# <sup>13</sup>C NMR Study of *N*-Acyl- and *N*-Sulphonyl-isatins and their Ring-Opened Derivatives

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The complete assignments of the <sup>13</sup>C NMR resonances of 1-acetyl-, 1-benzoyl-, 1-benzenesulphonyl-, 1methanesulphonyl- and 1-(2-nitrobenzenesulphonyl)-isatins and their ring-opened glyoxylic derivatives are reported. The <sup>13</sup>C assignments of acylisatins were derived from isatin as a model, while assignments of glyoxylic derivatives were made by complete assignment of compound 8b together with its use as a model.

KEY WORDS <sup>13</sup>C NMR 1-Acyl-2,3-dihydroindole-2,3-diones 2-Acylamidophenylglyoxylic acid derivatives

## **INTRODUCTION**

1-Acyl-2,3-dihydroindole-2,3-diones (acylisatins) are an important class of compounds which behave as activated imides. They undergo ring-opening reactions with a variety of nucleophiles to yield 2acylamidophenylglyoxylic acid derivatives.<sup>1,2</sup> These compounds have been used in the synthesis of quinoline and quinazoline derivatives.<sup>3,4</sup> Although <sup>13</sup>C NMR chemical shift assignments of isatins have been reported,<sup>5</sup> there is a scarcity of information regarding <sup>13</sup>C NMR chemical shifts of acylisatins and acylamidophenylglyoxylic acid derivatives. We now report the analyses of <sup>13</sup>C NMR spectra of a large number of acylisatins and their phenylglyoxylic acid derivatives.

### EXPERIMENTAL

1-Acetyl-2,3-dihydroindole-2,3-dione was prepared by heating isatin with excess of acetic anhydride.<sup>6</sup> 1-Benzoyl-, 1-benzenesulphonyl-, 1-methanesulphonyl and 1-(2-nitrobenzenesulphonyl)-2,3-dihydroindole-2,3diones were prepared from sodium isatide and corresponding acid chlorides by the method previously described for 1-benzoylisatin.<sup>7</sup> The N-acylamidophenylglyoxylic acid derivatives were prepared by the reaction of N-acylisatins with alcohols, water and amines.

<sup>1</sup>H NMR spectra were recorded on a Bruker AM 500 spectrometer using CDCl<sub>3</sub> or  $(CD_3)_2SO$  as solvent and internal deuterium lock. The chemical shifts were read from the solvent [CHCl<sub>3</sub>,  $\delta$  7.26 pm;  $(CD_3)_2SO$ ,  $\delta$  2.5 ppm]. The <sup>13</sup>C NMR spectra were recorded in the pulsed Fourier transform mode (16K data points for the

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FID) at 298 K on a Bruker AM 500 spectrometer (125.76 MHz) using  $\text{CDCl}_3 \delta$  77.00 ppm. All <sup>13</sup>C measurements were carried out using the DEPT heteronuclear multi-pulse program with a pulse repetition time of 3 s.

The acquisition parameters were AQ 0.297 s,  $D_1 = 2$  s, SI = 16K,  $S_2 = 18$ H,  $D_2 = 3.12$  µs,  $PO = 45^{\circ}$ ,  $90^{\circ}$ ,  $135^{\circ}$ .

#### **RESULTS AND DISCUSSION**

The <sup>13</sup>C NMR spectral assignments of the parent acylisatins (1 and 2) and the parent sulphonamides (3-5) were easily made from the broad-band decoupled and DEPT spectra using isatin<sup>5</sup> as a model, and are presented in Table 1. The assignments of glyoxylic derivatives were made from the complete assignment of 8b, which was subsequently used as the primary model system for the <sup>1</sup>H and <sup>13</sup>C NMR assignments. The 500 MHz <sup>1</sup>H NMR spectrum of derivative **8b** in CDCl<sub>3</sub> shows seven distinct resonances. Assignments of two upfield singlets  $(SO_2CH_3, OCH_3)$  and the slightly broadened downfield singlet (NH) were easily made on the basis of chemical shifts of the related acylisatin derivatives 6b and 7b. The four signals between 7.79 and 7.18 ppm were assigned on the basis of decoupling and NOE difference experiments. One doublet (7.79 ppm, H-7) and one triplet (7.18 ppm, H-5) were sharp whereas the other doublet (7.73 ppm, H-4) and triplet (7.66 ppm, H-6) appeared to be broadened. The broadening of the two signals is apparently due to unresolved meta coupling between protons H-4 and H-6. Their relative orientation was established by decoupling experiments. Irradiation at 7.18 ppm (H-5) caused the doublet at 7.73 ppm (H-4) to collapse to a singlet while the triplet at 7.66 ppm (H-6) collapsed to a doublet. Irradiation at 7.73 ppm (H-4) resulted in sharpening of the broadened triplet at 7.66 ppm (H-6) and the triplet at 7.18 ppm

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(H-5) collapsed to a doublet. Irradiation of the doublet at 7.79 ppm (H-7) reduced the triplet at 7.66 ppm (H-6) to a doublet. The spectral orientation of protons H-4 and H-7 relative to the *ortho* substituent was established by a NOE difference experiment. Irradiation at 7.79 ppm (H-7) gave a NOE at 10.51 (NH) and 7.66

ppm (H-6), whereas irradiation at 7.73 ppm (H-4) gave a NOE at 7.18 (H-5).

The <sup>13</sup>C NMR broad-band decoupled and DEPT spectra of **8b** allowed the ready assignment of all carbons except C-4, C-5, C-6 and C-7. However, a  ${}^{13}C/{}^{1}H$  heteronuclear correlation experiment allowed

			Parent con	npound		
Carbon	1	2	3	4	5	Isatin
C-2	158.13	157.09	156.8	156.1	156.0	159.2
C-3	180.10	180.17	179.2	178.5	177.9	184.3
C-3a	119.70	119.89	119.7	120.0	120.0	117.8
C-4	125.49	125.23	125.5 <sup>b</sup>	125.6 <sup>b</sup>	125.9 <sup>b</sup>	123.8
C-5	124.32	124.38	125.4 <sup>b</sup>	125.3 <sup>6</sup>	125.7 <sup>6</sup>	122.7
C-6	137.77	137.38	138.2	138.2	138.1	138.3
C-7	117.16	116.06	114.5	114.5	115.1	112.5
C-7a	147.85	147.98	147.0	146.3	146.4	151.8
C-9	169.76	167.94				
C-10		133.68		137.7	130.1	
C-11		128.02°		129.9°	147.4	
C-12		129.58°		128.0°	125.3 <sup>b</sup>	
C-13		132.96		135.3	136.9	
C-14		129.58°		128.0°	133.6	
C-15		128.02°		129.9°	132.5	
CH₃	25.88		41.7			
In ppm <sup>,</sup> ° Signa	from TMS	at 0.0 ppm. rtical column	may be int	erchanged.		

Table 1.	<sup>13</sup> C NM	R chemical	shifts*	for	isatin, <sup>5</sup>	acylisatins	(1	and	2)
	and sulph	onylisatins	(3-5) in	DN	4SO	-			

their assignment. The experiment clearly showed the  ${}^{13}C$  signals at 137.2 (C-6), 134.2 (C-4), 122.7 (C-5) and 117.7 ppm (C-7) to be correlated with the  ${}^{1}H$  NMR signals at 7.66 (H-6), 7.73 (H-4), 7.18 (H-5) and 7.79 ppm (H-7), respectively. This cumulative data allowed the complete  ${}^{13}C$  NMR assignments for **8b**, which subse-

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quently served as a primary model system for the assignments of compounds 6-10, (Tables 2 and 3).

It can be noted that different chemical shifts are observed for C-16 and C-20 and also for C-17 and C-19 in the spectra of the morpholine and piperidine amide derivatives, resulting from slightly restricted rotation.

			Acids <sup>b</sup>					Methyl ester	's <sup>c</sup>			Anil	ides	
Carbon	6a	7a	<b>8</b> a	9a	10a	6b	7b	8b	9b	10b	6c <sup>b</sup>	7c⁵	<b>8c</b> °	9c <sup>c.f</sup>
C-2	165.0	165.2 <sup>d</sup>	165.6	165.4	165.2	163.7	163.7	163.4	163.2	163.4	162.1	162.5	160.2	159.7
C-3	189.4	190.8	191.5	190.8	191.0	190.1	190.5	189.7	189.6	189.1	190.3	191.7	190.9	190.7
C-3a	123.8	122.6	123.0	126.6	124.2	116.8	117.3	117.2	117.5	118.4	124.7	123.4	118.7	119.8
C-4	131.7	132.5°	133.3	132.4	133.3ª	133.4	133.7	134.2	133.8 <sup>d</sup>	134.0	131.6	132.7 <sup>d</sup>	134.9	134.4
C-5	124.2	124.4	124.7	125.8	125.7°	122.4	122.7	122.7	122.8	123.4	124.6 <sup>d</sup>	124.8°	122.6	123.0
C-6	135.0	135.8	136.4	135.3	136.0	137.0	137.4	137.2	136.7	136.8	134.2	135.2	136.5	135.9
C-7	122.3	122.4	121.2	123.0	121.8	120.5	120.7	117.7	118.4	118.3	122.0	122.3	117.9	119.6
C-7a	139.1	140.1	140.1	138.3 <sup>d</sup>	137.9	142.5	143.0	141.9	141.2	140.2	138.4°	139.8	141.2	140.3
C-9	169.5	165.8 <sup>d</sup>				169.5	165.9				169.2	165.9		
C-10		134.2		138.9 <sup>d</sup>	130.9		134.2		138.7	132.2		134.2		138.3
C-11		127.7		129.7°	148.0		127.4		129.1	148.0		127.7		129.0 <sup>d</sup>
C-12		129.2		127.3°	125.6°		128.8		127.0	125.6		129.1		126.8
C-13		132.6°		133.8	135.7		132.3		133.3 <sup>d</sup>	134.6		132.6ª		133.2
C-14		129.2		127.3°	133.2 <sup>d</sup>		128.8		127.0	132.7		129.1		126.8
C-15		127.7		129.7°	131.1		127.4		129.1	131.2		127.7		129.0 <sup>d</sup>
C-16											138.2°	138.1	136.4	136.3
C-17											120.4	120.4	120.1	120.0
C-18											129.1	129.1	128.9	128.8 <sup>d</sup>
C-19											124.0 <sup>d</sup>	124.3 <sup>e</sup>	125.3	125.1
C-20											129.1	129.1	128.9	128.8 <sup>d</sup>
C-21											120.4	120.4	120.1	120.0
CH <sub>3</sub>	24.3		40.2			25.3		40.4			24.1		40.0	
OCH3						52.8	53.0	53.1	52.9	53.1				
<sup>a</sup> In ppm <sup>b</sup> DMS( <sup>c</sup> CDCl <sub>3</sub>	from TM ) as solve as solve	/IS at 0.0 j ent. nt.	opm.											

<sup>d,e</sup> Signals in any vertical column may be interchanged.

<sup>f</sup>Values for the pairs C-11, C-15 and C-18, C-20 must be identical.

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Table 3.	<sup>13</sup> C NMR	chemical	l shifts" fc	r amide d	erivatives	in CDCI	e												
		Σ	orpholine am	ide			-N	Methyl amid	9			N,N-Dimeth	yl amide			Pip	eridine amide		
	ę	РĹ	<b>9</b> 8	<b>B</b>	10d	°е	76	8	8	10e	6f	7	8f	96	ç	79	80	<b>6</b> 6	109
C-2	163.9	164.3	163.9	163.7	163.7	164.5	163.8	163.3	163.0	162.6	165.8	166.0	165.3	165.1	163.8	164.0	163.5	163.5	163.6
C-3	194.7	195.5	194.5	194.6	194.0	191.8	192.6	191.6	191.4	190.6	195.6	196.3	195.2	195.3	195.5	196.2	195.3	195.5	194.9
C-3a	117.3	118.1	117.8	118.4	118.8	124.5	118.8	118.9	119.7	120.9	117.5	118.1	117.5	118.1	117.4	118.0	117.6	118.4	118.8
C-4	132.9	133.5	134.0	133.6 <sup>b</sup>	133.9	131.9	134.7	135.1	134.5	134.4 <sup>b</sup>	133.2	133.6	134.0	133.6	133.0	133.4	113.8	133.6 <sup>b</sup>	133.9
C-5	122.2	122.8	122.7	123.1	123.4	123.8	122.7	122.7	122.9	123.6	122.5	122.9	122.6	123.1	122.2	122.7	122.5	123.0	123.3
C-6	136.4	137.1	137.0	136.6	136.6	134.3	136.7	136.5	135.9	136.0	136.5	137.0	136.8	136.3	136.2	136.7	136.6	136.2	136.4
C-7	120.0	120.7	117.4	118.9	117.9	121.9	120.6	117.8	119.1	119.0	120.3	120.6	117.2	118.6	120.0	120.4	117.1	118.7	117.8
C-7a	141.8	146.6	141.6	141.0	140.0	138.8	142.5	141.2	140.4	139.4	142.1	142.7	141.4	140.7	141.8	142.4	141.3	140.8	140.0
C-9	168.8	165.8				169.1	166.0				169.3	166.0			169.0	165.7			
C-10		134.1		139.0	132.0		134.4		138.6	132.2		134.2		138.8		134.0		139.0	132.2
C-11		127.3		129.1	147.8		127.4		129.0	147.9		127.4		129.0		127.1		129.0	147.9
C-12		128.7		127.0	125.4		128.8		126.9	125.5		128.8		126.8		128.6		126.7	125.4
C-13		132.2		133.2 <sup>b</sup>	134.6		132.2		133.2	134.7 <sup>b</sup>		132.2		133.2		132.0		133.1 <sup>b</sup>	134.5
C-14		128.7		127.0	132.6		128.8		126.9	132.6		128.8		126.8		128.6		126.7	132.6
C-15		127.3		129.1	131.2		127.4		129.0	131.4		127.4		129.0		127.1		129.0	131.1
C-16	45.7	46.2	46.2	46.0	46.1										46.6	46.8	46.8	46.7	47.0
C-17	66.0 <sup>b</sup>	66.4	66.4 <sup>b</sup>	66.3	66.4 <sup>b</sup>										25.6°	25.8 <sup>b</sup>	25.7 <sup>b</sup>	25.8°	25.9 <sup>6</sup>
C-18															25.0 <sup>b</sup>	25.1 <sup>b</sup>	25.0 <sup>b</sup>	25.0°	25.2 <sup>b</sup>
C-19	65.9°	66.4	66.3 <sup>b</sup>	66.3	66.3 <sup>b</sup>										23.7	23.9	23.9	23.9	24.0
C-20	41.0	41.5	41.6	41.4	41.6										41.6	41.9	41.9	41.9	42.1
C-21																			
сн <sub>э</sub>	24.9		40.4			24.3		40.1			25.2		40.2		24.9		40.1		
осн																			
NHCH <sub>3</sub>						25.8	26.2	26.1	25.9	26.2									
N(CH <sub>3</sub> ) <sub>2</sub>											36.8 33.7	37.0 33.9	36.9 33.8	36.6 33.6					
<sup>a</sup> In ppm fr	om TMS a	t 0.0 ppm.																	
<sup>b.c</sup> Signals	in any vert	tical colun	nn may be	interchang	jed.													ļ	

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## REFERENCES

- G. Heller, Ber. Dtsch. Chem. Ges. 55, 2681 (1922).
  F. J. Meyers, Chem. Ber. 99, 3060 (1966).
  C. R. Wetzel, J. R. Shanklin, Jr, and R. E. Lutz, J. Med. Chem. 16, 528 (1973).
- J. Bergman, B. Egestad and J.-O. Lindström, *Tetrahedron Lett*. 2625 (1977).
- V. Galasso, G. Pellizer and G. C. Pappalardo, Org. Magn. Reson. 9, 401 (1977).
  J. Büchi, H. Hurni and R. Lieberherr, Helv. Chim. Acta 32, 1806
- (1949).
- 7. G. Tacconi, P. P. Righetti and G. Desimoni, *J. Prakt. Chem.* **315**, 339 (1973).