

Concise Synthesis of a Hexasaccharide Related to the Adhesin Receptor of *Streptococcus oralis* ATCC 55229

Samir Ghosh and Anup Kumar Misra

Division of Molecular Medicine, Bose Institute, A.J.C. Bose Centenary Campus, P-1/12, C.I.T. Scheme VII-M, Kolkata 700054, India

A concise synthesis of a hexasaccharide related to the adhesin receptor of *Streptococcus oralis* ATCC 55229 (previously characterized as *Streptococcus sanguis* H1) has been achieved in excellent yield. A general glycosylation condition has been used throughout the synthetic scheme. All glycosylation steps and protecting group functionalization steps are high yielding and suitable for scale-up preparation.

Keywords Hexasaccharide; *Streptococcus oralis*; Glycosylation; Adhesin

INTRODUCTION

Most microbial infections initiate with protein-carbohydrate interactions, which play a key role in the effective adhesion of bacteria, viruses, or protozoa to the host.^[1] Microbial cell-wall glycolipids,^[2] glycoproteins,^[3] and capsular polysaccharides^[4] are known to contain receptors for carbohydrate-binding proteins playing crucial roles at the initial stage of microbial adhesion to the host. Dental plaque formation is a common phenomenon in the human oral environment due to the aggression of a diverse range of microbes.^[5] It has been well established that specific interactions between bacterial adhesins and carbohydrates play significant roles in the initiation and maturation of dental plaque formation.^[6] In the past, a number of studies have been carried out

Received May 15, 2009; accepted August 15, 2009.

Address correspondence to Anup Kumar Misra, Division of Molecular Medicine, Bose Institute, A.J.C. Bose Centenary Campus, P-1/12, C.I.T. Scheme VII-M, Kolkata 700054, India. E-mail: akmisra69@rediffmail.com

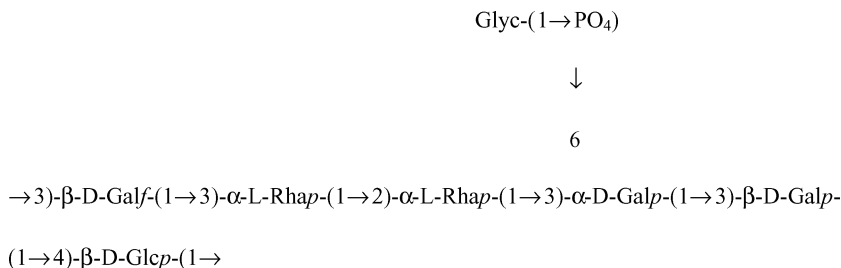


Figure 1: Structure of the adhesin receptor of *Streptococcus oralis* ATCC 55229.

for the identification and biochemical characterization of specific molecular mediators of interbacterial aggregation responsible for plaque formations.^[7] The frequently found bacterial species acting as primary colonizers in dental plaque are *Streptococcus oralis*, *Actinomyces viscosus*, and *Actinomyces naeslundii*.^[8] To date, a few adhesin receptor polysaccharides have been identified and characterized to study their role in dental plaque formation. A number of carbohydrate receptor molecules having significant antigenicity have been isolated from *Streptococcus mitis* 522, *S. oralis* ATCC 10557, *S. oralis* 34, and *S. oralis* C104 strains, which are found in the matured dental plaque.^[9] The complete structure of an antigenic hexasaccharide repeating unit of the adhesin receptor polysaccharide from *S. oralis* ATCC 55229 (previously characterized as *Streptococcus sanguis* H1) has been reported by Glushka et al. (Fig. 1).^[10]

The development of glycoconjugate vaccines against microbial infections has been of considerable interest for a long time, which has been reflected in several reports in the recent past.^[11] Although oligosaccharides can be isolated from natural sources, their limited availability cannot always meet the required quantity for their extensive biological studies. Efficient chemical synthetic strategies could offer access to larger quantities of natural oligosaccharides and its several analogs. In order to determine the relation between structure and immunological specificity, it is quite logical to prepare oligosaccharides related to the repeating unit of the antigenic polysaccharide. The synthesized oligosaccharide could also be utilized as a molecular probe for studying the immunochemical behavior of the antigen. In this report, we describe a concise chemical synthesis of a hexasaccharide related to the repeating unit of the adhesin receptor polysaccharide of *S. oralis* ATCC 55229 as its

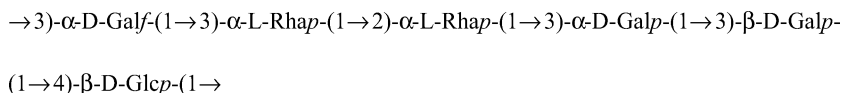


Figure 2: Structure of the synthesized hexasaccharide related to the adhesin receptor of *Streptococcus oralis* ATCC 55229 in which the D-Galp unit is linked through an α -linkage.



Figure 3. Chemical structure of the synthesized hexasaccharide as 4-Methoxyphenyl glycoside.

4-methoxyphenyl glycosides (Figs. 2 and 3). A 1,2-*cis* linked D-galactofuranose moiety is present in the nonreducing terminus in the synthesized hexasaccharide, whereas in the natural glycan repeating unit the D-galactofuranose moiety is 1,2-*trans* linked. The 4-methoxyphenyl group could serve as a temporary anomeric protecting group, which could be removed easily for the preparation of glycoconjugate molecule.

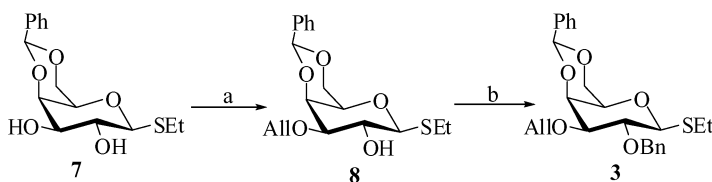
RESULTS AND DISCUSSION

The synthesis of the hexasaccharide was achieved by sequential glycosylation of suitably protected monosaccharide derivatives prepared from commercially available reducing sugars using literature-reported reaction conditions (Fig. 4).

4-Methoxyphenyl (4-*O*-acetyl-2,6-di-*O*-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside (**2**)^[12] was prepared from D-lactose following an earlier reported procedure. Ethyl 4,6-*O*-benzylidene-1-



Figure 4: Suitably protected mono- and disaccharide intermediates used for the synthesis of target hexasaccharide **1**.



Scheme 1: Reagents: (a) (i) Bu_2SnO , CH_3OH , 80°C , 3 h; (ii) allyl bromide, CsF , DMF , rt, 12 h; (b) benzyl bromide, NaOH , TBAB , THF , rt, 4 h, overall 92%.

thio- β -D-galactopyranoside (**7**),^[13] derived from D-galactose, was selectively 3-*O*-allylated via stannylidene acetal formation to give compound **8**, which was benzylated using benzyl bromide and sodium hydroxide to furnish compound **3** in 92% overall yield (Sch. 1). Starting from L-rhamnose, compounds **4**^[14] and **5**^[15] were prepared using similar reaction conditions reported earlier. Compound **6**^[16] was prepared by mercury (II) chloride-catalyzed cyclization of acyclic D-galactose diethyldithioacetal.

Stereoselective glycosylation of disaccharide acceptor **2** with thioglycoside derivative **3** in the presence of *N*-iodosuccinimide (NIS) and trimethylsilyl trifluoromethanesulfonate (TMSOTf)^[17] furnished trisaccharide derivative **9** in 89% yield. Formation of compound **9** was confirmed from its spectral analysis [signals at δ 5.15 (d, $J = 3.2$ Hz, H-1_C), 4.75 (d, $J = 8.7$ Hz, H-1_A), and 4.47 (d, $J = 7.8$ Hz, H-1_B) in ^1H and δ 102.9 (C-1_A), 102.4 (C-1_B), 100.8 (PhCH), and 93.6 (C-1_C) in the ^{13}C NMR spectra]. 1,2-*Cis* glycosylation of the D-galactose derivative was supported by the coupling constant ($J = 3.2$ Hz) of the H-1 of the D-galactose residue in compound **9**. Removal of allyl group using palladium chloride in sodium acetate buffer medium^[18] afforded trisaccharide acceptor **10** in 77% yield. Stereoselective glycosylation of compound **10** with thioglycoside derivative **4** using NIS-TMSOTf^[17] furnished tetrasaccharide derivative **11** in 92% yield, which was deacetylated using sodium methoxide to give tetrasaccharide acceptor **12** in 98% yield. Presence of signals at δ 102.8 (C-1_A), 102.4 (C-1_B), 100.6 (PhCH), 100.2 (C-1_D), and 93.5 (C-1_C) in the ^{13}C NMR spectra confirmed the formation of compound **11**. Exclusive formation of 1,2-*trans* glycoside was achieved due to the presence of the neighboring participating *O*-acetyl group in the C-2 position of the L-rhamnosyl thioglycoside derivative. NIS-TMSOTf-mediated^[17] stereoselective glycosylation of compound **12** with thioglycoside derivative **5** furnished pentasaccharide derivative **13** in 83% yield. Presence of signals at δ 102.8 (C-1_B), 102.7 (C-1_A), 101.4 (C-1_E), 100.7 (PhCH), 99.1 (C-1_D), and 95.1 (C-1_C) in the ^{13}C NMR spectra supported the formation of compound **13**. In this case also, exclusive formation of 1,2-*trans* glycoside was achieved due to the presence of the neighboring participating *O*-acetyl group in the C-2 position of the L-rhamnosyl thioglycoside derivative. Oxidative removal of 4-methoxybenzyl group using 2,3-dichloro-5,6-dicyanobenzoquinone (DDQ)^[19] in a biphasic reaction condition resulted in the formation of

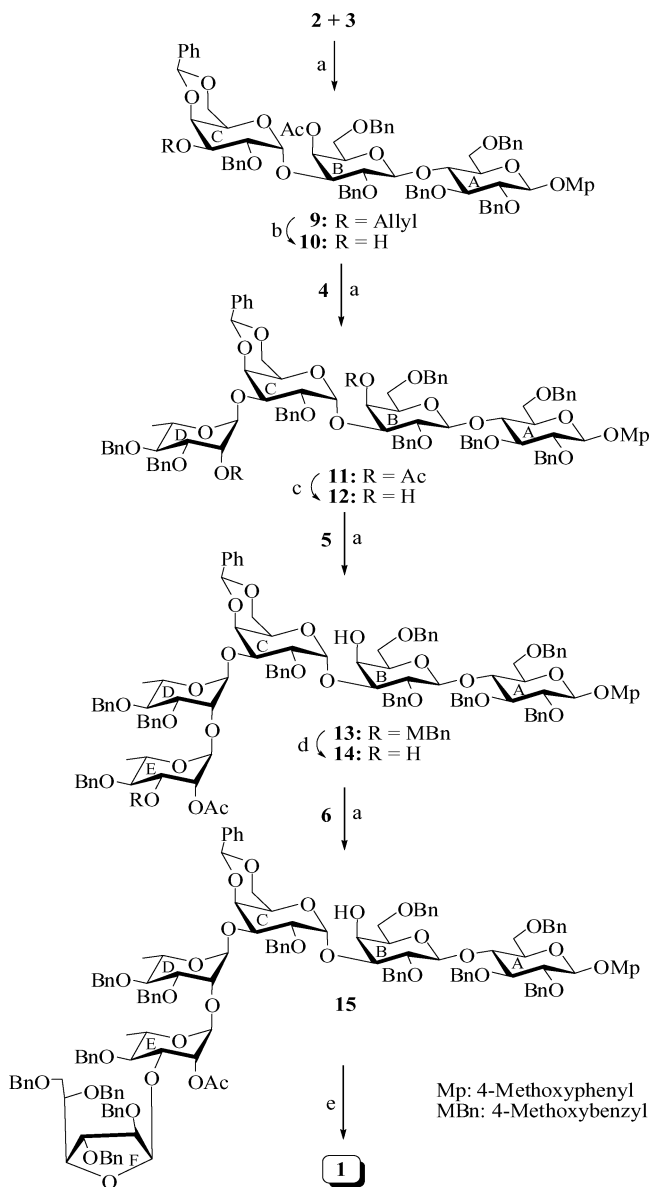
pentasaccharide acceptor **14** in 73% yield. Stereoselective glycosylation of compound **14** with thioglycoside derivative **6** using NIS-TMSOTf^[17] furnished hexasaccharide derivative **15** in 78% yield, which was confirmed from its spectral analysis. Presence of signals at δ 108.4 (C-1_F), 102.7 (C-1_A), 102.4 (C-1_B), 101.3 (2 C, C-1_D, C-1_E), 100.5 (PhCH), and 93.8 (C-1_C) in the ¹³C NMR spectra supported the formation of compound **15**. Although in the ¹H NMR spectrum of compound **15** H-1_F appeared in a multiplet, the presence of a signal at δ 108.4 in the ¹³C NMR spectrum unambiguously confirmed the formation of 1,2-*cis* glycosylation of D-galactofuranose moiety.^[16] Hydrogenolysis of the hexasaccharide derivative **15** over Pearlman's catalyst^[20] followed by deacetylation furnished pure hexasaccharide **1** in overall 76% yield. Formation of compound **1** was confirmed from its 1D and 2D NMR and mass spectral analysis. Presence of signals at δ 5.16 (br s, 2 H, H-1_E, H-1_F), 5.10 (br s, H-1_C), 4.97 (d, J = 7.8 Hz, H-1_A), 4.92 (br s, H-1_D), and 4.47 (d, J = 7.6 Hz, H-1_B) in the ¹H NMR and at δ 109.1 (C-1_F), 102.8 (C-1_B), 101.8 (C-1_E), 101.1 (C-1_A), 100.5 (C-1_D), and 95.5 (C-1_C) in the ¹³C NMR spectra unambiguously confirmed the formation of target hexasaccharide **1** (Sch. 2).

In summary, synthesis of a hexasaccharide related to the hexasaccharide adhesin receptor of *S. oralis* ATCC 55229 as its 4-methoxyphenyl glycoside has been achieved in a concise manner using a sequential stereoselective glycosylation strategy. Most of the glycosylation steps are highly stereoselective and reproducible for scale-up preparation. The 4-methoxyphenyl group has been chosen as the temporary protecting group at the reducing end for its easy removal whenever required for the preparation of glycoconjugates.

EXPERIMENTAL

General Procedure

All reactions were monitored by thin layer chromatography over silica gel-coated TLC plates. The spots on TLC were visualized by warming ceric sulphate (2% Ce(SO₄)₂ in 2N H₂SO₄)-sprayed plates in a hot plate. Silica gel 230–400 mesh was used for column chromatography. ¹H and ¹³C NMR, 2D COSY, and HSQC spectra were recorded on a Bruker Advance DPX 300 MHz using CDCl₃ and D₂O as solvents and TMS as internal reference unless stated otherwise. Chemical shift value is expressed in δ ppm. ESI-MS data were recorded on a MICROMASS QUATRO II triple quadrupole mass spectrometer. Elementary analysis was carried out on a Carlo ERBA-1108 analyzer. Optical rotations were measured at 25°C on a Rudolf Autopol III polarimeter. Commercially available grades of organic solvents of adequate purity are used in many reactions.



Scheme 2: Reagents: (a) *N*-iodosuccinimide, trimethylsilyl trifluoromethane sulfonate, MS 4Å, CH₂Cl₂, −40°C, 1 h, 89% for **9**, 92% for **11**, 83% for **13**, 78% for **15**; (b) PdCl₂, NaOAc, AcOH–H₂O, rt, 12 h, 77%; (c) CH₃ONa, CH₃OH, rt, 3 h, 98%; (d) DDQ, CH₂Cl₂, H₂O, rt, 2 h, 73%; (e) (i) CH₃ONa, CH₃OH, rt, 3 h; (ii) H₂, 20% Pd(OH)₂-C, rt, 24 h, 76%.

Ethyl 3-O-allyl-2-O-benzyl-4,6-O-benzylidene-1-thio-β-D-galactopyranoside (**3**)

To a solution of compound **7** (5 g, 16 mmol) in anhydrous CH₃OH (120 mL) was added dibutyltin oxide (4.8 g, 19.3 mmol), and the reaction mixture

was allowed to stir at 80°C for 3 h. The solvents were removed under reduced pressure and the crude mass was dissolved in anhydrous DMF (50 mL). To the reaction mixture were added cesium fluoride (2.5 g, 16.45 mmol) and allyl bromide (2.5 mL, 28.9 mmol) and the reaction mixture was allowed to stir at rt for 12 h. The solvents were removed under reduced pressure and the crude residue was dissolved in CH₂Cl₂ (150 mL). The organic layer was washed with 1 N HCl and water in succession, dried (Na₂SO₄), and concentrated. To a solution of the crude product in anhydrous THF (70 mL) were added powdered NaOH (1.8 g, 45 mmol), benzyl bromide (3.8 mL, 32 mmol), and tetrabutylammonium bromide (200 mg, 0.62 mmol) and the reaction mixture was allowed to stir briskly at rt for 4 h. The reaction was quenched by addition of satd. NH₄Cl and concentrated under reduced pressure. The crude mass was dissolved in CH₂Cl₂ (150 mL) and the organic layer was washed with water, dried (Na₂SO₄), and concentrated. The crude product was purified over SiO₂ using hexane-EtOAc (7:1) as eluant to give pure compound **3** (6.5 g, 92%). Colorless oil; IR (neat): 3448, 2986, 1752, 1375, 1235, 1090, 1057, 919, 719 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 7.52–7.24 (m, 10 H, Ar-H), 6.0–5.88 (m, 1 H, CH=CH₂), 5.50 (s, 1 H, PhCH), 5.32–5.15 (m, 2 H, CH=CH₂), 4.87–4.70 (2 d, *J*=10.2 Hz, 2 H, PhCH₂), 4.42 (d, *J*=9.6 Hz, 1 H, H-1), 4.33 (d, *J*=12.3 Hz, 1 H, H-6_a), 4.22–4.18 (m, 3 H, H-4, OCH₂-CH=CH₂), 4.0 (d, *J*=12.3 Hz, H-6_b), 3.81 (t, *J*=9.3 Hz, 1 H, H-2), 3.50 (dd, *J*=9.2, 3.4 Hz, 1 H, H-3), 3.37 (br s, 1 H, H-5), 2.86–2.70 (m, 2 H, SCH₂CH₃), 1.33 (t, *J*=7.4 Hz, 3 H, SCH₂CH₃); ¹³C NMR (75 MHz, CDCl₃): δ 138.4–137.9 (Ar-C), 134.9 (CH=CH₂), 129.0–126.5 (Ar-C), 117.4 (CH=CH₂), 101.5 (PhCH), 84.4 (C-1), 80.9 (C-5), 76.8 (C-3), 75.6 (PhCH₂), 74.2 (OCH₂CH=CH₂), 71.2 (C-4), 69.8 (C-2), 69.4 (C-6), 23.8 (SCH₂CH₃), 15.0 (SCH₂CH₃); ESI-MS: 465.2 [M+Na]⁺; Anal. Calcd. for C₂₅H₃₀O₅S (442.18): C, 67.85; H, 6.83; found: C, 67.69; H, 7.00.

4-Methoxyphenyl (3-O-allyl-2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(4-O-acetyl-2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (9**)**

To a solution of compound **2** (4 g, 4.25 mmol) and compound **3** (2.8 g, 6.33 mmol) in anhydrous CH₂Cl₂ (50 mL) was added MS-4 (5 g) and the reaction mixture was allowed to stir at rt for 1 h under argon. The reaction mixture was cooled to -40°C and *N*-iodosuccinimide (NIS; 1.7 g, 7.55 mmol) and TM-SOTf (25 μ L) were added to it. After stirring at same temperature for 1 h, the reaction mixture was quenched with Et₃N (0.2 mL), filtered through a Celite bed, and washed with CH₂Cl₂ (100 mL). The organic layer was washed with aq. Na₂S₂O₃ and water in succession, dried (Na₂SO₄), and concentrated under reduced pressure to give the crude product, which was purified over SiO₂ using

hexane-EtOAc (5:1) as eluant to furnish pure **9** (5 g, 89%). Colorless solid; m.p. 110–112°C; IR (KBr): 3018, 2925, 2361, 1742, 1505, 1453, 1364, 1217, 1099, 1062, 758, 668 cm⁻¹; $[\alpha]_D^{25} +140$ (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.36–7.06 (m, 35 H, Ar-H), 6.90 (d, $J=9.0$ Hz, 2 H, Ar-H), 6.66 (d, $J=9.0$ Hz, 2 H, Ar-H), 5.94–5.81 (m, 1 H, CH=CH₂), 5.42 (d, $J=2.9$ Hz, 1 H, H-4_B), 5.20 (s, 1 H, PhCH), 5.15 (d, $J=3.2$ Hz, 1 H, H-1_C), 5.26–5.06 (m, 2 H, CH=CH₂), 4.94–4.83 (m, 3 H, PhCH₂), 4.75 (d, $J=8.7$ Hz, 1 H, H-1_A), 4.70–4.49 (m, 6 H, PhCH₂), 4.47 (d, $J=7.8$ Hz, 1 H, H-1_B), 4.39–4.32 (m, 2 H, PhCH₂), 4.21 (d, $J=12.0$ Hz, 1 H, PhCH₂), 4.10–4.06 (m, 2 H, OCH₂-CH=CH₂), 4.01–3.93 (m, 2 H, H-3_A, H-6_{aC}), 3.86 (dd, $J=9.2, 3.4$ Hz, 1 H, H-2_C), 3.77–3.69 (m, 4 H, H-4_C, H-5_C, H-6_{abA}), 3.68–3.62 (m, 2 H, H-3_B, H-3_C), 3.66 (s, 3 H, OCH₃), 3.55–3.48 (m, 3 H, H-2_A, H-2_B, H-5_A), 3.45–3.36 (m, 3 H, H-4_A, H-5_B, H-6_{bC}), 3.29–3.20 (m, 2 H, H-6_{abB}), 1.58 (s, 3 H, COCH₃); ¹³C NMR (75 MHz, CDCl₃): δ 169.7 (COCH₃), 155.3–137.9 (Ar-C), 135.4 (CH=CH₂), 128.7–118.5 (Ar-C), 116.6 (CH=CH₂), 114.5 (Ar-C), 102.9 (C-1_A), 102.4 (C-1_B), 100.8 (PhCH), 93.6 (C-1_C), 82.7 (C-5_A), 81.4 (C-2_B), 79.3 (C-2_A), 76.1 (C-3_A), 75.5 (2 C, C-4_A, C-5_C), 75.4 (2 C, 2 PhCH₂), 75.3 (C-3_C), 75.2 (PhCH₂), 74.5 (C-4_C), 74.1 (PhCH₂), 73.6 (2 C, 2 PhCH₂), 73.1 (C-2_C), 72.3 (C-5_B), 70.9 (OCH₂CH=CH₂), 69.2 (C-6_C), 68.4 (C-6_A), 67.7 (C-6_B), 64.8 (C-4_B), 62.2 (C-3_B), 55.5 (OCH₃), 20.3 (COCH₃); ESI-MS: 1343.5 [M+Na]⁺; Anal. Calcd. for C₇₉H₈₄O₁₈ (1320.57): C, 71.80; H, 6.41; found: C, 71.64; H, 6.58.

4-Methoxyphenyl (2-O-benzyl-4,6-O-benzylidene- α D-galactopyranosyl)-(1 \rightarrow 3)-(4-O-acetyl-2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (10**)**

To a solution of compound **9** (4 g, 3.03 mmol) in AcOH-H₂O (60 mL; 20:1 v/v) were added NaOAc·3H₂O (1.6 g, 16 mmol) and PdCl₂ (380 mg, 2.14 mmol) and the reaction mixture was allowed to stir at rt for 12 h. The solvents were removed under reduced pressure and the crude product was purified over SiO₂ using hexane-EtOAc (4:1) as eluant to give pure **10** (3 g, 77%). Colorless oil; IR (neat): 3423, 3020, 2359, 1741, 1593, 1504, 1426, 1368, 1216, 1058, 761, 670 cm⁻¹; $[\alpha]_D^{25} +118$ (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.41–7.14 (m, 35 H, Ar-H), 6.97 (d, $J=9.0$ Hz, 2 H, Ar-H), 6.75 (d, $J=9.0$ Hz, 2 H, Ar-H), 5.50 (d, $J=3.0$ Hz, 1 H, H-4_B), 5.27 (d, $J=3.1$ Hz, 1 H, H-1_C), 5.23 (s, 1 H, PhCH), 5.01–4.89 (m, 3 H, PhCH₂), 4.83–4.80 (m, 2 H, H-1_A, PhCH₂), 4.77–4.70 (m, 2 H, PhCH₂), 4.65–4.51 (m, 4 H, H-1_B, PhCH₂), 4.48–4.41 (m, 2 H, PhCH₂), 4.30 (d, $J=12.0$ Hz, 1 H, PhCH₂), 4.10–3.92 (m, 3 H, H-3_A, H-6_{aA}, H-6_{aC}), 3.85–3.83 (m, 2 H, H-4_C, H-6_{bA}), 3.78–3.68 (m, 3 H, H-2_C, H-3_B, H-3_C), 3.73 (s, 3 H, OCH₃), 3.62–3.57 (m, 4 H, H-2_A, H-2_B, H-5_A, H-5_C), 3.51–3.41 (m, 3 H, H-4_A, H-5_B, H-6_{bC}), 3.34–3.28 (m, 2 H, H-6_{abB}), 1.72 (s, 3 H, COCH₃); ¹³C NMR

(75 MHz, CDCl_3): δ 169.9 (COCH_3), 155.2–114.4 (Ar-C), 102.8 (C-1_A), 102.4 (C-1_B), 100.8 (PhCH), 93.1 (C-1_C), 82.7 (C-5_A), 81.5 (C-2_B), 79.3 (C-2_A), 76.5 (C-3_A), 76.1 (C-4_A), 75.9 (C-5_C), 75.6 (C-4_C), 75.5 (PhCH₂), 75.3 (PhCH₂), 75.1 (PhCH₂), 73.6 (3 C, 3 PhCH₂), 73.2 (C-2_C), 72.3 (C-5_B), 69.0 (C-6_C), 68.5 (C-3_C), 68.4 (C-6_A), 67.6 (C-6_B), 64.9 (C-4_B), 62.0 (C-3_B), 55.5 (OCH_3), 20.4 (COCH_3); ESI-MS: 1303.5 $[\text{M}+\text{Na}]^+$; Anal. Calcd. for $\text{C}_{76}\text{H}_{80}\text{O}_{18}$ (1280.53): C, 71.23; H, 6.29; found: C, 71.05; H, 6.50.

4-Methoxyphenyl (2-O-acetyl-3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(4-O-acetyl-2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (11)

To a solution of compound **10** (2.8 g, 2.18 mmol) and compound **4** (1.2 g, 2.79 mmol) in anhydrous CH_2Cl_2 (30 mL) was added MS-4 (3 g) and the reaction mixture was allowed to stir at rt for 1 h under argon. The reaction mixture was cooled to -40°C and NIS (750 mg, 3.33 mmol) and TMSOTf (20 μL) were added to it. After stirring at same temperature for 1 h, the reaction mixture was quenched with Et_3N (0.2 mL) and filtered through a Celite bed and washed with CH_2Cl_2 (100 mL). The organic layer was washed with aq. $\text{Na}_2\text{S}_2\text{O}_3$ and water in succession, dried (Na_2SO_4), and concentrated under reduced pressure to give the crude product, which was purified over SiO_2 using hexane-EtOAc (4:1) as eluant to furnish pure **11** (3.3 g, 92%). Colorless solid; IR (neat): 3461, 2924, 2856, 2362, 1744, 1605, 1505, 1454, 1368, 1232, 1063, 739, 697 cm^{-1} ; $[\alpha]_{\text{D}}^{25} +212$ (c 1.2, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 7.42–7.16 (m, 45 H, Ar-H), 6.97 (d, J = 9.0 Hz, 2 H, Ar-H), 6.76 (d, J = 9.0 Hz, 2 H, Ar-H), 5.47 (d, J = 2.9 Hz, 1 H, H-4_B), 5.44–5.43 (m, 1 H, H-2_D), 5.24 (s, 1 H, PhCH), 5.22 (d, J = 3.1 Hz, 1 H, H-1_C), 5.06 (br s, 1 H, H-1_D), 5.02–4.90 (m, 4 H, PhCH₂), 4.86–4.80 (m, 2 H, H-1_A, PhCH₂), 4.78–4.58 (m, 5 H, PhCH₂), 4.56–4.45 (m, 5 H, H-1_B, PhCH₂), 4.40–4.25 (m, 2 H, PhCH₂), 4.10–3.96 (m, 4 H, H-3_A, H-5_D, H-6_{AA}, H-6_{AC}), 3.91–3.80 (m, 5 H, H-2_C, H-3_D, H-4_C, H-6_{BA}, H-6_{BC}), 3.77–3.70 (m, 2 H, H-3_B, H-3_C), 3.74 (s, 3 H, OCH_3), 3.65–3.57 (m, 3 H, H-2_A, H-2_B, H-5_A), 3.54–3.46 (m, 2 H, H-4_A, H-5_C), 3.39 (t, J = 9.3 Hz each, 1 H, H-4_D), 3.35–3.24 (m, 3 H, H-5_B, H-6_{AB}), 2.15, 1.59 (2 s, 6 H, 2 COCH_3), 1.25 (d, J = 6.1 Hz, 3 H, CCH_3); ^{13}C NMR (75 MHz, CDCl_3): δ 169.9, 169.8 (2 COCH_3), 155.3–114.5 (Ar-C), 102.8 (C-1_A), 102.4 (C-1_B), 100.6 (PhCH), 100.2 (C-1_D), 93.5 (C-1_C), 82.6 (C-5_A), 81.4 (C-2_B), 79.8 (C-2_A), 79.2 (C-4_D), 77.7 (C-3_C), 77.4 (C-3_D), 76.3 (C-4_A), 75.9 (C-5_C), 75.5 (PhCH₂), 75.4 (C-4_C), 75.2 (PhCH₂), 75.0 (PhCH₂), 74.7 (PhCH₂), 74.6 (C-2_C), 74.2 (PhCH₂), 73.5 (2 C, 2 PhCH₂), 73.1 (C-5_D), 72.3 (C-5_B), 71.6 (PhCH₂), 69.1 (C-2_D), 68.9 (C-6_C), 68.4 (C-6_A), 68.0 (C-3_A), 67.6 (C-6_B), 64.6 (C-4_B), 61.8 (C-3_B), 55.4 (OCH_3), 21.0, 20.3 (2 COCH_3), 18.1 (CCH_3);

ESI-MS: 1671.7 $[M+Na]^+$; Anal. Calcd. for $C_{98}H_{104}O_{23}$ (1648.70): C, 71.34; H, 6.35; found: C, 71.18; H, 6.50.

4-Methoxyphenyl (3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (12**)**

A solution of compound **11** (3 g, 1.82 mmol) in 0.1 M CH_3ONa in CH_3OH (60 mL) was allowed to stir at rt for 4 h and neutralized with Dowex-50W X8 (H^+) resin. The reaction mixture was filtered and concentrated under reduced pressure to give the crude product, which was purified over SiO_2 using hexane-EtOAc (2:1) as eluant to give pure **12** (2.8 g, 98%). White solid, m.p. 96–98°C; IR (KBr): 3486, 3032, 2924, 2858, 2363, 1507, 1454, 1365, 1225, 1065, 828, 748, 698 cm^{-1} ; $[\alpha]_D^{25} +68$ (c 1.2, $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$): δ 7.44–7.21 (m, 45 H, Ar-H), 6.98 (d, $J = 9.0$ Hz, 2 H, Ar-H), 6.74 (d, $J = 9.0$ Hz, 2 H, Ar-H), 5.29 (s, 1 H, PhCH), 5.05 (br s, 1 H, H-1_D), 5.02–4.88 (m, 3 H, PhCH₂), 4.84 (d, $J = 8.7$ Hz, 1 H, H-1_A), 4.80–4.76 (m, 3 H, H-1_C, PhCH₂), 4.73–4.57 (m, 6 H, PhCH₂), 4.51–4.36 (m, 6 H, H-1_B, PhCH₂), 4.07–4.02 (m, 2 H, H-3_A, H-5_D), 3.98–3.96 (m, 1 H, H-2_D), 3.94–3.90 (m, 3 H, H-4_B, H-6_{abA}), 3.88–3.79 (m, 5 H, H-2_C, H-3_D, H-4_C, H-6_{abC}), 3.73 (s, 3 H, OCH₃), 3.71–3.68 (m, 1 H, H-3_C), 3.63–3.57 (m, 3 H, H-2_A, H-2_B, H-5_A), 3.52–3.45 (m, 4 H, H-3_B, H-4_A, H-4_D, H-5_C), 3.42–3.32 (m, H-5_B, H-6_{abB}), 1.25 (d, $J = 6.3$ Hz, 3 H, CCH₃); ^{13}C NMR (75 MHz, $CDCl_3$): δ 155.3–114.5 (Ar-C), 102.8 (C-1_B), 102.6 (C-1_A), 101.9 (C-1_D), 100.7 (PhCH), 95.1 (C-1_C), 82.8 (C-5_A), 81.5 (C-2_B), 79.9 (C-4_D), 79.7 (C-2_A), 78.8 (C-3_C), 78.1 (C-3_D), 77.9 (C-4_A), 76.4 (2 C, C-4_C, C-5_C), 75.6 (PhCH₂), 75.5 (C-2_C), 75.3 (PhCH₂), 75.1 (PhCH₂), 74.8 (PhCH₂), 74.5 (PhCH₂), 74.3 (C-5_D), 73.5 (PhCH₂), 73.4 (PhCH₂), 73.0 (C-5_B), 72.2 (PhCH₂), 69.0 (C-6_C), 68.9 (C-2_D), 68.5 (C-6_A), 68.2 (C-6_B), 67.8 (C-3_A), 64.6 (C-4_B), 62.5 (C-3_B), 55.4 (OCH₃), 18.0 (CCH₃); ESI-MS: 1587.7 $[M+Na]^+$; Anal. Calcd. for $C_{94}H_{100}O_{21}$ (1564.68): C, 72.10; H, 6.44; found: C, 71.93; H, 6.65.

4-Methoxyphenyl [2-O-acetyl-4-O-benzyl-3-O-(4-methoxybenzyl)- α -L-rhamnopyranosyl]-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (13**)**

To a solution of compound **12** (2.5 g, 1.6 mmol) and compound **5** (900 mg, 1.95 mmol) in anhydrous CH_2Cl_2 (30 mL) was added MS-4 (3 g) and the reaction mixture was allowed to stir at rt for 1 h under argon. The reaction mixture was cooled to $-40^\circ C$ and NIS (500 mg, 2.22 mmol) and TMSOTf (10 μL) were

added to it. After stirring at same temperature for 1 h, the reaction mixture was quenched with Et₃N (0.1 mL) and filtered through a Celite bed and washed with CH₂Cl₂ (80 mL). The organic layer was washed with aq. Na₂S₂O₃ and water in succession, dried (Na₂SO₄), and concentrated under reduced pressure to give the crude product, which was purified over SiO₂ using hexane-EtOAc (3:1) as eluant to furnish pure **13** (2.6 g, 83%). Colorless oil; IR (neat): 3453, 3020, 2360, 1596, 1424, 1216, 1050, 762, 670 cm⁻¹; [α]_D²⁵ +191 (*c* 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.41–7.20 (m, 52 H, Ar-H), 6.98 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.81–6.73 (m, 4 H, Ar-H), 5.51–5.47 (m, 1 H, H-2_E), 5.27 (s, 1 H, PhCH), 5.04 (d, *J* = 12.1 Hz, 1 H, PhCH₂), 4.99–4.95 (m, 3 H, H-1_D, H-1_E, PhCH₂), 4.90–4.79 (m, 5 H, H-1_A, PhCH₂), 4.73–4.66 (m, 3 H, H-1_C, PhCH₂), 4.64–4.55 (m, 5 H, PhCH₂), 4.53–4.31 (m, 8 H, H-1_B, PhCH₂), 4.06–3.95 (m, 1 H, H-3_A), 3.98–3.87 (m, 5 H, H-2_D, H-5_D, H-5_E, H-6_{abA}), 3.86–3.76 (m, 6 H, H-2_C, H-3_E, H-4_B, H-4_C, H-6_{abC}), 3.75, 3.73 (2 s, 6 H, 2 OCH₃), 3.71–3.68 (m, 1 H, H-3_C), 3.66–3.54 (m, 4 H, H-2_A, H-2_B, H-3_D, H-5_A), 3.52–3.44 (m, 3 H, H-4_A, H-4_D, H-4_E), 3.41–3.36 (m, 3 H, H-3_B, H-5_B, H-5_C), 3.33–3.27 (m, 2 H, H-6_{abB}), 2.13 (s, 3 H, COCH₃), 1.28–1.24 (m, 6 H, 2 CCH₃); ¹³C NMR (75 MHz, CDCl₃): δ 169.8 (COCH₃), 159.3–113.8 (Ar-C), 102.8 (C-1_B), 102.7 (C-1_A), 101.4 (C-1_E), 100.7 (PhCH), 99.1 (C-1_D), 95.1 (C-1_C), 82.9 (C-5_A), 81.5 (C-2_B), 80.0 (C-4_D), 79.9 (C-4_E), 79.6 (C-2_A), 78.9 (C-3_E), 77.9 (C-3_C), 77.3 (2 C, C-3_D, C-4_A), 76.4 (2 C, C-4_C, C-5_C), 75.7 (PhCH₂), 75.6 (PhCH₂), 75.5 (C-2_C), 75.4 (PhCH₂), 75.1 (PhCH₂), 74.9 (PhCH₂), 74.7 (C-5_E), 74.6 (PhCH₂), 74.5 (C-5_D), 73.6 (PhCH₂), 73.5 (PhCH₂), 73.1 (C-5_B), 72.3 (PhCH₂), 71.4 (PhCH₂), 69.0 (C-6_C), 68.9 (C-2_D), 68.6 (C-6_A), 68.5 (C-2_E), 68.4 (C-3_A), 68.3 (C-6_B), 64.5 (C-4_B), 62.5 (C-3_B), 55.5 (OCH₃), 21.1 (COCH₃), 18.2 (CCH₃), 18.1 (CCH₃); ESI-MS: 1985.8 [M+Na]⁺; Anal. Calcd. for C₁₁₇H₁₂₆O₂₇ (1962.85): C, 71.54; H, 6.47; found: C, 71.37; H, 6.70.

4-Methoxyphenyl (2-O-acetyl-4-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (14**)**

To a solution of compound **13** (2.2 g, 1.12 mmol) in CH₂Cl₂ and water (50 mL, 1:1) was added DDQ (380 mg, 1.67 mmol) and the reaction mixture was allowed to stir at rt for 2 h. The reaction mixture was diluted with CH₂Cl₂ (100 mL) and the organic layer was washed successively with satd. aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated under reduced pressure to give the crude product, which was purified over SiO₂ using hexane-EtOAc (5:1) to furnish pure **14** (1.5 g, 73%). Colorless oil; IR (neat): 3468, 2935, 2370, 1736, 1721, 1394, 1232, 1076, 769 cm⁻¹; [α]_D²⁵ +126 (*c* 1.2, CHCl₃); ¹H NMR (300

MHz, CDCl₃): δ 7.42–7.13 (m, 50 H, Ar-H), 6.98 (d, J = 9.0 Hz, 2 H, Ar-H), 6.76 (d, J = 9.0 Hz, 2 H, Ar-H), 5.32–5.29 (m, 1 H, H-2_E), 5.23 (s, 1 H, PhCH), 5.18 (d, J = 2.7 Hz, 1 H, H-1_C), 5.02 (br s, 1 H, H-1_E), 5.0–4.85 (m, 4 H, H-1_D, PhCH₂), 4.83–4.76 (m, 3 H, H-1_A, PhCH₂), 4.75–4.54 (m, 7 H, PhCH₂), 4.53–4.40 (m, 6 H, H-1_B, PhCH₂), 4.28 (d, J = 12.0 Hz, 1 H, PhCH₂), 4.22–4.13 (m, 1 H, H-3_A), 4.12–4.04 (m, 1 H, H-5_D), 4.02–3.89 (m, 5 H, H-5_E, H-6_{abA}, H-6_{abC}), 3.87–3.80 (m, 6 H, H-2_A, H-2_C, H-2_D, H-3_E, H-4_B, H-4_C), 3.75 (s, 3 H, OCH₃), 3.78–3.72 (m, 2 H, H-2_B, H-3_C), 3.70–3.67 (m, 1 H, H-3_D), 3.66–3.57 (m, 3 H, H-4_A, H-4_D, H-4_E), 3.56–3.42 (m, 3 H, H-3_B, H-5_A, H-5_C), 3.40–3.26 (m, 3 H, H-5_B, H-6_{abB}), 2.06 (s, 3 H, COCH₃), 1.25–1.22 (m, 6 H, 2 CCH₃); ¹³C NMR (75 MHz, CDCl₃): δ 169.6 (COCH₃), 102.9 (C-1_A), 102.5 (C-1_B), 101.5 (C-1_E), 101.1 (PhCH), 100.6 (C-1_D), 93.7 (C-1_C), 82.8 (C-5_A), 81.7 (C-2_B), 81.5 (C-4_D), 80.2 (C-4_E), 80.0 (C-2_A), 79.8 (C-3_E), 79.5 (C-3_C), 79.2 (C-3_D), 79.1 (C-4_A), 77.2 (C-5_C), 76.4 (C-4_C), 75.9 (C-2_C), 75.5 (PhCH₂), 75.4 (2 C, C-5_D, C-5_E), 75.1 (PhCH₂), 73.9 (PhCH₂), 73.6 (2 C, 2 PhCH₂), 73.4 (C-5_B), 72.8 (PhCH₂), 72.5 (PhCH₂), 72.4 (PhCH₂), 72.0 (PhCH₂), 70.4 (C-2_D), 69.7 (C-6_C), 69.0 (C-6_A), 68.4 (C-2_E), 68.0 (C-3_A), 67.7 (C-6_B), 64.9 (C-4_B), 61.9 (C-3_B), 55.5 (OCH₃), 20.3 (COCH₃), 18.3, 18.2 (2 CCH₃); ESI-MS: 1865.8 [M+Na]⁺; Anal. Calcd. for C₁₀₉H₁₁₈O₂₆ (1842.79): C, 70.99; H, 6.45; found: C, 70.82; H, 6.61.

4-Methoxyphenyl (2,3,5,6-tetra-O-benzyl- α -D-galactofuranosyl)-(1 \rightarrow 3)-(2-O-acetyl-4-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 2)-(3,4-di-O-benzyl- α -L-rhamnopyranosyl)-(1 \rightarrow 3)-(2-O-benzyl-4,6-O-benzylidene- α -D-galactopyranosyl)-(1 \rightarrow 3)-(2,6-di-O-benzyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (15)

To a solution of compound **14** (1.2 g, 0.65 mmol) and compound **6** (450 mg, 0.77 mmol) in anhydrous CH₂Cl₂ (20 mL) was added MS-4 (2 g) and the reaction mixture was allowed to stir at rt for 1 h under argon. The reaction mixture was cooled to –40°C and NIS (200 mg, 0.88 mmol) and TMSOTf (5 μ L) were added to it. After stirring at same temperature for 1 h, the reaction mixture was quenched with Et₃N (0.1 mL) and filtered through a Celite bed and washed with CH₂Cl₂ (100 mL). The organic layer was washed with aq. Na₂S₂O₃ and water in succession, dried (Na₂SO₄), and concentrated under reduced pressure to give the crude product, which was purified over SiO₂ using hexane-EtOAc (3:1) as eluant to furnish pure **15** (1.2 g, 78%). Colorless oil; IR (neat): 3460, 3030, 2400, 1572, 1447, 1234, 1076, 792, 680 cm^{–1}; [α]_D²⁵ +96 (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.42–7.13 (m, 70 H, Ar-H), 7.02 (d, J = 9.2 Hz, 2 H, Ar-H), 6.78 (d, J = 9.2 Hz, 2 H, Ar-H), 5.48–5.47 (m, 1 H, H-2_E), 5.44 (d, J = 3.4 Hz, 1 H, H-1_F), 5.25 (s, 1 H, PhCH), 5.17 (d, J = 3.2 Hz, 1 H, H-1_C), 5.05–5.03 (m, 2 H, H-1_D, H-1_E), 4.99–4.92 (m, 2 H, PhCH₂),

4.90–4.80 (m, 4 H, H-1_A, PhCH₂), 4.78–4.70 (m, 3 H, PhCH₂), 4.68–4.58 (m, 5 H, PhCH₂), 4.57–4.38 (m, 12 H, H-1_B, PhCH₂), 4.36–4.22 (m, 4 H, H-3_F, H-6_{aF}, PhCH₂), 4.16–4.13 (m, 1 H, H-3_A), 4.12–4.04 (m, 3 H, H-2_D, H-5_D, H-5_E), 4.03–3.98 (m, 2 H, H-4_F, H-6_{bF}), 3.97–3.90 (m, 2 H, H-2_F, H-5_F), 3.88–3.80 (m, 7 H, H-2_C, H-3_B, H-3_E, H-6_{abA}, H-6_{abC}), 3.75 (s, 3 H, OCH₃), 3.74–3.71 (m, 2 H, H-2_A, H-4_B), 3.68–3.60 (m, 5 H, H-2_B, H-3_C, H-3_D, H-4_A, H-4_C), 3.56–3.44 (m, 3 H, H-4_D, H-4_E, H-5_A), 3.38–3.29 (m, 4 H, H-5_B, H-5_C, H-6_{abB}), 2.05 (s, 3 H, COCH₃), 1.28–1.23 (m, 6 H, 2 CCH₃); ¹³C NMR (75 MHz, CDCl₃): δ 169.7 (COCH₃), 155.2–113.5 (Ar-C), 108.4 (C-1_F), 102.7 (C-1_A), 102.4 (C-1_B), 101.3 (2 C, C-1_D, C-1_E), 100.5 (PhCH), 93.8 (C-1_C), 89.2 (C-3_F), 83.6 (C-2_F), 82.8 (C-2_B), 82.7 (C-5_A), 81.4 (2 C, C-4_D, C-4_E), 80.9 (C-3_D), 80.5 (C-4_A), 80.1 (C-2_A), 80.0 (C-3_E), 78.9 (C-3_C), 77.2 (C-5_C), 76.3 (C-4_C), 75.8 (C-2_C), 75.3 (2 C, C-5_E, PhCH₂), 75.1 (PhCH₂), 74.7 (C-5_D), 73.7 (PhCH₂), 73.5 (4 C, 4 PhCH₂), 73.4 (C-4_F), 73.2 (PhCH₂), 73.1 (PhCH₂), 72.3 (2 C, C-5_B, C-5_F), 72.0 (PhCH₂), 71.9 (PhCH₂), 71.5 (PhCH₂), 71.4 (PhCH₂), 70.6 (C-6_F), 69.0 (C-6_C), 68.5 (C-2_D), 68.3 (C-6_A), 68.2 (C-2_E), 68.1 (C-3_A), 67.8 (C-6_B), 64.9 (C-4_B), 61.9 (C-3_B), 55.6 (OCH₃), 20.3 (COCH₃), 18.0, 17.9 (2 CCH₃); ESI-MS: 2387.0 [M+Na]⁺; Anal. Calcd. for C₁₄₃H₁₅₂O₃₁ (2365.03): C, 72.57; H, 6.47; found: C, 72.38; H, 6.70.

4-Methoxyphenyl (α-D-galactofuranosyl)-(1→3)-(α-L-rhamnopyranosyl)-(1→2)-(α-L-rhamnopyranosyl)-(1→3)-(α-D-galactopyranosyl)-(1→3)-(β-D-galactopyranosyl)-(1→4)-β-D-glucopyranoside (1)

A solution of compound **15** (1 g, 0.42 mmol) in 0.1 M CH₃ONa in CH₃OH (20 mL) was allowed to stir at rt for 2 h and neutralized with Amberlite IR-120 (H⁺) resin. The reaction mixture was filtered and evaporated to dryness. To a solution of the crude product in CH₃OH (20 mL) was added 20% Pd(OH)₂-C (200 mg) and the reaction mixture was allowed to stir at rt under a positive pressure of hydrogen for 24 h. The reaction mixture was filtered through a Celite bed and then washed with CH₃OH-H₂O (60 mL; 3:1 v/v). The combined filtrate was evaporated under reduced pressure to furnish compound **1**, which was purified through a Sephadex LH-20 column using CH₃OH-H₂O (4:1) as eluant to give pure compound **1** (340 mg, 76%). Glassy solid; [α]_D²⁵ +3.3 (c 1.0, H₂O); IR (KBr): 1605, 1472, 1357, 1046, 699 cm⁻¹; ¹H NMR (300 MHz, D₂O): δ 7.06 (d, *J* = 9.0 Hz, 2 H, Ar-H), 6.93 (d, *J* = 9.0 Hz, 2 H, Ar-H), 5.16 (br s, 2 H, H-1_E, H-1_F), 5.10 (br s, 1 H, H-1_C), 4.97 (d, *J* = 7.8 Hz, 1 H, H-1_A), 4.92 (br s, 1 H, H-1_D), 4.47 (d, *J* = 7.6 Hz, 1 H, H-1_B), 4.30–4.09 (m, 4 H, H-2_E, H-2_F, H-3_F, H-4_F), 4.08–4.0 (m, 4 H, H-2_D, H-3_C, H-4_B, H-4_C), 3.98–3.87 (m, 4 H, H-3_D, H-5_C, H-5_F, H-6_{aA}), 3.86–3.61 (m, 15 H, H-2_C, H-3_A, H-3_B, H-3_E, H-4_A, H-5_B, H-5_D, H-5_E, H-6_{aA}, H-6_{aB}, H-6_{aC}, H-6_{aF}), 3.75 (s, 3 H, OCH₃), 3.60–3.37 (m, 5 H, H-2_A, H-2_B, H-4_D, H-4_E, H-5_A), 1.25–1.21 (m, 6 H, 2 CCH₃); ¹³C NMR (75

MHz, D₂O): δ 154.7, 150.9, 118.2 (2 C), 114.9 (2 C) (Ar-C), 109.1 (C-1_F), 102.8 (C-1_B), 101.8 (C-1_E), 101.1 (C-1_A), 100.5 (C-1_D), 95.5 (C-1_C), 82.9 (2 C, C-3_F, C-4_F), 80.9 (C-2_F), 78.6 (C-2_D), 78.3 (C-4_A), 77.3 (C-3_E), 75.9 (C-3_C), 74.9 (C-2_A), 74.8 (3 C, C-3_A, C-5_A, C-5_B), 74.3 (C-4_D), 72.6 (2 C, C-4_E, C-5_F), 72.1 (C-3_B), 70.7 (C-3_D), 70.5 (2 C, C-5_D, C-5_E), 69.5 (C-5_C), 69.1 (C-2_B), 68.8 (C-2_E), 67.8 (C-4_B), 64.8 (C-2_C), 62.8 (C-4_C), 62.5 (C-6_C), 60.9 (C-6_F), 60.7 (C-6_B), 59.9 (C-6_A), 55.6 (OCH₃), 16.7, 16.6 (2 CCH₃); ESI-MS: 1087.3 [M+Na]⁺; Anal. Calcd. for C₄₃H₆₈O₃₀ (1064.38): C, 48.49; H, 6.44; found: C, 48.30; H, 6.70.

ACKNOWLEDGEMENTS

S.G. thanks UGC, New Delhi, for providing a Senior Research Fellowship. This work was supported by Ramanna Fellowship (AKM), Department of Science and Technology, New Delhi (SR/S1/RFPC-06/2006).

REFERENCES

1. (a) *Microbial Lectins and Agglutinins, Properties and Biological Activities*, Mirelman, D., Ed.; **1986**, John Wiley & Sons, New York; (b) Razin, S.; Yogev, D. In *Molecular Mechanisms of Microbial Adhesion*; Switalski, L.; Hook, M.; Beachey, E., Eds.; **1989**, Springer-Verlag, New York, pp. 52–76; (c) Weis, W.; Brown, J. H.; Cusack, S.; Paulson, J.C.; Skehel, J.J.; Wiley, D.C. Structure of the influenza virus haemagglutinin complexed with its receptor, sialic acid. *Nature* **1988**, *333*, 426–431.
2. (a) Leffler, H.; Svanborg-Eden, C. Glycolipids as receptors for *Escherichia coli* lectins. In *Microbial Lectins and Agglutinins, Properties and Biological Activity*; Mirelman, D., Ed.; **1986**, John Wiley & Sons, New York, pp. 83–112; (b) Karlsson, K.A. Animal alycosphingolipids as membrane attachment sites for bacteria. *Annu. Rev. Biochem.* **1989**, *58*, 309–350.
3. (a) Roberts, D.D.; Olson, L.D.; Barile, M.F.; Ginsburg, V.; Krivan, H.C. Sialic acid-dependent adhesion of *Mycoplasma pneumoniae* to purified glycoproteins. *J. Biol. Chem.* **1989**, *264*, 9289–9293; (b) Parkkinen, J.; Rogers, G.N.; Korhonen, T.; Dahr, W.; Finne, J. Identification of the O-linked sialyloligosaccharides of glycophorin A as the erythrocyte receptors for S-fimbriated *Escherichia coli*. *Infect. Immun.* **1986**, *54*, 37–42.
4. (a) Abeygunawardana, C.; Bush, C.A.; Cisar, J.O. Complete structure of the polysaccharide from *Streptococcus sanguis* J22. *Biochemistry* **1990**, *29*, 234–248; (b) Neeser, J.R.; Koellreutter, B.; Wuersch, P. Oligomannoside-type glycopeptides inhibiting adhesion of *Escherichia coli* strains mediated by type 1 pili: preparation of potent inhibitors from plant glycoproteins. *Infect. Immun.* **1986**, *52*, 428–436; (c) McIntire, F.C.; Crosby, L.K.; Vatter, A.E.; Cisar, J.O.; McNeil, M.R.; Bush, C.A.; Tjoa, S.S.; Fennessey, P.V. A polysaccharide from *Streptococcus sanguis* 34 that inhibits coaggregation of *S. sanguis* 34 with *Actinomyces viscosus* T14V. *J. Bacteriol.* **1988**, *170*, 2229–2235; (d) Cassels, F.J.; London, J. Isolation of a coaggregation-inhibiting cell wall polysaccharide from *Streptococcus sanguis* H1. *J. Bacteriol.* **1989**, *171*, 4019–4025.
5. Rosan, B.; Lamont, R.J. Dental plaque formation. *Microbes Infect.* **2000**, *2*, 1599–1607.
6. (a) Abeygunawardana, C.; Bush, C.A.; Cisar, J.O. Complete structure of the cell surface polysaccharide of *Streptococcus oralis* C104: a 600-MHz NMR study. *Biochemistry* **1991**, *30*, 8568–8577; (b) Kolenbrander, P.E. Intergeneric coaggregation among

human oral bacteria and ecology of dental plaque. *Annu. Rev. Microbiol.* **1988**, *42*, 627–656; (c) London, J.; Allen, J. Purification and characterization of a *Bacteroides loeschei* adhesin that interacts with procaryotic and eucaryotic cells. *J. Bacteriol.* **1990**, *172*, 2527–2534.

7. (a) McIntire, F.C.; Vatter, A.E.; Baros, J.; Arnold, J. Mechanism of coaggregation between *Actinomyces viscosus* T14V and *Streptococcus sanguis* 34. *Infect. Immun.* **1978**, *21*, 978–988; (b) Kolenbrander, P.E. Coaggregation: adherence in the human oral microbial ecosystem. In *Microbial Cell-Cell Interactions*, Dworkin, M., Ed.; **1991**, American Society of Microbiology, Washington, D.C., pp. 303–329; (c) Weiss, E.I.; London, J.; Kolenbrander, P.E.; Kagermeier, A.S.; Andersen, R.N. Characterization of lectinlike surface components on *Capnocytophaga ochracea* ATCC 33596 that mediate coaggregation with gram-positive oral bacteria. *Infect. Immun.* **1987**, *55*, 1198–1202.

8. (a) Kolenbrander, P.E.; Andersen, R.N.; Moore, L.V.H. Intrageneric coaggregation among strains of human oral bacteria: potential role in primary colonization of the tooth surface. *Appl. Environ. Microbiol.* **1990**, *56*, 3890–3894; (b) Kolenbrander, P.E.; Anderson, R.N. Cell to cell interactions of *Capnocytophaga* and *Bacteroides* species with other oral bacteria and their potential role in development of plaque. *J. Periodontal Res.* **1984**, *19*, 564–569; (c) Weiss, E.I.; Eli, I.; Shenitzki, B.; Smorodinsky, N. Identification of the rhamnose-sensitive adhesin of *Capnocytophaga ochracea* ATCC 33596. *Arch. Oral. Biol.* **1990**, *35*, 127S–130S.

9. (a) Yoshida, Y.; Yang, J.; Peaker, P.-E.; Kato, H.; Bush, C.A.; Cisar, J.O. Molecular and antigenic characterization of a *Streptococcus oralis* coaggregation receptor polysaccharide by carbohydrate engineering in *Streptococcus gordonii*. *J. Biol. Chem.* **2008**, *283*, 12654–12664; (b) Cassels, F.J.; Fales, H.M.; London, J.; Carlson, R.W.; van Halbeek, H. Structure of a streptococcal adhesin carbohydrate receptor. *J. Biol. Chem.* **1990**, *265*, 14127–14135.

10. Glushka, J.; Cassels, F.J.; Carlson, R.W.; van Halbeek, H. Complete structure of the adhesin receptor polysaccharide of *Streptococcus oralis* ATCC 55229 (*Streptococcus sanguis* H1). *Biochemistry* **1992**, *31*, 10741–10746.

11. (a) Verez-Bencomo, V.; Fernández-Santana, V.; Hardy, E.; Toledo, M.E.; Rodríguez, M.C.; Heynngnezz, L.; Rodriguez, A.; Baly, A.; Herrera, L.; Izquierdo, M.; Villar, A.; Valdés, Y.; Cosme, G.; Deler, M.L.; Montane, M.; Garcia, E.; Ramos, A.; Aguilar, A.; Medina, E.; Toraño, G.; Sosa, I.; Hernandez, I.; Martínez, R.; Muzachio, A.; Carmenates, A.; Costa, L.; Cardoso, F.; Campa, C.; Diaz, M.; Roy, R. A synthetic conjugate polysaccharide vaccine against *Haemophilus influenzae* type b. *Science* **2004**, *305*, 522–525; (b) Tamborini, M.; Werz, D.B.; Frey, J.; Pluschke, G.; Seeberger, P.H. Anti-carbohydrate antibodies for the detection of anthrax spores. *Angew. Chem. Int. Ed. Engl.* **2006**, *45*, 6581–6582; (c) Pozsgay, V. Synthetic shigella vaccines: a carbohydrate-protein conjugate with totally synthetic hexadecasaccharide haptens. *Angew. Chem. Int. Ed. Engl.* **1998**, *37*, 138–142; (e) Jansen, W.T.M.; Hogenboom, S.; Thijssen, M.J.L.; Kamerling, J.P.; Vliegthart, J.F.G.; Verhoef, J.; Snippe, H.; Verheul, A.F.M. Synthetic 6B di-, tri-, and tetrasaccharide-protein conjugates contain pneumococcal type 6A and 6B common and 6B-specific epitopes that elicit protective antibodies in mice. *Infect. Immun.* **2001**, *69*, 787–793.

12. Mandal, P.K.; Maiti, G.H.; Misra, A.K. Facile synthesis of a tetrasaccharide corresponding to the capsular polysaccharide of *Streptococcus pneumoniae* type 15B. *Arkivoc* **2009**, *ii*, 281–287.

13. Sato, S.; Ito, Y.; Ogawa, T. Stereo- and regio-controlled, total synthesis of the Le^b antigen, III⁴ FucIV²FucLcOSe₄ Cer. *Carbohydr. Res.* **1986**, *155*, C1–C5.

14. Sajtos, F.; Hajko, J.; Kover, K.E.; Liptak, A. Synthesis of the α -D-GlcP-A-(1 \rightarrow 3)- α -L-Rhap-(1 \rightarrow 2)-L-Rha trisaccharide isolated from the cell wall hydrolyzate of the green alga, *Chlorella vulgaris*. *Carbohydr. Res.* **2001**, *334*, 253–259.

15. Mukherjee, C.; Misra, A.K. Total synthesis of an antigenic heptasaccharide motif found in the cell-wall lipooligosaccharide of *Mycobacterium gordonae* strain 989. *Glycoconjugate J.* **2008**, *25*, 611–624.
16. (a) Mandal, P.K.; Misra, A.K. Synthesis of oligosaccharides corresponding to the polysaccharides of *Lactobacillus* and *Thermophilus* strains. *Synthesis* **2007**, 2660–2666; (b) Gelin, M.; Ferrieres, V.; Plusquellec, D. A general and diastereoselective synthesis of 1,2-cis-hexofuranosides from 1,2-trans-thiofuranosyl donors. *Eur. J. Org. Chem.* **2000**, 1423–1431.
17. (a) Veeneman, G.H.; van Leeuwen, S.H.; van Boom, J.H. Iodonium ion promoted reactions at the anomeric centre. II An efficient thioglycoside mediated approach toward the formation of 1,2-*trans* linked glycosides and glycosidic esters. *Tetrahedron Lett.* **1990**, *31*, 1331–1334; (b) Konradsson, P.; Udodong, U.E.; Fraser-Reid, B. Iodonium promoted reactions of disarmed thioglycosides. *Tetrahedron Lett.* **1990**, *31*, 4313–4316.
18. Ali, A.; Channe Gowda, D.; Vishwakarma, R.A. A new approach to construct full-length glycosylphosphatidylinositols of parasitic protozoa and [4-deoxy-Man-III]-GPI analogues. *Chem. Commun.* **2005**, 519–521.
19. Oikawa, Y.; Yoshioka, T.; Yonemitsu, O. Specific removal of *o*-methoxybenzyl protection by DDQ oxidation. *Tetrahedron Lett.* **1982**, *23*, 885–888.
20. Pearlman, W.M. Noble metal hydroxides on carbon nonpyrophoric dry catalysts. *Tetrahedron Lett.* **1967**, *8*, 1663–1664.

Copyright of Journal of Carbohydrate Chemistry is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.