there is an induction period (ca. 8 hours) in the catalysis under N₂ while no such period of time has been observed in the catalysis under H₂ (see Figure 2). In the presence of hydrogen (H2), complex 2c catalyzes the isomerization of tDC2B to give a mixture of cDC1B and tDC1B, and hydrogenation of tDC2B to give DCB (Figure 4). Again, the same results (as shown in Figure 4) have been obtained by using 1c in place of 2c under H2 (1c immediately reacts with H₂ to form **2c** under the catalytic conditions). The rate of \overline{DCB} formation from cDC1B (or tDC1B) with **2a** (or **2b**) (Figure 2) seems to be significantly faster than that from tDC2B with 2c (Figure 4). It is apparent in Figure 4 that DCB is formed by the hydrogenation of cDC1B and tDC1B which are produced by the isomerization of tDC2B although it is not clear whether DCB is also formed by the direct hydrogenation of tDC2B (without going through the isomerization).

Finally, it would be mentioned that the hydrogenation and isomerization of the dicyanoolefins (*t*DC2B, *c*DC1B, *t*DC1B) investigated in this study are significantly slower than those of the monocyanoolefins (*trans*-CH₃CH = CHCN, *cis*-CH₃CH = CHCN, CH₂ = CHCH₂CN) with [IrL(CO)(PPh₃)₂]ClO₄ (L = monocyanoolefin).²

Acknowledgement. We are grateful for the financial support of the Korea Science and Engineering Foundation and the Ministry of Education. We also thank Professor Wan Chul Joo for FT-NMR measurements.

Reference

1. M. K. Lee, I. B. Kim, and C. S. Chin, *J. Organomet. Chem.*, **290**, 115 (1985).

- S. H. Park, H.-K. Park, and C. S. Chin, *Inorg. Chem.*, 24, 1120 (1985).
- 3. F. Garcia-Alonso, V. Riera, and M. J. Misas, *Transition Met. Chem.*, 10, 19 (1985).
- J. Peone and L. Vaska, Angew. Chem., Int. Ed. Engl., 10, 511 (1971).
- 5. Molar conductance of a standard 1:1 electrolyte $(n-Bu)_4NClO_4$ in dichloromethane at 25°C is 34 $ohm^{-1}cm^2mol^{-1}$.
- S. J. Bryan, P. G. Huggett, K. Wade, J. A. Daniels, and J. R. Jenning, *Coord. Chem. Rev.*, 44, 149 (1982) and references therein.
- 7. M. Klinger, Adv. Organomet. Chem., 10, 115 (1971).
- 8. K. Nakamoto, "Infrared and Raman Spectra of Inorgnic and Coordination Compounds"; Wiley-Interscience: New York, (1978) P. 267.
- R. Brady, B. R. Flynn, G. L. Geoffroy, H. B. Gray, J. Peone, and L. Vaska, *Inorg. Chem.*, 15, 1485 (1976).
- (a) D. L. Thorn, Organometallics, 1, 197 (1982); (b) R. H. Crabtree and R. J. Uriate, Inorg. Chem., 22, 4152 (1983); (c) C. E. Johnson, B. J. Fisher, and R. Eisenberg, J. Am. Chem. Soc., 105, 7772 (1983); (d) R. H. Crabtree, G. G. Hlatky, C. P. Parnell, B. E. Segmüller, and R. J. Uriate, Inorg. Chem., 23, 354 (1984); (e) R. H. Crabtree, J. W. Faller, M. F. Mellea, and J. M. Quirk, Organometallics, 4, 1361 (1982); (f) M. Drouin and J. F. Harrod, Inorg. Chem., 22, 999 (1983); (g) M. Basato, F. Morandini, B. Longato, and S. Bresadola, Inorg. Chem., 23, 649 (1984); (h) B. Olgemöller and W. Beck, Inorg. Chem., 22, 997 (1983); (i) A. M. Mueting, P. Boyle, and L. H. Pignolet, Inorg. Chem., 23, 44 (1984).

Synthetic beta-Lactam Antibiotics

I. Synthesis and Antibacterial Activity of 7-Amino-3-[1-(halo-substituted phenyl)-1H-tetrazole-5-yl]thiomethyl-3-cepheme-4-carboxylic acids

Dongsoo Koh, Sang-Woo Park, and Youseung Kim*

Organic Chemistry Lab., Korea Advanced Institute of Science and Technology, Seoul 131 Received February 25, 1987

The syntheses of mercaptotetrazoles and cephalothin analogs are described. Their in vitro poetency was established. The compounds exhibited high antibacterial activity against Gram-positive bacteria and moderate activity against Gram-negative bacteria.

Introduction

The cephalosporin antibiotics represent the most important class of drugs against infectious diseases caused by bacteria. In recent years, several new cephalosporin antibiotics such as cefotaxime (1), ceftizoxime¹, and ceftazidime² with a broad spectrum of activity and increased activity against bacteria producing beta-lactamase have been developed³. Although these recently introduced cephalosporins are characterized by their excellent activity against a variety of Gram-positive and Gram-negative bacteria, they are relatively weak in anti-Staphylococcal activity as com-

pared to older cephalosporins such as cephalothin (2) and cefazoline⁴.

In the course of our extensive research on the modification of cephalosporin, our primary target in the present work was to obtain cephalothin analogs with an enhanced penetration ability into Gram-negative cells including *Pseudomonas* aeruginosa while retaining anti-Staphylococcal activity.

From the viewpoint of molecular modification, new cephalosporin derivatives having the fluoroaromatic group at the N_1 -position of the mercaptotetrazole were of particular interest to us since the introduction of a fluorine atom does influence biological properties such as antimicrobial activity, pharmacokinetics and metabolism of those cephalosporins as well as it has been known that electron-withdrawing groups on the 3-position of cephalosporins enhance antibacterial activity⁵.

To achieve this goal, starting from 7-aminocephalosporanic acid (7-ACA), several compounds were synthesized by introducing the heterocyclic-thio substituents containing a fluorine or chlorine atom at the 3-methyl position by displacement of the acetoxy group.

This paper describes the very simple and dependable preparation method of the mercaptotetrazol (5), the syntheses of the new cephalosporins represented by formular 9 and their in vitro antibacterial activities against 20 selected strains of Gram-positive and Gram-negative bacteria in comparison with cefotaxime (1).

Synthesis

The mercaptotetrazoles (5a-5c) were prepared by the method shown in scheme 1. The syntheses of the 1-alkyl or aryl substituted mercaptotetrazoles have been already reported in several literatures. ⁶⁻⁸ But the known procedures are usually unreproducible, unpleasant to handle, and the yield was found not to be satisfactory. We have found that the potassium salt of dithiocarbamates (4), prepared in quantitative yields by reaction of the anilines (3a-3c) and carbon disulfide in the presence of two equimolar amounts of potassium hydroxide at room temperature, was reacted with sodium azide at reflux for 6 hr to afford the mercaptotetrazoles (5a-5c) in high yields. The reaction was one pot reaction without isolation of the intermediate (4). We tried other bases such as potassium carbonate instead of potassium hydroxide without success.

The synthesis of cephalothin analogs is shown in scheme 2. The reaction was performed without any protection for the amino and carboxy groups of the 7-ACA (6).

Nucleophilic displacement of the 3'-acetoxy group of 6 by the mercaptotetrazoles (5a-5c) was carried out in anhydrous acetonitrile with the addition of borontrifluoride etherate at 50°C for 2hr to afford 7 in high yields.⁹ The reaction in aqueous solution conditions was not attempted because of a low yield. 3 .

For coupling reaction between 7 and thiophene acetic acid, we used the acid chloride method among several acylation reaction conditions at C-7 of the 7-ACA. 10-12 The thiopheneacetic acid was reacted with phosphorus pentachloride in methylene chloride at 0°C. Without isolation of the acid chloride (8), the 7-ACA derivatives (7a-7c) were reacted with 8 in the presence of excess amounts of N-(trimethylsilyl) acetamide at -20°C. This silylating agent can increase the solubility in solvents of low polarity such as methylene chloride and accelerate the reaction rate as well as protect the carboxy group of 7. Three new cephalothin analogs (9) were prepared in satisfactory yields.

Antibacterial Activity and Disscussion

Minimum inhibitory concentrations (MICs) of the cephalosporins were determined by the standard two fold agar dilution method. Mueller Hinton Agar was generally used for bacteria. The size of inoculum used for MIC determination was adjusted to Mcfarland No. 0.5. The lowest concentration inhibiting the visible growth after 18hr incubation at 37°C was expressed as the MIC.

The MIC values of this series of cephalosporins against 20 selected strains of Gram-positive and Gram-negative bacteria in comparison with cefotaxime are shown in Table 1

The data indicates that the cephalothin analogs (9a-9c) have excellent antimicrobial activity against Gram-positive bacteria such as *Bacillus subtilis* ATCC No. 6633, *Micrococcus luteus* ATCC No. 10240, *Staphylococcus aureus* ATCC No. 6538p (penicillin G sensitive) compared with cefotaxime. These cephalothin analogs showed moderate activity against *Escherichia coli*, *Klebsiella pneumoniae*, and *Proteus mirabilis*. None of the compound prepared was as active as cefotaxime against Gram-negative bacteria including *Pseudomonas aeruginosa*. These three compounds had about the same range of antibacterial activity or better as cephalothin^{13,14} against Gram-negative bacteria and improved activity against Gram-positive bacteria.

Among three compounds, no significant difference in activity against 20 species was observed. It can therefore be presumed that a fluorine atom does not influence an-

Table 1. Antibacterial Activity of Three Cephalosporins in Comparison with Cefotaxime

No.		ATCC No.	1	9a	9b	9c
1	Bacillus cereus	27348	. 256	64	64	64
2	Bacillus subtilis	6633	4	0.032	0.032	0.032
3	Micrococcus luteus	9341	0.062	32	32	32
4	Micrococcus luteus	10240	1	0.125	0.125	0.125
5	Staphylococcus aureus	6538P	4	0.125	0.062	0.125
6	Staphylococcus epidermidis	12228	2	64	64	64
7	Acinetobacter calcoaceticus	15473	16	256	256	256
8	Bordetella bronchiseptica	4617	128	256	256	256
9	Enterobacter aerogenes	29751	8	256	256	256
10	Escherichia coli	10536	0.016	16	16	16
11	Escherichia coli	25922	0.032	32	16	32
12	Escherichia coli	31030	0.25	64	128	128
13	Klebsiella pneumoniae	10031	0.016	2	1	0.5
14	Proteus mirabilis	25933	0.016	8	16	8
15	Providencia rettgeri	9919	16	128	64	128
16	Pseudomonas aeruginosa	10145	16	256	256	256
17	Pseudomonas aeruginosa	25619	0.5	256	256	256
18	Pseudomonas aeruginosa	27853	128	256	256	256
19	Salmonella typhimurium	14028	0.25	64	32	64
20	Serratia marcescens	27117	0.25	256	256	256

timicrobial activity in this series.

Although the variations of the side chain of the mercaptotetrazole did not cause significant change in antibacterial activity against Gram-negative bacteria, further modifications to optimize activity in this series and alternative approaches to improve the biological activity of the cephalosporins are under investigation and will be the subject of further publications.

Experimental

Melting points were determined using a Thomas-Hoover capillary melting point apparatus and are uncorrected. NMR-spectra were recorded at 60 MHz on JEOL PMX 60SI NMR and at 200 MHz on Bruker AM 200 NMR using TMS as an internal standard. IR spectra were taken on Analect Instrument fx-6160 FT-IR. Mass spectra were obtained by use of Hewlett-Packard 5985 GC-Mass spectroscopy. Elementary analysis were carried out at KAIST. No effort was made to improve the yields.

1-(4'-Chlorophenyl)-1H-tetrazole-5-thiol (5a): To a solution of 4-chloroaniline (12.7g, 0.10 mol) (3a) in 60 ml of water were added potassium hydroxide (11g, 0.20 mol) and carbon disulfide (7.6g, 0.10 mol) at 20°C. After being stirred for 1hr at the same temperature, sodium azide (7.8g, 0.12 mol) in 20 ml of water was added to the above solution. The reaction mixture was refluxed for 6hr. The aqueous solution was washed with ethyl acetate once and acidified to pH 2 with 1N HCL. The precipitate was filtered and washed with water and acetone. The crude solid was recrystallized from diisopropyl ether to afford 17.3g (81%) of 5a; mp 156°C; NMR (CF₃COOD) ppm 7.1-8.0 (4H,m).

1-(2'-Fluorophenyl)-1H-tetrazole-5-thiol (5b): 5b was obtained (83%) from 2-fluoroaniline (3b) by a similar procedure as described for the preparation of 5a; mp 146°C (lit⁶. 148-148.5°C); MS 196(M+); NMR (CF₃COOD) ppm 7.2-7.9

(4H,m).

1-(4'-Fluorophenyl)-1H-tetrazole-5-thiol (5c): 5c was obtained (88%) from 4-fluoroaniline (3c) by a similar procedure as described for the preparation of 5a; mp 157°C (lit⁶. 157-158°C); MS 196(M $^+$); NMR(CF₃COOD) ppm 7.2-7.9 (4H,m).

7-Amino-3-[1-(4'-chlorophenyl)-1H-tetrazole-5-yl]thiomethyl-3-cepheme-4-carboxylic acid (7a): To a stirred solution of boron trifluoride etherate (7.85g, 55.5 mmol) in 100 ml of anhydrous acetonitrile under N2 were added successively 3.6g (18.5 mmol) of 5a and 5.0g (18.5 mmol) of 7amino-3-acetoxymethyl-3-cephem-4-carboxylic acid (6, 7-ACA) at room temperature. The resulting solution was allowed to react at 50°C for 2hr. After cooling it, the residue in the solution was filtered off and washed with 25 ml of acetonitrile. The filtrate was diluted with 50 ml of water and adjusted to pH 4.0 by addition of ammonium hydroxide solution (28%) at 0°C. The precipitate was filtered and washed with water and acetone. The solid was dried in vacuo to give 6.5g (83%) of 7a; mp 199-201°C (decom); IR(KBr) 1804, 1617 cm⁻; NMR(CF₃COOD) ppm 7.5-7.6(4H, m), 5.4(2H, brs), 4.5(2H, s), 3.9(2H, s).

Anal Calcd for $C_{15}H_{13}N_6CLO_3S_2$: C 42.4, H 3.08, N 19.8. Found : C 42.2, H 3.05, N 19.7.

7-Amino-3-[1-(2'-fluorophenyl)-1H-tetrazole-5-yl] thiomethyl-3-cepheme-4-carboxylic acid (7b): 7b was obtained (93%) from 5b and the 7-ACA by a similar procedure as described for the preparation of 7a; mp 210-211°C (decom); IR (KBr.) 1802, 1617 cm $^{-1}$; NMR (CF $_3$ COOD) ppm 7.1-7.6 (4H, m), 5.4 (2H, brs), 4.5 (2H, s), 3.9 (2H, s).

Anal Calcd for $C_{15}H_{13}N_6FO_3S_2$: C 44.1, H 3.18, N 20.5. Found : C 43.7, H 3.17, N 20.3.

7-Amino-3-[1-(4'-fluorophenyl)-1H-tetrazole-5-yl] thiomethyl-3-cepheme-4-carboxylic acid (7c): 7c was obtained

(87%) from 5c and the 7-ACA by a similar procedure as described for the preparation of 7a; mp 205-210°C (decom); IR (KBr) 1804, 1617 cm⁻¹; NMR (CF₃COOD) ppm 7.1-7.6 (4H, m), 5.4 (2H, brs), 4.5 (2H, s), 3.9 (2H, s).

Anal Calcd for $C_{15}H_{13}N_6FO_3S_2$: C 44.1, H 3.18, N 20.5. Found : C 43.6, H 3.17, N 20.2.

7ß-[2-Thiophene)acetamido]-3-[1-(4'-chlorophenyl)-1H-tetrazol-5-yl] thiomethyl-3-cephem-4-carboxylic acid (9a): To 2-thiopheneacetic acid (0.28 g, 2.0 mmol) in 20 ml of methylene chloride was added phosphorus pentachloride (0.5 g, 2.4 mmol) at 0°C and the mixture was stirred for 1 hr at the same temperature. The resultant solution was evaporated in vacuo. The residue was dissolved in 5 ml of methylene chloride to produce the acid chloride solution (8) of thiopheneacetic acid. The above acid chloride solution was added dropwise to the mixture of 7a (0.85 g, 2.0 mmol) and N-(trimethylsilyl) acetamide (1.5 g, 12 mmol) in 20 ml of methylene chloride at -20 °C. The reaction mixture was stirred for 2 hr at -15°C. After removing the solvent, the residue was dissolved in a mixture of water and ethyl acetate. The organic layer was separated and the aqueous layer was washed with ethyl acetate once. To the combined ethyl acetate solution was added water and the solution was adjusted to pH 7.0 with a saturated sodium bicarbonate solution. The aqueous solution was separated and acidified to pH 2.0 with 10% hydrochloric acid solution. The organic compound was extracted with ethyl acetate three times. The combined ethyl acetate solution was washed with water, brine and dried over magnesium sulfate. After evaporation of the solvent, the crude product was triturated with n-hexane to afford 0.58g (53%) of 9a; mp 92-93°C (decom); IR(KBr) 1786, 1718, 1683, 1512 cm⁻; NMR (DMSO-d₆) 9.0-9.15 (1H, d), 7.3-7.9 (4H, m), 6.9-7.0 (2H, m), 5.7-5.9 (1H, m), 5.0 (1H, d), 4.4-4.5 (2H, d), 3.9 (2H, s) 3.8 (2H, s).

7ß-[(2-Thiophene)acetamido]-3-[1-(2'-fluorophenyl)-1H-tetrazol-5-yl] thiomethyl-3-cepheme-4-carboxylic acid (9b): 9b was obtained (61%) from 7b and 8 by a similar procedure as described for the preparation of 9a. The crude product was triturated with ethyl ether; mp 107-109°C (decom); IR(KBr) 1782, 1722, 1685 cm⁻¹; NMR (DMSO-d₆) 9.0-9.1(1H, d), 7.3-7.7 (4H, m), 6.9-7.0 (2H, d), 5.7 (CH, q), 5.0 (1H, q), 4.4-4.5 (2H, d), 3.9 (3H, s), 3.8 (2H, s).

7ß-[(2-Thiophene)acetamido]-3-[1-(4'-fluorophenyl)-1H-

tetrazol-5-yl] thiomethyl-3-cepheme-4-carboxylic acid (9c): was obtained (48%) from 7c and 8 by a similar procedure as described for the preparation of 9a; mp 124-126°C (decom); IR(KBr) 1778, 1718, 1511 cm⁻¹ NMR (DMSO-d₆) 9.0-9.15 (1H, d), 7.4-7.7 (4H, m), 6.9-7.0 (2H, d), 5.6-5.8 (1H, q), 5.0-5.1 (1H, d), 4.4-4.5 (2H, d), 3.9 (2H, s), 3.7 (2H, 2).

Acknowledgements. We wish to express our deep gratitude to Dr. Joong-Hyup Kim at KAIST for his valuable advice and encouragement throughout this work and for supplying the 7-ACA. We would also like to thank Dr. Cheolsoo Shin at KRICT for carring out the biological evaluation. This work was fully supported by Ministry of Science and Technology.

References

- R.B. Morin and M. Gorman: Chemistry and Biology of beta-Lactam Antibiotics, Vol. 1: Penicillins and Cephalosporins P. 422, Academic Press, New York 1982.
- 2. E.M. Wilson, Chem. Ind. (London) 217 (1984).
- 3. W. Dürckheimer, J. Blumbach, R. Lattrell, and K.H. Scheunemann, *Angew. Chem. Int. Ed. Engl.*, **24**, 180 (1985).
- 4. Ref. 1 p. 376.
- 5. R.B. Hermann, J. Antibiot., 26, 233 (1973).
- J.C. Kauer and W.A. Sheppard, J. Org. Chem., 32, 3580 (1967).
- 7. T. Kamiya, K. Tanak, T. Teraji, Y. Shiokawa, and K. Henmi, *Japan kokai* 76-68, 568, Jun. 14 (1976).
- 8. T. Tsuji, H. Satoh, M. Narisada, Y. Hamashima, and T. Yoshida, *J. Antibiot.*, **38(4)**, 466 (1985).
- I. Saikawa, S. Takano, K. Momonoi, I. Takakura, K. Tanaka, and C. Kutani, *Chem. Pharm. Bull.*, 33(12), 5534 (1985).
- 10. Ref. 1 and Ref. 3.
- 11. J. Goto, K. Sakane, Y. Nakai, T. Teraji, and T. Kamiya, *J. Antibiot.*, **37**, 546 (1984).
- T. Takaya, H. Takasugi, T. Masugi, Hiromukochi, and H. Nakano, J. Antibiot., 34, 1290 (1981).
- G.S. Lewis and P.N. Nelson, J. Med. Chem., 22(10), 1214 (1979).
- T. Naito, J. Okumura, H. Kamachi, H. Hoshi, and H. Kawaguchi, J. Antibiot., 30(9), 705 (1977).