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Kakeromamide A, a new cyclic pentapeptide inducing astrocyte differentiation isolated from the marine cyanobacterium *Moorea bouillonii*

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Introduction

Marine cyanobacteria are known to be a rich source of peptides such as lyngbyabellin B¹ and dolastatin 10.^{2.3} In this study, a new cyclic peptide named kakeromamide A (**1**) was isolated from the marine cyanobacterium *Moorea bouillonii* collected at Kakeroma Island in Kagoshima prefecture of the southern part of Japan. This peptide shares the common amino acid sequence with that of a cyclic depsipeptide ulongamide D⁴ in which valine was replaced by hydroxy isovaleric acid. Ulongamide D showed cytotoxicity against KB and LoVo cells with the IC₅₀ values of 1 and 5 μ M, respectively, while compound **1** showed only a moderate cytotoxicity against HeLa cells with the IC₅₀ value of 10 μ M. However, we found a unique biological activity in compound **1** to induce differentiation of neural stem cells into astrocytes at $10 \,\mu\text{M}$ in the *in vitro* differentiation model using mouse ES cells.^{5,6} In this letter, we report the isolation, structure elucidation, and biological activities of kakeromamide A (**1**).

The frozen specimen of *Moorea bouillonii* (128 g wet weight), collected by hand using SCUBA at Kakeroma Island in Kagoshima prefecture (N 28° 04.67', E 129° 18.42'), was extracted with MeOH. The combined methanolic extract was evaporated *in vacuo* and partitioned between H₂O and CHCl₃. The organic layer was subjected to ODS flash chromatography and followed by the reversed-phase HPLC, yielding 1.2 mg of kakeromamide A (1) as the colorless amorphous solid (9.4×10^{-6} % yield based on the wet weight).

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ABSTRACT

Kakeromamide A (1), a new cyclic pentapeptide encompassing a thiazole ring moiety and a β -amino acid, was isolated from the marine cyanobacterium *Moorea bouillonii*. Its structure was elucidated by the spectral analysis and the modified Marfey's method. Compound **1** induced differentiation of neural stem cells into astrocytes at the concentration of 10 μ M.

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Table 1				
NMR spectral data for kakeromamide A (1) in	CD ₃ CN (+	400 MH	lz a



Fig. 1. Substructure of kakeromamide A (1).

Unit	Position	δ_{C}	$\delta_{\rm H}$ mult. (J in Hz)	COSY	HMBC
Val	1	176.8			
	2	57.2	4.30 dd (9.8, 7.3)	NH-2, H-3	
	3	32.0	1.80 dgg (9.8, 6.8, 6.7)	H-2, H-4, H-5	
	4	18.9	0.79 d (6.8)	Н-3	C-2, C-5
	5	20.2	0.89 d (6.7)	H-3	C-2, C-4
	2-NH		6.53 d (7.3)	H-2	C-32 ^a
N,O-diimethyl-Tyr-1	6	174.0			
	7	52.3	5.53 dd (11.3, 4.6)	H-8	C-6 ^a , C-8 ^a
	8	33.4	1.38 dd (16.4, 4.6)	H-7	C-7 ^a , C-9
			2.70 dd (16.4, 11.3)	H-7	,
	9	130.1			
	10/14	130.0	6.88 d (8.7)	H-11/13	C-8, C-12
	11/13	114.9	6.77 d (8.7)	H-10/14	C-9
	12	159.2			
	N-CH ₃	31.7	3.05 s		C-1, C-7
	O-CH ₃	55.9	3.73 s		C-12
N,O-dimethyl-Tyr-2	15	169.9			
	16	63.7	5.25 dd (9.7, 5.2)	H-17	C-15 ^a , C-17 ^a
	17	34.6	2.61 dd (14.4, 9.7)	H-16	C-16 ^a . C-18
			2.97 dd (14.4, 5.2)	H-16	· · , · · ·
	18	131.0			
	19/23	131.3	6.99 d (8.6)	H-20/22	C-17, C-21
	20/22	115.2	6.56 d (8.6)	H-19/23	C-18
	26	159.5			
	N-CH ₃	29.7	2.86 s		C-6, C-16
	O-CH ₃	55.7	3.48 s		C-21
Val-thz-ca	24	161.4			
	25	150.4			
	26	123.6	8.01 s		C25 ^a , C-27 ^a
	27	170.0			
	28	57.2	5.33 dd (9.3, 5.5)	NH-28, H-29	C-27 ^a , C-29
	29	36.8	2.01 dqq (5.5, 6.8, 6.8)	H-28, H-30, H-31	
	30	17.8	0.78 d (6.8)	H-29	C-28, C-31
	31	20.8	0.94 d (6.8)	H-29	C-28, C-30
	28-NH		8.60 d (9.3)	H-28	
Amha	32	173.3			
	33	44.6	2.60 dq (3.4, 7.0)	H-34, H-38	
	34	52.8	4.09 dddd(12.0, 10.4, 3.4, 2.5)	H-33, H-35, NH-34	
	35	31.9	1.07 m, 1.70 dddd (14.1, 9.2, 7.0, 2.5)	H-34, H-36	C-33
	36	20.2	1.27 m. 1.45 m	H-35, H-37	
	37	14.6	0.97 t (7.5)	H-36	C-35, C-36
	38	14.4	1.09 d (7.0)	H-33	C-32, C-33, C-34
	34-NH	• •	851 d(104)	H_34	,, - • •

^a observed only in the spectrum recorded on the spectrometer (600 MHz).

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Fig. 2. Key COSY and HMBC correlations for kakeromamide A (1).

The ¹H NMR spectrum of compound **1** (Table 1) measured in CD₃CN indicated its peptidal nature, i.e. three amide protons ($\delta_{\rm H}$ 6.53, 8.51, 8.60), two singlets for N-methyl protons ($\delta_{\rm H}$ 2.86, 3.05) and signals for four α -protons ($\delta_{\rm H}$ 4.30, 5.25, 5.33, 5.53), as well as an α -proton of the β -amino acid residue at $\delta_{\rm H}$ 2.60. ¹H-¹H COSY analysis revealed the presence of seven spin systems (Fig. 1). Analysis of HMQC⁷ and HMBC⁸ spectra identified three α -amino acid residues; valine (Val) and two N,O-dimethyl-tyrosines (N,O-diMeTyr-1, 2). The fourth amino acid, 2-(1-amino-2methylpropyl)thiazole-4-carboxylic acid (Val-thz-ca), biogenetically derived from valine and cysteine, was deduced as follows; a downfield shifted singlet proton signal at $\delta_{\rm H}$ 8.01 and the sp² carbon resonances at $\delta_{\rm C}$ 123.6, 150.8, 161.4, and 170.0 were characteristic of a 2-alkylthiazole-4-carboxylic acid unit (C-24 to C-27).⁹ A characteristic ${}^{1}J_{CH}$ value of 194 Hz between H-26 and C-26 supported the existence of a thiazole ring.¹⁰ HMBC correlations between H-26/C-27 and H-28/C-27 connected the thiazole unit to Val-like unit, to from a Val-thz-ca residue. The



Fig. 3. Astrocyte differentiation assay. (a): The schematic drawings of the astrocyte differentiation assay. (b): Fluorescent microscopy photograph of differentiated cells from neural stem cells (NSCs). Cells were treated with DMSO (left) or 10 μ M compound **1** (right, magnification \times 20, blue: fluorescent labeled nucleus by Hoechst 33342, green: fluorescent labeled Glial Fibrillary Acidic Protein by chomeo 488, scale bar: 50 μ m). (c): Rates for NSCs differentiating into astrocytes (calculated by counting of GFAP-positive cells) compared to that of control (n = 4, mean ± S.D., *p < 0.05). d: Percentages of cell numbers after sample treatment (n = 4, mean ± S.D.). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

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remaining β -amino acid was deduced as 3-amino-2-methylhexanoic acid (Amha) by an HMBC cross peak from H-38 of the spin system (CH₃-38 to CH₃-37) to C-32.

Connectivities among these amino acids were established on the basis of inter-residual HMBC cross peaks (Figs. S6 and S7). Correlation between 2-NH ($\delta_{\rm H}$ 6.53) and C-32 ($\delta_{\rm C}$ 173.3) clearly indicated the amide bond formation between Val (NH) and Amha (CO). An HMBC cross-peak between *N*-methyl protons at $\delta_{\rm H}$ 2.86 (*N*,O-diMeTyr-2) and $\delta_{\rm C}$ 174.0 (C-26) connected *N*,O-diMeTyr-2 to *N*,*O*-diMeTyr-1, while an HMBC correlation between the *N*-methyl signal at $\delta_{\rm H}$ 3.05 (*N*,O-diMeTyr-1) and C-1 ($\delta_{\rm C}$ 176.2) established the linkage between N,O-diMeTyr-1 and Val. Although no interresidual HMBC cross-peak was observed among Val-thz-ca and other residues, the lower field shifted δ values for H-28 ($\delta_{\rm H}$ 5.33) and H-34 ($\delta_{\rm H}$ 4.09) as well as typical values for amide protons of 28-NH ($\delta_{\rm H}$ 8.60) and 34-NH ($\delta_{\rm H}$ 8.51) strongly suggested that the nitrogen atoms at 28 and 34 form amide bonds. Therefore, position of Val-thz-ca was between Amha and N,O-diMeTyr-2, completing the planar structure of kakeromamide A (1) (Fig. 2). The detailed NMR spectral data are listed in Table 1.

Absolute configuration of 5 amino acid residues in kakeromamide A (1) could be established by the modified Marfey's method.^{11,12} Amino acid residues (Val, *N*,O-diMeTyr-1 and 2, Valthz-ca, and Amha) obtained by hydrolysis of compound 1 were derivatized with L-FDLA. LC-MS analysis of the L-FDLA derivatives using reversed phase column (C_{18}) revealed the absolute configuration of each residue.

Kakeromamide A (1) exhibited a moderate cytotoxicity with an IC₅₀ value of 10 µM against HeLa cells, while it showed no cytotoxicity against P388 cells at 10 µM. Cyclic depsipeptides ulongamide D⁴ encompassing the similar amino acid sequence in which Val is replaced by hydroxylisovaleric acid, showed cytotoxicity against KB and LoVo cells with lower IC_{50} values of 1 μM and 5 μM , respectively. In the in vitro differentiation model using mouse ES cells, compound **1** induced differentiation from neural stem cells (NSCs)⁵ into astrocytes in the concentration dependent manner (2.5-10 μ M), while no cell death was observed (Fig. 3b-d). In addition, compound **1** suppressed the differentiation from NSCs into neurons which was monitored as the decreased number of the differentiated neurons stained by NeuO¹³ (Fig. S8). Since effects on NSC differentiation by marine cyclic peptide have never reported, this is the first example of the marine cyclic peptides inducing NSC differentiation. Compound 1 is expected as the lead compound for regenerative medicine or the chemical probe to study NSC differentiation system.

In conclusion, a new cyclic peptide kakeromamide A (1) containing unique amino acid residues of Val-thz-ca and β -amino acid (Amha) was isolated from the marine cyanobacterium *Moorea bouillonii*. Compound 1 induced differentiation of neural stem cells into astrocytes without marked cytotoxicity. The detailed mechanism for this biological activity is under the investigation.

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A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at https://doi.org/10.1016/j.bmcl.2018.04.067.

References

- 1. Luesch H, Yoshida WY, Moore RE, Paul VJ. J Nat Prod. 2000;63:1437-1439.
- 2. Pettit GR, Kamano Y, Herald CL, et al. J Am Chem Soc. 1987;109:6883–6885.
- 3. Luesch H, Moore RE, Paul VJ, Mooberry SL, Corbett TH. J Nat Prod. 2001;64:907–910.
- Luesch H, Williams PG, Yoshida WY, Moore RE, Paul VJ. J Nat Prod. 2002;65:996–1000.
- 5. Iwata T, Otsuka S, Tsubokura K, et al. Chem Eur J. 2016;22:14707-14716.
- 6. Nakayama T, Inoue N. Methods Mol Biol. 2006;330:1–13.
- 7. Summers MF, Marzilli LG, Bax A. J Am Chem Soc. 1986;108:4285–4294.
- Bax A, Aszalos A, Dinya Z, Sudo K. J Am Chem Soc. 1986;108:8056–8063.
 Williamus PG, Yoshida WY, Moore RE, Paul VJ, Liu JO. J Nat Prod.
- 9. Winfamus PG, Yoshida WY, Moore KE, Paul VJ, Liu JO. J Nat Prod. 2002;65:29–31.
- 10. Crews P, Kakou Y, Quiñoà E. J Am Chem Soc. 1988;110:4365-4368.
- 11. Fujii K, Ikai Y, Mayumi T, Oka H, Suzuki M, Harada K. Anal Chem. 1997;69:3346–3352.
- 12. Fujii K, Ikai Y, Oka H, Suzuki M, Harada K. Anal Chem. 1997;69:5146–5151.
- 13. Er JC, Leong C, Teoh CL, et al. Angew Chem Int Ed. 2015;54:2442-2446.