

Article

Subscriber access provided by La Trobe University Library

Chemical Resolution of C,N-Unprotected #-Substituted-#-Amino Acids Using Stable and Recyclable Proline-derived Chiral Ligands

Shuni Wang, Yong Nian, Shengbin Zhou, Jiang Wang, and Hong Liu

J. Org. Chem., Just Accepted Manuscript • DOI: 10.1021/acs.joc.8b01336 • Publication Date (Web): 13 Jul 2018

Downloaded from <http://pubs.acs.org> on July 15, 2018**Just Accepted**

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



ACS Publications

is published by the American Chemical Society, 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

1
2
3 **Chemical Resolution of C,N-Unprotected α -Substituted- β -Amino Acids Using**
4 **Stable and Recyclable Proline-derived Chiral Ligands**
5
6
7

8 Shuni Wang,^{†,‡,§} Yong Nian,^{†,‡,§} Shengbin Zhou,^{†,‡} Jiang Wang^{†,‡,*} and Hong Liu^{†,‡,*}
9
10

11 [†] State Key Laboratory of Drug Research and CAS Key Laboratory of Receptor Research,
12 Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 Zuchongzhi Road,
13 Shanghai 201203, China.
14

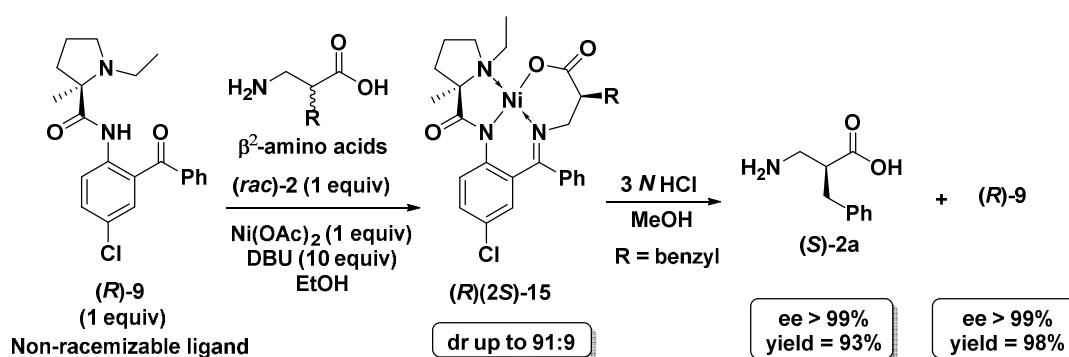
15 [‡] University of Chinese Academy of Sciences, No.19A Yuquan Road, Beijing 100049, China.
16

17 [§] These authors contributed equally.
18
19

20 E-mail: hliu@simm.ac.cn (Hong Liu); jwang@simm.ac.cn (Jiang Wang)
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table of Contents



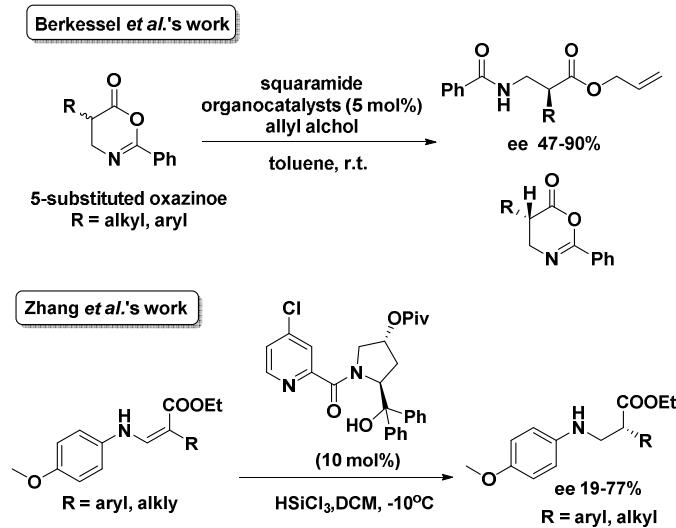
Abstract

We report the first purely chemical method for the resolution of *C,N*-unprotected racemic α -substituted- β -amino acids (β^2 -AAs) using thermodynamically stable and recyclable chiral proline-derived ligands. The ligands and racemic β^2 -AAs along with Ni(II) could form a pair of Ni(II) complex diastereoisomers with desirable diastereoselectivity (dr up to 91:9). Enantiomerically pure *C,N*-unprotected β^2 -AAs could be obtained by simple hydrolysis of isolated favored-Ni(II) complex. The method featured unique versatility compared with enzymatic approaches, and characterized by its broad synthetic generality, good stereochemical outcome and mild reaction conditions, thus making it a powerful supplement in the field of chemical resolution of β^2 -AAs.

Introduction

α -Substituted- β -amino acids (β^2 -AAs) are being widely used increasingly in the pharmaceutical and healthcare industries,¹⁻⁸ and play as the vital building blocks in the pharmaceuticals and natural products.⁹⁻¹¹ Preparation of structurally varied β^2 -AAs in enantiomerically pure forms has been a research focus in synthetic organic chemistry, leading to several ingenious asymmetric synthesis approaches.¹²⁻¹³ However, current asymmetric synthesis methods to prepare enantiomerically pure β^2 -AAs are relatively cumbersome, involving expensive, dangerous or toxic reagents, and requiring multiple steps to prepare substrates and elaborate reaction conditions.¹⁴⁻¹⁶ By contrast, the resolution of racemic β^2 -AAs is still used frequently in

practice, because there are a variety of methods to prepare racemic β^2 -AAs.¹⁷⁻¹⁸ However, the enzymatic resolution of racemic β^2 -AA derivatives is limited to only a few examples.¹⁹⁻²⁵ For example, Berkessel *et al.* reported the enzymatic dynamic kinetic resolution of oxazinones¹⁹ or β -lactams²⁴ to obtain optically pure β^2 -AAs. These enzymatic approaches require high throughput screening for selective enzymes and multi-step preparation for special substrates. Similarly, the exploration of purely chemical methods for the resolution of racemic β^2 -AAs is almost omitted with limited literatures. Berkessel *et al.* reported the kinetic resolution of 5-substituted oxazinone substrates with chiral squaramide organocatalysts to afford *N*-protected β^2 -amino esters *via* alcoholysis ring opening.²⁶ Zhang *et al.* reported chiral Lewis base organocatalyzed asymmetric hydrosilylation of α -substituted β -enamino ester to afford enantioenriched β^2 -amino esters *via* dynamic kinetic resolution.²⁷ Both methods require preparation of special β^2 -AAs derivatives and resulted products are *C,N*-protected or substituted β^2 -AAs, thus unfriendly to following applications (Scheme 1). Therefore, a purely chemical resolution of *C,N*-unprotected β^2 -AAs, starting from simple and cheap starting materials, to obtain enantiomerically pure *C,N*-unprotected β^2 -AAs, remains an important challenge for synthetic organic chemists.



Scheme 1. Reported chemical resolution methods to obtain chiral β^2 -AAs or their derivatives.

Various enantiomerically pure non-natural amino acids, including α -AAs and β^3 -AAs, could be obtained by dynamic resolution of racemic AAs *via* forming Ni(II) complexes with chiral ligands (Figure 1).²⁸ On the other hand, we have previously synthesized a series of racemic

α -aryl-/alkyl-substituted β -amino acids using Ni(II) complexed with the achiral PBP ligand.^{18, 29-30} Encouraged by these works, we proposed to explore the possibility of chemical resolution of *C,N*-unprotected racemic β^2 -AAs *via* a chiral ligand by forming Ni(II) complex to obtain the enantiomerically pure forms (Scheme 2). Unlike α -amino acids (α -AAs),³¹⁻³³ the kinetic rate of chelation reaction of β^2 -AAs with the chiral ligand and Ni(II) is comparatively lower, since the resulted flexible six-membered ring goes against the Ni(II)-centered quadrilateral plane.

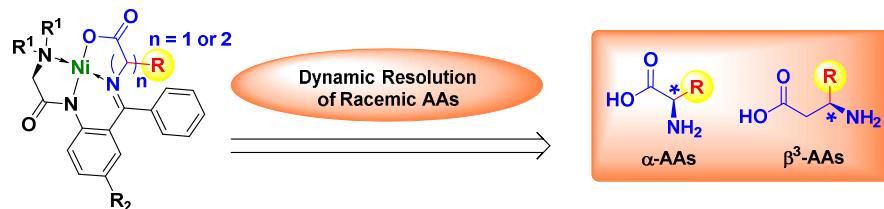
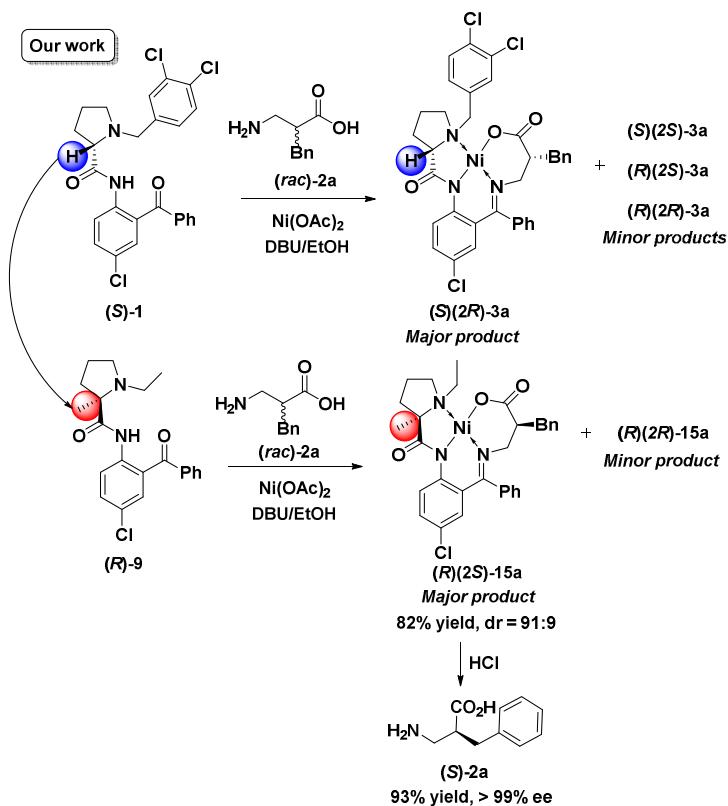


Figure 1. Dynamic resolution of racemic amino acids using Ni(II) complex



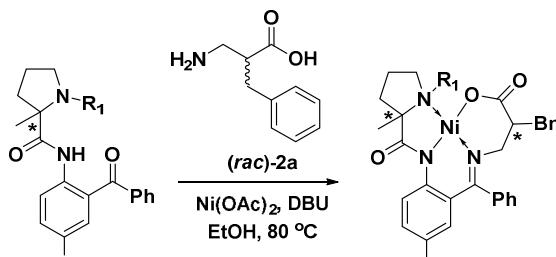
Scheme 2. Chemical resolution of *C,N*-unprotected racemic β^2 -AAs by Ni(II) complexes.

Results and Discussion

In initial stage of the attempts, we selected the chiral proline-derived ligand **1** for the dynamic resolution of *C,N*-unprotected β^2 -AAs, which has been successfully applied in the chemical resolution of α -AAs and β -substituted- β -amino acids (β^3 -AAs).^{32,34} Interestingly, the reaction of racemic α -benzyl- β -alanine (*rac*)-**2a** (1 equiv) with (*S*)-**1** (1 equiv) generated two expected Ni(II) complex products, (*S*)(*2R*)-**3a** and (*S*)(*2S*)-**3a**, and two unexpected products, (*R*)(*2S*)-**3a** and (*R*)(*2R*)-**3a** (Scheme 1, Figures S1 and S2). Theoretically, racemization was able to occur at the α -position of both proline and β^2 -AA moieties in the structure of the Ni(II) complex **3a** under base-catalyzed epimerization. According to above results, we could reasonably assume that the pK_a of α -H of the proline moiety is comparable with that of the β^2 -AA moiety in the structure of the Ni(II) complex **3a**, which suggested that ligand (*S*)-**1** is inappropriate as a chiral auxiliary for the thermodynamic resolution of racemic β^2 -AAs. To solve the thermodynamic instability of ligand **1**, we then designed a ligand, (*S*)-**4**, with a quaternary carbon stereogenic center by introducing a methyl at the α -position of the proline moiety in the structure of ligand **1**.

To confirm our ideas, we then reacted (*S*)-**4** with (*rac*)-**2a** under the standard reaction conditions: heating an ethanol solution of the ligand (*S*)-**4**, (*rac*)-**2a**, anhydrous Ni(OAc)₂ and DBU. In this case, as expected, we did not observe (*R*)(*2R*)-**10a** and (*R*)(*2S*)-**10a** (Figures S3 and S4), and the target nickel(II) complexes, (*S*)(*2R*)-**10a** and (*S*)(*2S*)-**10a**, were obtained at a yield of 76% and in a ratio of 81:19 (Table 1, entry 1). These results were encouraging, as they established that the direct chemical resolution of unprotected β^2 -AAs by a simple reaction with (*S*)-**6** was feasible. Fortunately, we could easily got the diastereoisomers (*S*)(*2R*)-**10a** and (*S*)(*2S*)-**10a** by column chromatography on silica gel (v/v, dichloromethane/methanol = 20/1). After the determination of dr value of the diastereoisomers by ¹H NMR, the enantiomerically pure (*S*)(*2R*)-**10a** could be obtained by column chromatography on silica gel once again (v/v, dichloromethane/acetone = 5/1, R_f ((*S*)(*2R*)-**10a**) = 0.4, R_f ((*S*)(*2S*)-**10a**) = 0.25). The definite and absolute configurations of Ni(II) complexes (*S*)(*2R*)-**10a** and (*S*)(*2S*)-**10a** were determined by X-ray analysis (Figure S5).³⁵

Table 1. Optimization of chiral ligands for chemical resolution of unprotected racemic α -benzyl- β -alanine **2a**.^a



- (S)-4 $R_1 = 3,4\text{-dichlorobenzyl}$
 (S)-5 $R_1 = \text{benzyl}$
 (S)-6 $R_1 = 3,4\text{-dibromobenzyl}$
 (S)-7 $R_1 = 3,4\text{-dimethoxybenzyl}$
 (S)-8 $R_1 = \text{phenylpropyl}$
 (R)-9 $R_1 = \text{ethyl}$
- (S)(2R)-10a
 (S)(2R)-11a
 (S)(2R)-12a
 (S)(2R)-13a
 (S)(2R)-14a
 (R)(2S)-15a

Entry	Ligand	Ni(II)-complex	yield (%) ^b	dr ^c
1	(S)-4	(S)(2R)-10a	76	81/19
2	(S)-5	(S)(2R)-11a	74	83/17
3	(S)-6	(S)(2R)-12a	89	68/32
4	(S)-7	(S)(2R)-13a	83	69/31
5	(S)-8	(S)(2R)-14a	69	82/18
6	(R)-9	(R)(2S)-15a	82	91/9

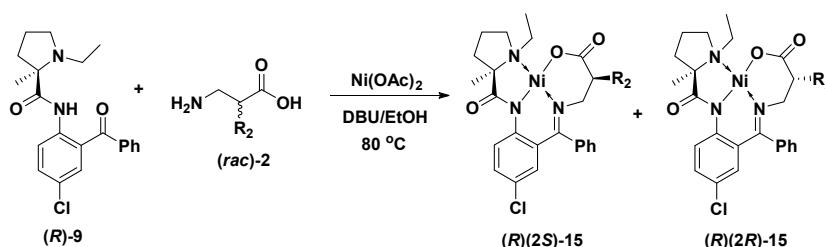
^a Reaction condition: Ligand (0.1 mmol), (rac)-2a (0.1 mmol), anhydrous Ni(OAc)₂ (0.1 mmol) and DBU (1 mmol) were refluxed in ethanol (2 mL) at 80 °C for 96 h. ^b Combined yield of isolated products 10a-15a. ^c dr was determined by ¹H NMR analysis of isolated products 10a-15a.

To further improve the stereochemical outcome, various attempts at modifying of ligand 4 were conducted (Table 1, entries 2-6). The synthesis method of the proline-derived ligands 4-9 was extremely similar with the previous reported method, and ligands 4-5 and 8-9 have been well characterized.³⁶ When we substituted 3,4-dichlorobenzyl with a benzyl moiety on proline, it gave a comparable yield and diastereoselectivity (Table 1, entry 2). An apparent increase in yield and decrease in diastereoselectivity were observed when introducing electron-donating groups on the benzyl moiety (Table 1, entries 3-4). Prolonging benzyl to phenylpropyl gave a comparative diastereoselectivity but an undesired decrease in yield (Table 1, entry 5). Finally, it was found that ligand (R)-9 with an ethyl substitution on proline showed a relatively good yield (82%) and diastereoselectivity (dr = 91:9) of the Ni(II) complex (Table 1, entry 6), which has been also selected as the best proline-derived ligand in the resolution of racemic β^3 -AAs.³⁶ Further

1
2
3 investigation on the reaction time of ligand (*R*)-**9** with (*rac*)-**2a** revealed that the final
4 thermodynamic control stopped at 96 h was the best choice. And with the longer reaction time,
5 diastereoisomer (*R*)(2*R*)-**15a** could surely be interconverted to the thermodynamically more stable
6 diastereoisomer (*R*)(2*S*)-**15a** while accompanied with the decomposition of Ni(II) complex **15a**
7 (Table S1).
8
9

10 Furthermore, we investigated the generality of (*R*)-**9** in the resolution of β^2 -AAs (Table 2).
11 First, the α -benzyl-substituted β^2 -AAs were applied successfully with satisfactory yields ($\geq 70\%$)
12 and good diastereoselectivity ($dr \geq 84:16$) (Table 2, entries 1-5). Importantly, for these
13 benzyl-substituted β^2 -AAs, the electronic property of the substituents on the benzyl group did not
14 show any apparent effects on the stereochemical outcome of the reactions. In addition, β^2 -AAs
15 bearing aliphatic substituents, such as methyl, *i*-butyl, *i*-propyl, and cyclohexyl also exhibited
16 good yield ($\geq 80\%$) and a slight decrease in diastereoselectivity compared with the
17 benzyl-substituted β^2 -AAs (Table 2, entries 6-9). For the sterically hindered *t*-butyl-substituted
18 β^2 -AA, the Ni(II) complexes **15j** were isolated with a moderate yield and diastereochemistry ($dr =$
19 62:38) (Table 2, entry 10). Unfortunately, the α -aryl-substituted β -AAs (*rac*)-**2k-2n** (Table 2,
20 entries 11-14), bearing electron-withdrawing as well as electron-donating groups, and the
21 sterically hindered 2-naphthyl, all resulted in products with poor diastereoselectivities ($dr \leq 62 :$
22 38).
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37

38 **Table 2.** Chemical resolution of various *C,N*-unprotected β^2 -AAs (*rac*)-**2** using ligand (*R*)-**9**.^a



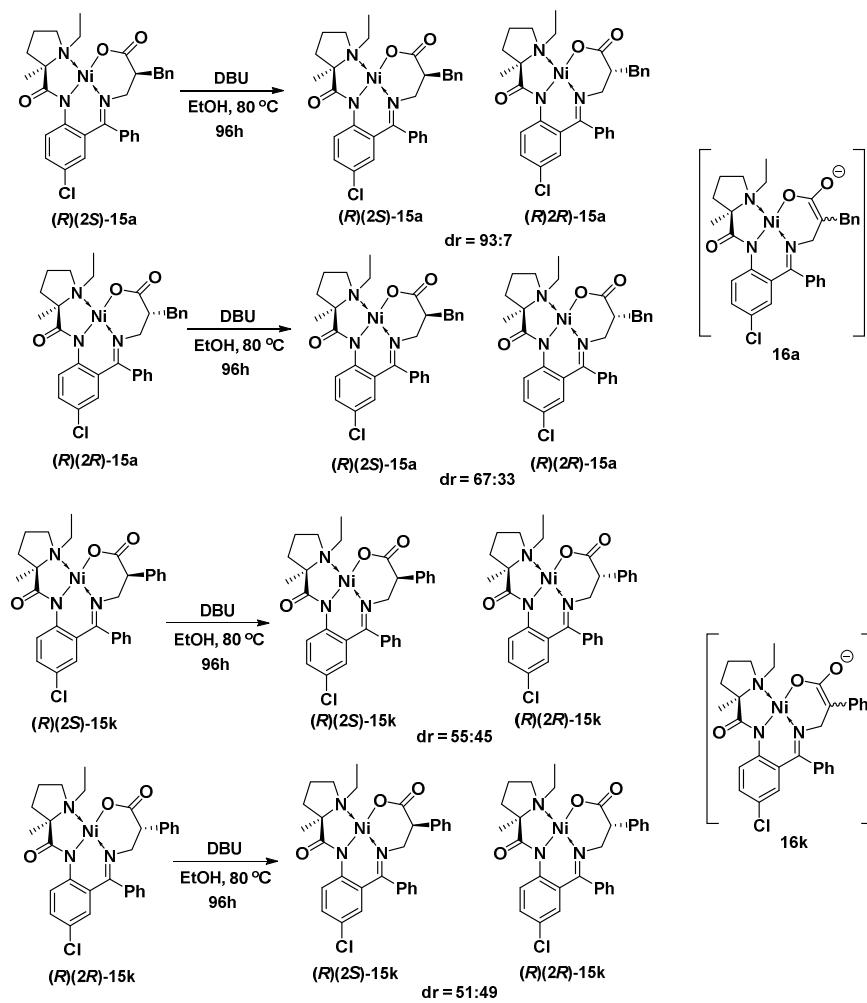
Entry	R ₂	Ni(II)-complex	Yield (%) ^b	dr ^c
1	benzyl	(R)(2S)-15a	82	91/9
2	4-OMe-benzyl	(R)(2S)-15b	83	88/12
3	4-Me-benzyl	(R)(2S)-15c	70	84/16
4	4-F-benzyl	(R)(2S)-15d	76	89/11

5	4-CF ₃ -benzyl	(R)(2S)-15e	72	89/11
6	methyl	(R)(2S)-15f	83	73/27
7	i-butyl	(R)(2S)-15g	80	83/17
8	i-propyl	(R)(2S)-15h	81	76/24
9	cyclohexyl	(R)(2S)-15i	84	83/17
10	t-butyl	(R)(2S)-15j	69	62/38
11	phenyl	(R)(2S)-15k	73	51/49
12	4-MeO-phenyl	(R)(2S)-15l	65	54/46
13	4-Cl-phenyl	(R)(2S)-15m	69	62/38
14	2-naphthyl	(R)(2S)-15n	76	56/44

^a Reaction condition: (R)-9 (0.1 mmol), (rac)-2 (0.1 mmol), anhydrous Ni(OAc)₂ (0.1 mmol) and DBU (1 mmol) were refluxed in ethanol (2 mL) at 80 °C for 96 h. ^b Combined yield of (R)(2S)-15 and (R)(2R)-15. ^c dr was determined by ¹H NMR analysis of the mixtures of (R)(2S)-15 and (R)(2R)-15.

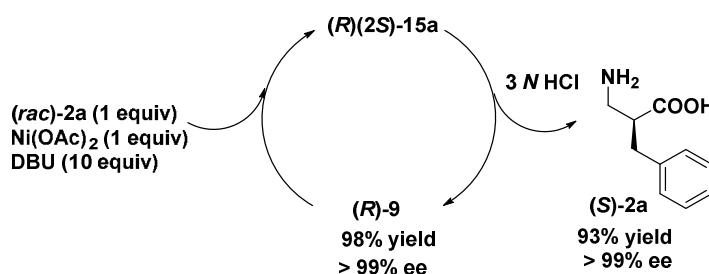
As we can see in Table 2, the yield of (R)(2S)-15a was 75%, exceeding more than 50%, convincingly indicating that the resolution of racemic β²-AAs was a dynamic process. To further understand this process, we isolated enantiomerically pure (R)(2S)-15a and (R)(2R)-15a, respectively, and treated with DBU (10 equiv) in EtOH. After heating for 96 h, 7% of pure (R)(2S)-15a was converted to (R)(2R)-15a, and in contrast, 67% of pure (R)(2R)-15a was transformed to (R)(2S)-15a, fully illustrating the two diastereoisomers could be interconverted via keto-enol tautomerism (enol 16a as the possible intermediate), and (R)(2S)-15a was undoubtedly identified as the thermodynamically favored diastereoisomer (Scheme 3). When it came to Ni(II) complex 15k, whether (R)(2S)-15k or (R)(2R)-15k as the starting diastereoisomer, the ratio of (R)(2S)-15k and (R)(2R)-15k were both nearly 50:50 after the thermodynamic equilibrium transformation (Scheme 2), which consisted with the resolution result (Table 2, entry 11). One may agreed that the (R)(2S)-15k and (R)(2R)-15k featured with comparable intrinsic thermodynamical stability, thus exhibiting low diastereoselectivity in the resolution process. According to the previous report, the phenyl ring of the benzyl group could shield the Nickel atom

to form extra metal-phenyl attraction, which possibly illustrated the better diastereoselectivity for resolution of **2a** than **2k**.³⁷



Scheme 3. Thermodynamic equilibrium transformation of **15a** and **15k**.

To achieve the final goal of this study, we selected **2a** as an example of resolution of β^2 -AAs to obtain the enantiomerically pure forms. After simple purification by column chromatography, diastereoselectively pure *(R)(2S)-15a* was subjected to the disassembly procedure presented in Scheme 4. The disassembly procedure was also conducted under simple reaction conditions, thus producing the target α -benzyl- β -amino acid (*S*)-**2a** with an excellent chemical yield (93%) and enantioselectivity (ee > 99%). Furthermore, *(R)-9* could be recycled easily in almost quantitative yield (98%) with high enantiomeric purity (ee > 99%). Thus, the recovered *(R)-9* could be reused for repetitive thermodynamic resolution of other racemic β^2 -AAs.



Scheme 4. Disassembly of Nickel(II) complex (R)(2S)-15a to afford (S)-2a.

Conclusion

In conclusion, we have developed the first purely chemical method for the resolution of *C,N*-unprotected racemic β^2 -AAs using stable, and recyclable proline-derived ligands with a quaternary carbon stereogenic center. This new approach is more versatile compared with biocatalytic methods, especially without requiring multi-step preparation for resolution substrates. The method showed a broad synthetic generality for various α -alkyl-substituted *C,N*-unprotected β -AAs with good to moderate diastereoselectivities, featuring with simple operation and mild reaction conditions.

Experimental Section

General Information

The commercially available chemicals were used without further purification. Anhydrous nickel acetate was prepared by heating reagent grade tetrahydrate for 2 h at 110°C in vacuum. ¹H and ¹³C NMR spectra was recorded on a 400 MHz or 500 MHz Bruker AV400 instrument. Chemical shifts were reported in parts per million (ppm, δ) downfield from tetramethylsilane. Proton coupling patterns are described as singlet (s), doublet (d), triplet (t), quartet (q), multiplet (m), and broad (br). High-resolution mass spectra (HRMS) was measured on Micromass Ultra Q-TOF spectrometer. The determination of dr was performed *via* ¹H NMR or LC/MS analysis using Agilent 6120 spectrometer. Optical rotations were measured using a 1 mL cell with a 1 dm path length on an Autopol VI automatic polarimeter and are reported as follows: $[\alpha]^{20}_D$ (c: g/100 mL, in solvent).

General Procedures

1
2
3 **Nickel(II)-(S)-N-(2-benzoyl-4-chlorophenyl)-1-(3,4-dichlorobenzyl)-2-methylpyrrolidine-2-ca
4 rboxamide/(R)-3-amino-2-benzylpropanoic acid Schiff Base Complex ((S)(2R)-10a)**
5
6

7 (*S*)-4 (50.20 mg, 0.10 mmol), 3-amino-2-benzylpropanoic acid **2a** (17.93 mg, 0.10 mmol),
8 Ni(OAc)₂ (17.68 mg, 0.10 mmol), were dissolved in EtOH (2 mL) followed by DBU (149.31 μ L,
9 1.00 mmol) being added. After being refluxed for 96 h, the reaction was terminated by ice water of
10 5% acetic acid (10 mL). The mixture was extracted with dichloromethane (10 mL \times 3). The
11 combined organic layers were dried with Na₂SO₄, and then concentrated and purified by column
12 chromatography on silica gel (v/v, dichloromethane/methanol = 20/1) to give the crude products
13 **10a** (55.1 mg, yield 76%) for analysis (dr = 81/19). The crude product **10a** was purified again by
14 column chromatography on silica gel (v/v, dichloromethane/acetone = 5/1, R_f((*S*)(2*R*)-**10a**) = 0.4,
15 R_f((*S*)(2*S*)-**10a**) = 0.25) to give the major pure diastereomer (*S*)(2*R*)-**10a** as a brown solid. mp
16 128–130°C. [α]²⁰_D = +2426.5 (c = 0.034, CHCl₃). ¹H NMR (400 MHz, Methanol-*d*₄) δ 8.91 (d, *J* =
17 2.1 Hz, 1H), 8.32 (dd, *J* = 8.3, 2.1 Hz, 1H), 7.74 (d, *J* = 9.2 Hz, 1H), 7.48 (d, *J* = 8.2 Hz, 1H), 7.39
18 – 7.25 (m, 2H), 7.23 – 7.12 (m, 4H), 7.06 (dd, *J* = 9.2, 2.6 Hz, 1H), 7.03 – 6.95 (m, 3H), 6.64 (dt,
19 *J* = 7.6, 1.6 Hz, 1H), 6.42 (d, *J* = 2.6 Hz, 1H), 3.98 – 3.83 (m, 2H), 3.77 (d, *J* = 13.3 Hz, 1H), 3.58
20 – 3.48 (m, 1H), 3.40 (d, *J* = 13.3 Hz, 1H), 3.27 – 3.18 (m, 1H), 3.14 – 3.02 (m, 2H), 2.68 – 2.62
21 (m, 1H), 2.51 – 2.10 (m, 4H), 1.43 (s, 3H). ¹³C NMR (125 MHz, Methanol-*d*₄) δ 182.5, 180.4,
22 173.1, 141.3, 139.8, 138.2, 134.9, 134.6, 133.9, 133.5, 133.1, 132.7, 132.4, 132.2, 131.2, 131.0,
23 130.3, 130.2, 129.9, 129.6, 128.1, 127.7, 127.6, 126.8, 125.9, 74.9, 56.7, 56.3, 55.3, 49.3, 42.8,
24 37.2, 22.2, 18.6. LRMS (ESI+APCI) *m/z*: 718.0. HRMS (ESI-TOF) *m/z*: [M + H]⁺ Calcd for
25 C₃₆H₃₃Cl₃N₃NiO₃ 718.0935; Found 718.0940.

26
27
28
29 **(S)-N-(2-benzoyl-4-chlorophenyl)-1-(3,4-dibromobenzyl)-2-methylpyrrolidine-2-carboxamid
30 e ((S)-6)**
31

32 Yellow oil (3.7 g, yield 90%). [α]²⁰_D = -26.2 (c = 0.118, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ
33 11.72 (s, 1H), 8.63 (d, *J* = 8.7 Hz, 1H), 7.83 (d, *J* = 1.9 Hz, 1H), 7.76 (dd, *J* = 8.2, 1.4 Hz, 2H),
34 7.65 – 7.62 (m, 1H), 7.55 – 7.47 (m, 4H), 7.31 (d, *J* = 8.2 Hz, 1H), 7.18 (d, *J* = 8.2 Hz, 1H), 3.77
35 (d, *J* = 13.5 Hz, 1H), 3.38 (d, *J* = 13.5 Hz, 1H), 3.16 (t, *J* = 8.1 Hz, 1H), 2.40 (q, *J* = 8.1 Hz, 1H),
36 2.23 – 2.13 (m, 1H), 1.91 – 1.75 (m, 3H), 1.39 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 197.0,
37 176.5, 140.0, 138.4, 137.9, 133.8, 133.4, 133.3, 133.0, 132.2, 130.1, 128.8, 128.5, 127.2, 125.9,

1
2
3 124.7, 122.9, 122.8, 68.8, 53.4, 51.3, 40.1, 22.6, 16.4. LRMS (ESI+APCI) *m/z*: 590.8. HRMS
4 (ESI-TOF) *m/z*: [M + H]⁺ Calcd for C₂₆H₂₄Br₂ClN₂O₂ 588.9888; Found 588.9877.
5
6
7
8
9

10 **(S)-*N*-(2-benzoyl-4-chlorophenyl)-1-(3,4-dimethoxybenzyl)-2-methylpyrrolidine-2-carboxamido**
11 **(*S*)-7**

12 Yellow oil (3.5 g, yield 92%). $[\alpha]^{20}_D = -34.3$ (*c* = 0.103, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ
13 11.64 (s, 1H), 8.56 (d, *J* = 9.0 Hz, 1H), 7.73 – 7.69 (m, 2H), 7.64 – 7.59 (m, 1H), 7.52 – 7.48 (m,
14 3H), 7.45 (d, *J* = 2.5 Hz, 1H), 7.08 (d, *J* = 2.0 Hz, 1H), 6.83 (dd, *J* = 8.2, 1.9 Hz, 1H), 6.58 (d, *J*=
15 8.2 Hz, 1H), 3.79 (s, 3H), 3.77 (s, 3H), 3.37 (d, *J* = 13.0 Hz, 1H), 3.14 (td, *J* = 8.1, 7.6, 4.1 Hz,
16 1H), 2.43 – 2.36 (m, 1H), 2.18 – 2.08 (m, 1H), 1.85 – 1.69 (m, 4H), 1.39 (s, 3H). ¹³C NMR (125
17 MHz, CDCl₃) δ 196.8, 176.9, 148.9, 147.9, 138.2, 137.9, 133.2, 132.9, 131.8, 131.5, 130.0, 128.5,
18 127.3, 126.6, 123.2, 120.6, 111.8, 110.7, 68.5, 55.8, 54.0, 51.1, 40.2, 22.6, 16.2. LRMS
19 (ESI+APCI) *m/z*: 493.0. HRMS (ESI-TOF) *m/z*: [M + H]⁺ Calcd for C₂₈H₃₀ClN₂O₄ 493.1889;
20 Found 493.1875.
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

**Nickel(II)-(S)-*N*-(2-benzoyl-4-chlorophenyl)-1-benzyl-2-methylpyrrolidine-2-carboxamide/
(R)-3-amino-2-benzylpropanoic acid Schiff Base Complex ((*S*)(2*R*)-11a)**

Brown solid (48.1 mg, yield 74%). mp 128–130 °C. $[\alpha]^{20}_D = +2739.1$ (*c* = 0.046, CHCl₃). ¹H NMR
(400 MHz, Methanol-*d*₄) δ 8.58 – 8.50 (m, 2H), 7.59 (d, *J* = 9.1 Hz, 1H), 7.40 – 7.26 (m, 4H),
7.20 – 7.11 (m, 5H), 7.01 – 6.96 (m, 4H), 6.66 (dt, *J* = 7.3, 1.7 Hz, 1H), 6.40 (d, *J* = 2.6 Hz, 1H),
3.97 – 3.81 (m, 2H), 3.76 (d, *J* = 13.1 Hz, 1H), 3.65 – 3.53 (m, 1H), 3.40 (d, *J* = 13.1 Hz, 1H),
3.30 – 3.21 (m, 1H), 3.12 – 3.04 (m, 2H), 2.65 – 2.59 (m, 1H), 2.47 – 2.11 (m, 4H), 1.46 (s, 3H).
¹³C NMR (125 MHz, Methanol-*d*₄) δ 182.8, 180.5, 172.9, 141.6, 139.9, 137.1, 135.0, 132.8, 132.6,
132.3, 131.2, 131.1, 130.2, 130.1, 130.1, 130.0, 129.6, 129.5, 128.1, 127.7, 127.6, 126.8, 126.5,
75.1, 57.9, 56.8, 55.3, 49.1, 42.9, 37.2, 22.6, 18.6. LRMS (ESI+APCI) *m/z*: 650.1. HRMS
(ESI-TOF) *m/z*: [M + H]⁺ Calcd for C₃₆H₃₅ClN₃NiO₃ 650.1715; Found 650.1721.

**Nickel(II)-(S)-*N*-(2-benzoyl-4-chlorophenyl)-1-(3,4-dibromobenzyl)-2-methylpyrrolidine-2-ca
rboxamide/(R)-3-amino-2-benzylpropanoic acid Schiff Base Complex ((*S*)(2*R*)-12a)**

1
2
3 Brown solid (57.2 mg, yield 89%). mp 132–134 °C. $[\alpha]^{20}_D = +2723.3$ ($c = 0.048$, CHCl_3). ^1H NMR
4 (500 MHz, Methanol- d_4) δ 9.34 (d, $J = 2.1$ Hz, 1H), 8.08 (dd, $J = 8.2, 2.1$ Hz, 1H), 7.77 (d, $J = 9.2$
5 Hz, 1H), 7.64 (d, $J = 8.2$ Hz, 1H), 7.38 (t, $J = 7.5$ Hz, 1H), 7.33 (t, $J = 7.4$ Hz, 1H), 7.23 – 7.15 (m,
6 4H), 7.09 (dd, $J = 9.2, 2.6$ Hz, 1H), 7.03 (d, $J = 7.9$ Hz, 1H), 7.00 (dd, $J = 6.5, 2.9$ Hz, 2H), 6.67
7 (d, $J = 7.6$ Hz, 1H), 6.45 (d, $J = 2.5$ Hz, 1H), 3.93 (td, $J = 12.6, 5.0$ Hz, 1H), 3.86 (t, $J = 12.4$ Hz,
8 1H), 3.78 (d, $J = 13.3$ Hz, 1H), 3.60 – 3.48 (m, 1H), 3.40 (d, $J = 13.3$ Hz, 1H), 3.29 – 3.23 (m, 1H),
9 3.14 (dd, $J = 13.8, 3.7$ Hz, 1H), 3.05 (dd, $J = 12.8, 3.8$ Hz, 1H), 2.66 (dd, $J = 13.8, 9.8$ Hz, 1H),
10 2.49 – 2.36 (m, 2H), 2.33 – 2.24 (m, 1H), 2.22 – 2.13 (m, 1H), 1.45 (s, 3H). ^{13}C NMR (125 MHz,
11 Methanol- d_4) δ 181.1, 179.1, 171.8, 139.8, 138.5, 137.6, 136.7, 134.3, 133.6, 131.9, 131.5, 131.2,
12 129.8, 129.5, 128.9, 128.8, 128.6, 128.2, 126.7, 126.3, 126.2, 125.5, 125.0, 124.5, 124.3, 78.1,
13 73.5, 55.3, 54.8, 53.9, 41.4, 35.8, 20.9, 17.3. LRMS (ESI+APCI) m/z : 809.7. HRMS (ESI-TOF)
14 m/z: [M + H]⁺ Calcd for $\text{C}_{36}\text{H}_{33}\text{Br}_2\text{ClN}_3\text{NiO}_3$ 805.9925; Found 805.9912.
15
16
17
18
19
20
21
22
23
24
25
26
27 Nickel(II)-(S)-N-(2-benzoyl-4-chlorophenyl)-1-(3,4-dimethoxybenzyl)-2-methylpyrrolidine-2-
28 carboxamide/(R)-3-amino-2-benzylpropanoic acid Schiff Base Complex ((S)(2R)-13a)
29
30 Brown solid (53.8 mg, yield 83%). mp 130–132 °C. $[\alpha]^{20}_D = +2690.6$ ($c = 0.050$, CHCl_3). ^1H
31 NMR (500 MHz, Methanol- d_4) δ 8.37 (d, $J = 2.1$ Hz, 1H), 7.88 (dd, $J = 8.2, 2.1$ Hz, 1H), 7.70 (d,
32 $J = 9.2$ Hz, 1H), 7.39 (tt, $J = 7.5, 1.3$ Hz, 1H), 7.33 (t, $J = 7.3$ Hz, 1H), 7.24 – 7.17 (m, 4H), 7.04 –
33 6.97 (m, 4H), 6.90 (d, $J = 8.2$ Hz, 1H), 6.68 (d, $J = 7.6$ Hz, 1H), 6.41 (d, $J = 2.5$ Hz, 1H), 3.96 (t, J
34 = 12.5 Hz, 1H), 3.93 – 3.90 (m, 1H), 3.89 (s, 3H), 3.76 (d, $J = 13.3$ Hz, 1H), 3.71 (s, 3H), 3.65 –
35 3.56 (m, 1H), 3.23 (ddd, $J = 11.2, 8.2, 1.5$ Hz, 1H), 3.11 (dd, $J = 13.8, 3.8$ Hz, 1H), 3.08 (dd, $J =$
36 12.9, 3.9 Hz, 1H), 2.70 (dd, $J = 13.8, 9.7$ Hz, 1H), 2.50 – 2.36 (m, 3H), 2.34 – 2.25 (m, 1H), 2.18
37 (td, $J = 10.9, 8.3$ Hz, 1H), 1.48 (s, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 181.8, 179.2, 171.5,
38 149.3, 149.0, 140.4, 138.4, 133.6, 131.2, 130.9, 129.8, 129.8, 128.8, 128.8, 128.6, 128.2,
39 126.8, 126.2, 125.1, 125.0, 123.7, 114.6, 111.7, 78.1, 73.3, 55.7, 55.4, 55.0, 54.7, 54.1, 41.8, 35.9,
40 21.0, 17.2. LRMS (ESI+APCI) m/z : 710.7. HRMS (ESI-TOF) m/z: [M + H]⁺ Calcd for
41 $\text{C}_{38}\text{H}_{39}\text{ClN}_3\text{NiO}_5$ 710.1926; Found 710.1926.
42
43
44
45
46
47
48
49
50
51
52
53
54
55 Nickel(II)-(S)-N-(2-benzoyl-4-chlorophenyl)-2-methyl-1-(3-phenylpropyl)pyrrolidine-2-carbo
56 xamide/(R)-3-amino-2-benzylpropanoic acid Schiff Base Complex ((S)(2R)-14a)
57
58
59
60

1
2
3 Brown solid (46.8 mg, yield 69%). mp 103–105 °C. $[\alpha]^{20}_D = +2756.3$ ($c = 0.048$, CHCl_3). ^1H NMR
4 (400 MHz, Methanol- d_4) δ 7.99 (d, $J = 9.1$ Hz, 1H), 7.38 – 7.29 (m, 2H), 7.25 – 7.10 (m, 10H),
5 7.02 – 6.93 (m, 3H), 6.73 – 6.68 (m, 1H), 6.52 (d, $J = 2.6$ Hz, 1H), 3.95 – 3.82 (m, 2H), 3.69 –
6 3.56 (m, 1H), 3.20 – 3.04 (m, 5H), 2.85 – 2.78 (m, 1H), 2.67 – 2.43 (m, 4H), 2.36 – 2.19 (m, 3H),
7 1.88 – 1.81 (m, 1H), 1.14 (s, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.3, 180.6, 173.1, 142.1,
8 141.8, 140.0, 135.0, 133.5, 133.1, 131.7, 131.1, 130.2, 130.1, 129.9, 129.6, 129.4, 129.4, 128.1,
9 127.8, 127.6, 127.1, 126.9, 126.5, 75.5, 57.0, 54.7, 54.3, 49.0, 41.7, 37.2, 34.7, 31.9, 23.4, 18.1.
10 LRMS (ESI+APCI) m/z : 678.2. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for $\text{C}_{38}\text{H}_{39}\text{ClN}_3\text{NiO}_3$
11 678.2028; Found 678.2035.

12
13
14
15
16 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
17 3-amino-2-benzylpropanoic acid Schiff Base Complex ((R)(2S)-15a)

18 Brown solid (48.2 mg, yield 82%). mp 120–122 °C. $[\alpha]^{20}_D = -2726.5$ ($c = 0.034$, CHCl_3). ^1H NMR
19 (400 MHz, Methanol- d_4) δ 8.14 (d, $J = 9.2$ Hz, 1H), 7.38 – 7.34 (m, 2H), 7.26 (dd, $J = 9.1, 2.6$ Hz,
20 1H), 7.17 – 7.11 (m, 4H), 7.04 – 6.92 (m, 3H), 6.81 – 6.70 (m, 1H), 6.53 (d, $J = 2.6$ Hz, 1H), 3.95
21 (t, $J = 12.4$ Hz, 1H), 3.86 – 3.61 (m, 2H), 3.23 – 3.07 (m, 3H), 2.66 – 2.41 (m, 3H), 2.39 – 2.24 (m,
22 3H), 1.93 – 1.86 (m, 4H), 1.24 (s, 3H). ^{13}C NMR (100 MHz, Methanol- d_4) δ 183.7, 180.6, 173.0,
23 141.9, 140.0, 135.0, 133.5, 133.0, 131.7, 131.1, 130.2, 130.1, 129.9, 129.6, 128.1, 127.9, 127.6,
24 126.9, 126.5, 74.9, 57.0, 54.6, 49.7, 48.9, 41.9, 37.2, 23.4, 18.1, 15.5. LRMS (ESI+APCI) m/z :
25 588.2. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for $\text{C}_{31}\text{H}_{33}\text{ClN}_3\text{NiO}_3$ 588.1558; Found 588.1564.

26
27
28
29 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
30 3-amino-2-(4-methoxybenzyl)propanoic acid Schiff Base Complex ((R)(2S)-15b)

31 Brown solid (51.3 mg, yield 83%). mp 127–129 °C. $[\alpha]^{20}_D = -3618.8$ ($c = 0.032$, CHCl_3). ^1H NMR
32 (400 MHz, Methanol- d_4) δ 8.14 (d, $J = 9.1$ Hz, 1H), 7.41 – 7.32 (m, 2H), 7.26 (d, $J = 9.1$ Hz, 1H),
33 7.15 (t, $J = 8.5$ Hz, 1H), 6.99 (d, $J = 7.6$ Hz, 1H), 6.91 – 6.84 (m, 2H), 6.77 (d, $J = 7.6$ Hz, 1H),
34 6.72 – 6.66 (m, 2H), 6.53 (d, $J = 2.6$ Hz, 1H), 3.94 (t, $J = 12.4$ Hz, 1H), 3.79 (s, 3H), 3.78 – 3.71
35 (m, 1H), 3.20 – 3.02 (m, 3H), 2.64 – 2.49 (m, 2H), 2.45 – 2.26 (m, 4H), 1.91 (t, $J = 7.4$ Hz, 3H),
36 1.24 (s, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.7, 180.7, 173.0, 159.8, 142.0, 135.1, 133.5,
37 133.0, 131.9, 131.7, 131.0, 131.0, 130.2, 130.2, 128.1, 127.9, 126.9, 126.6, 115.0, 75.0, 57.0, 55.7,

1
2
3 54.58, 49.7, 49.3, 42.0, 36.3, 23.4, 18.1, 15.5. LRMS (ESI+APCI) m/z : 618.1. HRMS (ESI-TOF)
4 m/z: [M + H]⁺ Calcd for C₃₂H₃₅ClN₃NiO₄ 618.1664; Found 618.1667.
5
6
7
8
9

10 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
11 **3-amino-2-(4-methylbenzyl)propanoic acid Schiff Base Complex ((R)(2S)-15c)**

12 Brown solid (42.1 mg, yield 70%). mp 124-126 °C. $[\alpha]^{20}_D = -3277.3$ ($c = 0.034$, CHCl₃). ¹H NMR
13 (400 MHz, Methanol-d₄) δ 8.13 (d, $J = 9.1$ Hz, 1H), 7.40 – 7.32 (m, 2H), 7.28 – 7.21 (m, 1H),
14 7.15 – 7.09 (m, 1H), 6.99 – 6.93 (m, 3H), 6.85 (d, $J = 8.0$ Hz, 2H), 6.77 (d, $J = 7.2$ Hz, 1H), 6.53
15 (d, $J = 2.6$ Hz, 1H), 3.93 (t, $J = 12.4$ Hz, 1H), 3.84 – 3.68 (m, 2H), 3.20 – 3.06 (m, 3H), 2.62 –
16 2.53 (m, 2H), 2.44 – 2.39 (m, 1H), 2.39 – 2.34 (m, 1H), 2.32 (s, 3H), 2.31 – 2.22 (m, 2H), 1.92 –
17 1.87 (m, 4H), 1.24 (s, 3H). ¹³C NMR (125 MHz, Methanol-d₄) δ 183.7, 180.6, 173.0, 142.0, 137.1,
18 136.9, 135.1, 133.5, 133.0, 131.7, 130.8, 130.3, 130.1, 129.9, 128.1, 127.9, 126.9, 126.6, 75.0,
19 57.0, 54.7, 49.69, 42.0, 36.8, 23.4, 21.1, 18.1, 15.5. LRMS (ESI+APCI) m/z : 601.8. HRMS
20 (ESI-TOF) m/z: [M + H]⁺ Calcd for C₃₂H₃₅ClN₃NiO₃ 602.1715; Found 602.1705.
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

30 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
31 **3-amino-2-(4-fluorobenzyl)propanoic acid Schiff Base Complex ((R)(2S)-15d)**

32 Brown solid (46.1 mg, yield 76%). mp 123-124 °C. $[\alpha]^{20}_D = -3310.5$ ($c = 0.038$, CHCl₃). ¹H NMR
33 (400 MHz, Methanol-d₄) δ 8.15 (d, $J = 9.1$ Hz, 1H), 7.41 – 7.33 (m, 2H), 7.26 (dd, $J = 9.1, 2.6$ Hz,
34 1H), 7.21 – 7.13 (m, 1H), 7.03 – 6.94 (m, 3H), 6.90 – 6.82 (m, 2H), 6.78 (dt, $J = 6.6, 1.9$ Hz, 1H),
35 6.53 (d, $J = 2.6$ Hz, 1H), 3.96 (t, $J = 12.3$ Hz, 1H), 3.86 – 3.73 (m, 2H), 3.22 – 3.06 (m, 3H), 2.62
36 – 2.29 (m, 6H), 1.94 – 1.86 (m, 4H), 1.24 (s, 3H). ¹³C NMR (125 MHz, Methanol-d₄) δ 183.7,
37 180.3, 173.0, 163.9, 162.0, 141.9, 136.1, 136.1, 135.1, 133.4, 133.0, 131.7, 131.6, 131.6, 130.9,
38 130.2, 130.2, 128.1, 127.8, 126.9, 126.6, 116.3, 116.1, 75.0, 56.9, 54.5, 49.7, 49.1, 41.9, 36.3, 23.4,
39 18.1, 15.5. LRMS (ESI+APCI) m/z : 606.1. HRMS (ESI-TOF) m/z: [M + H]⁺ Calcd for
40 C₃₁H₃₂ClFN₃NiO₃ 606.1464; Found 606.1476.
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

53 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
54 **3-amino-2-(4-(trifluoromethyl)benzyl)propanoic acid Schiff Base Complex ((R)(2S)-15e)**

1
2
3 Brown solid (47.2 mg, yield 72%). mp 137–139 °C. $[\alpha]^{20}_D = -3950.5$ ($c = 0.046$, CHCl_3). ^1H NMR
4 (400 MHz, Methanol- d_4) δ 8.14 (d, $J = 9.1$ Hz, 1H), 7.45 (d, $J = 8.0$ Hz, 2H), 7.35 – 7.25 (m, 3H),
5 7.17 (d, $J = 8.1$ Hz, 2H), 7.05 – 6.95 (m, 2H), 6.78 (d, $J = 7.6$ Hz, 1H), 6.52 (d, $J = 2.5$ Hz, 1H),
6 4.00 (t, $J = 12.3$ Hz, 1H), 3.88 – 3.78 (m, 2H), 3.30 – 3.26 (m, 1H), 3.21 (dd, $J = 11.1, 7.4$ Hz, 1H),
7 3.07 (dd, $J = 12.7, 3.4$ Hz, 1H), 2.61 – 2.44 (m, 3H), 2.39 – 2.30 (m, 3H), 1.94 – 1.87 (m, 4H),
8 1.25 (s, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.7, 179.9, 173.1, 145.1, 142.0, 135.1, 133.4,
9 133.1, 131.4, 131.0, 130.6, 130.1, 130.0, 129.8, 129.5, 128.0, 127.7, 126.9, 126.6, 126.5, 126.5,
10 75.0, 56.8, 54.5, 49.7, 42.0, 36.9, 23.4, 18.1, 15.5. LRMS (ESI+APCI) m/z : 655.8. HRMS
11 (ESI-TOF) m/z : [M + H]⁺ Calcd for $\text{C}_{32}\text{H}_{32}\text{ClF}_3\text{N}_3\text{NiO}_3$ 656.1432; Found 656.1431.
12
13
14
15
16
17
18
19
20
21
22 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
23 3-amino-2-methylpropanoic acid Schiff Base Complex ((R)(2S)-15f)
24
25 Brown solid (42.5 mg, yield 83%). mp 257–259 °C. $[\alpha]^{20}_D = -4854.5$ ($c = 0.044$, CHCl_3). ^1H NMR
26 (400 MHz, Methanol- d_4) δ 8.17 (d, $J = 9.1$ Hz, 1H), 7.66 – 7.49 (m, 3H), 7.45 – 7.35 (m, 1H),
27 7.29 (dd, $J = 9.1, 2.5$ Hz, 1H), 6.99 (d, $J = 7.6$ Hz, 1H), 6.65 (d, $J = 2.6$ Hz, 1H), 4.26 – 3.53 (m,
28 3H), 3.29 – 3.18 (m, 1H), 3.12 – 2.90 (m, 1H), 2.64 – 2.18 (m, 5H), 2.06 – 1.76 (m, 4H), 1.26 (s,
29 3H), 0.98 (d, $J = 7.3$ Hz, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.7, 182.0, 172.7, 142.0,
30 135.8, 133.6, 133.1, 131.5, 130.6, 130.1, 128.7, 128.3, 126.9, 126.6, 75.1, 59.8, 54.4, 49.8, 42.1,
31 41.9, 23.4, 18.1, 15.5, 15.2. LRMS (ESI+APCI) m/z : 512.0. HRMS (ESI-TOF) m/z : [M + H]⁺
32 Calcd for $\text{C}_{25}\text{H}_{29}\text{ClN}_3\text{NiO}_3$ 512.1245; Found 512.1259.
33
34
35
36
37
38
39
40 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
41 2-(aminomethyl)-4-methylpentanoic acid Schiff Base Complex ((R)(2S)-15g)
42
43 Brown solid (44.3 mg, yield 80%). mp 120–122 °C. $[\alpha]^{20}_D = -4383.3$ ($c = 0.048$, CHCl_3). ^1H NMR
44 (400 MHz, Methanol- d_4) δ 8.19 (d, $J = 9.1$ Hz, 1H), 7.60 – 7.52 (m, 3H), 7.44 – 7.42 (m, 1H),
45 7.29 (dd, $J = 9.1, 2.6$ Hz, 1H), 6.99 (dd, $J = 7.7, 1.7$ Hz, 1H), 6.67 (d, $J = 2.6$ Hz, 1H), 3.95 – 3.73
46 (m, 3H), 3.25 – 3.14 (m, 2H), 2.62 – 2.53 (m, 1H), 2.41 – 2.32 (m, 3H), 2.24 – 2.17 (m, 1H), 1.94
47 – 1.86 (m, 4H), 1.74 – 1.67 (m, 1H), 1.26 (s, 3H), 1.25 – 1.05 (m, 2H), 0.78 (dd, $J = 6.5, 3.8$ Hz,
48 6H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.6, 181.9, 172.9, 142.0, 135.9, 133.5, 133.1, 131.4,
49 131.3, 130.7, 130.2, 128.5, 128.4, 126.9, 126.6, 75.1, 58.0, 54.3, 49.8, 45.2, 41.9, 40.8, 26.8, 23.6,
50
51
52
53
54
55
56
57
58
59
60

1
2
3 23.4, 22.2, 18.1, 15.5. LRMS (ESI+APCI) m/z : 554.2. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for
4 C₂₈H₃₅ClN₃NiO₃ 554.1715; Found 554.1720.
5
6
7
8
9

10 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
11 2-(aminomethyl)-3-methylbutanoic acid Schiff Base Complex ((R)(2S)-15h)
12
13

14 Brown solid (43.7 mg, yield 81%). mp 130-132 °C. $[\alpha]^{20}_D = -3241.0$ ($c = 0.034$, CHCl₃). ¹H NMR
15 (400 MHz, Methanol-*d*₄) δ 8.18 (d, $J = 9.1$ Hz, 1H), 7.61 – 7.53 (m, 3H), 7.46 – 7.39 (m, 1H),
16 7.29 (dd, $J = 9.1$, 2.6 Hz, 1H), 7.00 (d, $J = 7.5$ Hz, 1H), 6.66 (d, $J = 2.6$ Hz, 1H), 4.03 – 3.87 (m,
17 2H), 3.85 – 3.74 (m, 1H), 3.24 – 3.16 (m, 2H), 2.60 – 2.46 (m, 2H), 2.41 – 2.32 (m, 3H), 2.14 –
18 2.08 (m, 1H), 1.95 – 1.87 (m, 4H), 1.26 (s, 3H), 0.72 (d, $J = 6.9$ Hz, 3H), 0.65 (d, $J = 7.1$ Hz, 3H).
19 ¹³C NMR (125 MHz, Methanol-*d*₄) δ 183.6, 181.0, 173.1, 142.1, 135.7, 133.4, 133.1, 131.5, 130.6,
20 130.2, 128.6, 128.5, 126.9, 126.6, 75.1, 54.3, 54.0, 53.1, 49.8, 42.0, 29.7, 23.4, 20.9, 18.1, 17.9,
21 15.5. LRMS (ESI+APCI) m/z : 539.9. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for
22 C₂₇H₃₃ClN₃NiO₃ 540.1558; Found 540.1566.
23
24
25
26

27 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
28 3-amino-2-cyclohexylpropanoic acid Schiff Base Complex ((R)(2S)-15i)
29
30

31 Brown solid (48.7 mg, yield 84%). mp 145-147 °C. $[\alpha]^{20}_D = -3161.8$ ($c = 0.034$, CHCl₃). ¹H NMR
32 (400 MHz, Methanol-*d*₄) δ 8.18 (d, $J = 9.1$ Hz, 1H), 7.66 – 7.51 (m, 3H), 7.42 – 7.37 (m, 1H),
33 7.28 (dd, $J = 9.1$, 2.6 Hz, 1H), 6.98 (d, $J = 7.6$ Hz, 1H), 6.65 (d, $J = 2.6$ Hz, 1H), 4.02 – 3.69 (m,
34 2H), 3.26 – 3.03 (m, 2H), 2.65 – 2.50 (m, 1H), 2.41 – 2.32 (m, 2H), 2.27 – 2.19 (m, 1H), 2.14 –
35 2.05 (m, 1H), 1.96 – 1.54 (m, 8H), 1.46 – 0.54 (m, 11H). ¹³C NMR (125 MHz, Methanol-*d*₄) δ
36 183.6, 180.9, 173.0, 142.0, 135.7, 133.5, 133.0, 131.5, 131.4, 130.6, 130.2, 128.6, 128.4, 126.9,
37 126.6, 75.1, 54.8, 54.3, 52.9, 49.9, 41.9, 40.2, 32.5, 29.3, 27.8, 27.6, 27.2, 23.3, 18.0, 15.5. LRMS
38 (ESI+APCI) m/z : 580.2. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for C₃₀H₃₇ClN₃NiO₃ 580.1871;
39 Found 580.1876.
40
41
42
43
44
45
46
47
48

49 Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-
50 2-(aminomethyl)-3,3-dimethylbutanoic acid Schiff Base Complex ((R)(2S)-15j)
51
52

53 Brown solid (38.2 mg, yield 69%). mp = 136-137 °C. $[\alpha]^{20}_D = -3670.0$ ($c = 0.048$, CHCl₃). ¹H
54 NMR (400 MHz, Methanol-*d*₄) δ 8.21 (d, $J = 9.1$ Hz, 1H), 7.63 – 7.58 (m, 2H), 7.56 – 7.52 (m,
55 1H), 7.42 – 7.37 (m, 1H), 7.29 (dd, $J = 9.1$, 2.6 Hz, 1H), 6.99 (d, $J = 7.6$ Hz, 1H), 6.67 (d, $J = 2.6$
56
57
58
59
60

1
2
3 Hz, 1H), 4.27 (t, J = 12.5 Hz, 1H), 4.07 – 3.97 (m, 1H), 3.93 – 3.84 (m, 1H), 3.33 (d, J = 4.7 Hz,
4 1H), 3.13 (dd, J = 10.9, 7.8 Hz, 1H), 2.66 (dq, J = 15.0, 7.5 Hz, 1H), 2.44 – 2.34 (m, 2H), 2.28 (dq,
5 J = 14.1, 7.1 Hz, 1H), 2.07 (dd, J = 12.5, 4.7 Hz, 1H), 2.03 – 1.95 (m, 1H), 1.89 (t, J = 7.3 Hz, 3H),
6 1.24 (s, 3H), 0.84 (s, 9H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.8, 181.5, 173.1, 142.3, 135.6,
7 133.5, 133.2, 131.7, 131.7, 130.6, 130.1, 128.8, 128.6, 126.9, 126.5, 74.9, 57.8, 56.4, 55.0, 49.5,
8 42.2, 33.9, 28.5, 23.7, 18.1, 15.5. LRMS (ESI+APCI) m/z : 655.8. HRMS (ESI-TOF) m/z : [M +
9 H]⁺ Calcd for C₂₈H₃₅ClN₃NiO₃ 554.1715; Found 554.1728.

10
11
12
13
14
15
16 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
17
18 **3-amino-2-phenylpropanoic acid Schiff Base Complex ((R)(2S)-15k)**

19 Brown solid (41.9 mg, yield 73%). mp 150–151 °C. $[\alpha]^{20}_D$ = -3648.0 (c = 0.05, CHCl₃). ^1H NMR
20 (400 MHz, Methanol- d_4) δ 8.20 (d, J = 9.1 Hz, 1H), 7.66 – 7.52 (m, 3H), 7.46 (t, J = 7.6 Hz, 1H),
21 7.31 (dd, J = 9.1, 2.6 Hz, 1H), 7.28 – 7.16 (m, 3H), 7.10 – 7.03 (m, 2H), 6.91 (d, J = 7.7 Hz, 1H),
22 6.69 (d, J = 2.6 Hz, 1H), 4.34 (t, J = 13.0 Hz, 1H), 3.89 – 3.86 (m, 2H), 3.53 (dd, J = 11.8, 3.5 Hz,
23 1H), 3.29 – 3.97 (m, 1H), 3.15 (dd, J = 13.0, 3.6 Hz, 1H), 2.65 – 2.56 (m, 1H), 2.46 – 2.29 (m,
24 3H), 1.99 – 1.90 (m, 4H), 1.27 (s, 3H). ^{13}C NMR (125 MHz, Methanol- d_4) δ 183.7, 180.1, 173.3,
25 142.2, 139.1, 135.8, 133.7, 133.2, 131.6, 131.4, 130.6, 130.1, 129.8, 129.1, 128.5, 128.4, 128.4,
26 126.9, 126.7, 75.2, 60.1, 54.5, 54.3, 49.9, 41.9, 23.5, 18.1, 15.5. LRMS (ESI+APCI) m/z : 574.1.
27 HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for C₃₀H₃₁ClN₃NiO₃ 574.1402; Found 574.1405.

28
29
30
31
32
33
34
35
36
37
38 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
39
40 **3-amino-2-(4-methoxyphenyl)propanoic acid Schiff Base Complex ((R)(2S)-15l)**

41 Brown solid (39.3 mg, yield 65%). mp = 146–148 °C. $[\alpha]^{20}_D$ = -3052.4 (c = 0.042, CHCl₃). ^1H
42 NMR (400 MHz, Methanol- d_4) δ 8.19 (d, J = 9.1 Hz, 1H), 7.68 – 7.52 (m, 3H), 7.48 (t, J = 7.5 Hz,
43 1H), 7.31 (dd, J = 9.1, 2.6 Hz, 1H), 7.01 – 6.95 (m, 2H), 6.93 (d, J = 7.7 Hz, 1H), 6.84 – 6.77 (m,
44 2H), 6.69 (d, J = 2.6 Hz, 1H), 4.29 (t, J = 12.9 Hz, 1H), 3.91 – 3.78 (m, 2H), 3.73 (s, 3H), 3.48 (dd,
45 J = 11.7, 3.6 Hz, 1H), 3.29 – 3.23 (m, 1H), 3.12 (dd, J = 13.0, 3.6 Hz, 1H), 2.64 – 2.56 (m, 1H),
46 2.44 – 2.34 (m, 3H), 1.95 (q, J = 10.3, 8.7 Hz, 4H), 1.27 (s, 3H). ^{13}C NMR (125 MHz,
47 Methanol- d_4) δ 183.7, 180.6, 173.3, 160.4, 142.2, 135.8, 133.7, 133.2, 131.6, 131.4, 131.0, 130.6,
48 130.1, 128.5, 128.5, 126.9, 126.6, 115.2, 75.2, 60.2, 55.7, 54.4, 53.5, 49.9, 41.9, 23.5, 18.1, 15.5.
49 LRMS (ESI+APCI) m/z : 604.1. HRMS (ESI-TOF) m/z : [M + H]⁺ Calcd for C₃₁H₃₃ClN₃NiO₄
50 604.1508; Found 604.1518.

1
2
3 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
4 **3-amino-2-(4-chlorophenyl)propanoic acid Schiff Base Complex ((R)(2S)-15m)**

5
6 Brown solid (42.0 mg, yield 69%). mp = 130-132 °C. $[\alpha]^{20}_D = -3312.5$ (c = 0.048, CHCl₃). ¹H
7 NMR (400 MHz, Methanol-d₄) δ 8.20 (d, J = 9.1 Hz, 1H), 7.65 – 7.44 (m, 4H), 7.31 (dd, J = 9.1,
8 2.6 Hz, 1H), 7.28 – 7.23 (m, 2H), 7.08 – 7.01 (m, 2H), 6.93 (d, J = 7.7 Hz, 1H), 6.69 (d, J = 2.6
9 Hz, 1H), 4.35 (t, J = 12.5 Hz, 1H), 3.97 – 3.81 (m, 2H), 3.54 (dd, J = 12.0, 3.4 Hz, 1H), 3.29 –
10 3.27 (m, 1H), 3.12 (dd, J = 12.9, 3.5 Hz, 1H), 2.63 – 2.50 (m, 1H), 2.46 – 2.31 (m, 3H), 1.98 –
11 1.90 (m, 4H), 1.27 (s, 3H). ¹³C NMR (125 MHz, Methanol-d₄) δ 183.7, 179.6, 173.4, 142.2, 137.8,
12 135.7, 134.2, 133.7, 133.3, 131.6, 131.4, 130.8, 130.7, 130.1, 129.8, 128.6, 128.4, 127.0, 126.7,
13 75.2, 59.8, 54.4, 49.5, 41.9, 23.5, 18.1, 15.5. LRMS (ESI+APCI) m/z: 608.0. HRMS (ESI-TOF)
14 m/z: [M + H]⁺ Calcd for C₃₀H₃₀Cl₂N₃NiO₃ 608.1012; Found 608.1027.

15
16 **Nickel(II)-(R)-N-(2-benzoyl-4-chlorophenyl)-1-ethyl-2-methylpyrrolidine-2-carboxamide/(S)-**
17 **3-amino-2-(naphthalen-1-yl)propanoic acid Schiff Base Complex ((R)(2S)-15n)**

18 Brown solid (47.4 mg, yield 76%). mp = 146-147 °C. $[\alpha]^{20}_D = -3440.0$ (c = 0.046, CHCl₃). ¹H
19 NMR (400 MHz, Methanol-d₄) δ 8.21 (d, J = 9.1 Hz, 1H), 7.80 – 7.72 (m, 3H), 7.65 (d, J = 4.6 Hz,
20 2H), 7.58 – 7.37 (m, 5H), 7.32 (dd, J = 9.1, 2.6 Hz, 1H), 7.18 (dd, J = 8.5, 1.8 Hz, 1H), 6.89 (dd, J
21 = 7.8, 1.4 Hz, 1H), 6.70 (d, J = 2.6 Hz, 1H), 4.45 (t, J = 12.9, 11.8 Hz, 1H), 3.92 – 3.79 (m, 2H),
22 3.71 (dd, J = 11.7, 3.5 Hz, 1H), 3.38 – 3.32 (m, 1H), 3.22 (dd, J = 13.0, 3.5 Hz, 1H), 2.69 – 2.53
23 (m, 1H), 2.48 – 2.27 (m, 3H), 2.02 – 1.87 (m, 4H), 1.26 (s, 3H). ¹³C NMR (125 MHz, Methanol-d₄)
24 δ 183.7, 180.1, 173.4, 142.2, 136.4, 135.8, 134.9, 134.0, 133.7, 133.3, 131.6, 131.4, 130.6, 130.1,
25 129.5, 128.7, 128.6, 128.5, 128.5, 128.2, 127.3, 127.0, 127.0, 126.9, 126.7, 75.2, 60.0, 54.4, 54.4,
26 49.9, 41.9, 23.5, 18.1, 15.5. LRMS(ESI+APCI) m/z: 624.1. HRMS (ESI-TOF) m/z: [M + H]⁺
27 Calcd for C₃₄H₃₃ClN₃NiO₃ 624.1558; Found 624.1576.

28 **(S)-3-Amino-2-benzylpropanoic acid ((S)-2a)**

29 A solution of (R)(2S)-15a (1.50 g, 2.55 mmol) in MeOH (20 mL) was added to a stirring solution
30 of 3N HCl in MeOH (1/1, v/v) 40 mL at 50 °C. The mixture was gently heated under reflux for 30
31 min, the solution was evaporated to dryness. Water (50 mL) was added, and the resultant mixture
32 was treated with an excess of concentrated NH₄OH and extracted with CH₂Cl₂. The CH₂Cl₂
33 extracts were dried (Na₂SO₄) and evaporated under vacuum to afford the free chiral ligand (R)-9
34 (930 mg, yield 98%). The aqueous phase was evaporated under vacuum and dissolved in a
35 36 37 38 39 40 41 42 43 44

minimum amount of H₂O and loaded on a Dowex 50×2 100 ion-exchange column, which was washed with H₂O until neutral. The column was then washed with 10% aq NH₄OH. The first fraction (400 mL) was collected and evaporated under vacuum to afford the corresponding amino acid (*S*)-**2a** as a white solid (425 mg, yield 93%). mp = 208–210 °C. [α]_D²⁰ = -16.5 (c = 1, 1 M HCl). [ref.[38] [α]_D²⁵ = -16.0 (c = 1, 1 M HCl)]. ¹H NMR (400 MHz, D₂O) δ 7.39 – 7.16 (m, 5H), 3.08 – 2.69 (m, 5H). ¹³C NMR (100 MHz, D₂O) δ 181.5, 140.4, 130.8, 130.5, 128.6, 49.0, 42.5, 38.0. LRMS (ESI+APCI) *m/z*: 180.1. HRMS (ESI-TOF) *m/z*: [M + H]⁺ Calcd for C₁₀H₁₄NO₂ 180.1019; Found 180.1014.

Supporting Information

X-ray crystal structures of (*S*)(2*R*)-**10a** and (*S*)(2*S*)-**10a**.

¹H NMR spectra for dr value determination of **10a-15a** and **15b-15n**.

HPLC spectra for ee value determination of (*S*)-**2a** and (*R*)-**9**.

¹H and ¹³C NMR spectra for all compounds.

Acknowledgments

We gratefully acknowledge financial support from the National Natural Science Foundation of China (81620108027, 21632008, 21672231, and 21472209), the Major Project of Chinese National Programs for Fundamental Research and Development (2015CB910304).

References

1. Liu, M.; Sibi, M. P., Recent advances in the stereoselective synthesis of β-amino acids. *Tetrahedron* **2002**, 58 (40), 7991–8035.
2. Seebach, D.; Beck, A. K.; Capone, S.; Deniau, G.; Groselj, U.; Zass, E., Enantioselective Preparation of β²-Amino Acid Derivatives for β-Peptide Synthesis. *Synthesis* **2009**, (1), 1–32.
3. Zeng, W.C.; Xu, R.A.; Zeng, Q.Y., Research progress in the synthesis of β-amino acids. *Hecheng Huaxue* **2013**, 21 (5), 634–644.
4. Lelais, G.; Seebach, D., β²-amino acids—Syntheses, occurrence in natural products, and components of β-peptides. *Biopolymers* **2004**, 76 (3), 206–243.

- 1
2
3 5. Ma, J. A., Recent developments in the catalytic asymmetric synthesis of alpha- and beta-amino
4 acids. *Angew. Chem. Int. Ed.* **2003**, *42* (36), 4290-4299.
5
6 6. Lukaszuk, A.; Demaegdt, H.; Szemenyei, E.; Toth, G.; Tymecka, D.; Misicka, A.; Karoyan, P.;
7 Vanderheyden, P.; Vauquelin, G.; Tourwe, D., beta-homo-amino acid scan of angiotensin IV. *J.
8 Med. Chem.* **2008**, *51* (7), 2291-2296.
9
10 7. Shih, C.; Gossett, L. S.; Gruber, J. M.; Grossman, C. S.; Andis, S. L.; Schultz, R. M.; Worzalla,
11 J. F.; Corbett, T. H.; Metz, J. T., Synthesis and biological evaluation of novel cryptophycin
12 analogs with modification in the beta-alanine region. *Bioorg. Med. Chem. Lett.* **1999**, *9* (1), 69-74.
13
14 8. Seebach, D.; Matthews, J. L., beta-peptides: a surprise at every turn. *Chem. Commun.* **1997**,
15 (21), 2015-2022.
16
17 9. Pavlov, N.; Gilles, P.; Didierjean, C.; Wenger, E.; Naydenova, E.; Martinez, J.; Calmes, M.,
18 Asymmetric Synthesis of beta(2)-Tryptophan Analogues via Friedel-Crafts Alkylation of Indoles
19 with a Chiral Nitroacrylate. *J. Org. Chem.* **2011**, *76* (15), 6116-6124.
20
21 10. Kim, D.; Wang, L. P.; Beconi, M.; Eiermann, G. J.; Fisher, M. H.; He, H. B.; Hickey, G. J.;
22 Kowalchick, J. E.; Leiting, B.; Lyons, K.; Marsilio, F.; McCann, M. E.; Patel, R. A.; Petrov, A.;
23 Scapin, G.; Patel, S. B.; Roy, R. S.; Wu, J. K.; Wyvratt, M. J.; Zhang, B. B.; Zhu, L.; Thornberry,
24 N. A.; Weber, A.; E.,
25 (2R)-4-Oxo-4-[3-(trifluoromethyl)-5,6-dihydro[1,2,4]triazolo[4,3-alpha]pyrazin-7(8H)-yl]-1-(2,4,
26 5-trifluorophenyl)butan-2-amine: A potent, orally active dipeptidyl peptidase IV inhibitor for the
27 treatment of type 2 diabetes. *J. Med. Chem.* **2005**, *48* (1), 141-151.
28
29 11. Snyder, O.; Wahl, A.; Swanson, M.; Spagnuolo, R. A.; Garcia, J. V., The Role of Semen on
30 Vaginal HIV-1 Transmission and on the Efficacy of Maraviroc as a Topically Applied
31 Microbicide. *Aids Res. Hum. Retrov.* **2014**, *30*, A50-A50.
32
33 12. Juaristi, E.; Lopez-Ruiz, H., Recent advances in the enantioselective synthesis of beta-amino
34 acids. *Curr. Med. Chem.* **1999**, *6* (10), 983-1004.
35
36 13. Weiner, B.; Szymanski, W.; Janssen, D. B.; Minnaard, A. J.; Feringa, B. L., Recent advances
37 in the catalytic asymmetric synthesis of beta-amino acids. *Chem. Soc. Rev.* **2010**, *39* (5),
38 1656-1691.
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 14. Huang, H. M.; Liu, X. C.; Deng, J.; Qiu, M.; Zheng, Z., Rhodium-catalyzed enantioselective
4 hydrogenation of beta-phthalimide acrylates to synthesis of beta(2)-amino acids. *Org. Lett.* **2006**,
5 8 (15), 3359-3362.
6
7 15. Martin, N. J. A.; Cheng, X.; List, B., Organocatalytic asymmetric transferhydrogenation of
8 beta-nitroacrylates: Accessing beta(2)-amino acids. *J. Am. Chem. Soc.* **2008**, 130 (42),
9 13862-13863.
10
11 16. Beddow, J. E.; Davies, S. G.; Ling, K. B.; Roberts, P. M.; Russell, A. J.; Smith, A. D.;
12 Thomson, J. E., Asymmetric synthesis of beta(2)-amino acids: 2-substituted-3-aminopropanoic
13 acids from N-acryloyl SuperQuat derivatives. *Org. Biomol. Chem.* **2007**, 5 (17), 2812-2825.
14
15 17. Moumne, R.; Lavielle, S.; Karoyan, P., Efficient synthesis of beta(2)-amino acid by
16 homologation of alpha-amino acids involving the Reformatsky reaction and Mannich-type
17 imminium electrophile. *J. Org. Chem.* **2006**, 71 (8), 3332-3334.
18
19 18. Lin, D. Z.; Lv, L.; Wang, J.; Ding, X.; Jiang, H. L.; Liu, H., Preparation of
20 alpha-Alkyl-beta-Amino Acids via beta-Alanine Ni(II) Complex. *J. Org. Chem.* **2011**, 76 (16),
21 6649-6656.
22
23 19. Berkessel, A.; Jurkiewicz, I.; Mohan, R., Enzymatic Dynamic Kinetic Resolution of
24 Oxazinones: A New Approach to Enantiopure β 2-Amino Acids. *ChemCatChem* **2011**, 3 (2),
25 319-330.
26
27 20. Fitz, M.; Forró, E.; Vigóczki, E.; Lázár, L.; Fülöp, F., Lipase-catalysed N-acylation of
28 β^2 -amino esters. *Tetrahedron: Asymmetry* **2008**, 19 (9), 1114-1119.
29
30 21. Ma, D.Y.; Wang, D.X.; Zheng, Q.Y.; Wang, M.X., Nitrile biotransformations for the practical
31 synthesis of highly enantiopure azido carboxylic acids and amides, ‘click’ to functionalized chiral
32 triazoles and chiral β -amino acids. *Tetrahedron: Asymmetry* **2006**, 17 (16), 2366-2376.
33
34 22. Yokomatsu, T.; Takada, K.; Yasumoto, A.; Yuasa, Y.; Shibuya, S., Chemo-enzymatic
35 synthesis of novel β -amino acids substituted by (thymin-1-yl)methyl functional group at the
36 α -position. *Heterocycles* **2002**, 56 (1-2), 545-552.
37
38 23. Solymár, M.; Liljeblad, A.; Lázár, L.; Fülöp, F.; Kanerva, L. T., Lipase-catalysed kinetic
39 resolution in organic solvents: an approach to enantiopure α -methyl- β -alanine esters. *Tetrahedron:*
40 *Asymmetry* **2002**, 13 (17), 1923-1928.
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 24. Gianolio, E.; Mohan, R.; Berkessel, A., Enantiopure N-Benzylloxycarbonyl-beta(2)-amino
4 Acid Allyl Esters from Racemic beta-Lactams by Dynamic Kinetic Resolution using Candida
5 antarctica Lipase B. *Adv. Synth. Catal.* **2016**, *358* (1), 30-33.
6
7 25. Salamonczyk, G. M.; Han, K.; Guo, Z. W.; Sih, C. J., Total synthesis of cryptophycins via a
8 chemoenzymatic approach. *J. Org. Chem.* **1996**, *61* (20), 6893-6900.
9
10 26. Eroksuz, S.; Neudorfl, J. M.; Berkessel, A., Kinetic Resolution of 5-Substituted Oxazinones
11 with Bifunctional Chiral Base/Squaramide Organocatalysts. *Synlett* **2017**, *28* (11), 1278-1281.
12
13 27. Shu, C.; Hu, X. Y.; Li, S. S.; Yuan, W. C.; Zhang, X. M., Lewis Base Catalyzed Asymmetric
14 Hydrosilylation of alpha-Substituted beta-Enamino Esters: Facile Access to Enantioenriched
15 beta(2)-Amino Esters via Dynamic Kinetic Resolution. *Synlett* **2014**, *25* (13), 1879-1882.
16
17 28. Wang, Y. B.; Song X. H.; Wang J.; Moriwaki H.; Soloshonok V. A.; Liu H., Recent
18 approaches for asymmetric synthesis of α -amino acids via homologation of Ni(II) complexes.
19 *Amino Acids* **2017**, *49*, 1487-1520.
20
21 29. Ding, X.; Ye, D. J.; Liu, F.; Deng, G. H.; Liu, G. N.; Luo, X. M.; Jiang, H. L.; Liu, H.,
22 Efficient Synthesis of alpha-Aryl-/Heteroaryl-Substituted beta-Amino Acids via Ni(II) Complex
23 through the Suzuki Coupling Reaction. *J. Org. Chem.* **2009**, *74* (15), 5656-5659.
24
25 30. Lin, D. Z.; Deng, G. H.; Wang, J.; Ding, X.; Jiang, H. L.; Liu, H., Efficient Synthesis of
26 Symmetrical alpha,alpha-Disubstituted beta-Amino Acids and alpha,alpha-Disubstituted
27 Aldehydes via Dialkylation of Nucleophilic beta-Alanine Equivalent. *J. Org. Chem.* **2010**, *75* (5),
28 1717-1722.
29
30 31. Wang, S.; Zhou, S.; Wang, J.; Nian, Y.; Kawashima, A.; Moriwaki, H.; Acena, J. L.;
31 Soloshonok, V. A.; Liu, H., Chemical Dynamic Thermodynamic Resolution and S/R
32 Interconversion of Unprotected Unnatural Tailor-made alpha-Amino Acids. *J. Org. Chem.* **2015**,
33 *80* (20), 9817-9830.
34
35 32. Nian, Y.; Wang, J.; Zhou, S. B.; Wang, S. N.; Moriwaki, H.; Kawashima, A.; Soloshonok, V.
36 A.; Liu, H., Recyclable Ligands for the Non-Enzymatic Dynamic Kinetic Resolution of
37 Challenging -Amino Acids. *Angew. Chem. Int. Ed.* **2015**, *54* (44), 12918-12922.
38
39 33. Takeda, R.; Kawamura, A.; Kawashima, A.; Sato, T.; Moriwaki, H.; Izawa, K.; Akaji, K.;
40 Wang, S.; Liu, H.; Acena, J. L.; Soloshonok, V. A., Chemical Dynamic Kinetic Resolution and
41

- 1
2
3 *S/R* Interconversion of Unprotected alpha-Amino Acids. *Angew. Chem. Int. Ed.* **2014**, *53* (45),
4 12214-12217.
5
6
7 34. Zhou, S. B.; Wang, J.; Chen, X.; Acena, J. L.; Soloshonok, V. A.; Liu, H., Chemical Kinetic
8 Resolution of Unprotected beta-Substituted beta-Amino Acids Using Recyclable Chiral Ligands.
9 *Angew. Chem. Int. Ed.* **2014**, *53* (30), 7883-7886.
10
11
12 35. CCDC numbers of crystallographic structure of Ni(II) complexes **(S)(2R)-10a** and **(S)(2S)-10a**
13 are 1477075 and 1477074, respectively.
14
15
16 36. Zhou, S.; Wang, S.; Wang, J.; Nian, Y.; Peng, P.; Soloshonok, V. A.; Liu, H.,
17 Configurationally Stable (*S*)- and (*R*)- α -Methylproline-Derived Ligands for the Direct Chemical
18 Resolution of Free Unprotected β^3 -Amino Acids. *Eur. J. Org. Chem.* **2018**, *2018* (15), 1821-1832.
19
20
21 37. Belokon, Y. N.; Maleyev, V. I.; Vitt, S. V.; Ryzhov, M. G.; Kondrashov, Y. D.; Golubev, S.
22 N.; Vauchskii, Y. P.; Kazika, A. I.; Novikova, M. I.; Krasutskii, P. A.; Yurchenko, A. G.;
23 Dubchak, I. L.; Shklover, V. E.; Struchkov, Y. T.; Bakhmutov, V. I.; Belikov, V. M.,
24 Enantioselectivity of Nickel(II) and Copper(II) Complexes of Schiff-Bases Derived from
25 Amino-Acids and (*S*)-*Ortho*-[(*N*-Benzylprolyl)Amino]-Acetophenone or
26
27 (*S*)-*Ortho*-[(*N*-Benzylprolyl)Amino]Benzaldehyde- Crystal and Molecular-Structures of
28
29 [Ni((*S*)-Bap-(*S*)-Val)] and [Cu((*S*)-Bap-(*S*)-Val)]. *J. Chem. Soc. Dalton Trans.* **1985**, (1), 17-26.
30
31
32
33
34 38. Guzman-Mejia, R.; Reyes-Rangel, G.; Juaristi, E., Preparation of chiral derivatives of β -Ala
35 containing the alpha-phenylethyl group: useful starting materials for the asymmetric synthesis of
36 β -amino acids. *Nat. Protoc.* **2007**, *2* (11), 2759-2766.
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60