

A NEW FLAVANONE GLUCOSIDE FROM THE FLOWERS OF *Carthamus tinctorius* AND ASSIGNMENT OF ABSOLUTE CONFIGURATION

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A new flavanone glucoside was isolated from the flowers of Carthamus tinctorius. The structural elucidation was performed mainly by mean of FAB-MS, NMR, and acid hydrolysis; the absolute configuration was also determined by circular dichroism spectroscopy.

Keywords: *Carthamus tinctorius*, Compositae, flavanone glucoside.

Carthamus is one of the most important genera in the Compositae family. The dried flower of *Carthamus tinctorius* (safflower) has long been used traditionally in China and southeast Asia. It showed therapeutic potential for treating coronary heart disease, stroke, gynecological disease, angina, and hypertension [1–7]. Many studies concerning the chemical constituents of *Carthamus tinctorius* have been carried out, which have led to the isolation of a series of secondary metabolites, including flavonoids, alkaloids, lignans, and fatty acids. These constituents were found to be responsible for the overall curative effects of safflower [8–18]. In this article, we present the isolation and structure elucidation of a new flavanone glucoside based on extensive spectroscopic and chemical studies.

Compound **1** was obtained as white needles, $[\alpha]_D^{30} -20.2^\circ$ in MeOH; its IR spectrum (KBr) showed absorption bands for hydroxyl (3390 cm^{-1}), conjugated carbonyl (1645 cm^{-1}) groups, and aromatic rings (1616 and 1520 cm^{-1}). HR-FAB-MS showed an m/z 473.1062 ($M + Na$)⁺ corresponding to the molecular formula $C_{21}H_{22}O_{11}$.

The ^1H NMR spectrum of compound **1** exhibited doublets at δ 5.80 and 5.89 (each 1H, d, $J = 2.10\text{ Hz}$) due to meta coupling of two protons of the A-ring. The specific dihydropyrone core of a flavanone aglycone was confirmed by the ^1H NMR spectrum, in which the signals due to the H-2, H-3a, and H-3b protons were observed as an ABX type at δ 5.44 (dd, $J_{XA} = 12.36$, $J_{XB} = 3.42\text{ Hz}$), 2.71 (dd, $J_{AB} = 17.16$, $J_{BX} = 3.42\text{ Hz}$, *cis*), and 3.21 (dd, $J_{AB} = 17.16$, $J_{AX} = 12.36\text{ Hz}$, *trans*), respectively. The ^1H NMR data also showed signals attributed to an anomeric proton at δ 4.69 (1H, d, $J = 7.56\text{ Hz}$, H-1''), together with five glycosyl protons in the range δ 3.8–3.4. The ^{13}C NMR and DEPT spectra of **1** displayed 21 carbon signals consisting of six carbons of the glycosyl moiety. All of the above data revealed that **1** is a flavanone monoglycoside with a β sugar unit; acid hydrolysis of compound **1** also indicates that the monosaccharide moiety was a D-glucose.

The signals in the ^1H and ^{13}C NMR spectra were unambiguously assigned by HMQC, HMBC, H–H COSY, and NOESY experiments (Fig. 1). Long-range correlation from the anomeric proton δ 4.69 (d, $J = 7.56\text{ Hz}$) to C-4' (δ 145.4) in the HMBC spectrum and the correlation between δ 7.12 (d, $J = 8.22\text{ Hz}$, H-5') and 4.69 (d, $J = 7.56\text{ Hz}$, H-1'') in the NOESY spectrum unequivocally established that the sugar linkage to the aglycone in **1** was at C-4'.

Additionally, the absolute configuration at C-2 was found to be *S* as it showed a negative Cotton effect at 290 nm ($\Delta\epsilon = -11.2$, $c = 11.10 \times 10^{-5}\text{ M}$) and positive Cotton effect ($\Delta\epsilon = +8.2$, $c = 11.10 \times 10^{-5}\text{ M}$) at 334 nm in the CD spectrum. Accordingly, the structure of **1** was determined as (2*S*)-5,7,3'-trihydroxyflavanone-4'-*O*- β -D-glucopyranoside [19].

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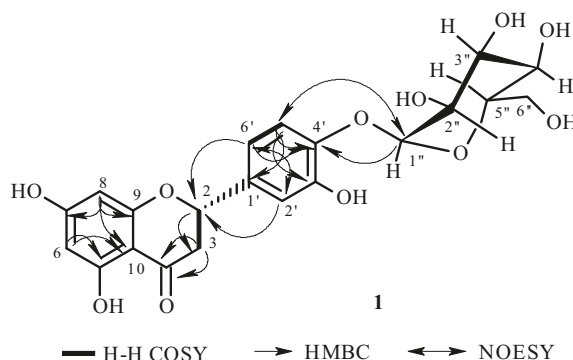


Fig. 1. Structure of compound **1**.

EXPERIMENTAL

General. NMR spectra were recorded on a Varian Inova AS 600 system; CD spectra were measured on a Jasco J-720 W spectrometer; FAB-MS spectra were recorded with a JEOL JMS-SX 102A mass spectrometer. Diaion HP-20 (Mitsubishi Chemical Corporation) and Chromatorex ODS DM1020T (100–200 mesh, Fuji Silysia Chemical Ltd.) were used for column chromatography. Preparative HPLC: Shimadzu LC-10AD with refractive index detector; preparative column: YMC-Pack ODS-A (250 × 20 mm i.d., S-5 μ m, 12 nm); sugar identification HPLC: optical rotation detector (Shodex OR-2), Kaseisorb LC NH₂-60-5, 250 × 4.6 mm.

Plant Material. The flowers of *Carthamus tinctorius* were collected from Xinjiang, China in 2011. A voucher specimen of this plant was deposited at the College of Agronomy, Sichuan Agricultural University.

Extraction and Isolation. The flowers of *Carthamus tinctorius* (2 kg) were extracted three times with MeOH under reflux for 3 h. Evaporation of the solvent under reduced pressure provided the MeOH extract (40.1 g). The MeOH extract was passed through a Diaion HP-20 column chromatograph and sequentially eluted twice with H₂O and MeOH to give an H₂O-eluted fraction (15.5 g) and an MeOH-eluted fraction (24.5 g). The MeOH-eluted fraction (24.5 g) was chromatographed on ODS columns using a gradient of MeOH–H₂O (20: 80→40: 60→60: 40→80: 20)→MeOH to give 10 fractions (1–10). Fraction 4 (2.0 g) was repeatedly chromatographed on RP-C₁₈ [MeOH–H₂O (45:55)] to give **1** (6.0 mg).

Compound 1. White needles. ¹H NMR (600 MHz, DMSO-d₆, δ , ppm, J/Hz): 2.71 (1H, dd, J = 3.42, 17.16, H-3a), 3.21 (1H, dd, J = 12.36, 17.16, H-3b), 3.40 (2H, m, H-4'', 5''), 3.46 (2H, m, H-2'', 3''), 3.71 (2H, dd, J = 3.42, 11.7, H-6''), 4.69 (1H, d, J = 7.56, H-1''), 5.44 (1H, dd, J = 3.42, 12.36, H-2), 5.80 (1H, d, J = 2.10, H-6), 5.89 (1H, d, J = 2.10, H-8), 6.85 (1H, dd, J = 2.04, 8.22, H-6'), 6.95 (1H, d, J = 2.04, H-2'), 7.12 (1H, d, J = 8.22, H-5'). ¹³C NMR (150 MHz, DMSO-d₆, δ , ppm): 78.1 (C-2), 42.0 (C-3), 196.1 (C-4), 163.4 (C-5), 95.8 (C-6), 166.6 (C-7), 95.0 (C-8), 162.7 (C-9), 101.8 (C-10), 133.2 (C-1'), 114.4 (C-2'), 146.7 (C-3'), 145.4 (C-4'), 116.6 (C-5'), 117.7 (C-6'), 102.2 (C-1''), 73.3 (C-2''), 75.8 (C-3''), 69.8 (C-4''), 77.2 (C-5''), 60.7 (C-6'').

REFERENCES

1. X. Qiao, J. Han, M. Xu, Z.-C. Zhang, S.-f. Sun, H.-f. Li, B.-R. Wang, M. Ye, and D.-a. Guo, *Planta Med.*, **74**, 1720 (2008).
2. P.-H. Nie, L. Zhang, W.-H. Zhang, W.-F. Rong, and J.-M. Zhi, *J. Ethnopharmacol.*, **139**, 746 (2012).
3. M. Sugano, I. Ikeda, K. Wakamatsu, and T. Oka, *Br. J. Nutr.*, **72**, 775 (1994).
4. H. He, X. Yang, M. Shi, X. Zeng, J. Yang, L. Wu, and L. Li, *J. Pharm. Pharmacol.*, **60**, 115 (2008).
5. Y. Zou, L. Lin, Y. Ye, J. Wei, N. Zhou, Y. Liang, H. Gong, L. Li, J. Wu, Y. Li, Z. Jia, Y. Wu, J. Zhou, and J. Ge, *J. Cardiovasc. Pharmacol.*, **59**, 268 (2012).
6. Y. Aoki, M. Suzuki, and Y. Kasahara, *Yamagata-ken Eisei Kenkyushoho*, **33**, 9 (2000).
7. H.-B. Zhu, L. Zhang, Z.-H. Wang, J.-W. Tian, F.-H. Fu, K. Liu, and C.-L. Li, *J. Asian Nat. Prod. Res.*, **7**, 607 (2005).
8. M. N. Kim, F. L. Scao-Bogaert, and M. Paris, *Planta Med.*, **58**, 285 (1992).

9. S. M. Aslanov and S. S. Mamedov, *Chem. Nat. Compd.*, **23**, 251 (1987).
10. K. M. Ahmed, M. S. Marzouk, E. A. el-Khrisy, S. A. Wahab, and S. S. el-Din, *Pharmazie*, **55**, 621 (2000).
11. Y. Z. Zhou, H. Chen, L. Qiao, N. Xu, J. Q. Cao, and Y. H. Pei, *J. Asian Nat. Prod. Res.*, **10**, 429 (2008).
12. H. H. Yoo, J. H. Park, and S. W. Kwon, *Biosci. Biotechnol., Biochem.*, **70**, 2783 (2006).
13. N. Sakakibara, T. Nakatsubo, S. Suzuki, D. Shibata, M. Shimada, and T. Umezawa, *Org. Biomol. Chem.*, **5**, 802 (2007).
14. S. M. Aslanov and A. A. Radzhabov, *Chem. Nat. Compd.*, **24**, 744 (1988).
15. N. T. Ul'chenko, S. D. Gusakova, and S. K. Khidoyatova, *Chem. Nat. Compd.*, **49**, 88 (2013).
16. O. S. Otroshchenko, N. N. Stepanichenko, S. D. Gusakova, and S. Z. Mukhamedzhanov, *Chem. Nat. Compd.*, **15**, 236 (1979).
17. G. Zhang, M.-L. Guo, R.-P. Li, Y. Li, H.-M. Zhang, and Z.-W. Su, *Chem. Nat. Compd.*, **45**, 398 (2009).
18. T. A. Suleimanov, *Chem. Nat. Compd.*, **40**, 13 (2004).
19. D. Slade, D. Ferreira, and J. P. Marais, *Phytochemistry*, **66**, 2177 (2005).