

Development of Spiroligomer-Peptoid Hybrids

Justin D Northrup, Giulia Mancini, Claire R. Purcell, and Christian E. Schafmeister

J. Org. Chem., **Just Accepted Manuscript** • DOI: 10.1021/acs.joc.7b01956 • Publication Date (Web): 21 Nov 2017

Downloaded from <http://pubs.acs.org> on November 24, 2017

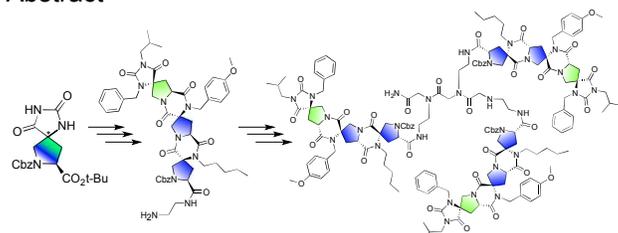
Just Accepted

“Just Accepted” manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides “Just Accepted” as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. “Just Accepted” manuscripts appear in full in PDF format accompanied by an HTML abstract. “Just Accepted” manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). “Just Accepted” is an optional service offered to authors. Therefore, the “Just Accepted” Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the “Just Accepted” Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these “Just Accepted” manuscripts.



DEVELOPMENT OF SPIROLOGOMER-PEPTOID HYBRIDSJustin D. Northrup,[‡] Giulia Mancini,[§] Claire R. Purcell,[‡] and Christian E. Schafmeister^{‡*}[‡]Department of Chemistry, Temple University, Philadelphia, Pennsylvania 19122, United States of America[§]Department of Chemistry, University of the Sciences, Philadelphia, Pennsylvania 19104, United States of America

*corresponding author: meister@temple.edu

Abstract

Creating functional macromolecules that possess the diversity and functionality of proteins poses an enormous challenge, as this requires large, preorganized macromolecules to facilitate interactions. Peptoids have been shown to interact with proteins, and combinatorial libraries of peptoids have been useful in discovering new ligands for protein binding. We have created spirologomer-peptoid hybrids that have a spirocyclic core that preorganizes functional groups in three-dimensional space. By utilizing spirologomers, we can reduce the number of rotatable bonds between functional groups, while increasing the stereochemical diversity of the molecules. We have synthesized 15 new spirologomer monomer amines that contain two stereocenters and three functional groups (67-84% yields from a common hydantoin starting material), as well as a spirologomer trimer **25** with six stereocenters and five functional groups. These 16 amines were used to synthesize five first-generation spirologomer-peptoids hybrids.

Introduction

To create functional macromolecules with the protein binding, catalytic, and membrane channel activities of folded proteins, it is necessary to create macromolecules that organize many diverse functional groups in pre-organized three-dimensional constellations. Many approaches to this challenge have been developed including peptide synthesis, stapled-peptides,^{1,2} beta- and gamma-peptides,^{3,4} triazolamers,^{5,6} and N-amino peptides,^{7,8} among many others. Two other

1
2
3 approaches are spiroligomers and peptoids, which are both peptidomimetic oligomers
4 synthesized from readily available building blocks, with spiroligomers being synthesized from a
5 unique set of bis-amino acid monomers derived from *trans*-4-hydroxy-*L*-proline,^{9,10} while peptoids
6 are typically synthesized from bromoacetic acid and a primary amine via a submonomer
7 method.¹¹
8
9

10
11
12 Peptoids have been studied for decades and have been utilized for a wide variety of
13 purposes: Peptoids have been demonstrated as biologically active molecules ranging from
14 protein secondary structure mimetics¹²⁻¹⁶ to antimicrobial agents¹⁷⁻¹⁹ and other therapeutics;²⁰
15 they have been used to construct nanomaterials such as nanotubes^{21,22} and nanosheets;^{23,24} and
16 metal binding peptoids.²⁵⁻²⁸ Peptoids are assembled using a very convenient submonomer
17 approach, which allows for efficient synthesis of long oligomers with excellent yield and purity.
18 Compared to peptides, the side chain is moved from the α -carbon to the backbone nitrogen
19 creating a tertiary amide linkage. This removes the backbone chirality, any inter- or intrachain
20 hydrogen bonding of the backbone, and leads to increased flexibility of the peptoid backbone
21 relative to peptides due to rotational freedom around the ω dihedral.²⁹⁻³¹ To increase the degree
22 of pre-organization, Kirshenbaum,³² and others, have developed a macrocyclization strategy for
23 the peptoid core.^{18,19,33-38} These macrocycles range from only three residues in the peptoid core,³³
24 to six, eight, or more residues.³² Peptoid containing macrocycles have been utilized for many
25 applications, including protein inhibitors,^{39,40} antimicrobials,¹⁸ and antifreeze agents.⁴¹
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40

41 Spiroligomers are excellent at pre-organizing functional groups, as the fused-spirocyclic
42 structures do not allow any free rotation throughout the backbone.^{9,42,43} Spiroligomers also benefit
43 from having a large pool of side chains to utilize, as almost any aldehyde, ketone, alkyl halide,
44 isocyanate, or amino acid can be incorporated to provide a functional group.^{44,45} Functionalized,
45 preorganized spiroligomers have been utilized for a variety of applications such as a
46 transesterification catalyst,⁴⁶ a proline-aldol catalyst,⁴⁷ a spiroligomer that binds MDM2,⁴⁸ and a
47 spiroligomer that forms supramolecular metal binding complexes,⁴⁹ among many others.
48
49
50
51
52
53
54

55 The submonomer synthesis of peptoids allows the incorporation of a large variety of
56 functional groups as primary amines. The first peptoids that incorporated large side chains were
57
58
59
60

1
2
3 glucose-based glycopeptide mimetics,⁵⁰ with interest in these glycopeptoids ongoing.^{51,52} Large
4 side chains have also been incorporated into peptoids using a 3+2 Huisgen's cyclization (a click
5 reaction),⁵³ after the displacement step with propargyl amine.⁵⁴⁻⁵⁶
6
7
8

9 Herein, we investigate linking complex, stereochemically-pure spiroligomers that display
10 multiple functional groups through a peptoid backbone. This approach would provide new
11 molecules for combinatorial screening of protein-protein interaction inhibitors and catalysts. To
12 achieve this, we need to incorporate a protected primary amine in the spiroligomer synthesis, and
13 we need to develop efficient conditions for integrating these complex amines into peptoids. In
14 doing so, we can then combine any number of uniquely functionalized spiroligomers into any
15 sequence of a peptoid for a variety of applications.
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

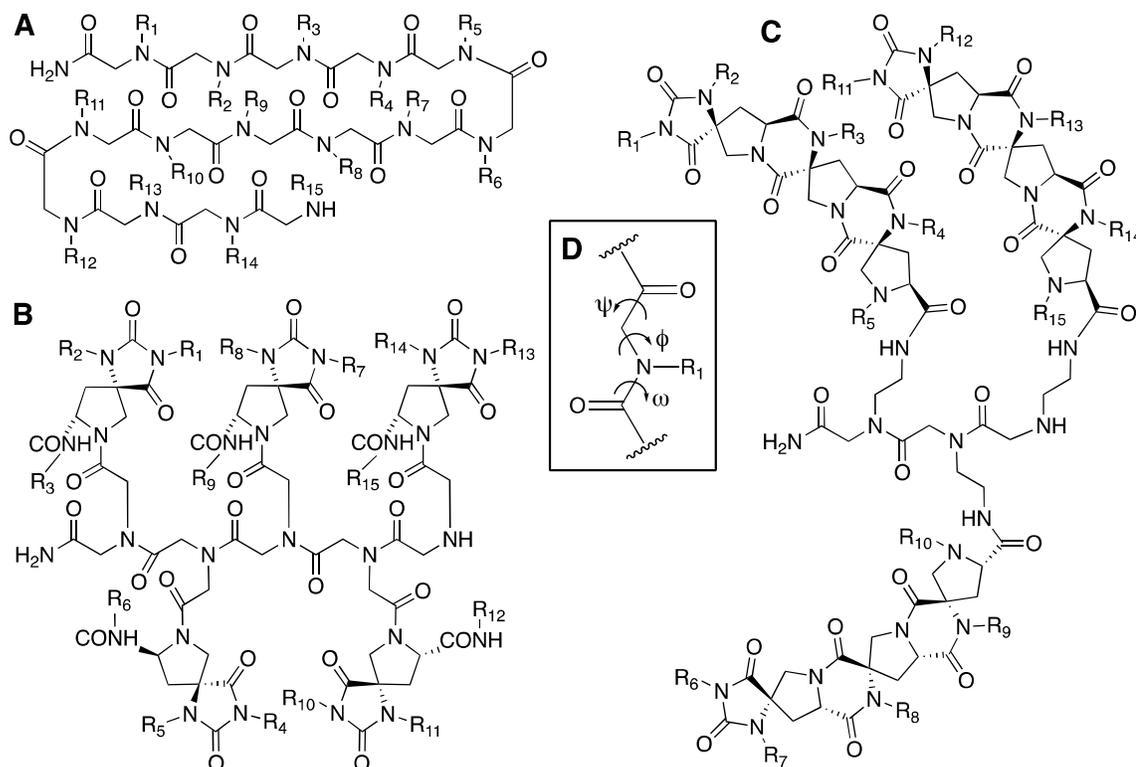


Figure 1. Three theoretical molecules displaying identical functional groups with increasing preorganization. (A) A theoretical peptoid 15-mer (B) A theoretical spirologomer-peptoid hybrid 5-mer, incorporating five spirologomers which each contain three functional groups and two stereocenters (15 total groups, 10 total stereocenters) (C) A spirologomer-peptoid hybrid 3-mer, incorporating three spirologomers which each contain five functional groups and six stereocenters (15 total groups, 18 total stereocenters). (D) Depiction of a peptoid showing omega, phi, and psi dihedral angles.

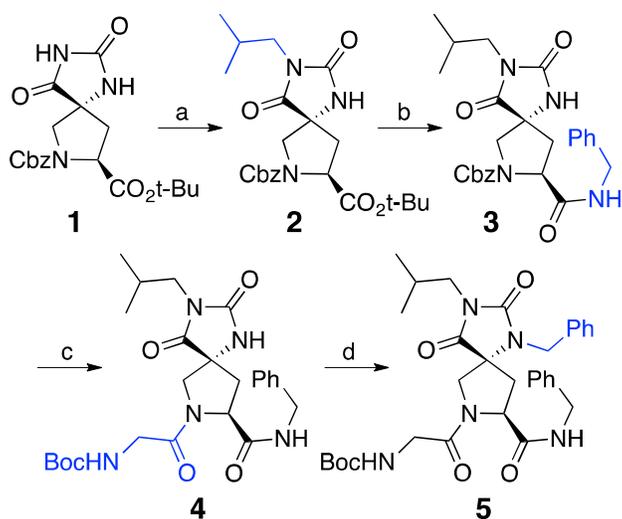
As shown in Figure 1A, a prototypical linear peptoid 15-mer has a very high ratio of rotatable bonds to functional groups. Each peptoid residue has a ϕ , ψ , and ω rotatable bond (Figure 1D), which means there are 42 rotatable bonds in the peptoid backbone. In comparison, a spirologomer-peptoid hybrid like the one shown in Figure 1B incorporates fifteen functional groups and ten stereo-centers, yet it contains only 27 rotatable bonds in the peptoid backbone (a 35% reduction compared to the prototypical peptoid). Spirologomers similar in size and functional group display to the amines for Figure 1B have previously been used by our group as small, organic catalysts.⁴⁷ If we were to incorporate a spirologomer trimer into a peptoid (Figure 1C), fifteen functional groups can be incorporated with just three spirologomer trimers containing eighteen total stereo-centers, and only 21 rotatable bonds in the backbone (a 50% reduction compared to the prototypical peptoid). For instance, between groups R₁-R₅ on the peptoid there are 12

1
2
3 rotatable bonds between residues in the peptoid backbone, whereas between groups R_1 - R_5 on
4
5 the spirologomer, there are no rotatable bonds in the spirologomer core. Spirologomers similar in
6
7 size and functional group display to the amines for Figure 1C have been shown to bind in the P53
8
9 groove of HDM2.⁴⁸ These hypothetical spirologomer-peptoid hybrids represent a wide range of
10
11 functional group and stereochemical diversity relative to flexibility, with increasing diversity and
12
13 increasing pre-organization as the size of the spirologomer is increased. Each of these
14
15 functionalized spirologomers approximates the structural complexity of a small-molecule fused-
16
17 ring natural product containing two, four, six or more fused rings and several functional groups in
18
19 precise three-dimensional constellations. Three to five of these spirologomer domains can be
20
21 displayed – in close proximity – on a peptoid chain and provide a molecule with a great deal of
22
23 preorganization that has a large surface area which could potentially interact with other
24
25 molecules, and at the same time display flexibility between the domains to accommodate different
26
27 protein surfaces. One could imagine molecules like these binding to multiple shallow grooves on
28
29 a protein surface through many non-covalent contacts. By incorporating the structural rigidity and
30
31 stereochemical diversity of spirologomers with the modular linking chemistry provided by peptoid
32
33 synthesis, this should facilitate the rapid discovery of new molecules of interest for catalysis,
34
35 disrupting protein-protein interactions, and exploring large surface area host-guest interactions.
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

RESULTS AND DISCUSSION

Synthesis of Spiroligomer Primary Amines

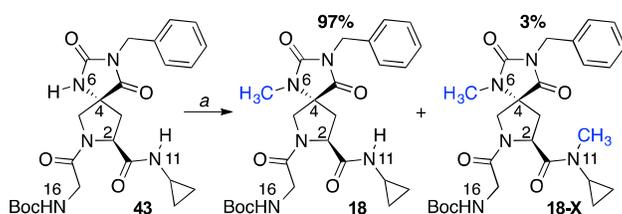
To facilitate the synthesis of spiroligomer-peptoid hybrids, we must first synthesize a variety of spiroligomers displaying primary amines. The building blocks of spiroligomers are the four stereoisomers of the proline hydantoin intermediate in spiroligomer bis-amino acid synthesis that we routinely synthesize on a 600 mmol scale (230g of mixed diastereomers and we have synthetic access to all four stereoisomers).¹⁰ We have recently developed chemistry that allows us to directly functionalize the hydantoin by exploiting the varying reactivity between the imide and amide nitrogen atoms of the hydantoin.⁴⁵ As shown in Scheme 1, we can install a functional



Scheme 1: Synthesis of spiroligomer amine **5** from (2*S*, 4*R*) proline hydantoin **1**. a) i. DMF, K₂CO₃, 1-iodo-2-methylpropane; ii. Et₂NH; b) 1. TFA; 2. i. HOAT, EDCI, DMF, DCM; ii. benzylamine; c) 1. 1:1 HBr/DCM; 2. DMF, DCM, DIPEA, Boc-Gly-OAT; d) i. DMF, K₂CO₃, benzylbromide; ii. Et₂NH

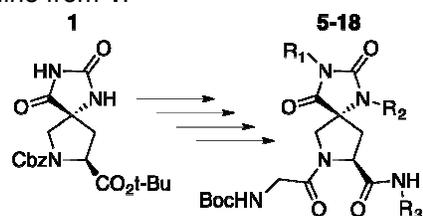
group onto the imide of hydantoin **1** via direct alkylation with either alkyl, allyl, or benzyl halides to form **2**. We then remove the *tert*-butyl protecting group with TFA, and couple a primary amine to form **3**. This is followed by removal of the Cbz protecting group and coupling an N-Boc protected amino acid to make **4**. Finally, another alkylation with either an allyl or benzyl halide to install a functional group on the amide of the hydantoin provides the final spiroligomer N-Boc protected amine **5**. We observe that the hydantoin amide is functionalized selectively in the presence of the secondary amide and secondary carbamate on **4**. When a small, highly reactive electrophile such

1
2
3 as methyl iodide is used, we observe less than 3% alkylation to what is most likely the secondary
4 amide N11 (Scheme 2). The evidence that the alkylation is taking place selectively on the
5 hydantoin amide was determined from the ^1H NMR for the alkylated products. If N6 were
6 unalkylated, we would expect to see a triplet for NH16 and a singlet for NH6, yet we see two
7 amide NH signals that show either a triplet for NH16 and a triplet for NH11 or a triplet for NH16
8 and a doublet for NH11, depending on whether the carbon attached to N11 is a methylene or
9 methyne, respectively. This chemistry was used with varying electrophiles and amines to form the
10 spirooligomer amines **6** to **18** shown in Table 1.
11
12
13
14
15
16
17
18
19



27
28 Scheme 2. a) DMF, 2 equiv $\text{CH}_3\text{-I}$, 2 equiv K_2CO_3 , 24 hours. Methylation of N6 is the only
29 alkylation that led to the formation of another product, which has the same mass as **18-X** and we
30 presume that it has the structure shown above (this byproduct was not recovered)
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 1. Spiroligomer amines synthesized from proline hydantoin **1**. Yield represents total yield of the amine from **1**.



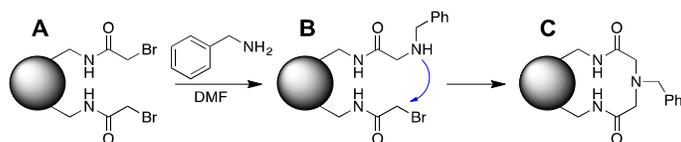
Cmpd	R ₁	R ₂	R ₃	Yield
5				84%
6				69%
7				75%
8				67%
9				73%
10				76%
11				75%
12				73%
13				79%
14				76%
15				77%
16				75%
17				75%
18				71%

Testing the Minimum Requirements for Peptoid Chemistry

1
2
3 To make efficient use of these complex and synthetically derived amines we carried out a
4 series of trials to determine the minimum amount of the amines required to achieve peptoid
5 couplings while minimizing cross-linking (C-L) of the nascent peptoids on solid support. We varied
6 the number of equivalents of amine relative to the resin loading on solid support and the
7 concentration of amine required for peptoid chemistry to function reliably. We utilized
8 commercially available benzylamine to obtain the baseline conditions for peptoid chemistry
9 shown in Table 2, below. Varying the concentration of amine in the peptoid submonomer
10 synthesis,^{11,26} we determined that 3 equivalents of amine relative to resin loading (Rink Amide
11 polystyrene, 0.63 mmol/g) with one equivalent of exogenous base (DIPEA; extra base lead to
12 uncharacterized side products), at a concentration of 150 mM of benzylamine in DMF were the
13 minimum equivalents and concentration that achieved efficient peptoid coupling and suppressed
14 cross-linking to below 2%. Cross-linked products were observed and quantitated using HPLC-
15 MS/UV-VIS. Any further decrease in equivalents or concentration resulted in significant cross-
16 linking of the resin, whereby a secondary amine on resin would react with another bromoacetate
17 on resin before an amine in solution could react, thereby terminating the sequence (shown in
18 Scheme 3). On Rink Amide Tentagel® resin (loading 0.37 mmol/g), we found that we could lower
19 the required concentration of amine to 125 mM, but the minimum number of equivalents required
20 remained at 3 equivalents. A 2-Cl-Trityl chloride polystyrene resin (loading 1.1 mmol/g) required
21 an increase in both the equivalents and concentration of benzylamine (5 equiv and 250 mM,
22 respectively) to ensure efficient coupling and suppress cross-linking to less than 2%. These
23 results were achieved at room temperature with four hour couplings.
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 2. Minimum equivalents required for peptoid couplings on various resins

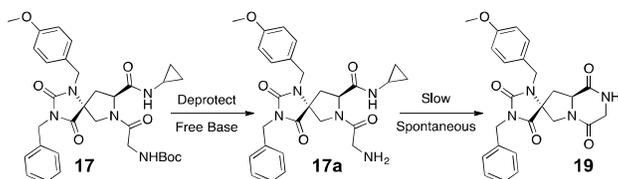
Trial	Equiv. Amine	Equiv. Base	Amine Conc. (mM)	Resin Loading (mmol/g)	% C-L (UV)
1	20	0	2000	0.63	< 1%
2	3	2	150	0.63	Side Product
3	3	1	150	0.63	< 2%
4	3	1	10	0.63	6-8%
5	3	1	30	0.63	6-8%
6	3	1	100	0.63	2-3%
7	2	1	100	0.63	2-3%
8	2	1	150	0.63	2-3%
9	3	1	125	0.37	< 2%
10	3	1	75	0.37	6-8%
11	3	1	150	1.1	8-10%
12	5	1	250	1.1	< 2%



Scheme 3. Benzylamine is combined with the resin (A); however, low concentration of amine in solution allows for an amine on resin to displace a neighboring bromine (B). This results in a crosslinked resin, and the termination of two peptoid sequences (C). C-L(UV) stands for cross-linking as determined using HPLC with UV detection.

Having determined the requirements of peptoid chemistry at sub molar concentrations, we proceeded to synthesize several variants of spiro oligomer-peptoid hybrids. Any primary amine we use would first need to be Boc deprotected, as shown in Scheme 4, which was achieved with a 50% TFA / DCM mix, followed by concentration *in vacuo*. The free base of the amine **17** was obtained by extracting the trifluoroacetate salt with EtOAc and 1M KOH, followed by two washes with an equal volume of 1M KOH, which were back extracted with EtOAc. The Boc-protected amines are stable at room temperature for several years; however, if the free amine is left dry at room temperature or in solution, we observed slow, spontaneous formation of the diketopiperazine (DKP) **19**. To avoid this, the amine free-base was concentrated under reduced

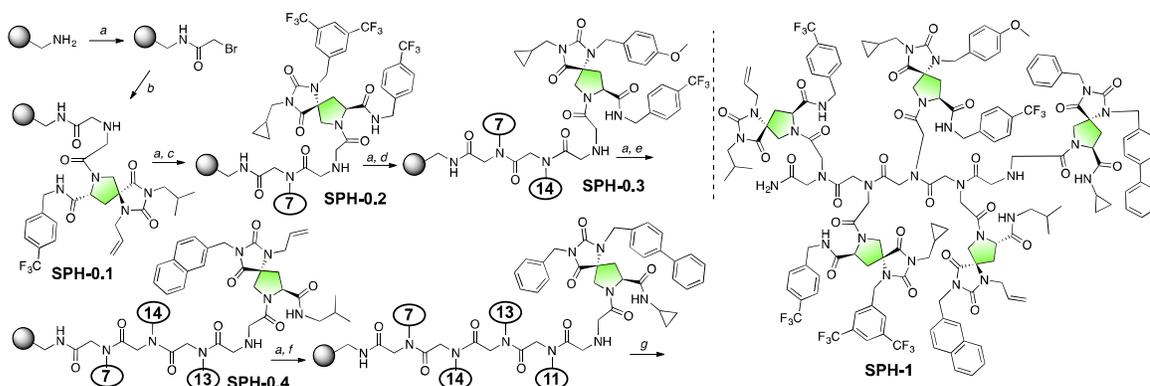
pressure to yield a foamy solid, which is stable for over a year (dry, -20 °C). Unreacted amine in the peptoid coupling can be recovered, protected, and stored.



Scheme 4. Generation of the spiroligomer free amine **17a** from **17**. Spontaneous DKP formation of spiroligomer free amines is possible at room temperature in solution.

Synthesis of Spiroligomer Peptoid Hybrids

Scheme 5 demonstrates the synthesis of spiroligomer-peptoid hybrid **1 (SPH-1)** on Rink Amide polystyrene resin using standard submonomer synthesis and spiroligomer primary amines. This



Scheme 5. Peptoid synthesis of the spiroligomer-peptoid hybrid, **SPH-1** (a) Bromoacetic acid, DIC, DMF; (b) DMF, 150 mM **7**, 1 equiv DIPEA (c) DMF, 150 mM **14**, 1 equiv DIPEA; (d) DMF, 150 mM **15**, 1 equiv DIPEA; (e) DMF, 150 mM **11**, 1 equiv DIPEA; (f) DMF, 150 mM **16**, 1 equiv DIPEA; (g) TFA, neat.

spiroligomer-peptoid hybrid incorporates 15 functional groups and ten stereocenters on pre-organized spiroligomer side chains. HPLC chromatograms documenting each elongation of **SPH-1** are highlighted in Figure 2. There is a solvent injection peak (denoted I) and a resin impurity (denoted as II) that are present in each chromatogram (Figure 2A-D, test cleavages) which do not change. The intended product III is the only other peak in the test cleavages. Figure 2E shows the crude HPLC chromatogram for **SPH-1**, which indicates a high level of fidelity for each coupling step. The peak around 2.8 min is from the final spiroligomer amine, which indicated longer washing steps were needed to recover unreacted amine from the resin. The spiroligomer-

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

peptoid hybrid can then be purified to remove the small impurities present in the crude material (Figure 2F-G).

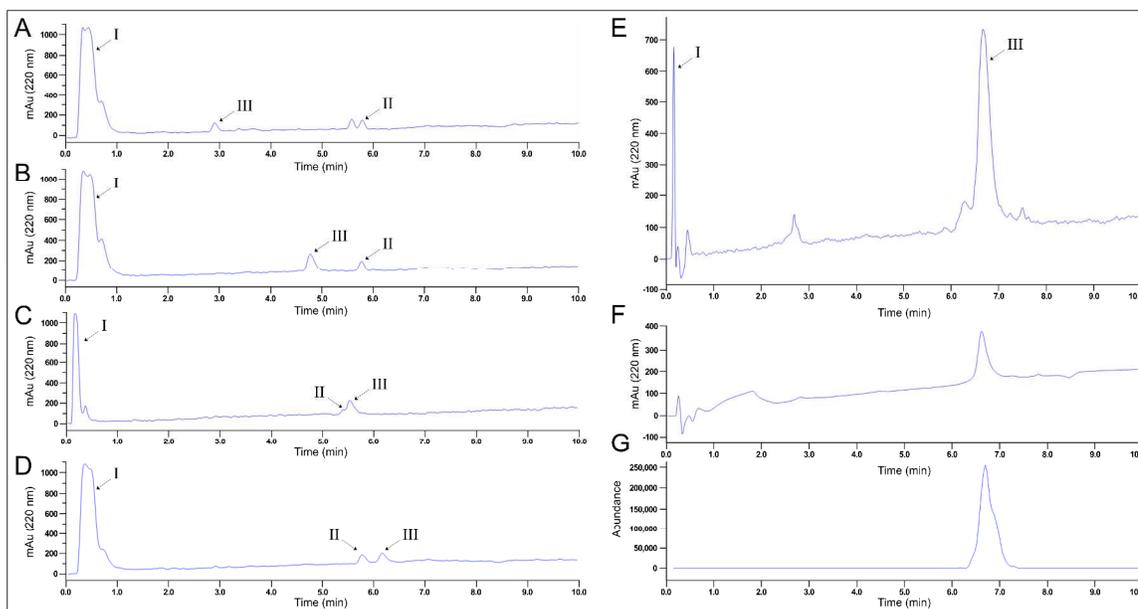


Figure 2. HPLC Chromatograms (A-F) following the synthesis of **SPH-1** (I = DMSO for A-D, 50% ACN/Water for E-G, II = resin impurity, III = desired product): (A) Test Cleavage of **SPH-0.1** (B) Test cleavage of **SPH-0.2** (C) Test cleavage of **SPH-0.3** (D) Test cleavage of **SPH-0.4** (E) Full resin cleavage for **SPH-1**, crude (F, G) HPLC-MS for purified **SPH-1**.

To test the fidelity of a spiroligomer-peptoid hybrid with increasing chain length, we synthesized both a 9-mer (**SPH-2**, 58% yield) and a 12-mer (**SPH-3**, 57%) incorporating 3 and 6 different amines, respectively (Figure 3A, B). Other spiroligomer-peptoid hybrids that we have synthesized include custom linkers incorporating propargyl (post-peptoid synthesis modification potential) or bromobenzyl side chains (the bromobenzyl group provides an easily identified mass spectroscopy signature due to the natural isotopic abundance of bromine). Installation of a c-terminal methionine residue to facilitate CNBr mediated cleavage from resin has been tested successfully on various spiroligomer-peptoid hybrids (Figure 3C, **SPH-4**, 22% yield). These types of hybrids would be most beneficial to those interested in on-resin screening for compounds, as it allows for the deprotection of any side chains, while leaving the peptoid still attached to the resin.

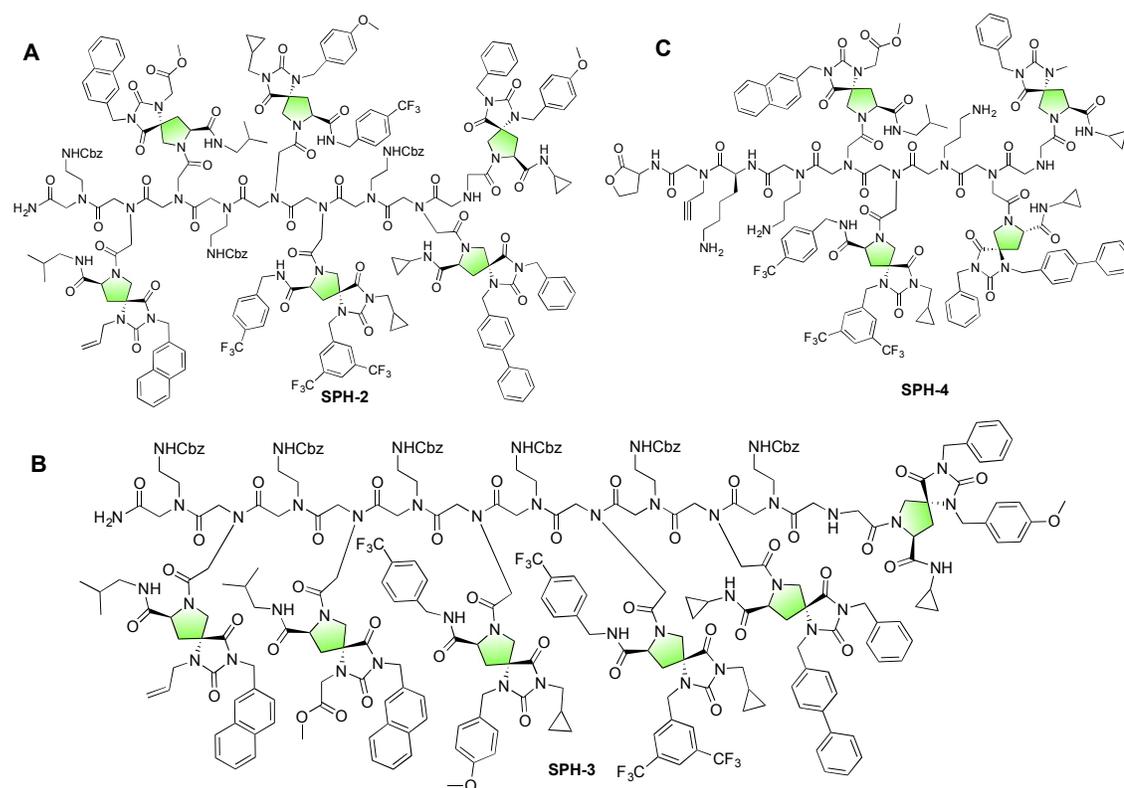
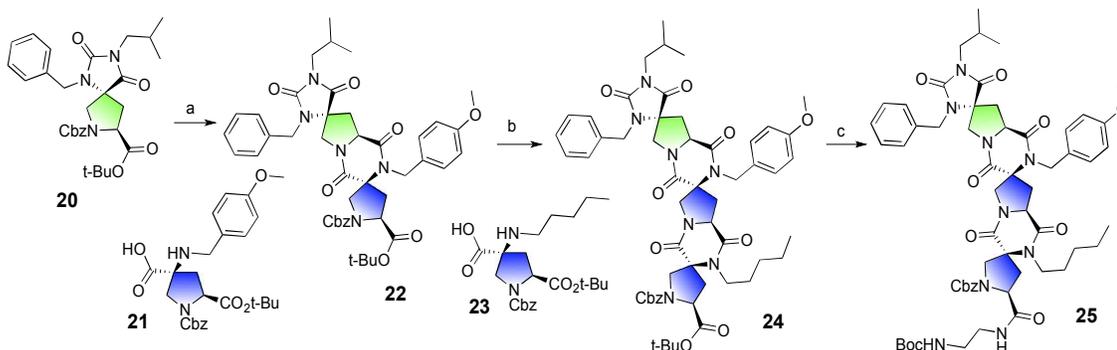


Figure 3. Three spirologomer-peptoid hybrids of varying length, functional groups, and resin linker. (A) **SPH-2**, a peptoid 9-mer incorporating 6 spirologomers (B) **SPH-3**, a peptoid 12-mer incorporating 6 spirologomers (C) **SPH-4**, a hybrid peptoid containing 8 residues, four of which are spirologomers, and an initial methionine residue for CNBr mediated cleavage.

Synthesis of a spirologomer-peptoid hybrid containing a spirologomer trimer

To further reduce the number of rotatable bonds in the peptoid backbone, we have synthesized a spirologomer-peptoid hybrid containing three copies of a highly preorganized, spirologomer trimer. Spirologomer trimers have been demonstrated within our lab as designed catalysts⁴⁶ or for protein binding.⁴⁸ Incorporating large, structured macromolecules like these will be used to create even more functional, preorganized macromolecules. The trimer **25** was synthesized from a bifunctionalized proline hydantoin **20** and two bis-amino acids **21** and **23**, as shown in Scheme 6. The Cbz and *t*-Bu protected, dialkylated proline hydantoin **20** was treated with 1:1 DCM/(33% HBr/AcOH) for 30 minutes, after which the solvent was removed *in vacuo*, and the molecule left on a high vacuum pump overnight. Bis-amino acid **21** was preactivated

under inert conditions with EDC and HOAt in a 1:1 mix of anhydrous DMF/DCM for 1.5 h. This preactivation is achieved without base so the bis-amino acid does not react with itself. Deprotected compound **20** was dissolved in a minimal amount of anhydrous DMF, and added to the preactivated bis-amino acid **21** along with DIPEA. After stirring overnight, more EDC was added to the reaction to close the diketopiperazine ring to afford the spirologomer dimer **22**. Spirologomer dimer **22** is purified via flash chromatography (gradient from 0-100% ethyl acetate in hexanes) in 46% yield.



Scheme 6. Synthesis of the spirologomer trimer **25** (a) i. 1:1 DCM/(33% HBr/AcOH); ii. (Preactivated **21**: HOAT, EDC, 1:1 dry DMF/DCM, 1.5 h), DMF, DIPEA; (b) i. 1:1 DCM/(33% HBr/AcOH); ii. (Preactivated **23**: HOAT, EDC, 1:1 dry DMF/DCM, 1.5 h), DMF, DIPEA; (c) i. 95:4:1 TFA/H₂O/TIPS, 1 h; ii. EDC, HOAT, 1:1 dry DMF/DCM, 1.5 h; iii. N-boc-ethylenediamine

Dimer **22** was deprotected with 1:1 DCM/(33% HBr/AcOH) for 30 minutes, the solvent removed *in vacuo*, and held under high vacuum overnight. Bis-amino acid **23** was pre-activated under inert conditions with EDC and HOAt in a 1:1 mix of anhydrous DMF/DCM for 1.5 h. This preactivation is achieved without base so the bis-amino acid does not react with itself. Dimer **22** was dissolved in a minimal amount of anhydrous DMF, and added along with DIPEA to the preactivated bis-amino acid **23** to afford the spirologomer trimer **24**. Spirologomer trimer **24** is then purified via flash chromatography (gradient from 0-100% ethyl acetate in hexanes) in 21% yield. Spirologomer **24** can then be treated with a 95:4:1 TFA/H₂O/TIPS mixture to remove the *t*-Bu protecting group, then following the removal of the solvent, this spirologomer is preactivated using EDC and HOAt in a 1:1 mix of dry DMF/DCM for 1.5 h. The preactivated spirologomer was combined with N-Boc-ethylenediamine to provide spirologomer trimer **25**, which was utilized to

make **SPH-5** shown in Figure 4 (25% purified yield) using standard peptoid synthesis. This spirooligomer trimer is, to the best of our knowledge, the largest amine utilized for the displacement step of a peptoid synthesis, containing five unique functional groups and six stereocenters per residue. This means the spirooligomer-peptoid hybrid **SPH-5** contains 15 functional groups and 18 stereocenters, over all of which we have total control.

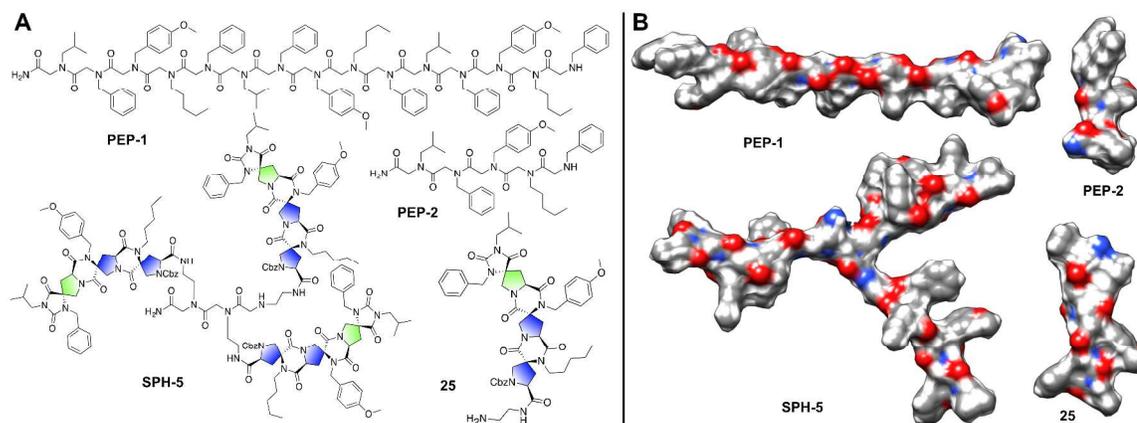


Figure 4. Comparisons of theoretical peptoids **PEP-1** (15mer) and **PEP-2** (5mer) vs **SPH-5** and amine **25**. (A) Chemdraws for **PEP-1**, **PEP-2**, **SPH-5**, and **25**. (B) CANDO models with solvent-excluded surface for **PEP-1** (surface area = 1743 Å²), **PEP-2** (surface area = 628.3 Å²), **SPH-5** (surface area = 2385 Å²), and **25** (surface area = 774.6 Å²).

By incorporating these large preorganized amines, we have successfully reduced the number of peptoid-backbone rotatable bonds by 50%. This was accomplished while increasing the potential surface area available for interactions as shown in Figure 4B, which depicts the solvent-excluded surface for two theoretical peptoids **PEP-1** and **PEP-2**, as well as hybrid **SPH-5** and its component amine **25**. When compared to a theoretical peptoid (**PEP-1**) with the same functional groups, **SPH-5** increases the total surface area by 33%. Similarly, when compared to a theoretical peptoid (**PEP-2**) with the same functional groups, amine **25** increases the total surface area by 23%. This shows that preorganization of functional groups on a spirooligomer-peptoid hybrid will not only reduce the number of backbone rotatable bonds, but it will also increase the potential surface area available for host-guest or protein-protein interactions. These large spirooligomer trimers, like the smaller monomers shown previously, can be used sequentially, or interspersed with smaller units, indicating that they would make excellent candidate amines for

1
2
3 future work. Furthermore, large spirologomer trimers preorganize many more groups than the
4 spirologomer monomer amines or regular peptoid amines, thus reducing any entropic penalty that
5 would be associated when binding a protein for example.
6
7
8
9

10 11 12 **Conclusion**

13
14 We have successfully synthesized 16 new spirologomers containing three to five functional
15 groups, two to six stereocenters, and a protected primary amine, which we have used to
16 incorporate the spirologomers into peptoids of varying lengths. To the best of our knowledge, this
17 is the first reported synthesis which utilized large, preorganized, spirocyclic amines during the
18 displacement step of peptoid synthesis. Notably, this work featured the use of spirologomer trimer
19 **25**, which organizes five functional groups on a spirologomer backbone containing six
20 stereocenters and no interior rotatable bonds. This spirologomer was used to make the
21 spirologomer-peptoid hybrid **SPH-5**, which contains 18 stereocenters and 15 functional groups
22 across three peptoid residues. A linear peptoid with the same number of functional groups would
23 have 42 rotatable bonds in the backbone, whereas this hybrid has only 6 in the backbone and 15
24 in the linkers, a 50% reduction in the number of rotatable bonds for the same number of
25 functional groups. This preorganization of functional groups will help facilitate the rapid discovery
26 of novel structures for host-guest interactions, protein-protein interactions, and catalysis. Now that
27 we can synthesize these new hybrid peptidomimetics, further work will be focused on developing
28 spirologomer-peptoid hybrids that display the appropriate side chains for these important
29 interactions.
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

48 **Experimental Details**

49 50 51 **General Procedure 1: Mono alkylation of P4srZBHyd**

52 To a stirred mixture of **1** in DMF [100 mM] was added 0.75-0.85 equiv of alkyl, allyl, or benzyl
53 halide (equiv dependent on salt content of the specific batch **1**, which can be determined utilizing
54 an internal standard) along with 1.5 equiv of K₂CO₃. The reaction proceeded at room temperature
55
56
57
58
59
60

1
2
3 for 2-24 h, dependent on electrophile, and the progress checked via LCMS. The reaction was
4 diluted with four times the reaction volume of EtOAc and washed with water, saturated
5 ammonium chloride solution, and brine. The organic layer was dried with Na₂SO₄, and
6 concentrated *in vacuo* to yield **26-29** as foamy off-white to yellow solids. ¹H / ¹³C NMR spectra for
7 compounds **26-29** are available in the supporting information.
8
9
10
11
12

13
14
15 **Compound 26 - 7-benzyl 8-(tert-butyl) (5R,8S)-3-(naphthalen-2-ylmethyl)-2,4-dioxo-1,3,7-**
16 **triazaspiro[4.4]nonane-7,8-dicarboxylate** - Compound **26** was synthesized using General
17 Procedure 1, with **1** [(2S, 4R) 7.5 mmol] and 2-(bromomethyl)naphthalene [6 mmol (0.80 equiv)].
18 Recovered yield was quantitative and the product used without purification. ¹H NMR (500 MHz,
19 CDCl₃) 1.42 (9H, s, rotameric), 2.36 (1H, m), 2.48 (1H, m), 3.81 (2H, m), 4.45 (1H, m), 4.78 (1H,
20 s, rotameric), 5.07 (2H, m), 7.25-7.84 (13H), 9.08 (1H, s, rotameric); ¹³C NMR (125 MHz, CDCl₃,
21 rotamers present) 27.8, 34.1, 39.6, 40.6, 42.8, 55.7, 58.4, 65.9, 66.8, 67.6, 82.4, 125.8, 126.2,
22 126.3, 126.4, 126.6, 126.8, 127.6, 127.8, 127.9, 128.0, 128.1, 128.2, 128.5, 128.6, 128.7, 132.9,
23 133.0, 133.2, 135.1, 135.8, 136.0, 154, 7, 156.3, 169.8, 172.9; HRMS (ESI/Q-TOF) m/z: (M+K)⁺
24 calcd for C₃₀H₃₁N₃O₆K 568.1844, found 568.1840
25
26
27
28
29
30
31
32
33
34
35

36
37 **Compound 27 - 7-benzyl 8-(tert-butyl) (5R,8S)-3-benzyl-2,4-dioxo-1,3,7-**
38 **triazaspiro[4.4]nonane-7,8-dicarboxylate** - Compound **27** was synthesized using General
39 Procedure 1, with **1** [(2S, 4R) 15 mmol] and benzyl bromide [12 mmol (0.80 equiv)]. Recovered
40 yield was quantitative and the product used without purification. ¹H NMR (500 MHz, CDCl₃) 1.43
41 (9H, s, rotameric), 2.31 (1H, dd, rotameric, J = 14.5, 8.1), 2.49 (1H, ddd, J = 18.0, 13.4, 9.2), 3.84
42 (2H, m), 4.43 (1H, dt, J = 24.4, 8.6), 4.63 (2H, s), 5.08 (2H, m), 7.30 (11H, m); ¹³C NMR (125
43 MHz, CDCl₃, rotamers present) 27.9, 39.7, 40.7, 42.5, 55.7, 58.9, 65.5, 66.2, 67.6, 82.4, 127.7,
44 127.9, 128.1, 128.2, 128.5, 128.6, 128.8, 135.7, 136.0, 154.5, 155.9, 169.7, 171.7; HRMS
45 (ESI/Q-TOF) m/z: (M+K)⁺ calcd for C₂₆H₂₉N₃O₆K 518.1688, found 518.1699
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Compound 28 - 7-benzyl 8-(tert-butyl) (5R,8S)-3-(cyclopropylmethyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-7,8-dicarboxylate - Compound **28** was synthesized using General Procedure 1, with **1** [(2S, 4R) 7.5 mmol] and (bromomethyl)cyclopropane [6.0 mmol (0.80 equiv)]. Recovered yield was quantitative and the product used without purification. ¹H NMR (500 MHz, CDCl₃) 0.33 (2H, d, J = 5.8), 0.49 (2H, d, J = 7.8), 1.13 (1H, m), 1.45 (9H, s, rotameric), 2.40 (1H, dd, rotameric, J = 14.3, 8.2), 2.54 (1H, ddd, J = 18.2, 13.4, 9.0), 3.35 (2H, dd, J = 7.2, 3.5), 3.81 (2H, m), 4.47 (1H, dt, J = 24.8, 8.3), 5.11 (2H, m), 7.32 (6H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 3.7, 10.2, 27.9, 40.7, 43.7, 55.8, 58.9, 66.1, 67.6, 82.4, 127.9, 128.2, 128.5, 136.0, 154.5, 156.8, 169.7, 172.2 HRMS (ESI/Q-TOF) m/z: (M+K)⁺ calcd for C₂₃H₂₉N₃O₆K 482.1688, found 482.1693

Compound 29 - 7-benzyl 8-(tert-butyl) (5R,8S)-3-isobutyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-7,8-dicarboxylate - Compound **29** was synthesized using General Procedure 1, with **1** [(2S, 4R) 15 mmol] and 1-iodo-2-methylpropane [18 mmol (1.2 equiv)]. Recovered yield was quantitative and the product used without purification. ¹H NMR (500 MHz, CDCl₃) 0.88 (6H, d, J = 6.7), 1.45 (9H, s, rotameric), 2.05 (1H, m), 2.38 (1H, m), 2.53 (1H, ddd, J = 18.2, 13.3, 9.2), 3.31 (2H, dd, J = 7.3, 3.1), 3.78 (2H, m), 4.48 (1H, dt, J = 24.4, 8.6), 5.11 (2H, m), 7.35 (6H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 20.9, 27.8, 27.9, 39.8, 40.8, 46.1, 55.8, 56.0, 58.4, 58.9, 65.2, 65.9, 67.6, 82.4, 127.8, 127.9, 128.2, 128.5, 135.9, 136.0, 154.2, 154.5, 156.6, 156.8, 169.7, 169.9, 172.2, 172.4; HRMS (ESI/Q-TOF) m/z: (M+K)⁺ calcd for C₂₃H₃₁N₃O₆K 484.1844, found 484.1861

General Procedure 2: Deprotection of C2 t-Bu group and coupling

Each of **26-29** was placed in a RB flask and then treated with 95% TFA / TIPS for 1 h, with the reaction progress checked via LCMS. Upon successful deprotection of the *tert*-butyl group, the solvent was removed *in vacuo*, and placed on a high vacuum pump overnight. To an inert atmosphere RB flask containing the free acid in dry DMF/DCM (1:1 ratio, [100 mM]) was added 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC, 1.5 equiv) and 1-Hydroxy-7-

1
2
3 azabenzotriazole (HOAT, 3 equiv), then stirred for 1 h. An amine (3 equiv) and DIPEA (2 equiv)
4 were then added, and the reaction proceeded for 2 h, at which point the progress was checked
5 via LCMS. Upon completion, the reaction was diluted with four times the reaction volume of
6 EtOAc and washed with saturated ammonium chloride solution, saturated sodium bicarbonate
7 solution, and brine. The organic layer was dried with Na₂SO₄, and concentrated *in vacuo* to yield
8 **30-35** as foamy yellow solids. ¹H / ¹³C NMR spectra for compounds **30-35** are available in the
9 supporting information.
10
11
12
13
14
15
16
17
18

19 **Compound 30 benzyl (5R,8S)-8-(isobutylcarbamoyl)-3-(naphthalen-2-ylmethyl)-2,4-dioxo-**
20 **1,3,7-triazaspiro[4.4]nonane-7-carboxylate** – Compound **30** was synthesized using General
21 Procedure 2 and compound **26**, with HOAt (22.5 mmol), EDC (11.25 mmol), Isobutylamine (22.5
22 mmol), DIPEA (15 mmol), and 75 mL of 1:1 DMF/DCM - Recovered yield was quantitative and
23 the product used without purification. ¹H NMR (500 MHz, CDCl₃) 0.82 (6H, d, *J* = 6.4), 1.89 (1H,
24 m), 2.25 (1H, m), 2.64 (1H, m), 2.99 (1H, m), 3.05 (1H, sep, *J* = 6.5), 3.68 (1H, m), 3.84 (1H, d, *J*
25 = 11.3), 4.59 (1H, m), 4.78 (2H, s, rotameric), 5.05 (2H, m), 7.02 (1H, m), 7.21 (5H, m), 7.44 (3H,
26 m), 7.66 (1H, m), 7.78 (4H, m); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers present) 20.0, 28.0,
27 28.1, 40.6, 41.5, 41.7, 46.0, 56.1, 56.5, 58.7, , 59.2, 65.1, 65.8, 66.2, 125.4, 125.5, 125.7, 125.8,
28 126.0, 126.3, 127.1, 127.5, 127.6, 127.7, 127.8, 128.2, 128.3, 132.2, 132.8, 134.0, 136.5, 136.6,
29 153.5, 152.7, 155.4, 155.5, 170.4, 170.7, 172.4; HRMS (ESI/Q-TOF) *m/z*: (M+H)⁺ calcd for
30 C₃₀H₃₃N₄O₅ 529.2445, found 529.2446
31
32
33
34
35
36
37
38
39
40
41
42
43
44

45 **Compound 31 - benzyl (5R,8S)-3-benzyl-8-(cyclopropylcarbamoyl)-2,4-dioxo-1,3,7-**
46 **triazaspiro[4.4]nonane-7-carboxylate** - Compound **31** was synthesized using General
47 Procedure 2 and compound **27**, with HOAt (22.5 mmol), EDC (11.25 mmol), cyclopropylamine
48 (22.5 mmol), DIPEA (15 mmol), and 75 mL of 1:1 DMF/DCM. Recovered yield was quantitative
49 and the product used without purification. ¹H NMR (500 MHz, CDCl₃) 0.30 (1H, m), 0.41 (1H, M),
50 0.58 (2H, dd, rotameric, *J* = 23.2, 7.1), 2.21 (1H, m), 2.33 (1H, m), 2.60 (1H, m), 3.64 (1H, dd,
51 rotameric, *J* = 15.5, 11.4), 3.81 (1H, d, *J* = 11.3), 4.30 (1H, m), 4.55 (2H, d, *J* = 12.5), 5.05 (2H,
52
53
54
55
56
57
58
59
60

1
2
3 m), 7.30 (10H, m), 8.20 (1H, m), 9.16 (1H, m); ^{13}C NMR (125 MHz, DMSO- d_6 , rotamers present)
4 5.4, 5.5, 5.6, 5.7, 22.3, 40.4, 41.3, 41.4, 56.0, 56.4, 58.6, 59.0, 65.1, 65.7, 66.2, 127.1, 127.2,
5 127.4, 127.5, 127.7, 127.8, 128.2, 128.4, 128.5, 128.6, 136.5, 136.6, 153.4, 153.6, 155.3, 155.4,
6 171.5, 171.8, 172.3; HRMS (ESI/Q-TOF) m/z : (M+Na) $^+$ calcd for $\text{C}_{25}\text{H}_{26}\text{N}_4\text{O}_5\text{Na}$ 485.1795, found
7 485.1790
8
9
10
11

12
13
14
15 **Compound 32 - benzyl (5R,8S)-3-benzyl-8-(isobutylcarbamoyl)-2,4-dioxo-1,3,7-**
16 **triazaspiro[4.4]nonane-7-carboxylate** - Compound **32** was synthesized using General
17 Procedure 2 and compound **27**, with HOAt (22.5 mmol), EDC (11.25 mmol), Isobutylamine (22.5
18 mmol), DIPEA (15 mmol), and 75 mL of 1:1 DMF/DCM. Recovered yield was quantitative and the
19 product used without purification. ^1H NMR (500 MHz, CDCl_3) 0.76 (6H, d, rotameric, $J = 6.7$) 1.62
20 (1H, sep, rotameric, $J = 6.7$), 2.22 (1H, dt, $J = 12.8, 10.0$), 2.38 (1H, m), 2.87 (2H, m), 3.66 (1H,
21 dd, $J = 17.0, 11.5$), 3.84 (1H, d, rotameric, $J = 11.5$), 4.43 (1H, dd, rotameric, $J = 10.0, 7.5$), 4.56
22 (2H, d, $J = 11.9$), 5.05 (1H, d, $J = 7.6$), 5.07 (1H, d, $J = 5.2$), 7.30 (10H, m), 8.14 (1H, t, rotameric,
23 $J = 5.9$), 9.22 (1H, s, rotameric); ^{13}C NMR (125 MHz, DMSO- d_6 , rotamers present) 19.42, 27.4,
24 27.5, 40.0, 40.9, 45.4, 55.5, 55.9, 58.1, 58.6, 64.5, 65.1, 65.6, 126.5, 126.6, 126.8, 126.9, 127.0,
25 127.2, 127.6, 127.8, 128.0, 135.9, 136.0, 152.9, 153.1, 154.8, 154.9, 169.8, 170.1, 171.8; HRMS
26 (ESI/Q-TOF) m/z : (M+Na) $^+$ calcd for $\text{C}_{26}\text{H}_{30}\text{N}_4\text{O}_5\text{Na}$ 501.2108, found 501.2110
27
28
29
30
31
32
33
34
35
36
37
38
39

40
41 **Compound 33 - benzyl (5R,8S)-3-(cyclopropylmethyl)-2,4-dioxo-8-((4-**
42 **(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonane-7-carboxylate** Compound
43 **33** was synthesized using General Procedure 2 and compound **28**, with HOAt (22.5 mmol), EDC
44 (11.25 mmol), 4-(trifluoromethyl)benzylamine (22.5 mmol), DIPEA (15 mmol), and 75 mL of 1:1
45 DMF/DCM. Recovered yield was quantitative and the product used without purification. ^1H NMR
46 (500 MHz, CDCl_3) 0.25 (2H, m), 0.43 (2H, m), 1.04 (1H, m), 2.26 (1H, ddd, $J = 13.0, 10.0, 7.2$),
47 2.38 (1H, m), 3.23 (2H, dd, $J = 11.0, 7.3$), 3.67 (1H, d, rotameric, $J = 11.2$), 4.33 (1H, dd, $J =$
48 12.5, 5.8), 4.40 (1H, t, $J = 6.1$), 4.50 (1H, dd, rotameric, $J = 9.9, 7.5$), 5.07 (2H, m), 7.34 (6H, m),
49 7.50 (2H, m), 7.65 (1H, d, $J = 7.9$), 8.83 (1H, m), 9.10 (1H, s, rotameric); ^{13}C NMR (125 MHz,
50
51
52
53
54
55
56
57
58
59
60

1
2
3 DMSO-*d*₆, rotamers present) 3.4, 9.9, 40.3, 41.3, 41.7, 42.5, 56.1, 56.4, 58.9, 59.4, 64.9, 65.5,
4
5 66.3, 123.2, 125.0, 127.0, 127.3, 127.5, 127.6, 127.8, 128.2, 128.3, 136.5, 144.0, 153.7, 155.6,
6
7 155.7, 170.9, 171.1, 172.3; HRMS (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₂₇H₂₇F₃N₄O₅Na
8
9 567.1826, found 567.1827
10

11
12
13 **Compound 34 - benzyl (5*R*,8*S*)-3-isobutyl-2,4-dioxo-8-((4-**
14 **(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonane-7-carboxylate** Compound

15 **34** was synthesized using General Procedure 2 and compound **29**, with HOAt (22.5 mmol), EDC
16 (11.25 mmol), 4-(trifluoromethyl)benzylamine (22.5 mmol), DIPEA (15 mmol), and 75 mL of 1:1
17 DMF/DCM. Recovered yield was quantitative and the product used without purification. ¹H NMR
18 (500 MHz, CDCl₃) 0.83 (6H, d, *J* = 6.8), 1.94 (1H, m), 2.26 (1H, ddd, *J* = 13.0, 10.1, 8.1), 2.41
19 (1H, ddd, rotameric, *J* = 20.6, 13.1, 7.3), 3.18 (2H, dd, *J* = 10.2, 7.5), 3.65 (1H, d, rotameric, *J* =
20 11.4), 3.82 (1H, d, rotameric, *J* = 11.2), 4.33 (1H, dd, *J* = 10.8, 6.0), 4.40 (1H, t, *J* = 5.6), 4.48
21 (1H, dd, rotameric, *J* = 9.9, 7.5), 5.07 (2H, m), 7.34 (6H, M), 7.51 (2H, m), 7.66 (1H, d, rotameric,
22 *J* = 8.2), 8.83 (1H, m), 9.10 (1H, s, rotameric); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers present)
23 19.8, 26.9, 40.4, 41.4, 41.7, 41.8, 45.3, 56.5, 58.9, 59.4, 64.8, 65.4, 66.3, 123.2, 123.3, 125.0,
24 125.1, 125.4, 127.0, 127.3, 127.6, 127.7, 127.9, 128.3, 128.4, 136.5, 136.6, 144.1, 144.3, 153.7,
25 153.8, 155.7, 155.8, 171.0, 171.2, 172.6; HRMS (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for
26 C₂₇H₂₉F₃N₄O₅Na 569.1982, found 569.1986
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42

43 **Compound 35 - benzyl (5*R*,8*S*)-8-(benzylcarbamoyl)-3-isobutyl-2,4-dioxo-1,3,7-**
44 **triazaspiro[4.4]nonane-7-carboxylate** - Compound **35** was synthesized using General
45 Procedure 2 and compound **29**, with 22.5 mmol HOAt, 11.25 mmol EDC, 22.5 mmol
46 benzylamine, 15 mmol DIPEA, and 75 mL of 1:1 DMF/DCM. Recovered yield was quantitative
47 and the product used without purification. ¹H NMR (500 MHz, DMSO-*d*₆) 0.82 (6H, dd, *J* = 6.7,
48 4.6), 1.94 (1H, m), 2.24 (1H, m), 2.39 (1H, m), 3.18 (2H, dd, *J* = 10.4, 7.3), 3.66 (1H, m), 3.81
49 (1H, m), 4.27 (2H, m), 4.46 (1H, ddd, *J* = 17.5, 8.9, 7.5), 5.07 (2H, m), 7.20 (10H, m), 8.72 (1H,
50 m), 9.10 (1H, s, rotameric); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers present) 19.9, 26.9, 40.5,
51
52
53
54
55
56
57
58
59
60

1
2
3 41.4, 42.0, 42.1, 45.3, 56.5, 58.9, 59.3, 64.8, 65.4, 66.3, 126.7, 127.0, 127.1, 127.6, 127.7, 127.9,
4
5 128.2, 128.3, 128.4, 136.6, 139.2, 153.6, 153.7, 155.7, 155.8, 170.7, 170.9, 172.6; HRMS
6
7 (ESI/Q-TOF) m/z: (M+Na)⁺ calcd for C₂₆H₃₀N₄O₅Na 501.2108, found 501.2113
8
9

10 11 **General Procedure 3: Deprotection of Proline Cbz group**

12
13
14
15 Each of **30-35** was added to a RB flask and reacted with 1:1 DCM/(33% HBr in AcOH) for 30 min,
16
17 and the reaction progress checked via LCMS. Upon successful deprotection of the Cbz group,
18
19 the solvent was removed *in vacuo* with the aid of toluene, and placed on a high vacuum pump
20
21 overnight to afford **36-41**. ¹H / ¹³C NMR spectra for compounds **36-41** are available in the
22
23 supporting information.
24
25

26
27 **Compound 36 - (5R,8S)-N-isobutyl-3-(naphthalen-2-ylmethyl)-2,4-dioxo-1,3,7-**
28
29 **triazaspiro[4.4]nonane-8-carboxamide** - Compound **36** was synthesized using General
30
31 Procedure 3 and compound **30**. Recovered yield was quantitative and the product used without
32
33 purification. ¹H NMR (500 MHz, DMSO-*d*₆) 08.4 (6H, d, *J* = 6.7), 1.72 (1H, m), 2.23 (1H, dd, *J* =
34
35 13.4, 11.9), 2.69 (1H, dd, *J* = 13.4, 6.7), 2.93 (1H, m), 3.02 (1H, m), 3.53 (1H, d, *J* = 12.8), 3.67
36
37 (1H, d, *J* = 12.8), 4.53 (1H, dd, *J* = 11.6, 6.7), 4.72 (2H, m), 7.41 (2H, m), 7.50 (1H, m), 7.76 (1H,
38
39 m), 7.90 (3H, m), 8.66 (1H, m), 9.01 (1H, s), 9.31 (1H, s), 9.91 (1H, s); ¹³C NMR (125 MHz,
40
41 DMSO-*d*₆, rotamers present) 20.0, 27.9, 34.6, 40.7, 42.0, 46.4, 48.6, 52.5, 55.0, 59.3, 65.7,
42
43 125.6, 125.9, 126.1, 126.4, 127.6, 127.7, 128.3, 128.7, 129.3, 132.3, 132.8, 133.8, 155.2, 165.9,
44
45 172.7; HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₂₂H₂₇N₄O₃ 395.2078, found 395.2084
46
47

48
49 **Compound 37 (5R,8S)-3-benzyl-N-cyclopropyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-8-**
50
51 **carboxamide** - Compound **37** was synthesized using General Procedure 3 and compound **31**.
52
53 Recovered yield was quantitative and the product used without purification. ¹H NMR (500 MHz,
54
55 DMSO-*d*₆) 0.47 (2H, m), 0.66 (2H, m), 2.21 (1H, dd, *J* = 13.4, 11.6), 2.56 (1H, dd, *J* = 13.3, 6.9),
56
57 2.7 (1H, tq, *J* = 7.3, 3.9) 3.49 (1H, d, *J* = 13.1), 3.62 (1H, d, *J* = 12.8), 4.42 (1H, dd, *J* = 11.6, 6.7),
58
59
60

1
2
3 4.55 (2H, s), 7.28 (5H, m), 8.73 (1H, d, $J = 4.3$), 8.96 (1H, s), 9.30 (1H, s), 9.88 (1H, s); ^{13}C NMR
4 (125 MHz, $\text{DMSO-}d_6$) 5.6, 21.1, 22.6, 40.5, 41.7, 48.6, 52.5, 59.2, 65.5, 127.3, 127.5, 128.6,
5
6 136.2, 150.1, 166.9, 172.6 HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{H})^+$ calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_3$ 329.1608,
7
8 found 329.1602
9
10

11
12
13 **Compound 38** (**5R,8S**)-3-benzyl-*N*-isobutyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-8-
14 **carboxamide** - Compound **38** was synthesized using General Procedure 3 and compound **32**.
15 Recovered yield was quantitative and the product used without purification. ^1H NMR (500 MHz,
16 $\text{DMSO-}d_6$) 0.86 (6H, d, $J = 7.6$), 1.74 (1H, m), 2.22 (1H, dd, $J = 13.4, 11.6$), 2.69 (1H, dd, $J =$
17 $13.4, 7.0$), 2.95 (1H, m), 3.03 (1H, m), 3.54 (1H, d, $J = 12.8$), 3.66 (1H, d, $J = 13.1$), 4.57 (2H, s),
18 4.58 (1H, dd, obscured, $J = 11.7, 6.7$), 7.31 (5H, m), 8.70 (1H, t, $J = 5.8$), 9.01 (1H, s), 9.29 (1H,
19 s), 9.96 (1H, s); ^{13}C NMR (125 MHz, $\text{DMSO-}d_6$) 20.0, 20.1, 27.9, 35.6, 40.7, 41.7, 46.4, 48.6,
20 52.4, 59.3, 65.6, 127.3, 127.5, 128.3, 128.6, 136.2, 155.1, 165.9, 172.7; HRMS (ESI/Q-TOF) m/z :
21 $(\text{M}+\text{H})^+$ calcd for $\text{C}_{18}\text{H}_{25}\text{N}_4\text{O}_3$ 345.1921, found 345.1920
22
23
24
25
26
27
28
29
30
31

32
33 **Compound 39** - (**5R,8S**)-3-(cyclopropylmethyl)-2,4-dioxo-*N*-(4-(trifluoromethyl)benzyl)-1,3,7-
34 **triazaspiro[4.4]nonane-8-carboxamide** - Compound **39** was synthesized using General
35 Procedure 3 and compound **33**. Recovered yield was quantitative and the product used without
36 purification. ^1H NMR (500 MHz, $\text{DMSO-}d_6$) 0.24 (2H, m), 0.43 (2H, m), 1.02 (1H, m), 2.27 (1H, dd,
37 $J = 13.4, 11.6$), 2.67 (1H, dd, $J = 13.3, 6.9$), 3.22 (2H, d, $J = 7.0$), 3.49 (1H, d, $J = 13.1$), 3.62 (1H,
38 d, $J = 13.1$), 4.46 (2H, m), 4.65 (1H, dd, $J = 11.4, 6.9$), 7.51 (2H, d, $J = 8.2$), 7.69 (2H, d, $J = 7.9$),
39 8.89 (1H, s), 9.36 (1H, t, $J = 6.0$), 9.46 (1H, s), 9.80 (1H, s); ^{13}C NMR (125 MHz, $\text{DMSO-}d_6$) 3.5,
40 9.8, 40.4, 42.1, 42.8, 48.6, 52.5, 59.4, 65.4, 125.3, 125.4, 127.7, 127.9, 128.1, 143.3, 155.4,
41 166.3, 172.7; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{H})^+$ calcd for $\text{C}_{19}\text{H}_{22}\text{F}_3\text{N}_4\text{O}_3$ 411.1639, found
42 411.1638
43
44
45
46
47
48
49
50
51
52

53
54 **Compound 40** - (**5R,8S**)-3-isobutyl-2,4-dioxo-*N*-(4-(trifluoromethyl)benzyl)-1,3,7-
55 **triazaspiro[4.4]nonane-8-carboxamide** - Compound **40** was synthesized using General
56
57
58
59
60

1
2
3 Procedure 3 and compound **34**. Recovered yield was quantitative and the product used without
4 purification. ^1H NMR (500 MHz, CDCl_3) 0.77 (6H, d, $J = 6.4$), 1.88 (1H, m), 2.49 (1H, m), 2.83,
5 1H, m), 3.16 (2H, m), 3.29 (1H, m), 3.91 (2H, m), 4.36 (1H, d, $J = 11.0$), 4.56 (1H, d, $J = 9.2$),
6 1H, m), 5.36 (1H, m), 7.35 (2H, d, $J = 7.2$), 7.47 (2H, d, $J = 7.6$), 8.31 (1H, m), 8.68 (1H, m), 9.45 (1H, m);
7 ^{13}C NMR (125 MHz, CDCl_3) 19.8, 27.3, 33.6, 43.4, 46.5, 60.2, 65.9, 122.9, 125.0, 125.5, 127.7,
8 127.9, 128.8, 129.0, 129.6, 129.8, 141.2, 156.3, 167.3, 172.2; HRMS (ESI/Q-TOF) m/z : (M+H) $^+$
9 calcd for $\text{C}_{19}\text{H}_{24}\text{F}_3\text{N}_4\text{O}_3$ 413.1795, found 413.1794
10
11
12
13
14
15
16
17
18

19 **Compound 41 (5R,8S)-N-benzyl-3-isobutyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-8-**
20 **carboxamide** - Compound **41** was synthesized using General Procedure 3 and compound **35**.
21 Recovered yield was quantitative and the product used without purification. ^1H NMR (500 MHz,
22 DMSO- d_6) 0.82 (6H, d, $J = 6.7$), 1.92 (1H, m), 2.24 (1H, dd, $J = 13.3, 11.7$), 2.65 (1H, dd, $J =$
23 13.3, 6.9), 3.17 (2H, d, $J = 7.3$), 3.49 (1H, d, $J = 13.1$), 3.61 (1H, d, $J = 12.8$), 4.36 (2H, dd, $J =$
24 5.8, 3.1), 4.60 (1H, dd, $J = 11.6, 7.0$), 7.29 (6H, m), 8.87 (1H, s), 9.21 (1H, t, $J = 5.8$); ^{13}C NMR
25 (125 MHz, DMSO- d_6) 19.8, 26.9, 34.6, 40.5, 42.5, 45.5, 52.5, 59.3, 65.3, 127.1, 127.3, 128.4,
26 128.7, 129.3, 138.3, 155.4, 166.0, 172.9; HRMS (ESI/Q-TOF) m/z : (M+H) $^+$ calcd for $\text{C}_{18}\text{H}_{25}\text{N}_4\text{O}_3$
27 345.1921, found 345.1921
28
29
30
31
32
33
34
35
36
37
38

39 **General Procedure 4: Coupling of Boc-Gly-OH to 36-41**

40 To an inert atmosphere RB flask containing 2 equiv of Boc-Gly-OH in dry DMF/DCM (1:1 ratio,
41 [200 mM]) was added EDC (2 equiv) and HOAT (4 equiv) and stirred for 1 h. Afterwards, one of
42 **36-41** in DMF [200 mM] and DIPEA (4 equiv) were added and the reaction stirred for another 2 h,
43 at which point the progress was checked via LCMS. Upon completion, the reaction was diluted
44 with four times the reaction volume of EtOAc and washed with saturated ammonium chloride
45 solution, saturated sodium bicarbonate solution, and brine. The organic layer was then dried with
46 Na_2SO_4 , and concentrated *in vacuo* to yield **42-47** as foamy dark-yellow solids. These
47 compounds were then purified by normal phase flash chromatography (0-5% MeOH in DCM) to
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 yield off-white foamy solids. ^1H / ^{13}C NMR spectra are available for compounds 42-47 in the
4
5 supporting information.
6
7

8
9 **Compound 42 - tert-butyl (2-((5R,8S)-8-(isobutylcarbamoyl)-3-(naphthalen-2-ylmethyl)-2,4-**
10 **dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** – Compound **42** was
11 synthesized using General procedure 4, Boc-Gly-OH (15 mmol), HOAt (30 mmol), EDC (15
12 mmol), DIPEA (30 mmol), and compound **36**. Purified yield 3.23 g (78%); ^1H NMR (500 MHz,
13 CDCl_3) 0.80 (6H, m), 1.21 (9H, s, rotameric), 1.65 (1H, m), 2.24 (1H, dd, $J = 12.8, 7.6$), 2.47 (2H,
14 m), 3.24 (1H, m), 3.73 (1H, dd, $J = 17.1, 4.6$), 3.95 (1H, m), 4.08 (1H, dd, $J = 10.4$), 4.43 (1H, m),
15 4.69 (1H, d, $J = 14.6$), 4.79 (1H, m), 4.82 (1H, d, $J = 14.6$), 5.77 (1H, m), 7.32 (1H, m), 7.46 (3H,
16 m), 7.76 (4H, m), 8.18 (1H, s); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 20.0, 20.1, 25.3,
17 28.1, 28.3, 28.4, 40.9, 43.0, 46.7, 54.5, 59.7, 64.3, 66.5, 79.6, 126.4, 126.5, 127.7, 127.8, 128.1,
18 128.6, 132.9, 133.0, 133.1, 155.5, 156.5, 169.6, 171.5; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd
19 for $\text{C}_{29}\text{H}_{37}\text{N}_5\text{O}_6\text{Na}$ 574.2636, found 574.2633
20
21
22
23
24
25
26
27
28
29
30
31

32
33 **Compound 43 - tert-butyl (2-((5R,8S)-3-benzyl-8-(cyclopropylcarbamoyl)-2,4-dioxo-1,3,7-**
34 **triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Compound **43** was synthesized using
35 General procedure 4, Boc-Gly-OH (15 mmol), HOAt (30 mmol), EDC (15 mmol), DIPEA (30
36 mmol), and compound **37** - Purified yield 2.94 g (81%); ^1H NMR (500 MHz, CDCl_3 , rotamers
37 present) 0.44 (1H, m), 0.52 (2H, m), 0.67 (1H, m), 1.31 (9H, s, rotameric), 2.17 (1H, m), 2.43 (1H,
38 t, $J = 11.7$), 2.55 (1H, m), 3.76 (1H, dd, $J = 17.1, 5.5$), 3.84 (1H, d, $J = 10.4$), 4.06 (1H, d, $J =$
39 10.4), 4.22 (1H, m), 4.58 (1H, d, $J = 14.6$), 4.65 (1H, d, $J = 14.6$), 4.67 (1H, m), 5.80 (1H, s), 7.30
40 (5H, m), 7.45 (1H, m), 8.21 (1H, s); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 5.9, 6.7, 22.6,
41 28.2, 40.3, 42.7, 43.2, 54.9, 59.7, 66.4, 79.8, 128.1, 128.6, 128.8, 135.7, 155.6, 156.6, 171.1,
42 171.5; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{24}\text{H}_{31}\text{N}_5\text{O}_6\text{Na}$ 508.2167, found 508.2165
43
44
45
46
47
48
49
50
51
52

53
54 **Compound 44 - tert-butyl (2-((5R,8S)-3-benzyl-8-(isobutylcarbamoyl)-2,4-dioxo-1,3,7-**
55 **triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Compound **44** was synthesized using
56
57
58
59
60

1
2
3 General procedure 4, Boc-Gly-OH (15 mmol), HOAt (30 mmol), EDC (15 mmol), DIPEA (30
4 mmol), and compound **38**. Purified yield 2.87 g (78%); ¹H NMR (500 MHz, CDCl₃) 0.85 (6H, m),
5 1.29 (9H, s, rotameric), 1.70 (1H, m), 2.22 (1H, dd, *J* = 12.5, 7.6), 2.45 (1H, t, *J* = 11.9), 2.47 (1H,
6 m), 3.27 (1H, m), 3.75 (1H, dd, *J* = 17.1, 4.6), 3.99 (1H, m), 4.08 (1H, d, *J* = 9.8), 4.49 (1H, m),
7 4.55 (1H, d, *J* = 14.6), 4.67 (1H, d, *J* = 14.6), 4.82 (1H, dd, *J* = 10.1, 8.2), 5.79 (1H, m), 7.31 (6H,
8 m), 8.17 (1H, s); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 20.1, 20.2, 25.3, 28.2, 28.3, 28.4,
9 41.0, 42.8, 43.1, 46.7, 54.4, 59.7, 60.4, 66.5, 79.6, 128.2, 128.7, 128.8, 135.7, 155.5, 156.5,
10 169.5, 171.4 HRMS (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₂₅H₃₅N₅O₆ Na 524.2480, found
11 524.2487
12
13
14
15
16
17
18
19
20
21
22
23

24
25 **Compound 45 - *tert*-butyl (2-((5*R*,8*S*)-3-benzyl-8-(isobutylcarbamoyl)-2,4-dioxo-1,3,7-**
26 **triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Compound **45** was synthesized using
27 General procedure 4, Boc-Gly-OH (15 mmol), HOAt (30 mmol), EDC (15 mmol), DIPEA (30
28 mmol), and compound **39** - Purified yield 3.64 g (86% Yield); ¹H NMR (500 MHz, CDCl₃) 0.25
29 (2H, d, *J* = 4.6), 0.44 (2H, m), 1.04 (1H, m), 1.33 (9H, s, rotameric), 2.07 (1H, m), 2.45 (1H, t, *J* =
30 11.9), 3.22 (1H, dd, *J* = 14.0, 7.3), 3.33 (1H, dd, *J* = 14.0, 7.0), 3.83 (1H, dd, *J* = 17.1, 5.2), 3.92
31 (1H, m), 4.11 (1H, m), 4.34 (2H, m), 4.62 (1H, dd, *J* = 15.4, 6.6), 4.89 (1H, m), 5.82 (1H, s), 7.42
32 (2H, d, *J* = 8.2), 7.54 (2H, d, *J* = 8.2), 8.08 (1H, s), 8.30 (1H, s); ¹³C NMR (125 MHz, CDCl₃,
33 rotamers present) 3.6, 4.1, 10.1, 21.1, 25.3, 28.2, 42.9, 43.2, 43.8, 54.8, 60.1, 60.4, 66.3, 79.9,
34 123.0, 125.2, 125.4, 125.5, 127.6, 129.4, 129.7, 142.2, 156.1, 156.6, 170.3, 171.5; HRMS
35 (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₂₆H₃₂F₃N₅O₆ Na 590.2197, found 590.2196
36
37
38
39
40
41
42
43
44
45
46
47
48

49 **Compound 46 - *tert*-butyl (2-((5*R*,8*S*)-3-isobutyl-2,4-dioxo-8-((4-**
50 **(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** -
51 Compound **42** was synthesized using General procedure 4, Boc-Gly-OH (30 mmol), HOAt (60
52 mmol), EDC (30 mmol), DIPEA (60 mmol), and compound **40** - Purified yield 6.82 g (91%); ¹H
53 NMR (500 MHz, CDCl₃) 0.82 (6H, d, rotameric, *J* = 6.7), 1.33 (9H, s, rotameric), 1.96 (1H, m),
54
55
56
57
58
59
60

1
2
3 2.02 (1H, m), 2.44 (1H, t, $J = 11.9$), 3.12 (1H, dd, $J = 13.1, 8.2$), 3.29 (1H, dd, $J = 13.4, 6.7$), 3.78
4
5 (1H, dd, $J = 17.1, 5.2$), 3.92 (1H, m), 4.07 (1H, m), 4.17 (1H, m), 4.56 (1H, dd, $J = 15.1, 7.2$), 4.86
6
7 (1H, m), 5.81 (1H, s), 7.26 (5H, m), 7.85 (1H, m), 8.36 (1H, s); ^{13}C NMR (125 MHz, CDCl_3 ,
8
9 rotamers present) 19.7, 19.8, 25.3, 27.1, 28.2, 40.7, 43.1, 43.3, 45.8, 54.5, 59.9, 66.1, 79.7,
10
11 127.3, 127.3, 128.5, 138.0, 156.0, 156.5, 169.9, 170.7, 171.7; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$
12
13 calcd for $\text{C}_{25}\text{H}_{35}\text{N}_5\text{O}_6 \text{Na}$ 524.2480, found 524.2487
14
15
16

17 **Compound 47 - tert-butyl (2-((5R,8S)-8-(benzylcarbamoyl)-3-isobutyl-2,4-dioxo-1,3,7-**
18 **triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Compound **42** was synthesized using
19
20 General procedure 4, Boc-Gly-OH (15 mmol), HOAt (30 mmol), EDC (15 mmol), DIPEA (30
21
22 mmol), and compound **41** – Purified Yield 3.50 g (82%); ^1H NMR (500 MHz, CDCl_3) 0.84 (6H, d,
23
24 rotameric, $J = 6.7$), 1.32 (9H, s), 2.00 (2H, m), 2.16 (1H, s), 2.46 (1H, t, $J = 11.9$), 3.17 (1H, dd, J
25
26 = 13.1, 7.9), 3.31 (1H, dd, $J = 13.4, 7.0$), 3.81 (1H, dd, $J = 16.9, 5.2$), 4.08 (1H, d, $J = 10.7$), 4.32
27
28 (2H, m), 4.57 (1H, dd, $J = 15.6, 6.4$), 4.88 (1H, m), 5.72 (1H, s), 7.4 (2H, d, $J = 8.2$), 7.54 (2H, d, J
29
30 = 8.2), 7.99 (1H, m), 8.30 (1H, s); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 19.7, 19.8, 27.2,
31
32 28.2, 31.0, 40.3, 42.9, 43.3, 46.0, 54.9, 60.1, 66.1, 80.0, 123.0, 125.1, 125.4, 125.5, 127.5, 129.5,
33
34 129.7, 142.2, 156.1, 156.6, 170.4, 171.5; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for
35
36 $\text{C}_{26}\text{H}_{34}\text{F}_3\text{N}_5\text{O}_6 \text{Na}$ 592.2353, found 592.2354
37
38
39
40

41 **General Procedure 5: Alkylation of 42-47 to Synthesize 2-15**

42
43 To a stirred mixture of one of **42-47** in DMF [100 mM] was added 1.05 equiv of allyl or benzyl
44
45 halide along with 1.5 equiv of K_2CO_3 . The reaction proceeded at room temperature for 14-24 h,
46
47 and the progress checked via LCMS. The reaction was diluted with four times the reaction
48
49 volume of EtOAc and washed with water, saturated ammonium chloride solution, and brine. The
50
51 organic layer was dried with Na_2SO_4 , and concentrated *in vacuo* to yield **5-18** as tan to yellow
52
53 foamy solids. UHPLC (C18, 5-100% ACN in Water) and $^1\text{H} / ^{13}\text{C}$ NMR spectra are available in the
54
55 supporting information.
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Spiroligomer Amine 5 - *tert*-butyl (2-((5*R*,8*S*)-1-benzyl-8-(benzylcarbamoyl)-3-isobutyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate – Spiroligomer Amine **5** was synthesized using General Procedure 5, compound **46**, and benzyl bromide (1.05 equiv). Purified yield 1.10 g (93%). ¹H NMR (500 MHz, CDCl₃) 0.94 (6H, d, *J* = 6.7), 1.39 (9H, s, rotameric), 2.16 (1H, m), 2.36 (1H, dd, *J* = 14.5, 9.0), 2.44 (1H, dd, *J* = 17.1, 4.3), 3.03 (1H, dd, *J* = 14.6, 8.5), 3.27 (1H, dd, *J* = 16.9, 5.3), 3.4 (1H, t, *J* = 3.5), 3.44 (2H, d, rotameric, *J* = 7.6), 3.80 (1H, d, *J* = 11.6), 4.05 (1H, d, *J* = 16.2), 4.37 (1H, d, rotameric, *J* = 5.5), 4.46 (1H, d, rotameric, *J* = 6.4), 4.76 (1H, t, *J* = 8.7), 5.00 (1H, t, *J* = 4.4), 5.04 (1H, d, *J* = 16.2), 7.07 (1H, t, *J* = 5.8), 7.18 (2H, d, *J* = 7.3), 7.25 (3H, m), 7.33 (5H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 19.9, 20.0, 21.0, 24.8, 24.9, 27.3, 28.3, 36.5, 42.7, 43.7, 44.1, 46.8, 54.6, 60.0, 63.8, 69.3, 80.0, 80.8, 120.7, 127.1, 127.4, 127.5, 128.3, 128.7, 129.1, 129.3, 137.0, 137.8, 151.3, 156.1, 168.3, 169.0, 171.5 ; HRMS (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₃₂H₄₁N₅O₆Na 614.2949, found 614.2955

Spiroligomer Amine 6 - *tert*-butyl (2-((5*R*,8*S*)-3-benzyl-1-(3,4-dichlorobenzyl)-8-(isobutylcarbamoyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate - Spiroligomer Amine **6** was synthesized using General Procedure 5, compound **44**, and 3,4-dichlorobenzyl bromide (1.05 equiv). Purified yield 1.16 g (88%); ¹H NMR (500 MHz, CDCl₃) 0.87 (6H, d, *J* = 6.7), 1.43 (9H, s, rotameric), 1.75 (1H, m), 2.24 (1H, ddd, *J* = 14.7, 9.1, 1.2), 2.96 (1H, dd, *J* = 17.1, 4.6), 3.05 (3H, m), 3.45 (1H, dd, *J* = 17.1, 5.2), 3.52 (1H, d, *J* = 11.6), 3.84 (1H, d, *J* = 11.9), 4.21 (1H, d, *J* = 16.2), 4.63 (1H, t, *J* = 8.5), 4.75 (3H, m), 5.14 (1H, t, *J* = 4.7), 6.70 (1H, t, *J* = 5.5), 7.06 (1H, d, *J* = 6.4), 7.36 (7H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 20.1, 28.3, 28.4, 36.0, 43.1, 43.2, 43.3, 47.2, 54.5, 59.7, 69.5, 80.2, 126.6, 128.3, 128.6, 128.7, 128.9, 129.1, 129.2, 130.0, 131.1, 132.6, 133.3, 135.2, 135.4, 137.2, 151.7, 155.7, 168.2, 168.6, 170.9 ; HRMS (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₃₂H₃₉Cl₂N₅O₆Na 682.2170, found 682.2161

Spiroligomer Amine 7 - *tert*-butyl (2-((5*R*,8*S*)-1-allyl-3-isobutyl-2,4-dioxo-8-((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate - Spiroligomer Amine **7** was synthesized using General Procedure 5, compound **47**, and allyl

1
2
3 bromide (1.05 equiv). Purified yield 804 mg (92%); ^1H NMR (500 MHz, CDCl_3) 0.80 (6H, d, J =
4 6.7), 1.39 (9H, s, rotameric), 2.07 (1H, m), 2.44 (1H, dd, J = 14.3, 9.5), 2.95 (1H, dd, J = 14.5,
5 7.5), 3.36 (2H, d, rotameric, J = 7.3), 3.82 (4H, m), 3.97 (1H, d, J = 1.16), 4.14 (1H, dd, rotameric,
6 J = 16.5, 4.9), 4.46 (1H, d, rotameric, J = 5.8), 4.53 (1H, d, rotameric, J = 6.1), 4.83 (1H, t, J =
7 8.2), 5.18 (2H, m), 5.32 (1H, t, J = 4.6), 5.84 (1H, dddd, J = 16.9, 10.5, 6.5, 5.2), 7.32 (1H, t, J =
8 6.0), 7.40 (2H, d, J = 8.2), 7.57 (2H, d, J = 7.9); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present)
9 19.9, 27.3, 28.2, 36.1, 43.1, 43.6, 46.6, 54.3, 60.2, 68.8, 80.5, 118.1, 125.5, 125.6, 127.7, 133.4,
10 142.1, 156.3, 156.0, 168.7, 169.5, 172.1; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for
11 $\text{C}_{29}\text{H}_{38}\text{F}_3\text{N}_5\text{O}_6\text{Na}$ 632.2666, found 632.2655
12
13
14
15
16
17
18
19
20
21
22

23 **Spiroligomer Amine 8 - *tert*-butyl (2-((5*R*,8*S*)-1-(2-amino-2-oxoethyl)-3-isobutyl-2,4-dioxo-8-**
24 **((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-**

25 **oxoethyl)carbamate** - Spiroligomer Amine **8** was synthesized using General Procedure 5,
26
27 compound **47**, and 4-(trifluoromethyl)benzyl bromide (1.05 equiv). Purified yield 770 mg (82%);
28
29 ^1H CDCl_3 (500 MHz, CDCl_3) 0.87 (6H, t, J = 6.7), 1.31 (9H, s, rotameric), 2.02 (1H, tq, J = 13.4,
30 6.6), 2.16 (1H, m), 2.47 (1H, dd, J = 13.3, 9.6), 3.32 (2H, d, J = 7.3), 3.81 (4H, m), 4.19 (1H, d, J
31 = 11.3), 4.42 (3H, m), 4.83 (1H, m), 5.88 (1H, s), 6.62 (1H, s), 7.36 (2H, d, J = 7.6), 7.51 (2H, d, J
32 = 7.9), 7.83 (1H, s, rotameric), 8.23 (1H, s, rotameric); ^{13}C NMR (125 MHz, CDCl_3 , rotamers
33 present) 3.6, 9.9, 19.8, 25.3, 27.3, 28.2, 28.3, 37.2, 42.6, 42.9, 43.7, 46.8, 54.9, 60.1, 64.4, 69.1,
34 80.7, 122.0, 122.9, 125.1, 125.4, 127.5, 127.7, 129.6, 155.8, 157.0, 170.0, 170.3, 170.9; HRMS
35 (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{28}\text{H}_{37}\text{F}_3\text{N}_6\text{O}_7\text{Na}$ 649.2568, found 649.2566
36
37
38
39
40
41
42
43
44
45
46

47 **Spiroligomer Amine 9 - *tert*-butyl (2-((5*R*,8*S*)-1-(3,4-dichlorobenzyl)-3-isobutyl-2,4-dioxo-8-**
48 **((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-**

49 **oxoethyl)carbamate** - Spiroligomer Amine **9** was synthesized using General Procedure 5,
50
51 compound **47**, and 3,4-Dichlorobenzyl bromide (1.05 equiv). Purified yield 971 mg (89%). ^1H
52 NMR (500 MHz, CDCl_3) 0.93 (6H, d, J = 7.0), 1.37 (9H, s, rotameric), 2.12 (1H, m), 2.35 (1H, dd,
53 J = 14.2, 9.6), 2.52 (1H, q, J = 7.3), 2.98 (1H, m), 3.43 (3H, m), 3.53 (1H, m), 3.90 (1H, d, J =
54
55
56
57
58
59
60

1
2
3 11.6), 4.23 (1H, d, $J = 16.5$), 4.41 (1H, d, rotameric, $J = 5.8$), 4.50 (1H, d, rotameric, $J = 6.1$), 4.74
4 (1H, t, $J = 8.4$), 4.82 (1H, d, $J = 16.2$), 5.17 (1H, t, $J = 4.9$), 7.09 (1H, dd, $J = 8.2, 1.8$), 7.34 (4H,
5 m), 7.44 (1H, m), 7.56 (1H, d, $J = 7.9$); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 19.9, 27.3,
6 28.2, 28.3, 36.3, 43.0, 43.1, 43.2, 46.8, 46.9, 54.5, 56.5, 59.9, 69.2, 80.5, 123.0, 125.6, 126.6,
7 127.6, 128.1, 129.2, 130.1, 130.6, 131.2, 135.6, 133.3, 137.4, 142.0, 156.0, 156.1, 168.5, 169.2,
8 171.5; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{33}\text{H}_{38}\text{Cl}_2\text{F}_3\text{N}_5\text{O}_6\text{Na}$ 750.2043, found
9 750.2042
10
11
12
13
14
15
16
17

18
19 **Spiroligomer Amine 10 - *tert*-butyl (2-((5*R*,8*S*)-1-(3,5-bis(trifluoromethyl)benzyl)-8-**
20 **(isobutylcarbamoyl)-3-(naphthalen-2-ylmethyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-**
21 **2-oxoethyl)carbamate** - Spiroligomer Amine **10** was synthesized using General Procedure 5,
22 compound **42**, and 3,5-bis(trifluoromethyl)benzyl bromide (1.05 equiv). Purified yield 1.88 g
23 (97%); ^1H NMR (500 MHz, CDCl_3) 0.85 (6H, d, $J = 6.7$), 1.41 (9H, s, rotameric), 1.73 (1H, dquin,
24 $J = 13.5, 6.8$), 2.09 (1H, dd, $J = 14.5, 9.0$), 3.01 (2H, m), 3.12 (1H, dd, $J = 14.8, 7.8$), 3.19 (1H,
25 dd, $J = 17.1, 5.2$), 3.42 (1H, dd, $J = 17.1, 4.9$), 3.64 (1H, d, $J = 11.6$), 3.88 (1H, d, $J = 11.9$) 4.53
26 (1H, t, $J = 8.2$), 4.61 (1H, d, $J = 16.8$), 4.78 (1H, d, $J = 16.5$), 4.93 (2H, m), 5.14 (1H, t, $J = 4.7$),
27 6.71 (1H, t, $J = 5.8$), 7.50 (3H, m), 7.82 (7H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present)
28 20.0, 20.1, 28.2, 28.3, 35.8, 43.0, 43.5, 43.6, 47.2, 54.3, 59.4, 69.5, 80.3, 122.3, 122.9, 124.0,
29 126.1, 126.4, 126.5, 127.5, 127.7, 128.0, 128.9, 132.4, 132.6, 132.7, 133.1, 133.3, 139.9, 155.8,
30 156.0, 168.3, 168.4, 170.9; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{38}\text{H}_{41}\text{F}_6\text{N}_5\text{O}_6\text{Na}$
31 800.2853, found 800.2864
32
33
34
35
36
37
38
39
40
41
42
43
44
45

46
47 **Spiroligomer Amine 11 - *tert*-butyl (2-((5*R*,8*S*)-1-allyl-8-(isobutylcarbamoyl)-3-(naphthalen-**
48 **2-ylmethyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Spiroligomer
49 Amine **11** was synthesized using General Procedure 5, compound **42**, and allyl bromide (1.05
50 equiv). Purified yield 1.41 g (96%); ^1H NMR (500 MHz, CDCl_3) 0.89 (6H, d, $J = 6.7$), 1.42 (9H, s,
51 rotameric), 1.78 (1H, m), 2.34 (1H, dd, $J = 14.5, 9.0$), 3.06 (3H, m), 3.73 (2H, m), 3.84 (2H, m),
52 3.91 (1H, d, $J = 11.6$), 4.10 (1H, dd, $J = 16.5, 5.2$), 4.72 (1H, t, $J = 8.4$), 4.84 (2H, m), 5.15 (2H,
53
54
55
56
57
58
59
60

1
2
3 m), 5.30 (1H, t, $J = 4.4$), 5.81 (1H, ddd, $J = 16.5, 11.6, 6.3$), 6.73 (1H, t, $J = 5.8$), 7.48 (3H, m),
4
5 7.82 (4H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 20.1, 20.2, 28.3, 28.4, 36.1, 43.3,
6
7 43.6, 47.2, 54.4, 60.1, 69.2, 80.2, 118.2, 126.3, 126.4, 127.7, 128.0, 128.7, 133.0, 133.2, 133.3,
8
9 154.9, 155.8, 168.3, 168.9, 171.5; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{32}\text{H}_{41}\text{N}_5\text{O}_6\text{Na}$
10
11 614.2949, found 614.2951
12

13
14
15 **Spiroligomer Amine 12 - methyl 2-((5R,8S)-7-((tert-butoxycarbonyl)glycyl)-8-**
16
17 **(isobutylcarbamoyl)-3-(naphthalen-2-ylmethyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-1-**
18
19 **yl)acetate** - Spiroligomer Amine **12** was synthesized using General Procedure 5, compound **42**,

20
21 and methyl-bromoacetate (1.05 equiv). Purified yield 1.44 g (93%); ^1H NMR (500 MHz, CDCl_3)
22
23 0.88 (6H, d, $J = 6.7$), 1.41 (9H, s), 1.78 (1H, m), 2.41 (1H, dd, $J = 14.5, 9.0$), 2.99 (1H, m), 3.07
24
25 (2H, m), 3.73 (3H, s), 3.77 (2H, m), 3.90 (2H, m), 4.09 (1H, m), 4.34 (1H, d, $J = 18.3$), 4.68 (1H, t,
26
27 $J = 8.5$), 4.86 (2H, m), 5.24 (1H, t, $J = 4.3$), 6.76 (1H, t, $J = 5.8$), 7.48 (3H, m), 7.82 (4H, m),; ^{13}C
28
29 NMR (125 MHz, CDCl_3 , rotamers present) 20.1, 20.2, 28.3, 28.4, 36.0, 41.9, 43.4, 47.1, 52.9,
30
31 54.5, 59.6, 69.4, 80.2, 126.2, 126.3, 126.4, 127.7, 127.9, 128.1, 128.8, 132.7, 133.0, 133.2,
32
33 155.2, 155.9, 168.8, 168.9, 169.4, 170.9; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for
34
35 $\text{C}_{32}\text{H}_{41}\text{N}_5\text{O}_8\text{Na}$ 646.2847, found 646.2848
36
37
38
39

40
41 **Spiroligomer Amine 13 - tert-butyl (2-((5R,8S)-3-(cyclopropylmethyl)-1-(4-methoxybenzyl)-**
42
43 **2,4-dioxo-8-((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-**
44
45 **oxoethyl)carbamate** - Spiroligomer Amine **13** was synthesized using General Procedure 5,

46
47 compound **45**, and 4-methoxybenzyl chloride (1.05 equiv). Purified yield 1.58 g (92%); ^1H NMR
48
49 (500 MHz, CDCl_3) 0.39 (2H, m), 0.55 (2H, m), 1.21 (1H, m), 1.35 (9H, s), 2.45 (1H, dd, $J = 14.5,$
50
51 9.3), 2.63 (1H, dd, $J = 16.8, 4.3$), 2.97 (1H, dd, $J = 14.6, 7.9$), 3.34 (1H, dd, $J = 16.8, 5.5$), 3.47
52
53 (3H, m), 3.78 (3H, s), 3.83 (1H, d, $J = 11.6$), 4.05 (1H, d, $J = 15.9$), 4.42 (1H, d, rotameric, $J =$
54
55 5.8), 4.51 (1H, d, rotameric, $J = 6.1$), 4.81 (1H, t, $J = 8.5$), 4.97 (1H, d, $J = 16.2$), 5.04 (1H, t, $J =$
56
57 4.7), 6.87 (2H, d, $J = 8.5$), 7.13 (2H, d, $J = 8.5$), 7.29 (1H, t, $J = 5.8$), 7.37 (2H, d, $J = 8.2$), 7.56
58
59
60

1
2
3 (2H, d, $J = 8.2$); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 3.8, 3.9, 10.1, 28.2, 28.3, 36.3,
4
5 43.0, 43.1, 43.5, 44.4, 54.6, 55.4, 60.2, 69.4, 80.2, 114.4, 125.5, 125.6, 127.6, 128.5, 128.8,
6
7 142.1, 155.8, 155.9, 159.5, 168.5, 169.5, 171.7; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for
8
9 $\text{C}_{34}\text{H}_{40}\text{F}_3\text{N}_5\text{O}_7\text{Na}$ 710.2772, found 710.2778

10
11
12
13 **Spiroligomer Amine 14 - *tert*-butyl 2-((5*R*,8*S*)-1-(3,5-bis(trifluoromethyl)benzyl)-3-**
14 **(cyclopropylmethyl)-2,4-dioxo-8-((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-**

15 **triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Spiroligomer Amine **14** was synthesized
16 using General Procedure 5, compound **45**, and 3,5-bis(trifluoromethyl)benzyl bromide (1.05
17 equiv). Purified yield 1.74 g (88%); ^1H NMR (500 MHz, CDCl_3) 0.39 (2H, m), 0.56 (2H, m), 1.22
18 (1H, m), 1.38 (9H, s, rotameric), 2.22 (1H, dd, $J = 14.3, 9.2$), 3.04 (1H, dd, $J = 14.6, 7.3$), 3.21
19 (1H, dd, $J = 16.8, 5.2$), 3.46 (1H, dd, $J = 16.9, 5.2$) 3.49 (2H, d, $J = 7.3$), 3.67 (1H, d, $J = 11.6$),
20 3.93 (1H, d, $J = 11.9$), 4.42 (1H, d, rotameric, $J = 5.8$), 4.47 (1H, d, rotameric, $J = 6.1$), 4.60 (1H,
21 d, $J = 16.8$), 4.66 (1H, t, $J = 8.2$), 4.83 (1H, d, $J = 16.8$), 5.16 (1H, t, $J = 5.0$), 7.29 (1H, t, $J = 5.5$),
22 7.35 (2H, d, rotameric, $J = 7.9$), 7.56 (2H, d, $J = 7.9$), 7.72 (2H, s, rotameric), 7.84 (1H, s); ^{13}C
23 NMR (125 MHz, CDCl_3 , rotamers present) 3.8, 10.0, 28.1, 35.9, 43.0, 43.2, 43.3, 44.6, 54.2, 59.6,
24 69.3, 80.5, 121.8, 122.3, 123.0, 124.0, 125.5, 125.6, 127.4, 127.6, 128.2, 132.4, 132.7, 140.0,
25 142.0, 156.0, 156.3, 168.6, 169.0, 171.3; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for
26 $\text{C}_{35}\text{H}_{36}\text{F}_9\text{N}_5\text{O}_6\text{Na}$ 816.2414, found 816.2412

27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43 **Spiroligomer Amine 15 methyl 2-((5*R*,8*S*)-7-((*tert*-butoxycarbonyl)glycyl)-3-**
44 **(cyclopropylmethyl)-2,4-dioxo-8-((4-(trifluoromethyl)benzyl)carbamoyl)-1,3,7-**

45 **triazaspiro[4.4]nonan-1-yl)acetate** - Spiroligomer Amine **15** was synthesized using General
46 Procedure 5, compound **45**, and methyl bromoacetate (1.05 equiv). Purified yield 1.42 g (89%);
47 ^1H NMR (500 MHz, CDCl_3) 0.35 (2H, m), 0.52 (2H, m), 1.15 (1H, m), 1.36 (9H, s, rotameric), 2.50
48 (1H, dd, $J = 14.3, 9.2$), 2.94 (1H, dd, $J = 14.6, 7.6$), 3.42 (2H, d, $J = 7.3$), 3.72 (1H, m), 3.72 (3H,
49 s), 3.83 (3H, m), 3.95 (1H, d, $J = 11.6$), 4.15 (1H, $J = 11.3$), 4.40 (1H, d, $J = 18.3$), 4.46 (1H, d,
50 rotameric, $J = 5.8$), 4.52 (1H, d, rotameric, $J = 6.1$), 4.79 (1H, t, $J = 8.5$), 5.28 (1H, t, $J = 5.0$), 7.39
51
52
53
54
55
56
57
58
59
60

1
2
3 (2H, d, $J = 7.9$), 7.57 (2H, d, $J = 8.2$); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 3.8, 3.9,
4
5 10.0, 28.2, 35.9, 41.8, 43.1, 43.6, 44.4, 52.9, 54.5, 59.7, 69.1, 80.5, 123.6, 125.2, 125.5, 125.6,
6
7 127.6, 128.2, 142.1, 155.5, 156.2, 169.2, 169.5, 169.6, 171.4; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$
8
9 calcd for $\text{C}_{29}\text{H}_{36}\text{F}_3\text{N}_5\text{O}_8\text{Na}$ 662.2408, found 662.2415
10

11
12
13 **Spiroligomer Amine 16 - *tert*-butyl (2-((5*R*,8*S*)-1-([1,1'-biphenyl]-4-ylmethyl)-3-benzyl-8-**
14 **(cyclopropylcarbamoyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate -**

15 Spiroligomer Amine **16** was synthesized using General Procedure 5, compound **43**, and 4-
16 (bromomethyl)biphenyl (1.05 equiv). Purified yield 1.51 g (93%); ^1H NMR (500 MHz, CDCl_3) 0.50
17 (2H, m), 0.71 (2H, m), 1.34 (9H, s, rotameric), 2.38 (1H, dd, $J = 14.2, 9.0$), 2.58 (1H, dd, $J = 16.8,$
18 4.3), 2.66 (1H, tq, $J = 7.2, 3.7$), 2.96 (1H, dd, $J = 14.4, 8.6$), 3.32 (1H, dd, $J = 16.8, 5.5$), 3.45 (1H,
19 d, $J = 11.6$), 3.79 (1H, d, $J = 11.3$), 4.13 (1H, d, $J = 16.2$), 4.68 (1H, t, $J = 8.5$), 4.76 (2H, m), 4.96
20 (1H, t, $J = 4.9$), 5.01 (1H, d, $J = 16.2$), 6.84 (1H, m), 7.25 (2H, d, $J = 7.9$), 7.35 (4H, m), 7.42 (4H,
21 m), 7.55 (4H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 6.3, 6.4, 22.8, 28.2, 28.3, 36.3,
22 43.0, 43.3, 43.8, 54.5, 60.0, 69.5, 80.1, 126.8, 127.0, 127.7, 128.2, 128.5, 128.6, 128.7, 128.9,
23 135.6, 135.7, 140.0, 141.3, 155.6, 155.7, 168.2, 170.4, 171.2; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$
24 calcd for $\text{C}_{37}\text{H}_{41}\text{N}_5\text{O}_6\text{Na}$ 674.2949, found 674.2935
25
26
27
28
29
30
31
32
33
34
35
36
37
38

39 **Spiroligomer Amine 17 - *tert*-butyl (2-((5*R*,8*S*)-3-benzyl-8-(cyclopropylcarbamoyl)-1-(4-**
40 **methoxybenzyl)-2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate -**

41 Spiroligomer Amine **17** was synthesized using General Procedure 5, compound **43**, and 4-
42 methoxybenzyl chloride (1.05 equiv). Purified yield 1.39 g (92%). ^1H NMR (500 MHz, CDCl_3) 0.50
43 (2H, m), 0.71 (2H, m), 1.42 (9H, s, rotameric), 2.33 (1H, dd, $J = 14.5, 9.0$), 2.61 (1H, dd, $J = 16.8,$
44 4.3), 2.67 (1H, tq, $J = 7.2, 3.7$), 2.93 (1H, dd, $J = 14.7, 8.2$), 3.31 (1H, dd, $J = 16.8, 5.2$), 3.39 (1H,
45 d, $J = 11.6$), 3.76 (3H, s) 3.77 (1H, m), 4.03 (1H, d, $J = 15.9$), 4.65 (1H, t, $J = 8.5$), 4.73 (2H, m),
46 4.91 (1H, d, $J = 16.2$), 5.08 (1H, t, $J = 4.6$), 6.84 (3H, m), 7.09 (2H, d, $J = 8.5$), 7.34 (3H, m), 7.4
47 (2H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 6.3, 6.4, 22.8, 28.3, 2=36.2, 43.0, 43.2,
48 43.5, 54.4, 55.3, 60.0, 69.5, 80.0, 114.4, 128.2, 128.5, 128.7, 128.9, 135.6, 155.6, 159.5, 168.2,
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 170.4, 171.3; HRMS (ESI/Q-TOF) m/z: (M+Na)⁺ calcd for C₃₂H₃₉N₅O₇Na 628.2742, found
4
5 628.2734
6
7

8
9 **Spiroligomer Amine 18 - tert-butyl (2-((5R,8S)-3-benzyl-8-(cyclopropylcarbamoyl)-1-methyl-**
10 **2,4-dioxo-1,3,7-triazaspiro[4.4]nonan-7-yl)-2-oxoethyl)carbamate** - Spiroligomer Amine **18**
11 was synthesized using General Procedure 5, compound **43**, and iodomethane(1.05 equiv).
12 Purified yield 1.09 g (88%). ¹H NMR (500 MHz, CDCl₃, rotamers present) 0.54 (2H, m), 0.75 (2H,
13 m), 1.43 (9H, s), 2.35 (1H, dd, *J* = 14.3, 9.2), 2.66 (1H, d, *J* = 7.3), 2.70 (1H, tq, *J* = 7.2, 3.7), 2.89
14 (3H, s), 3.76 (2H, m), 3.90 (2H, m), 1.94 (2H, s), 4.72 (1H, m), 5.40 (1H, t, *J* = 4.7), 6.83 (1H, m),
15 7.32 (5H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present) 6.3, 6.4, 22.8, 25.7, 28.3, 35.2, 42.9,
16 43.4, 53.3, 59.8, 68.7, 80.3, 128.1, 128.4, 128.6, 128.8, 135.6, 154.8, 156.0, 168.6, 170.5, 171.6;
17
18
19
20
21
22
23
24
25 HRMS (ESI/Q-TOF) m/z: (M+Na)⁺ calcd for C₂₅H₃₃N₅O₆Na 522.2323, found 522.2325
26
27
28

29 **Synthesis of the spiroligomer trimer 4.21**

30
31
32
33 **Compound 20 - 7-benzyl 8-(tert-butyl) (5R,8S)-1-benzyl-3-isobutyl-2,4-dioxo-1,3,7-**
34 **triazaspiro[4.4]nonane-7,8-dicarboxylate**

35
36 To a stirred mixture of **1** [(2S, 4R) 5.0 mmol] in DMF [100 mM] was added 2-iodo-1-methyl-
37 propane (1.1 equiv) along with K₂CO₃ (2.0 equiv). The reaction proceeded at room temperature
38 for 24 h, and the progress checked via LCMS. After completion, benzyl bromide (1.0 equiv) was
39 added and the reaction stirred for another 24 hours. The reaction was diluted with four times the
40 reaction volume of EtOAc and washed with water, saturated ammonium chloride solution, and
41 brine. The organic layer was dried with Na₂SO₄, and concentrated *in vacuo* to yield **20** (2.21 g
42 purified, 84%) as a foamy off-white solid; ¹H NMR (500 MHz, CDCl₃); 0.93 (6H, d, *J* = 6.7), 1.42
43 (9H, s, rotameric), 2.11 (1H, dq, *J* = 13.5, 7.0), 2.38 (1H, dd, *J* = 9.2, 1.5, rotameric), 2.44 (1H, t, *J*
44 = 14.4, rotameric), 3.39 (2H, t, *J* = 7.2), 3.72-3.88 (2H, m), 4.35 (1H, m), 4.67-4.86 (2H, m), 5.06
45 (2H, m), 7.15-7.37 (10H, m); ¹³C NMR (125 MHz, CDCl₃, rotamers present); 19.9, 27.8, 27.9,
46 38.2, 39.4, 44.2, 46.6, 54.3, 58.8, 59.5, 67.4, 68.3, 69.2, 82.2, 127.0, 127.2, 127.3, 128.0, 128.1,
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 128.2, 128.3, 128.4, 128.5, 128.8, 128.9, 136.0, 136.9, 153.7, 156.0, 169.7, 172.1; HRMS
4
5 (ESI/Q-TOF) m/z: (M+Na)⁺ calcd for C₃₀H₃₇N₃O₆ Na 558.2575, found 558.2574
6
7
8

9 **Compound 52 - (5R,8S)-1-benzyl-3-isobutyl-2,4-dioxo-1,3,7-triazaspiro[4.4]nonane-8-**
10 **carboxylic acid**

11
12
13 Compound **20** was reacted with 25 mL of 1:1 DCM/(33% HBr in AcOH) for 30 min, and the
14 reaction progress checked via LCMS. Upon successful deprotection of the Cbz group, the solvent
15 was removed *in vacuo* with the aid of toluene, and placed on a high vacuum pump overnight to
16 afford **52**. Recovered yield was quantitative and the product used without purification; ¹H NMR
17 (500 MHz, CDCl₃) 0.85 (6H, d, *J* = 6.7), 1.97 (1H, dquin, *J* = 13.7, 7.0), 2.11 (1H, dd, *J* = 14.0,
18 7.6), 2.21-2.25 (1H, m), 3.04 (1H, d, *J* = 11.9), 3.21 (1H, d, *J* = 11.9), 3.25 (1H, d, *J* = 7.3), 3.76
19 (1H, dd, *J* = 8.5, 7.6), 4.55-4.67 (2H, m), 7.26-7.37 (5H, m); ¹³C NMR (125 MHz, CDCl₃) 20.3,
20 27.5, 38.6, 40.5, 43.2, 54.0, 60.3, 70.3, 127.5, 127.7, 129.0, 138.4, 156.2, 173.5, 175.5; HRMS
21 (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₁₈H₂₄N₃O₄ 346.1761, found 346.1764
22
23
24
25
26
27
28
29
30
31
32

33 **Compound 21 (3S,5S)-1-((benzyloxy)carbonyl)-5-(tert-butoxycarbonyl)-3-((4-**
34 **methoxybenzyl)amino)pyrrolidine-3-carboxylic acid**

35
36
37 (3S,5S)-3-amino-1-((benzyloxy)carbonyl)-5-(tert-butoxycarbonyl)pyrrolidine-3-carboxylic acid (13
38 mmol) was dissolved in 100 mL of MeOH along with *p*-anisaldehyde (10 mmol) and stirred for 1
39 hour, after which NaCNBH₃ (14 mmol) was added and reacted for 3 hours. After completion, the
40 solvent was removed *in vacuo*, the resulting solid was redissolved in deionized (DI) H₂O, and the
41 amino acid precipitated upon neutralization with dropwise addition of 2M HCl. Precipitate was
42 collected by vacuum filtration and dried on a lyophilizer (3.72 g (7.68 mmol) recovered, 77%
43 yield); ¹H NMR (500 MHz, DMSO-*d*₆) 1.28 (9H, s, rotameric), 1.43 (1H, d, rotameric, *J* = 3.4),
44 2.09 (1H, ddd, *J* = 18.1, 12.6, 7.4), 2.65 (1H, ddd, *J* = 20.9, 12.2, 8.2), 3.46 (1H, t, *J* = 10.2), 3.71
45 (5H, m), 3.92 (1H, dd, *J* = 10.5, 8.9), 4.22 (1H, t, rotameric, *J* = 7.8), 5.06 (2H, m), 6.87 (2H, m),
46 7.32 (8H, m); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers present) 27.9, 28.1, 28.1, 28.2, 37.5, 38.3,
47 48.3, 53.9, 54.5, 55.5, 59.4, 59.7, 63.0, 66.5, 66.7, 67.0, 67.9, 80.5, 81.0, 81.2, 114.0, 127.8,
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 127.9, 128.2, 128.3, 128.4, 128.7, 128.8, 128.9, 130.4, 130.5, 137.0, 137.3, 154.0, 154.3, 159.1,
4
5 159.2, 170.9, 171.3, 173.2, 173.3; HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₂₆H₃₄N₂O₇
6
7 485.2282, found 485.2287
8
9

10
11 **Compound 22** - 1''-benzyl 5''-(tert-butyl) (3'S,4R,5''S,8a'S)-3-benzyl-1-isobutyl-2'-(4-
12 methoxybenzyl)-1',2,4',5-tetraoxotetrahydro-4'H,6'H-dispiro[imidazolidine-4,7'-pyrrolo[1,2-
13 a]pyrazine-3',3''-pyrrolidine]-1'',5''-dicarboxylate
14
15

16
17 Compound **21** (700 mg) was preactivated with HOAt [979 mg (6 eq)] and EDC [253 mg (1.1 eq)]
18 in 21 mL of anhydrous 2:1 DCM/DMF for 1.5 h, after which all of **52** and DIPEA (0.63 mL)
19 dissolved in 7 mL of DMF were added and reacted overnight with stirring. Subsequently, 575 mg
20 (2.5 eq) of EDC-HCl were added and the reaction stirred for another 4 hours. The reaction was
21 diluted with EtOAc, and washed with saturated solutions of NH₄Cl, NaHCO₃, and brine, dried over
22 Na₂SO₄, and rotovapped. The foamy solid was then purified via normal phase flash
23 chromatography to afford 478 mg (46% yield) of **22**; ¹H NMR (500 MHz, CDCl₃); 0.96 (6H, d, J =
24 6.7), 1.47 (9H, rotameric), 2.08-2.19 (2H, m), 2.53 (1H, dd, J = 14.2, 8.7), 2.55-2.59 (1H, m), 2.89
25 (1H, dd, J = 14.3, 7.6), 3.40-3.42 (1H, m), 3.43 (2H, dd, J = 7.5, 0.8), 3.78 (3H, s), 3.85 (1H, t, J =
26 10.7), 4.07 (1H, dd, J = 13.6, 2.3), 4.11-4.16 (1H, m), 4.18-4.26 (2H, m), 4.40-4.49 (1H, m), 4.65
27 (1H, d, J = 4.1, rotameric), 4.72-4.78 (1H, m), 4.89-5.00 (1H, m), 5.03-5.11 (1H, m), 5.15-5.19
28 (1H, m), 6.80-6.85 (2H, m), 7.10-7.18 (2H, m), 7.27-7.42 (10H, m) ¹³C NMR (125 MHz, CDCl₃,
29 rotamers present) 20.0, 27.4, 27.7, 28.0, 36.5, 38.9, 39.3, 43.9, 45.7, 46.6, 50.2, 50.5, 51.8, 55.2,
30 57.4, 57.6, 57.7, 57.9, 65.66, 65.73, 66.4, 67.3, 67.45, 67.53, 82.3, 114.3, 114.4, 126.9, 127.0,
31 127.6, 127.9, 128.4, 128.6, 128.9, 129.3, 135.5, 136.2, 136.3, 136.6, 153.9, 154.5, 156.4, 159.0,
32 165.2, 167.7, 169.9, 170.1, 173.3; HRMS (ESI/Q-TOF) m/z: (M+Na)⁺ calcd for C₄₄H₅₁N₅O₉Na
33 816.3579, found 816.3589
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50

51
52 **Compound 53** - (3'S,4R,5''S,8a'S)-3-benzyl-1-isobutyl-2'-(4-methoxybenzyl)-1',2,4',5-
53 tetraoxotetrahydro-4'H,6'H-dispiro[imidazolidine-4,7'-pyrrolo[1,2-a]pyrazine-3',3''-
54 pyrrolidine]-5''-carboxylic acid
55
56
57
58
59
60

1
2
3 **22** was treated with 1:1 DCM/(33% HBr/AcOH) for 30 minutes, after which the solvent was
4 removed *in vacuo*, and the molecule left on a high vacuum pump overnight to give the
5 deprotected spiriligomer dimer; ¹H NMR (500 MHz, DMSO-*d*₆) 0.88 (6H, dd, *J* = 6.7, 1.8), 1.99
6 (1H, dquin, *J* = 13.7, 7.0), 2.14 (1H, t, *J* = 12.8), 2.60 (2H, m), 2.79 (1H, dd, *J* = 13.4, 6.7), 3.27
7 (2H, d, *J* = 7.3), 3.42 (3H, m), 3.53 (1H, dd, *J* = 11.9, 6.7), 3.72 (3H, s), 3.94 (1H, d, *J* = 12.8),
8 4.43 (1H, d, *J* = 16.8), 4.62 (1H, d, *J* = 16.5), 4.81 (2H, m), 4.96 (1H, d, *J* = 16.5), 6.86 (2H, d, *J* =
9 8.8), 7.17 (2H, d, *J* = 8.5), 7.28 (1H, m), 7.39 (4H, m); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers
10 present) 20.3, 20.4, 27.5, 35.6, 38.3, 42.9, 44.8, 46.2, 48.1, 51.3, 55.5, 57.0, 59.6, 65.7, 69.5,
11 114.4, 127.4, 127.9, 129.2, 130.2, 138.4, 156.3, 158.7, 165.5, 168.3, 168.8, 174.8; HRMS
12 (ESI/Q-TOF) *m/z*: (M+Na)⁺ calcd for C₃₂H₃₇N₅O₇Na 626.2585, found 626.2587
13
14
15
16
17
18
19
20
21
22
23

24
25 **Compound 23 - (3S,5S)-1-((benzyloxy)carbonyl)-5-(tert-butoxycarbonyl)-3-**
26 **(pentylamino)pyrrolidine-3-carboxylic acid**
27

28 (3S,5S)-3-amino-1-((benzyloxy)carbonyl)-5-(tert-butoxycarbonyl)pyrrolidine-3-carboxylic acid (13
29 mmol) was dissolved in 100 mL of MeOH along with valeraldehyde (10 mmol) and stirred for 1
30 hour, after which NaCNBH₃ (14 mmol) was added and reacted for 3 hours. After completion, the
31 solvent was removed *in vacuo*, the resulting solid was redissolved in DI H₂O, and the amino acid
32 precipitated upon neutralization with dropwise addition of 2M HCl. Precipitate was collected by
33 vacuum filtration and dried on a lyophilizer, 3.56 g (8.21 mmol) recovered, 82% yield; ¹H NMR
34 (500 MHz, DMSO-*d*₆) 0.84 (4H, m), 1.29 (16H, m), 1.46 (1H, m), 2.04 (1H, ddd, rotameric, *J* =
35 17.1, 12.2, 7.9), 2.62 (2H, m), 3.44 (1H, m), 3.87 (1H, dd, *J* = 10.1, 3.7), 4.28 (1H, t, rotameric, *J*
36 = 8.3), 5.04 (2H, m), 7.33 (5H, m); ¹³C NMR (125 MHz, DMSO-*d*₆, rotamers present) 13.8, 13.9,
37 14.0, 14.3, 19.0, 21.0, 21.8, 22.1, 27.2, 27.3, 27.4, 27.6, 27.7, 27.8, 28.3, 31.5, 31.7, 36.2, 36.9,
38 44.3, 53.5, 59.0, 59.4, 66.1, 66.2, 67.0, 80.5, 80.7, 127.3, 127.4, 127.7, 127.8, 128.2, 128.3,
39 128.4, 136.5, 136.8, 153.5, 153.8, 170.5, 170.9, 171.0; HRMS (ESI/Q-TOF) *m/z*: (M+H)⁺ calcd for
40 C₂₃H₃₅N₂O₆ 435.2490, found 435.2481
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
Compound 24 - 1'''-benzyl 5'''-(tert-butyl) (3'S,3''S,4R,5'''S,8a'S,8a''S)-3-benzyl-1-isobutyl-
2'-(4-methoxybenzyl)-1',1'',2,4',4'',5-hexaoxo-2''-pentyloctahydro-4'H,4''H,6'H,6''H-
trisp[ir]o[imidazolidine-4,7'-pyrrolo[1,2-a]pyrazine-3',7''-pyrrolo[1,2-a]pyrazine-3'',3'''-
pyrrolidine]-1''',5'''-dicarboxylate

11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Compound **23** [370 mg (1.2 equiv)] was preactivated with HOAt [563 mg (6 equiv)] and EDC [143 mg (1.1 equiv)] in 15 mL of anhydrous 2:1 DCM/DMF for 1.5 h, after which **53** [420 mg (1.0 equiv)] and DIPEA [366 μ L (5 equiv)] dissolved in 5 mL of DMF were added and reacted overnight with stirring. Subsequently, EDC [330 mg (2.5 eq)] was added and the reaction stirred for another 4 hours. The reaction was diluted with EtOAc, and washed with saturated solutions of NH_4Cl , NaHCO_3 , and brine, dried over Na_2SO_4 , and rotovapped. The foamy solid was then purified via normal phase flash chromatography to afford 148 mg (21% yield) of **20**; ^1H NMR (500 MHz, CDCl_3 , rotamers present) 0.92 (9H, m), 1.29 (10H, m), 1.46 (4H, m), 1.59 (1H, m), 1.98 (1H, m), 2.13 (2H, m), 2.20 (1H, m), 2.61 (3H, m), 2.85 (1H, dd, $J = 14.3, 7.3$), 3.12 (1H, m), 3.38 (1H, d, $J = 12.8$), 3.42 (2H, d, $J = 7.6$), 3.64 (1H, m), 3.75 (3H, m), 3.82 (1H, d, $J = 11.6$), 4.05 (4H, m), 4.25 (2H, m), 4.37 (2H, m), 4.56 (1H, d, $J = 16.2$), 4.83 (1H, d, $J = 16.5$), 4.94 (1H, m), 5.06 (1H, d, $J = 12.2$), 5.19 (2H, m), 6.82 (2H, m), 7.12 (2H, m), 7.33 (10H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 13.9, 14.0, 20.0, 22.2, 22.3, 22.6, 27.4, 27.7, 27.9, 29.0, 29.3, 36.2, 38.7, 39.7, 43.4, 43.7, 45.8, 46.6, 49.5, 51.1, 51.7, 55.2, 55.9, 57.4, 57.7, 65.4, 66.5, 67.5, 67.5, 82.1, 114.5, 127.1, 127.7, 127.9, 128.35, 128.46, 128.48, 128.52, , 128.9, 129.4, 136.0, 136.5, 154.1, 156.3, 159.1, 164.7, 165.5, 166.6, 167.1, 170.5, 173.6; HRMS (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{55}\text{H}_{67}\text{N}_7\text{O}_{11}\text{Na}$ 1024.4791, found 1024.4784

46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Compound 54 - (3'S,3''S,4R,5'''S,8a'S,8a''S)-3-benzyl-1'''-((benzyloxy)carbonyl)-1-isobutyl-
2'-(4-methoxybenzyl)-1',1'',2,4',4'',5-hexaoxo-2''-pentyloctahydro-4'H,4''H,6'H,6''H-
trisp[ir]o[imidazolidine-4,7'-pyrrolo[1,2-a]pyrazine-3',7''-pyrrolo[1,2-a]pyrazine-3'',3'''-
pyrrolidine]-5'''-carboxylic acid

Spiroligomer trimer **24** was treated with a 95:4:1 TFA/ H_2O /TIPS mixture to remove the *t*-Bu protecting group and afford compound **54**; ^1H NMR (500 MHz, CDCl_3) 0.85 (3H, m), 0.93 (6H, d,

1
2
3 $J = 6.6$), 1.26 (6H, m), 1.56 (1H, m), 2.12 (1H, tt, $J = 13.7, 6.9$), 2.22 (2H, m), 2.55 (1H, m), 2.65
4 (2H, m), 2.80 (1H, dd, $J = 13.2, 6.6$), 3.06 (1H, t, $J = 11.2$), 3.40 (3H, m), 3.63 (1H, m), 3.74 (3H,
5 m), 3.85 (1H, m), 4.02 (3H, m), 4.18 (1H, m), 4.26 (1H, m), 4.37 (3H, m), 4.59 (1H, m), 4.70 (1H,
6 m), 4.91 (1H, m), 5.14 (2H, m), 6.80 (2H, m), 7.09 (2H, d, $J = 8.2$), 7.32 (10 H, m); ^{13}C NMR (125
7 MHz, CDCl_3 , rotamers present) 10.2, 11.7, 12.2, 13.9, 14.1, 17.3, 17.6, 19.4, 19.94, 19.96, 22.17,
8 22.22, 27.4, 28.9, 29.3, 36.1, 38.0, 38.2, 43.6, 45.9, 46.6, 49.5, 51.5, 55.2, 56.0, 57.3, 57.6, 65.5,
9 66.6, 67.4, 68.1, 114.4, 127.1, 127.2, 127.4, 127.6, 127.67, 127.72, 128.2, 128.3, 128.4, 128.6,
10 128.9, 129.3, 135.8, 136.6, 155.6, 156.4, 159.1, 164.5, 165.6, 167.3, 167.6, 173.8; HRMS
11 (ESI/Q-TOF) m/z : $(\text{M}+\text{Na})^+$ calcd for $\text{C}_{51}\text{H}_{59}\text{N}_7\text{O}_{11}\text{Na}$ 968.4165, found 968.4133
12
13
14
15
16
17
18
19
20
21
22

23 **Compound 25 - benzyl (3'S,3''S,4R,5'''S,8a'S,8a''S)-3-benzyl-5''-(2-((tert-**
24 **butoxycarbonyl)amino)ethyl)carbamoyl)-1-isobutyl-2'-(4-methoxybenzyl)-1',1'',2,4',4'',5-**
25 **hexaexo-2''-pentyloctahydro-4'H,4''H,6'H,6''H-trispiro[imidazolidine-4,7'-pyrrolo[1,2-**
26 **a]pyrazine-3',7''-pyrrolo[1,2-a]pyrazine-3'',3'''-pyrrolidine]-1'''-carboxylate**
27
28
29
30

31 All of **54** was preactivated with HOAt [82 mg (4 eq)] and EDC [400 mg (2 eq)] in 6.0 mL of
32 anhydrous 2:1 DCM/DMF for 1.5 h, after which N-Boc-Ethylenediamine [143 μL (5 eq)] was
33 added and reacted overnight with stirring. The reaction was diluted with EtOAc, and washed with
34 saturated solutions of NH_4Cl , NaHCO_3 , and brine, dried over Na_2SO_4 , and rotovapped. The foamy
35 solid was then purified via normal phase flash chromatography to afford 127 mg (46% yield) of
36 **25**; ^1H NMR (500 MHz, CDCl_3) 0.87 (3H, t, $J = 6.8$), 0.92 (6H, d, $J = 6.6$), 1.30 (6H, m), 1.40 (9H,
37 s), 1.57 (1H, m), 2.11 (1H, dt, $J = 13.7, 7.0$), 2.18 (1H, dd, $J = 12.9, 11.3$), 2.40 (1H, dd, $J = 11.7,$
38 7.3), 2.57 (3H, m), 2.82 (1H, dd, $J = 14.0, 7.7$), 3.12 (5H, m), 3.37 (4H, m), 3.68 (5H, m), 4.01
39 (3H, m), 4.10 (1H, m), 4.23 (2H, d, $J = 16.4$), 4.35 (2H, m), 4.56 (1H, d, $J = 16.1$), 4.77 (1H, d, $J =$
40 16.4), 4.93 (1H, m), 5.17 (2H, m), 6.79 (2H, t, $J = 9.5$), 7.08 (2H, d, $J = 8.2$), 7.18 (1H, s, broad),
41 7.32 (10H, m); ^{13}C NMR (125 MHz, CDCl_3 , rotamers present) 14.0, 20.0, 22.3, 25.3, 28.4, 28.9,
42 19.4, 36.3, 38.6, 40.2, 43.4, 43.8, 45.8, 46.6, 49.4, 50.9, 51.8, 55.2, 55.9, 57.5, 58.9, 65.5, 66.6,
43 67.4, 67.9, 79.3, 114.4, 127.1, 127.2, 127.7, 127.9, 128.2, 128.49, 128.54, 128.9, 129.4, 136.0,
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 136.6, 156.0, 156.3, 159.1, 165.1, 165.3, 166.7, 167.3, 170.7, 173.6; HRMS (ESI/Q-TOF) m/z:
4
5 (M+H)⁺ calcd for C₅₈H₇₄N₉O₁₂ 1088.5451, found 1088.5495
6
7

8 9 **Peptoid Synthesis**

10 11 12 **Peptoid SPH-1**

13
14 54 mg of Rink amide (0.63 mmol / g) resin was swelled with DCM, and then treated with 20%
15
16 Piperidine in DMF 2x15 min, and subsequently rinsed with DCM and DMF. 800 uL of 1.3 M
17
18 Bromoacetic acid in DMF was reacted with 200 uL of DIC for 5 minutes, added to the resin, and
19
20 allowed to react for 30 min. The resin was drained and washed repeatedly with DCM and DMF,
21
22 after which 150 mM **7** (3 equiv, previously deprotected) in DMF was added to the resin and
23
24 reacted overnight with stirring. The bromoacetic acid addition and subsequent amine additions
25
26 were repeated for amines **14**, **13**, **11**, and **16**. After the final amine addition, the resin was rinsed
27
28 with DMF and DCM, and the resin treated with neat TFA to cleave the peptoid from the resin.

29
30 **SPH-1** was purified by reverse phase flash chromatography, recovered 59 mg (55% yield) HRMS
31
32 (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₁₅₂H₁₆₁F₁₅N₂₆O₂₆ 3049.1676; Found: 3049.1543
33
34

35 36 37 **Peptoid SPH-2**

38
39 60 mg of Rink amide (0.63 mmol / g) resin was swelled with DCM, and then treated with 20%
40
41 Piperidine in DMF 2x15 min, and subsequently rinsed with DCM and DMF. 800 uL of 1.3 M
42
43 Bromoacetic acid in DMF was reacted with 200 uL of DIC for 5 minutes, added to the resin, and
44
45 allowed to react for 30 min. The resin was drained and washed repeatedly with DCM and DMF,
46
47 after which N-(Z)-Ethylenediamine-HCl (10 equiv) in DMF (freebased with DIPEA) was reacted
48
49 with the resin for 1 h. The resin was drained and washed repeatedly with DCM and DMF,
50
51 followed by standard peptoid submonomer synthesis with 150 mM **11** (3 equiv, previously
52
53 deprotected) in DMF as the amine, and stirred overnight. The resin was subsequently rinsed with
54
55 DMF and DCM, and then the next round of peptoid coupling proceeded as the first, except with
56
57 the use amine **12** (3 equiv, previously deprotected). Another round of peptoid coupling follows,
58
59
60

1
2
3 with the use of N-(Z)-Ethylenediamine-HCl (10 equiv) as the amine for 1 h. The bromoacetic acid
4 addition and subsequent amine additions were repeated for amines **13**, **14**, **16**, and **17**, with the
5 N-(Z)-Ethylenediamine-HCl amine residue after every two spiroligomer amine additions. After the
6 final amine addition, the resin was rinsed with DMF and DCM, and the resin treated with neat
7 TFA to cleave the peptoid from the resin. **SPH-2** was purified by reverse phase flash
8 chromatography, recovered 102 mg (59% yield) HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for
9 C₂₂₀H₂₃₅F₁₂N₃₇O₄₃ 4310.7148; Found: 4310.6912
10
11
12
13
14
15
16
17
18

19 **Peptoid SPH-3**

20 60 mg of Rink amide resin (0.63 mmol / g) was swelled with DCM, and then treated with 20%
21 Piperidine in DMF 2x15 min, and subsequently rinsed with DCM and DMF. 800 uL of 1.3 M
22 Bromoacetic acid in DMF was reacted with 200 uL of DIC for 5 minutes, added to the resin, and
23 allowed to react for 30 min. The resin was drained and washed repeatedly with DCM and DMF,
24 after which N-(Z)-Ethylenediamine-HCl (10 equiv) in DMF (freebased with DIPEA) was reacted
25 with the resin for 1 h. The resin was drained and washed repeatedly with DCM and DMF,
26 followed by standard peptoid submonomer synthesis with 150 mM **11** (3 equiv, previously
27 deprotected) in DMF was added to the resin and reacted overnight with stirring. The resin was
28 subsequently rinsed with DMF and DCM, and then the next round of peptoid coupling proceeded
29 as the first, except with the use of N-(Z)-Ethylenediamine-HCl (10 equiv) as the amine for 1 h (10
30 equiv of DIPEA was used to freebase the amine prior to addition). The bromoacetic acid addition
31 and subsequent amine additions were repeated for amines **12**, **13**, **14**, **16**, and **17**, with the N-(Z)-
32 Ethylenediamine-HCl amine residue interspersed after each spiroligomer amine. After the final
33 amine addition, the resin was rinsed with DMF and DCM, and the resin treated with neat TFA to
34 cleave the peptoid from the resin. **SPH-3** was purified by reverse phase flash chromatography,
35 recovered 113 mg (57% yield) HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₂₅₆H₂₇₇F₁₂N₄₃O₅₂
36 Target Mass: 5013.0161; Found 5012.9707
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55

56 **Peptoid SPH-4**

57
58
59
60

1
2
3 100 mg of Tentagel-NH₂ (S) resin (0.37 mmol / g) was rinsed multiple times with DCM and DMF.
4
5 3 equiv of Fmoc-Met-OH and HATU in 0.6 mL of NMP with 6 equiv of DIPEA were then added to
6
7 the resin and stirred for 1 hour (Standard SPPS). The resin was treated with 20% piperidine in
8
9 DMF 2x15 min, and rinsed with DMF and DCM. 800 uL of 1.3 M Bromoacetic acid in DMF was
10
11 reacted with 200 uL of DIC for 5 minutes, added to the resin, and allowed to react for 30 min.
12
13 The resin was drained and washed repeatedly with DCM and DMF, after which 0.3 mL of a 1M
14
15 solution of propargylamine was added to the resin and stirred for 1 h (standard peptoid coupling).
16
17 Fmoc-Lys(Boc)-OH was coupled to the resin using standard SPPS, Fmoc-deprotected, and
18
19 followed by an N-substituted, boc protected ornithine derivative (N-Orn-Boc) with standard
20
21 peptoid coupling. The resin was drained and washed repeatedly with DCM and DMF, after which
22
23 150 mM **12** (3 equiv, previously deprotected) in DMF was added to the resin and reacted
24
25 overnight with stirring. This was repeated for addition of amine **14**. Standard peptoid coupling was
26
27 used to add another (N-Orn-Boc), and finally two more round of spirilogomer peptoid reactions
28
29 using amines **16** and **18**. The resin was washed repeatedly with DMF and DCM, then treated with
30
31 1:1 TFA/DCM to remove the Boc protecting groups. The resin was washed exhaustively with
32
33 water, then treated with a 7:3 Formic Acid/H₂O mixture containing 30 mg of cyanogen bromide to
34
35 cleave the peptoid from the resin. **SPH-4** was purified by reverse phase flash chromatography, 19
36
37 mg recovered (22% yield). HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₁₄₂H₁₆₃F₉N₂₈O₂₈
38
39 2879.2048; Found: 2879.1845
40
41
42

43 **Peptoid SPH-5**

44
45 15 mg of Rink amide resin (0.63 mmol / g) was swelled with DCM, and then treated with 20%
46
47 Piperidine in DMF 2x15 min, and subsequently rinsed with DCM and DMF. 400 uL of 1.3 M
48
49 Bromoacetic acid in DMF was reacted with 100 uL of DIC for 5 minutes, added to the resin, and
50
51 allowed to react for 30 min. The resin was drained and washed repeatedly with DCM and DMF,
52
53 after which 150 mM **25** (3 equiv, previously deprotected) in DMF was added to the resin and
54
55 stirred overnight. The resin was drained into a small vial containing 1.05 equiv of Boc₂O (relative
56
57 to the mmol of amine) to reprotect the spirilogomer trimer. This peptoid coupling was repeated
58
59
60

with amine **25** two more times, the resin washed exhaustively with DMF and DCM, then treated with neat TFA, followed by 1:1 TFA/DCM, and finally DCM to give 7.5 mg of purified **SPH-5** (25% yield). HRMS (ESI/Q-TOF) m/z: (M+H)⁺ calcd for C₁₆₅H₁₉₈N₂₈O₃₃ 3099.4676; Found: 3099.4803

Acknowledgements

The project or effort depicted is sponsored by the Department of the Defense, Defense Threat Reduction Agency (HDTRA1-16-1-0047). The content of the information does not necessarily reflect the position or the policy of the federal government, and no official endorsement should be inferred.

Supporting Information

Supporting information containing ¹H NMR, ¹³C NMR, and HPLC data is available free of charge at pubs.acs.org.

References

- (1) Verdine, G. L.; Hilinski, G. J. *Stapled Peptides for Intracellular Drug Targets*, 1st ed.; Elsevier Inc., 2012; Vol. 503, pp 3–33.
- (2) Walensky, L. D.; Bird, G. H. *Journal of Medicinal Chemistry* **2014**, *57* (15), 6275–6288.
- (3) Cheng, R. P.; Gellman, S. H.; DeGrado, W. F. *Chem. Rev.* **2001**, *101* (10), 3219–3232.
- (4) Guo, L.; Chi, Y.; Almeida, A. M.; Guzei, I. A.; Parker, B. K.; Gellman, S. H. *J. Am. Chem. Soc.* **2009**, *131* (44), 16018–16020.
- (5) Angelo, N. G.; Arora, P. S. *J. Am. Chem. Soc.* **2005**, *127* (49), 17134–17135.
- (6) Angelo, N. G.; Arora, P. S. *J. Org. Chem.* **2007**, *72* (21), 7963–7967.
- (7) Kang, C. W.; Sarnowski, M. P.; Elbatrawi, Y. M.; Del Valle, J. R. *J. Org. Chem.* **2017**, *82* (3), 1833–1841.
- (8) Sarnowski, M. P.; Kang, C. W.; Elbatrawi, Y. M.; Wojtas, L.; Del Valle, J. R. *Angew. Chem.* **2017**, *129* (8), 2115–2118.
- (9) Levins, C. G.; Schafmeister, C. E. *J. Am. Chem. Soc.* **2003**, *125* (16), 4702–4703.
- (10) Cheong, J. E.; Pfeiffer, C. T.; Northrup, J. D.; Parker, M. F. L.; Schafmeister, C. E. *Tetrahedron Lett.* **2016**, *57* (44), 4882–4884.
- (11) Zuckermann, R. N.; Kerr, J. M.; Kent, S. *J. Am. Chem. Soc.* **1992**, *114* (26), 10646–10647.
- (12) Kirshenbaum, K.; Barron, A. E.; Goldsmith, R. A.; Armand, P.; Bradley, E. K.; Truong, K. T.; Dill, K. A.; Cohen, F. E.; Zuckermann, R. N. *Proc. Natl. Acad. Sci.* **1998**, *95* (8), 4303–4308.
- (13) Lee, B.-C.; Zuckermann, R.; Dill, K. *J. Am. Chem. Soc.* **2005**, *127* (31), 10999–11009.
- (14) Lee, B.-C.; Chu, T. K.; Dill, K. A.; Zuckermann, R. N. *J. Am. Chem. Soc.* **2008**, *130* (27), 8847–8855.
- (15) Stringer, J. R.; Crapster, J. A.; Guzei, I. A.; Blackwell, H. E. *J. Am. Chem. Soc.* **2011**, *133* (39), 15559–15567.
- (16) Crapster, J. A.; Guzei, I. A.; Blackwell, H. E. *Angew. Chem. Int. Ed.* **2013**, *52* (19), 5079–5084.
- (17) Chongsiriwatana, N. P.; Patch, J. A.; Czyzewski, A. M.; Dohm, M. T.; Ivankin, A.; Gidalevitz, D.; Zuckermann, R. N.; Barron, A. E. *Proc. Natl. Acad. Sci.* **2008**, *105* (8), 2794–2799.

- 1
2
3 (18) Huang, M. L.; Shin, S. B. Y.; Benson, M. A.; Torres, V. J.; Kirshenbaum, K. *ChemMedChem* **2011**, *7* (1), 114–122.
- 4
5 (19) Huang, M. L.; Benson, M. A.; Shin, S. B. Y.; Torres, V. J.; Kirshenbaum, K. *Eur. J. Org. Chem.* **2013**, *2013* (17), 3560–3566.
- 6
7 (20) Zuckermann, R.; Kodadek, T. *Curr. Opin. Mol. Ther* **2009**, *11* (3), 299–307.
- 8 (21) Sun, J.; Jiang, X.; Lund, R.; Downing, K. H.; Balsara, N. P.; Zuckermann, R. N. *Proc. Natl. Acad. Sci.* **2016**, *113* (15), 3954–3959.
- 9
10 (22) Vollrath, S. B. L.; Hu, C.; Bräse, S.; Kirshenbaum, K. *Chem. Commun.* **2013**, *49* (23), 2317–2319.
- 11 (23) Robertson, E. J.; Battigelli, A.; Proulx, C.; Mannige, R. V.; Haxton, T. K.; Yun, L.; Whitelam, S.; Zuckermann, R. N. *Accounts Chem. Res.* **2016**, *49* (3), 379–389.
- 12 (24) Mannige, R. V.; Haxton, T. K.; Proulx, C.; Robertson, E. J.; Battigelli, A.; Butterfoss, G. L.; Zuckermann, R. N.; Whitelam, S. *Nature* **2015**, *526* (7573), 415–420.
- 13 (25) Nalband, D. M.; Warner, B. P.; Zahler, N. H.; Kirshenbaum, K. *Biopolymers* **2014**, *102* (5), 407–415.
- 14 (26) Zuckermann, R. N. *Biopolymers* **2010**, *96* (5), 545–555.
- 15 (27) Sun, J.; Zuckermann, R. *ACS nano* **2013**, *7* (6), 4715–4732.
- 16 (28) Rosales, A. M.; Segalman, R. A.; Zuckermann, R. N. *Soft Matter* **2013**, *9* (35), 8400–8414.
- 17 (29) Bradley, E. K.; Kerr, J. M.; Richter, L. S.; Figliozzi, G. M.; Goff, D. A.; Zuckermann, R. N.; Spellmeyer, D. C.; Blaney, J. M. *Mol. Divers.* **1997**, *3* (1), 1–15.
- 18 (30) Rosales, A. M.; Murnen, H. K.; Zuckermann, R. N.; Segalman, R. A. *Macromolecules* **2010**, *43* (13), 5627–5636.
- 19 (31) Rosales, A. M.; Murnen, H. K.; Kline, S. R.; Zuckermann, R. N.; Segalman, R. A. *Soft Matter* **2012**, *8* (13), 3673–3678.
- 20 (32) Shin, S. B. Y.; Yoo, B.; Todaro, L. J.; Kirshenbaum, K. *J. Am. Chem. Soc.* **2007**, *129* (11), 3218–3225.
- 21 (33) Culf, A. S.; Čuperlović-Culf, M.; Léger, D. A.; Decken, A. *Org. Lett.* **2014**, *16* (10), 2780–2783.
- 22 (34) Simpson, L. S.; Kodadek, T. *Tetrahedron Lett.* **2012**, *53* (18), 2341–2344.
- 23 (35) Hjelmgaard, T.; Faure, S.; Caumes, C.; De Santis, E.; Edwards, A. A.; Taillefumier, C. *Org. Lett.* **2009**, *11* (18), 4100–4103.
- 24 (36) Lee, J. H.; Kim, H.-S.; Lim, H.-S. *Org. Lett.* **2011**, *13* (19), 5012–5015.
- 25 (37) Kaniraj, P. J.; Maayan, G. *Org. Lett.* **2015**, *17* (9), 2110–2113.
- 26 (38) Lee, J. H.; Meyer, A. M.; Lim, H.-S. *Chem. Commun. (Camb.)* **2010**, *46* (45), 8615–8617.
- 27 (39) Liu, T.; Qian, Z.; Xiao, Q.; Pei, D. *ACS Combinatorial Science* **2011**, *13* (5), 537–546.
- 28 (40) Oh, M.; Lee, J. H.; Moon, H.; Hyun, Y.-J.; Lim, H.-S. *Angew. Chem.* **2015**, *128* (2), 612–616.
- 29 (41) Huang, M. L.; Ehre, D.; Jiang, Q.; Hu, C.; Kirshenbaum, K.; Ward, M. D. *Proc. Natl. Acad. Sci.* **2012**, *109* (49), 19922–19927.
- 30 (42) Levins, C. G.; Schafmeister, C. E. *J. Org. Chem.* **2005**, *70* (22), 9002–9008.
- 31 (43) Pornsuwan, S.; Bird, G.; Schafmeister, C. E.; Saxena, S. *J. Am. Chem. Soc.* **2006**, *128* (12), 3876–3877.
- 32 (44) Brown, Z. Z.; Alleva, J.; Schafmeister, C. E. *Peptide Science* **2011**, *96* (5), 578–585
- 33 (45) Northrup, J. D.; Purcell, C. R.; Schafmeister, C. E. *J. Org. Chem.* **2017**, *82* (6), 3223–3231.
- 34 (46) Kheirabadi, M.; Çelebi-Ölçüm, N.; Parker, M. F. L.; Zhao, Q.; Kiss, G.; Houk, K. N.; Schafmeister, C. E. *J. Am. Chem. Soc.* **2012**, *134* (44), 18345–18353.
- 35 (47) Zhao, Q.; Lam, Y.-H.; Kheirabadi, M.; Xu, C.; Houk, K. N.; Schafmeister, C. E. *J. Org. Chem.* **2012**, *77* (10), 4784–4792.
- 36 (48) Brown, Z. Z.; Akula, K.; Arzumanyan, A.; Alleva, J.; Jackson, M.; Bichenkov, E.; Sheffield, J. B.; Fietelson, M. A.; Schafmeister, C. E. *PLoS ONE* **2012**, *7* (10), e45948.
- 37 (49) Vaddypally, S.; Xu, C.; Zhao, S.; Fan, Y.; Schafmeister, C. E.; Zdilla, M. J. *Inorg. Chem.* **2013**, *52* (11), 6457–6463.
- 38 (50) Saha, U. K.; Roy, R. *Tetrahedron Lett.* **1995**, *36* (21), 3635–3638.
- 39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 (51) Seo, J.; Michaelian, N.; Owens, S. C.; Dashner, S. T.; Wong, A. J.; Barron, A. E.;
4 Carrasco, M. R. *Org. Lett.* **2009**, *11* (22), 5210–5213.
5 (52) Szekely, T.; Roy, O.; Faure, S.; Taillefumier, C. *Eur. J. Org. Chem.* **2014**, *2014* (26),
6 5641–5657.
7 (53) Amblard, F.; Cho, J. H.; Schinazi, R. F. *Chem. Rev.* **2009**, *109* (9), 4207–4220.
8 (54) Roy, O.; Faure, S.; Thery, V.; Didierjean, C.; Taillefumier, C. *Org. Lett.* **2008**, *10* (5),
9 921–924.
10 (55) Holub, J. M.; Garabedian, M. J.; Kirshenbaum, K. *QSAR Comb. Sci.* **2007**, *26* (11-12),
11 1175–1180.
12 (56) Jang, H.; Fafarman, A.; Holub, J. M.; Kirshenbaum, K. *Org. Lett.* **2005**, *7* (10), 1951–
13 1954.
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60