were enriched in the *Z*-isomer, and this *Z*-enriched

2

3

4

5

6 7 8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

Scheme 5. Second-generation synthesis of illudinine (5 steps, ca. 60% yield from 6; 7 steps, ca. 55% yield from dimedone)



material qualitatively outperformed the Z-depleted material in small-scale cycloisomerization reactions (not shown). This preliminary observation suggested that increasing the Z:E ratio might improve the oxidative cycloisomerization process, as would be expected if Z/E isomerization were not occurring (or were inefficient) during the high-temperature reactions.

Other concerns relate to the hypothetical conformational preferences of the ester (Figure 2, right). The ester of **11** is likely not well conjugated with the isoquinoline aromatic ring system, and we reasoned that there is similarly poor orbital overlap in rate-determining transition state structures associated with the cycloisomerization and bromide substitution processes.

With these concerns and considerations in mind, we focused on the small, linear nitrile as the functional precursor to the illudinine carboxylic acid. We formulated two hypotheses and identified two new concerns related to replacing the ester with a nitrile. The first hypothesis was that the tandem fragmentation / olefination would be more Z-selective with the smaller nitrile. The second was that the desired Br \rightarrow OMe S_NAr substitution (cf. $13 \rightarrow 14$. Scheme 5) would be better facilitated by the nitrile than the ester, owing to better π -conjugation with the aromatic ring. The two concerns were that the nitrile would be more resistant to hydrolysis in the last step than the ester, and the hypothetical impact of better π -conjugation of the nitrile in the inverse-demand Diels-Alder transition state was not assumed to be favorable to the reaction, given the general difficulties with inverse-demand Diels-Alder reactions cited above.33 Data and observations from subsequent experimental work support the two hypotheses and suggest that the concerns were largely unfounded.

Second-generation synthesis. The revised and refined synthesis of illudinine featuring the nitrile in place of the ester is outlined in Scheme 5. As noted earlier, hydroxy-cycloalkenyl triflate **6** is available in two steps, each >95% yield, from dimedone (Scheme 3, top). 4-Pyridinylacetonitrile (**7b**) is commercially available as the HCl salt, which was convenient to use as received with an extra equivalent (3.1 equiv) of base in the tandem fragmentation / olefination step. Enyne **8b** was obtained in 93% yield as a single observable isomer, which we attribute to the small steric profile of the (two-atom) nitrile.

Pyridyl-enyne **8b** is an excellent substrate for the oxidative cycloisomerization reaction. Whereas ester variant **8a** was not a viable substrate (0% yield of **10**), nitrile **8b** was converted into isoquinoline **12** in 85% yield upon heating at 200 °C. In this case, MW and CONV heating gave similar results, and even heating neat (solvent-free) delivered **12** with similar efficiency. This positive outcome is attributed to: i) the isomeric purity of **8b**; ii) better access to planar vinylpyridine conformations that are optimal for cycloaddition; and iii) strong π -conjugation between the electron-deficient diene component and the sp-hybridized (linear) nitrile. Some of the challenges of inverse-demand Diels–Alder reactions are associated with instability of electron-deficient dienes; as noted in our preliminary report,²⁹

these concerns are off-set by the extra stability of vinylarenebased 4π components.

Having observed favorable reactivity from enyne 8b, we elected to explore bromination of isoquinoline 12, rather than return to the original plan of brominating alkyne 8b. The paracyano group was now viewed as a liability in the electrophilic aromatic bromination, but N-bromo-succinimide (NBS) in sulfuric acid³⁹ at 0 °C introduced the desired bromine atom $(\rightarrow 13)$ in 88% yield without impacting the nitrile. In contrast to our efforts in the first-generation synthesis, sodium methoxide in refluxing methanol smoothly promoted the desired S_NAr substitution to methoxy-isoquinoline 14. Finally, potassium hydroxide in ethanol, now under refluxing conditions, hydrolyzed the nitrile and delivered illudinine in essentially quantitative yield. As noted previously, this second-generation manifestation of our approach based on tandem fragmentation / olefination and oxidative cycloisomerization provides illudinine in 7 steps and ca. 55% overall yield from dimedone.

Monoamine oxidase B (MAO-B) bioassay data. We were interested in exploring potential biological activity of illudinine. Its congeneric natural product illudalic acid is an inhibitor of LAR phosphatase,^{2,18,20j} but this activity is attributed to the *o*-formyl hydroxy-lactone that is not shared with illudinine. Illudinine and its synthetic precursors showed no LAR inhibitory activity in preliminary assays (not shown).

On the other hand, we recognized certain structural commonalities with the known monoamine oxidase inhibitors (MAOIs) rasagiline and M30 (Figure 3).⁴⁰ Rasagiline shares an indane core with illudinine, and M30 is a hydroxy-quinoline, not unlike the methoxy-isoquinoline core of illudinine. Rasagiline and M30 are irreversible submicromolar MAOIs, with rasagiline having selectivity for MAO-B. Therefore, we tested illudinine and its bromo-isoquinoline precursor **11** for MAO-B activity.



Figure 3. Structural commonalities between illudinine and known MAOIs rasagiline and M30.

Illudinine (1) and bromo-isoquinoline 11 were observed to inhibit MAO-B dose-dependently and with similar potency in our preliminary assays (Figure 4). We determined the IC₅₀ for illudinine to be $18.3 \pm 7.1 \ \mu$ M, whereas bromo-isoquinoline 11 had an IC₅₀ of $22.5 \pm 2.8 \ \mu$ M in our assays. This is the first biological activity reported for illudinine.

Having observed MAO-B inhibitory activity from illudinine and **11**, we screened a small sample of analogues as potential MAO-B inhibitors at a concentration of 22 μ M. (Figure 5). A few initial structure-activity relationship (SAR) observations

1

2

3 4

5

6

7

8

9

10

11

12

13

14

15 16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60



Figure 4. Dose-response curve for illudinine (blue) and bromoisoquinoline 11 (red) inhibition of human MOA-B.



Figure 5. MAO-B inhibitory activity of illudinine and a series of analogues at 22 μ M inhibitor concentration.

can be made. Nitriles 12 and 13 and ethyl ester 11 appear to be similar in potency to each other and to illudinine (1). Hydroxy-isoquinoline 15 was nearly inactive compared to its bromo-(13) or unsubstituted (12) isoquinoline counterparts. Deletion of the *gem*-dimethyl substitution on the indane core (compound 16) was deleterious to activity, which parallels observations made during medicinal chemistry efforts connected to alcyopterosin

A¹⁷ and illudalic acid,¹⁸ discussed above in the Introduction section. These preliminary data suggest potential opportunities to improve and refine MAOI activity in future analogues based on the illudinine cores structure.

CONCLUSIONS

We refined our "open and shut" (ring-opening fragmentation and ring-closing oxidative cycloisomerization) approach to the natural product illudinine, completing the total synthesis in 7 steps from dimedone (ca. 55% overall yield). Our firstgeneration route delivered the natural product in 8 steps and 14– 22% overall yield. The key innovation in this new synthesis is the use of the small, linear nitrile functional group as the precursor to the carboxylic acid of illudinine. Illudinine and analogous isoquinolines were observed to have modest inhibitory activity against monoamine oxidase B (MAO-B), which may merit further investigation.

EXPERIMENTAL SECTION

General Experimental Methods. All chemicals were used as received unless otherwise stated. Syringes were used in all protocols requiring the transfer of a liquid reactant of solvent. Tetrahydrofuran (THF) and dichloromethane (DCM) were dried under a column of molecular sieves in an argon atmosphere. A Schleck line was used in all reactions (except for the microwave reactions) to purge reaction vessels and provide an inert, nitrogen atmosphere. A CEM microwave reactor set in constant temperature mode was used for the reactions requiring microwave heating along with the microwave reaction vial, where the temperature was monitored using an external IR sensor and/or internal fiber-optic temperature probe. All purifications were executed using a Biotage Isolera One automated flash column system unless otherwise stated. Yields are reported as isolated yields considered to be \geq 95% pure by ¹H NMR following flash chromatography. All new compounds were characterized using a JEOL 400 spectrometer to conduct ¹H and ¹³C NMR spectroscopy in CDCl₃ (≥99.8 atom % D, contains 0.03% (v/v) TMS) purchased from Cambridge Isotope Laboratories. The chemical shifts (δ) are reported in parts per million (ppm) relative to the CHCl₃ peak (7.26 ppm and 77.16 ppm for ¹H and ¹³C, respectively). The coupling constants (J) are reported in Hertz (Hz). Mass spectra were recorded using electrospray ionization (ESI).

Monoamine oxidase assays. Illudinine and analogues were dissolved in 2% dimethyl sulfoxide (DMSO) buffer solution and added with purified MAO-B and kynuramine in a 96-well plate. After 30 minutes of incubation at 37 °C, a sodium chloride stop solution was added and the plate was analyzed using a plate reader. Excitation-emission fluorescence spectroscopy at 310 and 380 nm was used to determine the enzymatic activity relative to the negative control (no inhibitor added). MAO-B metabolized kynuramine to 4-hydroxyquinoline which can be quantified using fluorescence. A 2% DMSO buffer solution was prepared by mixing 19.6 mL of 0.1 M phosphate solution at pH 7.4 with 0.4 mL of DMSO. The kynuramine substrate solution was made by taking 19.15 µL of 25 mM kynuramine solution and adding it to 5.981 mL of the 2%-DMSO buffer. The MAO-B enzyme solution combines 75 µL of MAO-B with 5.928 mL of buffer. Using this method allow the concentration of DMSO to remain constant in each well. For the dose-response assays. the inhibitor concentration range started at 200 µM and finished at 0.274 μ M. IC₅₀ values were determined using an online IC₅₀ calculator.41

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

(Z)-5,5-Dimethyl-2-(pyridin-4-yl)oct-2-en-7-ynenitrile

(8b). To a solution of the reduced vinyl triflate 6 (1.17 g, 4.23) mmol, 1.1 equiv.) dissolved in 20 mL of THF at -78 °C, 12.1 mL of lithium (bistrimethylsilyl)amide (1.0 M in THF, 3.1 equiv.) was added dropwise. The solution was stirred for 5 minutes at -78 °C before adding 4-pyridinylacetonitrile hydrochloride (0.600 g, 3.88 mmol, 1.0 equiv.) as a solid. The reaction mixture was stirred for an additional 10 minutes at -78 °C, and was then allowed to come to room temperature. The reaction was stirred at room temperature for 30 minutes and then at 60 °C for 2 hours. The reaction was monitored by TLC using 30% EtOAc in hexanes as the eluent. Upon complete consumption of the starting material, the reaction mixture was cooled to room temperature and quenched with half-saturated ammonium chloride. The aqueous layer was extracted with EtOAc. The combined organic layers were washed with water and brine, dried over MgSO₄, and concentrated under vacuum. The product was purified by flash chromatography using a gradient from 10%-60% EtOAc in hexanes to give 8b (0.790 g, 93%) as a solid; m.p.: 41-42 °C. ¹H-NMR (400 MHz, CDCl₃): δ 8.67 (d, J= 4.2Hz, 2H), δ 7.44 (d, J= 6.2Hz, 2H), δ 7.16 (t, J= 8Hz, 1H), δ 2.68 (d, J= 8Hz, 2H), δ 2.21 (d, J= 2.5Hz, 2H), δ 2.09 (t, 3.6Hz, 1H), δ 1.12 (s, 6H). ¹³C{¹H}NMR (100 MHz, CDCl₃): δ 150.7, 147.8, 140.5, 119.9, 116.9, 116.2, 115.6, 81.3, 71.4, 43.8, 35.7, 32.0, 27.0. HRMS (ESI+) calculated for $C_{15}H_{16}N_2^+$ [(M+H)⁺]: 225.1386, found 225.1392.

7,7-Dimethyl-7,8-dihydro-6H-cyclopenta[g]isoquinoline-5-carbonitrile (12). Enyne 8b (300 mg, 1.34 mmol) was dissolved in 12 mL of ortho-dichlorobenzene in a 35-mL CEM microwave vial. The reaction mixture was stirred and heated to 200 °C by microwave radiation. The temperature was monitored using the (calibrated) IR sensor from the CEM instrument. After 24 hours, the solution was loaded onto a Biotage silica gel column. Three column volumes of hexanes were used to flush out the solvent. The product was purified using a gradient elution from 0-80% ethyl acetate in hexanes. The fractions were combined, and the solvent was removed under reduced pressure to give 12 (252.4 mg, 1.14 mmol, 85%) as a solid; m.p. 85-88 °C. ¹H-NMR (400 MHz, CDCl₃): δ 9.22 (s, 1H), δ 8.63 (d, J= 6Hz, 1H), δ 7.92, δ 7.92 (overlapping 2H), δ 3.12 (s, 2H), δ 2.97 (s, 2H), δ 1.25 (s, 6H).¹³C{¹H}NMR (100 MHz, CDCl₃): δ 155.5, 152.2, 144.8, 144.4, 138.9, 135.0, 127.8, 127.4, 117.6, 116.0, 104.8, 47.6, 47.3, 41.1, 28.3. HRMS (ESI+) calculated for $C_{15}H_{14}N_2^+$ [(M+H)+]: 223.1230, found 223.1230.

9-Bromo-7,7-dimethyl-7,8-dihydro-6H-

cyclopenta[g]isoquinoline-5-carbonitrile (13). Isoquinoline 12 (394.8 mg, 1.78 mmol, 1.0 equiv) was dissolved in 9 mL of concentrated sulfuric acid and stirred at 0° C for 10 minutes. N-Bromosuccinimide (695 mg, 3.91 mmol, 2.2 equiv) was added portion-wise over 2 minutes, and the flask was resealed. The reaction was stirred at 0° C for an additional 10 minutes and then at room temperature for 18 hours. The solution was cooled at 0 °C again before diluting carefully with 25 mL of water. The solution was adjusted to pH 8-9 using 30% ammonium hydroxide. The aqueous layer was extracted with ethyl acetate. The organic layer was washed with water, brine, dried over MgSO₄, and concentrated. The crude product was purified via flash chromatography using a 7-50% gradient elution of EtOAc in hexanes to give 13 (472 mg, 88%) as a solid; m.p.: 172-175 °C.1H-NMR (400MHz, CDCl3): 8 9.59 (s, 1H), 8 8.72 (d, J= 6Hz, 1H), δ 7.89 (d, J= 5.6Hz, 1H), δ 3.22 (s, 2H), δ 3.02 (s, 2H), δ 1.26 (s, 6H). ¹³C {¹H}NMR (100 MHz, CDCl₃): δ 155.0,

151.6, 145.6, 145.5, 136.4, 126.1, 125.0, 117.4, 115.5, 110.2, 104.5, 49.5, 48.8, 40.1, 28.6. HRMS (ESI+) calculated for $C_{15}H_{13}N_2Br^+$ [(M+H)⁺]: 301.0335, found 301.0335.

9-Methoxy-7,7-dimethyl-7,8-dihydro-6H-

cyclopenta[g]isoquinoline-5-carbonitrile (14). Isoquinoline 13 (103 mg, 0.342 mmol, 1.0 equiv.) was dissolved in 5 mL of anhydrous methanol, and then 30% sodium methoxide (0.761 mL, 3.42 mmol, 10 equiv.) was added. A jacketed condenser was connected to the flask and the solution was refluxed at 65°C for 4 hours. After completion of the reaction was confirmed by TLC, the reaction was cooled to room temperature and diluted with 10 mL of water. HCl (1M, 3 mL) was added dropwise to quench the reaction. The aqueous layer was extracted with ethyl acetate, and the organic layer was washed with water, brine, dried over MgSO₄, and concentrated in vacuo. The crude product was purified using flash chromatography with a 10-60% gradient elution of ethyl acetate in hexanes to give 14 (80.2 mg, 93%) as a solid; m.p.: 120-123 °C. 1H-NMR (100MHz, CDCl₃): δ 9.58 (s, 1H), δ 8.72 (d, J= 6Hz, 1H), δ 7.89 (d, J= 5.6Hz, 1H), δ 3.22 (s, 2H), δ 3.02 (s, 2H), δ 1.26 (s, 6H). ¹³C{¹H}NMR (400 MHz, CDCl₃): δ 159.4, 156.6, 147.2, 143.4, 137.1, 128.4, 121.8, 117.8, 116.0, 98.7, 60.6, 47.6, 46.1, 40.8, 28.3. HRMS (ESI+) calculated for $C_{16}H_{16}N_2O^+$ [(M+H)⁺]: 252.1223, found 252.1216.

Illudinine (1). Isoquinoline 14 (65.0 mg, 0.258 mmol, 1.0 equiv) was dissolved in 6 mL of ethanol, and KOH powder (144.5 mg, 2.58 mmol, 10.0 equiv.) was added quickly to the solution. The solution was heated at reflux for 72 hours before it was allowed to cool to room temperature. The reaction was quenched with 30 mL of pH-7 phosphate buffer. The aqueous layer was extracted with ethyl acetate. The combined organic layers were washed with water and brine, dried over MgSO4 and concentrated to give 69.1 mg of illudinine (99% yield) as a white powder; m.p.: 220-225 °C. 1HNMR (600 MHz, CD₃OD + 1 drop of TFA): δ 9.72 (s, 1H), 9.01 (d, J = 6.96Hz, 1H), 8.51 (d, J = 6.92Hz, 1H), 4.30 (s, 3H), 3.28 (s, 2H), 3.25 (s, 2H), 1.22 (s, 6H). ${}^{13}C{}^{1}H{}NMR$ (100MHz, CD₃OD + 1 drop of TFA): δ168.6, 163.7, 157.3, 143.4, 139.4, 133.7, 132.9, 123.9, 123.2, 120.8, 61.8, 50.8, 46.3, 41.3, 28.2. HRMS (ESI+) calculated for C₁₆H₁₈NO₃⁺ [(M+H)⁺]: 272.1287, found 272.1277.

9-Hydroxy-7,7-dimethyl-7,8-dihydro-6H-

cvclopenta[g]isoquinoline-5-carbonitrile (15). Isoquinoline 13 (100.3 mg, 0.33 mmol, 1.0 equiv.), acetohydroxamic acid (149.9 mg, 1.99 mmol, 6 equiv.), and K₂CO₃, (410.1 mg, 3.33 mmol, 10 equiv.) were dissolved in DMSO (1.1 mL). The brown, heterogeneous solution was allowed to stir overnight at 80 °C. After cooling to room temperature, the solution was placed in an ice bath without stirring where HCl (1 M) was added slowly dropwise to quench the reaction. During the HCl addition the solution formed a vellow tint, and after enough HCl was added the solid, crude product crashed out of solution. The solid was isolated by filtration and purified using flash chromatography (0-10% methanol in DCM) to provide 15 as a bright yellow solid (64.3 mg, 81%). ¹H-NMR (400 MHz, $CDCl_3$): δ 9.76 (s, 1H), 8.47 (d, J = 1.5 Hz, 1H), 7.94 (d, J = 1.6 Hz, 1H), 3.10 (s, 2H), 2.92 (s, 2H), 1.24 (s, 6H). ¹³C{¹H}NMR (100 MHz, DMSO-d6): 8158.0, 140.4, 138.4, 135.6, 121.6, 117.9, 114.3, 101.8, 99.5, 93.3, 45.4, 41.0 30.5, 24.6. (ESI+) calculated for $C_{15}H_{14}N_2O^+$ [(M+H)⁺]: 239.1179, found 239.1171.

9-Bromo-7,8-dihydro-6*H***-cyclopenta**[*g*]**isoquinoline-5**-**carbonitrile** (16). This didesmethyl analogue of compound 13

was made by analogy to **13**, starting from 1,3-cyclohexanedione in place of dimedone. 1,3-Cyclohexanedione was converted into 3-hydroxycyclohex-1-en-1-yl trifluoromethanesulfonate (the vinyl triflate analogous to **6**) as reported previously.⁴²

1

2

3 4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49 50

51

52

53

54

55

56

57 58

59

60

(a) By analogy to the preparation of **8b**, 3-hydroxycyclohex-1-en-1-yl trifluoromethanesulfonate⁴² (1.75 g, 7.12 mmol, 1.1 equiv.) was dissolved in THF at -78 °C, and lithium (bistrimethylsilyl)amide (20.1 mL, 1.0 M in THF, 3.1 equiv.) was added dropwise. The solution was stirred for 5 minutes at -78 °C before adding 4-pyridinylacetonitrile hydrochloride (1.0 g, 6.5 mmol, 1.0 equiv.) as a solid. The reaction mixture was stirred for an additional 10 minutes at -78 °C, then 30 min at room temperature, and then for 2 h at 60 °C. Workup and purification as described above for compound **8b** provided 1.00 g of a yellow solid, which was tentatively assigned as (*Z*)-2-(pyridin-4-yl)oct-2-en-7-ynenitrile (the didesmethyl analogue of **8b**, 79% yield) and used in the next step.

(b) By analogy to the preparation of **12**, (*Z*)-2-(pyridin-4-yl)oct-2-en-7-ynenitrile as prepared in the previous step (430 mg, 2.19 mmol) was dissolved in 12 mL of *ortho*-dichlorobenzene and heated at 200 °C by microwave radiation with stirring for 24 h. Purification as described for compound **12** provided 322 mg of a yellow solid, which was tentatively assigned as 7,8-dihydro-6*H*-cyclopenta[*g*]isoquinoline-5-carbonitrile (the didesmethyl analogue of **12**, 76% yield) and used in the next step.

(c) By analogy to the preparation of **13**, 7,8-dihydro-6*H*-cyclopenta[*g*]isoquinoline-5-carbonitrile as prepared in the previous step (136 mg, 0.70 mmol) was dissolved in 3 mL of concentrated sulfuric acid and stirred at 0 °C for 10 minutes. *N*-Bromosuccinimide (274 mg, 1.54 mmol, 2.2 equiv) was added in three portions over 2 minutes, and the flask was resealed. The reaction was stirred at 0° C for an additional 1 h and then at room temperature for 12 hours. Workup and purification as described above for compound **13** provided **16** as a white solid (167 mg, 87%). ¹H-NMR (400 MHz, CDCl₃): δ 9.62 (s, 1H), 8.73 (d, J = 5.6 Hz, 1H), 7.92 (d, 5.6 Hz, 1H), 3.44 (t, J = 7.6 Hz, 2H), 3.23 (t, J = 7.6 Hz, 2H), 2.31 (p, J = 7.6 Hz, 2H). ¹³C {¹H}NMR (100 MHz, CDCl₃): δ 155.4, 151.7, 146.2, 145.6, 136.4, 124.7, 117.3, 104.0, 100.1, 34.7, 29.7, 24.1. HRMS (ESI+) calculated for C₁₃H₁₀BrN₂⁺ 273.0022, found 273.0022

ASSOCIATED CONTENT

Supporting Information

Copies of NMR spectra for compounds prepared by the secondgeneration synthetic route and raw data from the monoamine oxidase inhibition assays (PDF). The Supporting Information is available free of charge on the ACS Publications website.

AUTHOR INFORMATION

Corresponding Author

* E-mail: gregory.dudley@mail.wvu.edu

ACKNOWLEDGMENT

This work was supported by grants from the National Science Foundation (CHE-1300722 and CHE-1834949). We gratefully acknowledge additional support from West Virginia University and the Eberly Family Foundation. We thank Prof. Brian Miller (Florida State University) for conducting preliminary LAR phosphatase inhibition assays and for helpful discussions.

REFERENCES

- (a) Anchel, M.; Hervey, A.; Robbins, W. J. Production of illudin M and of a fourth crystalline compound by *Clitocybe illudens. Proc. Natl. Acad. Sci.* 1952, 38, 927-928. (b) Nair, M .S. R.; Takeshita, H.; McMorris, T. C.; Anchel, M. Metabolites of *Clitocybe illudens.* IV. Illudalic acid, a sesquiterpenoid, and illudinine, a sesquiterpenoid alkaloid. *J. Org. Chem.* 1969, 34, 240– 243.
- (2) Yang, X.; Li, J.; Zhou, Y.; Shen, Q.; Chen, J.; Li, J. Discovery of novel inhibitor of human leukocyte common antigen-related phosphatase. *Biochim. Biophys. Acta* 2005, *1726*, 34–41.
- (3) (a) Kulas D. T.; Goldstein, B. J.; Mooney, R. A. The transmembrane protein-tyrosine phosphatase LAR modulates signaling by multiple receptor tyrosine kinases. J. Biol. Chem. 1996, 271, 748-754. (b) Zabolotny, J. M.; Kim, Y. B.; Peroni, O. D.; Kim, J. K.; Pani, M. A.; Boss, O.; Klaman, L. D.; Kamatkar, S.; Shulman, G. I.; Kahn, B. B.; Neel, B. G. Overexpression of the LAR (leukocyte antigen-related) protein-tyrosine phosphatase in muscle causes insulin resistance. Proc. Natl. Acad. Sci. 2001, 98, 5187-5192. (c) Mooney, R. A.; LeVea, C. M. The leukocyte common antigen-related protein LAR: candidate PTP for inhibitory targeting. Curr. Top. Med. Chem. 2003, 3, 809-819. (c) Chagnon, M. J.; Uetani, N.; Tremblay, M. L. Functional significance of the LAR receptor protein tyrosine phosphatase family in development and diseases. Biochem. Cell. Biol. 2004, 82, 664-675. (d) Takahashi. H.; Craig, A. M. Protein tyrosine phosphatases PTPo, PTPo, and LAR: presynaptic hubs for synapse organization. Trends Neurosci. 2013, 36, 522-534.
- (4) (a) Palermo, J. A.; Brasco, M. F.; Spagnuolo, C.; Seldes, A. M. Illudalane sesquiterpenoids from the soft coral Alcyonium paessleri: the first natural nitrate esters. *J. Org. Chem.* 2000, 65, 4482-4486. (b) Carbone, M.; Núñez-Pons, L.; Castelluccio, F.; Avila, C.; Gavagnin, M. Illudalane sesquiterpenoids of the alcyopterosin series from the Antarctic marine soft coral *Alcyonium grandis. J. Nat. Prod.* 2009, *72*, 1357-1360.
- (5) Zhan, Z. J.; Ying, Y. M.; Zhang, F. Y.; Li, C. P.; Shan, W. G. Three new illudalane sequiterpenoids from *Pteris semipinnata*. *Helv. Chim. Acta* **2010**, *93*, 550-554.
- (6) Li, H. T.; Tang, L.; Liu, T.; Yang, R.; Yang, Y.; Zhou, H.; Ding, Z. T. Polyoxygenated meroterpenoids and a bioactive illudalane derivative from a co-culture of *Armillaria* sp. and *Epicoccum* sp. Org. Chem. Front. 2019, 6, 3847-3853.
- (7) Suzuki, S.; Murayama, T.; Shiono, Y.; Illudalane sesquiterpenoids, echinolactones A and B, from a mycelial culture of *Echinodontium japonicum*. *Phytochemistry* 2005, *66*, 2329-2333.
- (8) Donnelly, D. M.; O'Reilly, J.; Polonsky, J.; Van Eijk, G. W. Fomajorin S and D from *Fomes annosus* (FR) cooke. *Tetrahedron Lett.* 1982, *23*, 5451-5452.

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57 58

- (9) Kokubun, T.; Scott-Brown, A.; Kite, G. C.; Simmonds, M. S. Protoilludane, Illudane, Illudalane, and Norilludane Sesquiterpenoids from *Granulobasidium vellereum*. J. Nat. Prod. 2016, 79, 1698-1701.
- (10) Nord, C. L.; Menkis, A.; Broberg, A. Cytotoxic illudalane sesquiterpenes from the wood-decay fungus *Granulobasidium vellereum* (Ellis & Cragin) Jülich. *Molecules* 2014, 19, 14195-14203.
- (11) Bunbamrung, N.; Intaraudom, C.; Dramae, A.; Boonyuen, N.; Veeranondha, S.; Rachtawee, P.; Pittayakhajonwut, P. Antimicrobial activity of illudalane and alliacane sesquiterpenes from the mushroom *Gloeostereum incarnatum* BCC41461. *Phytochemistry Lett.* **2017**, *20*, 274-281.
- (12) Banerji, A.; Ramakrishnan, G.; Chadha, M. S. Onitin and onitisin, new phenolic pterosins from the fern *Onychium auratum*. *Tetrahedron Lett.* **1974**, *15*, 1369–1370.
- (13) Yoshikawa, K.; Matsumoto, Y.; Hama, H.; Tanaka, M.; Zhai, H.; Fukuyama, Y.; Arihara, S.; Hashimoto, T. Russujaponols G-L, illudoid sesquiterpenes, and their neurite outgrowth promoting activity from the fruit body of *Russula japonica*. *Chem. Pharm. Bull.* **2009**, *57*, 311-314.
- (14) (a) Isaka, M.; Srisanoh, U.; Sappan, M.; Supothina, S.; Boonpratuang, T. Sterostreins F-O, illudalanes and norilludalanes from cultures of the Basidiomycete *Stereum ostrea* BCC 22955. *Phytochemistry* 2012, *79*, 116-120. (b) Li, J. F.; Qin, Y. K.; Tian, M. Q.; Zhang, K. Q.; Li, G. H. Two new sesquiterpenes from the fungus *Stereum* sp. NN048997. *Phytochemistry Lett.* 2014, *10*, 32-34.
- (15) Xie, S.; Wu, Y.; Qiao, Y.; Guo, Y.; Wang, J.; Hu, Z.; Zhang, Q.; Li, X.; Huang, J.; Zhou, Q.; Luo, Z. Protoilludane, illudalane, and botryane sesquiterpenoids from the endophytic fungus *Phomopsis* sp. TJ507A. *J. Nat. Prod.* **2018**, *81*, 1311-1320.
- (16) (a) Clericuzio, M.; Pan, F.; Han, F.; Pang, Z.; Sterner, O. Stearoyldelicone, an unstable protoilludane sesquiterpenoid from intact fruit bodies of *Russula delica*. *Tetrahedron Lett.* **1997**, *38*, 8237-8240. (b) Morisaki, N.; Furukawa, J.; Kobayashi, H.; Iwasaki, S.; Nozoe, S.; Okuda, S. Cyclobutyl cation rearrangements of 6-protoilluden-8α-ol, 7-protoilluden-6-ol and related compounds. *Chem. Pharm. Bull.* **1987**, *35*, 2678-2685.
- (17) Finkielsztein, L. M.; Bruno, A. M.; Renou, S. G.; Moltrasio Iglesias G. Y. Design, synthesis, and biological evaluation of alcyopterosin A and illudalane derivatives as anticancer agents. *Bioorg. Med. Chem.* **2006**, *14*, 1863-1870.
- (18) Ling, Q.; Huang, Y.; Zhou, Y.; Cai, Z.; Xiong, B.; Zhang, Y.; Ma, L.; Wang, X.; Li, X.; Li, J.; Shen, J. Illudalic acid as a potential LAR inhibitor: Synthesis, SAR, and preliminary studies on the mechanism of action. *Bioorg. Med. Chem.* **2008**, *16*, 7399–7409.
- (19) (a) Ling, Q.; Zhou, Y.-Y.; Cai, Z.-L.; Zhang, Y.-H.; Xiong, B.; Ma, L.-P.; Wang, X.; Li, X.; Li, J.; Shen, J.-K. Synthesis and LAR inhibition of 7-alkoxy analogues of illudalic acid. *Acta Pharmacol. Sin.* 2010, *45*, 1385–1397.
 (b) Uhl, G. R.; Martinez, M. J.; Paik, P.; Sulima, A.; Bi, G. H.; Iyer, M. R.; Gardner, E.; Rice, K. C.; Xi, Z. X. Cocaine reward is reduced by decreased expression of receptor-type protein tyrosine phosphatase D (PTPRD) and by a novel

PTPRD antagonist. Proc. Natl. Acad. Sci. U. S. A. 2018, 115, 11597–11602.

- (20) (a) Hoang, T. T.; Birepinte, M.; Kramer, N. J.; Dudley, G. B. Six-step synthesis of alcyopterosin A, a bioactive illudalane sesquiterpene with a gem-dimethylcyclopentane ring. Org. Lett. 2016, 18, 3470-3473. (b) Suresh, M.; Kumar, N.; Veeraraghavaiah, G.; Hazra, S.; Singh, R. B. Total Synthesis of Coprinol. J. Nat. Prod. 2016, 79, 2740-2743. (c) Suresh, M.; Kumari, A.; Das, D.; Singh, R. B. Total Synthesis of Onitin. J. Nat. Prod. 2018, 81, 2111-2114. (d) Melot, R.; Craveiro, M. V.; Bürgi, T.; Baudoin, O. Divergent Enantioselective Synthesis of (Nor)illudalane Sesquiterpenes via Pd⁰-Catalyzed Asymmetric C(sp3)-H Activation. Org. Lett. 2019, 21, 812-815. (e) Melot, R.; Craveiro, M. V.; Baudoin, O. Total Synthesis of (Nor)illudalane Sesquiterpenes Based on a C(sp³)-H Activation Strategy. J. Org. Chem. 2019, 84, 12933-12945. (f) Zeng, Z.; Zhao, Y.; Zhang, Y. Divergent total syntheses of five illudalane sesquiterpenes and assignment of the absolute configuration. Chem. Commun. 2019, 55, 4250-4253. (g) Xun, M. M.; Bai, Y.; Wang, Y.; Hu, Z.; Fu, K.; Ma, W.; Yuan, C. Synthesis of Four Illudalane Sesquiterpenes Utilizing a One-Pot Diels-Alder/Oxidative Aromatization Sequence. Org Lett. 2019, 21, 6879-6883. (h) Suresh, M.; Kumari, A.; Singh, R. B. Total synthesis of (±)-4, 8, 14-trihydroxyilludala-2, 6, 8triene. Tetrahedron 2019, 75, 3605-3608. (i) Elmehriki, A. A. H.; Gleason, J. L. A Spiroalkylation Method for the Stereoselective Construction of a-Quaternary Carbons and Its Application to the Total Synthesis of (R)-Puraquinonic Acid. Org. Lett. 2019, 21, 9729-9733. (j) McCullough, B. S.; Batsomboon, P.; Hutchinson, K. B.; Dudley, G. B.; Barrios, A. M. Synthesis and PTP Inhibitory Activity of Illudalic Acid and Its Methyl Ether, with Insights into Selectivity for LAR PTP over Other Tyrosine Phosphatases Under Physiologically Relevant Conditions. J. Nat. Prod. 2019, 12, 3386-3393.
- (21) Woodward, R. B.; Hoye, T. R. Total synthesis of illudinine, illudalic acid, and illudacetalic acid. J. Am. Chem. Soc. 1977, 99, 8007-8014.
- (22) Teske, J. A.; Deiters, A. Microwave-Mediated Nickel-Catalyzed Cyclotrimerization Reactions: Total Synthesis of Illudinine. *J. Org. Chem.* **2008**, *73*, 342-345.
- (23) Fleming, I.; de Marigorta, E. M. Silyl-cupration of an acetylene followed by ring-formation. J. Chem. Soc., Perkin Trans. 1 1999, 889-900.
- (24) (a) Schreiber, J.; Felix, D.; Eschenmoser, A.; Winter, M.; Gautschi, F.; Schulte-Elte, K. H.; Sundt, E.; Ohloff, G.; Kalovoda, J.; Kaufmann, H.; Wieland, P. Die Synthese von Acetylen-carbonyl-Verbindungen durch Fragmentierung von α, β-Epoxy-ketonen mit p-Toluolsulfonylhydrazin. Vorläufige Mitteilung. *Helv. Chim. Acta* **1967**, *50*, 2101-2108. (b) Felix, D.; Schreiber, J.; Ohloff, G.; Eschenmoser, A. α,β-Epoxyketon→Alkinon-Fragmentierung I: Synthese von exalton und rac-muscon aus cyclododecanon über synthetische methoden, 3. Mitteilung. *Helv. Chim. Acta*. **1971**, *54*, 2896–2912. (c) Fioroni, G.; Fringuelli, F.; Pizzo, F.; Vaccaro, L. Epoxidation of α,β-unsaturated ketones in water. An environmentally benign protocol. *Green Chem*. **2003**, *5*, 425-428. (d) Kürti, L.; Czakó, B. Eschenmoser– Tanabe fragmentation. In *Strategic Applications of Named*

Reactions in Organic Synthesis; Elsevier: New York, 2005; pp 158–159, and references cited.

(25) (a) Prantz, K.; Mulzer, J. Synthetic applications of the carbonyl-generating Grob fragmentation. *Chem. Rev.* 2010, *110*, 3741–3766. (b) Hoang, T. T.; Dudley, G. B.; Williams, L. J. Fragmentation Reactions. In *Comprehensive Organic Synthesis*, 2nd Edition; Molander, G., Knochel, P., Eds.; Elsevier: Oxford, 2014; Vol. 6, Chap. 30, 842–860. (c) Yang, J.; Hoang, T. T.; Dudley, G. B. Alkynogenic fragmentation. *Org. Chem. Front.* 2019, *6*, 2560–2569.

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56 57 58

59

60

- (26) (a) Kamijo, S.; Dudley, G. B. Tandem nucleophilic addition/fragmentation reactions and synthetic versatility of vinylogous acyl triflates. J. Am. Chem. Soc. 2006, 128, 6499-6507. (b) Tummatorn, J.; Dudley, G. B. Ring opening / fragmentation of dihydropyrones for the synthesis of homopropargyl alcohols. J. Am. Chem. Soc. 2008, 130, 5050-5051. (c) Jones, D. M.; Lisboa, M. P.; Kamijo, S.; Dudley, G. B. Ring opening of cyclic vinylogous acyl triflates using stabilized carbanion nucleophiles: Claisen condensations linked to carboncarbon bond cleavage. J. Org. Chem. 2010, 75, 3260-3267. (d) Lisboa, M. P.; Hoang, T. T.; Dudley, G. B. Tandem nucleophilic addition / fragmentation of vinylogous acyl triflates: 2-methyl-2-(1-oxo-5-heptynyl)-1,3-dithiane. Org. Synth. 2011, 88, 353-363. (e) Tummatorn, J.; Dudley, G. B. Generation of medium-ring cycloalkynes by ring expansion of vinylogous acyl triflates. Org. Lett. 2011, 13, 1572-1575.
- (27) Hoang, T. T.; Dudley, G. B. Synthesis of high-value 1,6-enynes by tandem fragmentation / olefination. *Org. Lett.* **2013**, *15*, 4026–4029.
- (28) (a) Morrison, A. E.; Hrudka, J. J.; Dudley, G. B. Thermal cycloisomerization of putative allenylpyridines for the synthesis of isoquinoline derivatives. *Org. Lett.* 2016, *18*, 4104–4107. (b) Kramer, N. J.; Hoang, T. T.; Dudley, G. B. Reaction discovery using neopentylene-tethered coupling partners: cycloisomerization/oxidation of electron-deficient dienynes. *Org. Lett.* 2017, *19*, 4636–4639.
- (29) Morrison, A. E.; Hoang, T. T.; Birepinte, M.; Dudley, G. B. Synthesis of illudinine from dimedone. *Org. Lett.* 2017, *19*, 858–861.
- (30) Duangkamol, C.; Batsomboon, P.; Stiegman, A. E.; Dudley, G. B. Microwave heating outperforms conventional heating for a thermal reaction that produces a thermally labile product: Observations consistent with selective microwave heating. *Chem.–Asian J.* 2019, 14, 2594–2597.
- (31) We have attempted to hydrolyze illudinine to the methyl ether of illudalic acid — the reverse of the condensation reaction with ammonia reported by Woodward and Hoye (Eq 1) — under a variety of conditions but to no avail.
- (32) (a) Kocsis, L. S.; Kagalwala, H. N.; Mutto, S.; Godugu, B.; Bernhard, S.; Tantillo, D. J.; Brummond, K. M. Mechanistic Insight into the Dehydro-Diels–Alder Reaction of Styrene–Ynes. J. Org. Chem. 2015, 80, 11686-11698. (b) Benedetti, E.; Kocsis, L. S.; Brummond, K. M. Synthesis and photophysical properties of a series of cyclopenta[b]naphthalene solvatochromic fluorophores. J. Am. Chem. Soc. 2012, 134, 12418-12421. (c) Kocsis, L. S.;

Benedetti, E.; Brummond, K. M. Thermal Dehydrogenative Diels–Alder Reaction of Styrenes for the Concise Synthesis of Functionalized Naphthalenes. *Org. Lett.* **2012**, *14*, 4430-4433.

- (33) (a) Spino, C.; Pesant, M.; Dory, Y. A New Look at the Diels–Alder Transition State. *Angew. Chem. Int. Ed.* 1998, 37, 3262-3265. (b) Bodwell, G. J.; Pi, Z.; Pottie, I. R. Electron Deficient Dienes. 2. One Step Synthesis of a Coumarin-Fused Electron Deficient Diene and its Inverse Electron Demand Diels-Alder Reactions with Enamines. *Synlett* 1999, 4, 477-479. (c) Domingo, L. R. Why do electron-deficient dienes react rapidly in diels-alder reactions with electron-deficient ethylenes? A density functional theory analysis. *Eur. J. Org. Chem.* 2004, 4788-4793.
- (34) Wu, W.; Jiang, H. Haloalkynes: A Powerful and Versatile Building Block in Organic Synthesis. Acc. Chem. Res. 2014, 47, 2483–2504.
- (35) (a) Prall, M.; Wittkopp, A.; Fokin, A. A.; Schreiner, P. R. Substituent effects on the Bergman cyclization of (Z)-1,5hexadiyne-3-enes: a systematic computational study. J. Comput. Chem. 2001, 22, 1605-1614. (b) Bent, H. A. An Appraisal of Valence-bond Structures and Hybridization in Compounds of the First-row elements. Chem. Rev. 1961, 61, 275-311.
- (36) (a) Dudley, G. B.; Stiegman, A. E. Changing perspectives on the strategic use of microwave heating in organic synthesis. *Chem. Rec.* 2018, *18*, 381–389. (b) Dudley, G. B.; Richert, R.; Stiegman, A. E. On the Existence of and Mechanism for Microwave-Specific Reaction Rate Enhancement. *Chem. Sci.* 2015, *6*, 2144–2152.
- (37) Fier, P. S.; Maloney, K. M. Direct Conversion of Haloarenes to Phenols under Mild, Transition-Metal-Free Conditions. *Org. Lett.* **2016**, *18*, 2244–2247.
- (38) (a) Aoyama, T.; Terasawa, S.; Sudo, K.; Shioiri, T. New methods and reagents in organic synthesis. 46. Trimethylsilyldiazomethane: a convenient reagent for the o-methylation of phenols and enols. *Chem. Pharm. Bull.* **1984**, *32*, 3759-3760. (b) Boger, D. L.; Takahashi, K. Total synthesis of granditropone, grandirubrine, imerubrine, and isoimerubrine. *J. Am. Chem. Soc.* **1995**, *117*, 12452-12459. (c) Xu, W.; Huang, L. F.; Bauer, L.; Bhargava, H. N.; Dunn, W. J. Synthesis and opiate receptor binding properties of 17-methyl-6, 7-dehydro-3, 14-dihydroxy-4, 5α-epoxy-6, 7: 4', 5'-pyrimidinomorphinans *Bioorg. Med. Chem. Lett.* **1999**, *23*, 3375-3380.
- (39) Brown, W. D.; Gouliaev, A. H. Bromination of isoquinoline, quinoline, quinazoline and quinoxaline in strong acid. *Synthesis* 2002, 83-86.
- (40) Finberg, J. P.; Rabey, J. M. Inhibitors of MAO-A and MAO-B in Psychiatry and Neurology. *Front. Pharmacol.* 2016, 7, 340 and references cited therein.
- (41) AAT Bioquest, Inc. (2020, Jan 10). "Quest Graph™IC50 Calculator." <u>https://www.aatbio.com/tools/ic50-</u> calculator
- (42) Kamijo, S.; Dudley, G. B. Cyclic vinylogous triflate hemiacetals as new surrogates for alkynyl aldehydes. *Tetrahedron Lett.* **2006**, *47*, 5629–5632.

Graphical Abstract:

